International Journal of Pharmacy and Pharmaceutical Sciences

ISSN- 0975-1491

Vol 6, Issue 10, 2014

Review Article

FAMILY ACANTHACEAE AND GENUS APHELANDRA: ETHNOPHARMACOLOGICAL AND PHYTOCHEMICAL REVIEW

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Received: 10 Aug 2014 Revised and Accepted: 15 Sep 2014

ABSTRACT

Aphelandra belong to family Acanthaceae. We have reviewed traditional uses, pharmacological potential and phytochemical study of family Acanthaceae and genus Aphelandra. Traditionally the most important part use in Acanthaceae is the leaves and they are used externally for wounds. We have found that Acanthaceae possess antifungal, cytotoxic, anti-inflammatory, anti-pyretic, anti-oxidant, insecticidal, hepatoprotective, immunomodulatory, Anti- platelet aggregation and anti-viral potential. Phytochemical reports on family Acanthaceae are glycosides, flavonoids, benzonoids, phenolic compounds, naphthoquinone and triterpenoids. We have also document genus Aphelandra, its phytochemical and pharmacological potential.

Keywords: Acanthaceae, Aphelandra, Ethnomedicinal, Phytochemistry, Wound healing potential,

INTRODUCTION

Majority of the population in developing world is struggling to raise living standards and improvement of health care delivery due to increasing poverty and population. According to an estimate, 70-80% of rising world is dependent on conventional plants obtained remedies as pharmaceuticals are high priced. From this reality, it can be retrieved that by data assembling and experimentation, valuable plus economical medicaments can be separated from different flora to satisfy requirements of evolving world. Hence requirements of officinal plants cannot be neglected[1]. Databases used to search for the literature were: Google scholar,PubMed, Tropicos (for plant taxonomy). Bold numeral in the text refers to chemical structures reported in Fig. 1. All the structures were reported from the literature and drawn by ChemDraw.

Ethnomedicinal importance of family acanthaceae

Ground leaves of *Adhatoda vesica* Nees. (Acanthaceae) are used in treatment of bronchial diseases. Ground fresh leaves of *Andrographispaniculata* are used to alleviate bites of poisonous insects and reptiles[2].

Infusions of leaves of *Aystasiaschimperi*, *Dyschoristeradicans*, *Acanthus eminens*, *Dyschoristethumbergiiflora*, *Lepidagathisscariosa* and *Thunbergiaalata* (Family: Acanthaceae) are used for cough, skin diseases, wounds, eye infections, anti-diarrhea, edema, pneumonia and backache. Paste of leaves of *Barberiagrandicalyx* (Acanthaceae) is used for snake bites. Ash of leaves of *Justiciabetonica*, *Acanthus pubescens* and *Justiciaflava* is used for dry cough, anti-diarrhea, flu and ulcers [3].

Leaves of *Blepharismaderaspatensis* belonging to family Acanthaceae are mixed with onion bulbs to form paste. This paste is applied externally for cuts and wounds. Leaves of *Hygrophilaauriculata* (Acanthaceae) are used to relieve from cough. Leaves of *Justiciatranquebariensis* (Acanthaceae) are used for poisonous bites [4].

Leaves of *Andrographispaniculata* Nees. And *Justiciagendrarussa* L. Which belongs to family Acanthaceae, are used for fever, headache, vertigo and wounds [5].

Powder of leaves of *Andrographispaniculata* (Acanthaceae) is used internally for diabetes. Juice of leaves of *Blepharismaderaspatensis* (Acanthaceae) is used externally for wounds. Paste of leaves of *Elytrariaacualis* (Acanthaceae) is used externally for hip pain. Decoction of whole plant of *Rungialinifolia* which belongs to family Acanthaceae is used internally for ulcers [6].

Leaves of *Justiciaadhatoda* which belongs to family Acanthaceae are used for rheumatism, fever, chest diseases, pneumonia, asthma, tuberculosis, antispasmodic, expectorant, diuretic, antiseptic and to reduce swelling [7].

Andrographispaniculata, Hygrophilaspinosa, Barleriaprionitis and *Adhatoda vasica* are members of the Acanthaceae family, which are used traditionally as antiviral, antipyretic, antiasthmatic and in respiratory diseases [8].

Pharmacological reports on family acanthaceae

Antibacterial activity

Aqueous extract of *Andrographispaniculata* belonging to family Acanthaceae showed significant antibacterial activity against *Bacillus subtilis, Staphylococcus aereus, Escherichia coli* and *Pseudomonas aeruginosa* in comparison to some known antibiotics[9].

Methanolic extract of *Asteracanthalongifolia* L. belonging to family Acanthaceae showed the most promising results against *Burkholderiapseudomallei* strain1and strain 2 and *Staphylococcus aureuss*[10].

Acetone crude extract of *Andrographisaffinis* Nees belongs to family Acanthaceae showed higher degree of inhibition against *Staphylococcus aureus*, *Proteus vulgaris*, *Escherichia coli*, *Klebsiellapneumoniae*, and *Pseudomonas aeruginosa*[11].

Chloroform crude extract of *Andrographispaniculata* (Acanthaceae) showed antibacterial activity against *S. aereus, E. coli,B. subtilis* and *P. vulgaris.* The effect produced by the extract was comparable with benzyl penicillin[12].

A large number of gram positive and gram negative bacteria exhibited vulnerability when exposed to extract of *Rhinacanthusnasutus* (Acanthaceae) [13].

Antifungal activity

The dichloromethane extract of the leaves of *Hypoestesserpens* (Acanthaceae) showed antifungal activity against *Cladosporiumcucumerinum* and *Candida albicans*. Fusicoserpenol A

and dolabeserpenoid acid A were isolated responsible for this activity [14].

The chloroform extract of *Androgrphispaniculata* (Acanthaceae) exhibited activity against *Aspergillusniger Penicillium chrysogenum*. The activity was found to be effective against all the organisms tested [15].

Antifungal activity was reported by aqueous extract of *Andrographispaniculata* belonging to family acanthaceae against *Candida albicans* in comparison to Nystatin[9]. Anti-fungal activity has been exhibited by the extract of *Rhinacanthusnasutus* (Acanthaceae) against *Aspergillusniger*, *Pyriculariaoryzae*,

Cryptococcus neoformans, Saccharomyces spp., Epidermophytesfloccosum, Candida albicans, Trichophytonrubrum, Tricophytonmentagagrophytes, Microsporumcanis and Microsporumgypeseum[13].

Cytotoxic activity

Elenoside**(1)** was reported in *Justiciahyssopifolia* belonging to family Acanthaceae. Elenoside showed cytotoxic activity to human cancer cell lines in range concentrations from 10^{-5} to 10^{-4} M. It has an LD₅₀ in mice, of 305mg/kg and central depression properties at doses of 25, 50, and 100mg/kg. Thus elenoside appears to be sedative with broad spectrum cytotoxicity [16].

Table 1: Traditional uses of family Acanthaceae

Species Part use Traditional uses	
Adhatoda vesica Nees Leaves Treatment of bronchial diseases ²	
Andrographispaniculata Leaves Alleviate bites of poisonous insects an	nd reptiles ²
Aystasiaschimperi, Dyschoristeradicans, Leaves Used for cough, skin diseases, wounds	s, eye infections, anti-diarrhea, edema,
Acanthus eminens, pneumonia and backache ³ .	
Dyschoristethumbergiiflora,	
Lepidagathisscariosa	
Thunbergiaalata	
Barberiagrandicalyx Paste of leaves Paste of leaves of (Acanthaceae) is use	ed for snake bites. ³
<i>Justiciabetonica, Acanthus pubescens and</i> Ash of leaves Ash of leaves of are used for dry cough	h, anti-diarrhea, flu and ulcers³
Justiciaflava	
Blepharismaderaspatens Mixed with This paste is applied externally for cut	ts and wounds. ⁴
onion bulbs to	
form paste	
Hygrophilaauriculata Leaves Leaves of Hygrophilaauriculata (Acam	thaceae) are used to relieve from cough. ⁴
Justiciatranquebariensis Leaves Leaves of Justiciatranquebariensis(Ac	anthaceae) are used for poisonous bites ⁴
Andrographispaniculata Nees. and Leaves Leaves of Andrographispaniculata Nee	es. and Justiciagendrarussa L. which
Justiciagendrarussa L belongs to family Acanthaceae, are use	ed for fever, headache, vertigo and
wounds ³	
Andrographispaniculata Powder of Powder of leaves of Andrographispan	iculata (Acanthaceae) is used internally
leaves for diabetes.	
Blepharismaderaspatensis Juice of leaves of Blepharismaderaspa	itensis (Acanthaceae) is used externally
Entreries englis	
Elytraridacualis Paste of leaves of Elytraridacualis (Ac	if a subject to family for hip pain
Rungiannijona Decocition of Whole plant of Rungianni	nona which belongs to family
whole plant Acatinaceae is used internally for ucc	ers o
Justiciaaanatoda Leaves Leaves of Justiciaaanatoda which belo	ongs to family Acanthaceae are used for
rneumatism, lever, chest diseases, pric	eumonia, astrinia, tuberculosis,
Andrographiangnigulata Ukaronhilagningga Laguaga Andrographiangnigulata Ukaronhilagningga	antiseptic and to reduce swelling
Antir ographispanicariau, nygrophilaspinosa, Leaves Antir ographispaniculata, nygrophilas	pillosa, barleriapilolillis allu Auffatoda
buile international validation and a set international and a set international and a set international set internation	ad in respiratory diseases 8

Justiflorinol(2), justicinol(3), patentiflorin A (4), patentiflorin B (5), 4"-O-acetylpatentiflorin B (6) and 4"-O-acetylmanathoside B (7) were separated from leaves and stems of *Justiciapatentiflora* (Acanthaceae) by bioassay-guided purification. These compounds display significant cytotoxic activity against cancer cell lines and arrest the cell cycle [17].

Methanol extract of *Andrographispaniculata*exhibited anti-cancer activity. Andrographolide**(8)**, 14-deoxyandrographolide **(9)** and 14-deoxy-11, 12-didehydroandrographolide **(10)** were separated from this extract [18]. Andrographolide**(8)** from the plant *Andrographispaniculata* (Acanthaecae) and its derivative 8, 17 - epoxy andrographolide**(11)** showed cytotoxic activity [19]. Rhinacanthin-A **(12)**, rhinacanthin-B **(13)**, rhinacanthin-C **(14)**, rhinacanthin-D **(15)**, rhinacanthin-G, rhinacanthin-H, rhinacanthin-I, rhinacanthin-K, rhinacanthin-M, rhinacanthin-N **(16)**, rhinacanthin-Q **(17)** and wogonin**(18)**separated from the roots of *Rhinacanthusnasutus* (Acanthaecae) showed cytotoxic activity. .The leaf and root extract of *Rhinacanthusnasutus* (Acanthaecae) showed antiproliferative activity against human cervix adenocarcinomaand other types of cancers [13].

Anti-inflammatory and antipyretic activity

Leaves extracts of *Hygrophilaspinosa* (Acanthaceae) exhibited antipyretic and anti-inflammatory activity [20]. Alcoholic extract of *Lepidegathisanobrya* of family acanthaceae showed analgesic and anti-inflammatory activities[21].

Anti-inflammatory and analgesic activities were reported by the methanol extract of *Diclipteraverticillata* (Acanthaceae)[22]. Ethanol extract of *Justciagendrarussa* (Acanthaceae) exhibited analgesic activity and anti-inflammatory activity[23].

The aqueous extract of roots of *Barleriaprionitis* L. (Acanthaceae) showed anti-inflammatory activity by using caragennan-induced rat paw edema method[24].

The methanol extract of leaves of *Andrographispaniculata* (Acanthaceae) was tested on inhibition of lipopolysaccharide induced and calcimycin induced mediators in diverse cell models. The results showed that extract was fairly potent in attenuating proinflammatory, inflammatory and allergic mediators[25].

Antiviral activity

Rhinacanthin-E and rhinacanthin-F separated from the aerial parts of *Rhinacanthusnasutus* (Acanthaceae) exhibited antiviral activity [13].

Antioxidant activity

Methanol and aqueous extracts of aerial parts of *Justiciaspicigera* of family acanthaceae exhibited antioxidant activity. DPPH radical

scavenge method was used. Water extract has lower activity than that of methanol [26].

Methanol extracts 100µg/ml) Ruelliakerrii fat of and Strohilanthesauriculata showed antioxidant activity. Justiciaprocumbens, Methanolextracts of Barleriacristata, Ruelliakerrii and Strobilanthesaureculata showed antioxidant activity [27].

The extract of *Rhinacanthusnasutus* (Acanthaceae) exhibited antioxidant activity. Anti-aging effect was produced by the cosmetics containing this extract. The mechanism involved is the removal of superoxide from the human body [13].

Antioxidant activity of the methanol fraction of *Blepharisedulis* (Forssk.) Pers. Family Acanthaceae was determined using the DPPH radical test [28].

Hepatoprotective activity

The extract of *Andrographispaniculata*(Acanthaceae) was examined for hepatoprotective effect by administration to male mice. Andrographolide**(8)** and its derivativesexhibited hepatoprotective activity [29].

Rhinacanthusnasutus root extracts showed hepatoprotective effect in rats treated with aflatoxin-B1[13].

Insecticidal activity

The alcoholic extract of the aerial parts of *Acanthus montanus* Nees (Family: Acanthaceae) exhibited a significant activity against adult *Aedesaegypti*. Phytochemical study of the plant has resulted in isolation of compounds which exhibit variable degrees of insecticidal activity. B-sitosterol-3-O- β -D-glucoside(**19**) exhibited mosquitocidal activity (100% mortality) against adult *Aedesaegypti* at 1.25µg/mg concentration, followed by palmitic acid (**20**) (90%), linaroside(**21**) (80%) and acetoside(**22**) (70%) respectively [30].

Immunomodulatory activity

The extract of *Rhinacanthusnasutus* of family Acanthaceae exhibited immunomodulatory activity [13].

Anti- platelet aggregation activity

The *Rhinacanthusnasutus* (Acanthaceae) plant extract showed the anti-platelet aggregation effect. Rhinacanthin-A **(12)**, rhinacanthin-B **(13)**, rhinacanthin-C **(14)**, rhinacanthin-G,rhinacanthin-H, rhinacanthin-I, rhinacanthin-K, rhinacanthin-M and rhinacanthin-Q **(17)** separated from the roots of *Rhinacanthusnasutus*. These compounds showed inhibition of rabbit platelet aggregation induced by arachidonic acid [13].

Phytochemical reports on family acanthaceae

Glycosides

From the aerial parts of Acanthus ilicifolicus, which is a member of Acanthaceae family, (2R)-2-O-β-D-gluco-pyranosyl-2H-1,4-(23), benzoxazin-3(4H)-one (2R)-2-*O*-β-*D*-glucopyranosyl-4-(2R)-2-*0*-βhydroxy-2H-1,4-benzoxazin-3(4H)-one (24), Dglucopyranosyl-7-hydroxy-2H-1,4-benzoxazin-3(4H)-one (25), 7chloro-(2R)-2-0-β-D-glucopyranosyl-2H-1,4-benzoxazin-3(4H)-one (26) and (2R)-2-O-β-D-glucopyranosyl-5-hydroxy-2H-1,4benzoxazin-3(4H)-one(27) have been isolated [31].

Salidroside (28), benzyl β -*D*-glucopyranoside (29), (6S,9R)roseoside (30), asysgangoside (31), ajugol (32), apigenin 7-*O*- β -*D*glucopyranoside (33), apigenin 7-*O*-neohesperidoside (34) and apigenin 7-*O*- β -Dglucopyranosy (1 \rightarrow 6) - β -*D*-glucopyranoside (35) have been reported in *Asystasiagangetica* which belongs to family Acanthaceae [32].

Olean-12-ene-1 β ,3 β ,11 α ,28-tetraol-28-*O*- β -*D*-glucopyranosyl-(1 \rightarrow 2) - β -D-glucopyranoside **(36)**, olean-12-ene-1 β ,3 β ,11 α ,28-tetraol-28-*O*- β -*D*-glucopyranosyl-(1 \rightarrow 2)- β -*D*-glucopyranosyl-(1 \rightarrow 2)- β -*D*-gluco pyranoside**(37)**, 11 α -methoxy-olean-12-ene-1 β ,3 β ,28-triol-28-*O*- β -*D*-glucopyranosyl-(1 \rightarrow 2)- β -*D*-glucopyranoside**(38)** and 11 α methoxy-olean-12-ene-1 β , 3 β , 28-triol 28-*O*- β -Dglucopyranosyl $(1\rightarrow 2)$ - β -D-glucopyranosyl- $(1\rightarrow 2)$ - β -D-glucopyranoside **(39)** were reported from *JusticiabetonicaL*. belonging to family Acanthaceae [33]. Three phenylethnoid glycosides, desrhamnosylacteoside**(40)**, acteoside **(41)** and poliumoside**(42)** were separated and identified from the callus cultures of *Barleriacristata L*. of family Acanthaceae [34].

phenylethanoid glycoside namely acanmontanoside(43) New with eight known identified together compounds, as decaffeoylverbascoside,verbascoside(44), isoverbascoside(45), leucosceptoside A (46), (2R)-2-0- β -D-glucopyranosyl-2H-1,4benzoxazin-3(4*H*)-one,(2*R*)-2-*O*-β-*D*-glucopyranosyl-4-hydroxy-2*H*-1,4-benzoxazin-3(4H)-one (47), (3R)-1-octen-3-ol-3-O-β-Dxylopyranosyl- $(1 \rightarrow 6)$ -O- β -D-glucopyranoside (48) and ebracteatoside B (49) have been reported from the methanol extract of aerial portion of Acanthus montanus[35].

From the genus *Asystasia* of Acanthaceae family following compounds have been reported: 6β-hydroxyantirrhide **(50)**,angeloside **(51)**,ajugol **(32)**, 6-*O*-α-L-rhamnopyranosyl-catapol **(52)**, 6-*O*-α-(3"-*O*-trans-caffeoyl)-*L*-rhamnopyranosyl-catapol **(53)**, 6-*O*-α-(3"-*O*-trans-*p*-coumaroyl)-*L*-rhamnopyranosyl-catapol **(54)**, sinuatol **(55)**, luteolin 7-*O*-*β*-*D*-glucopyranoside **(56)**, luteolin 7-*O*-rutinoside **(58)** and apigenin 7-*O*-*β*-*D*-glucuronide **(59)** [36].

Ethanol extract of leaves of *Barleriacristata* gave two iridoidal glycosides like barlerin(**60**) and shanshide methyl ester (**61**)[37].

Chemical investigation of the methanol fraction of the aerial parts of *Blepharisedulis* (Forssk.) Pers. Family Acanthaceae revealed the presence of phenylethnoids namely verbascoside(**44**), *cis*-verbascoside, isoverbascoside(**45**) and leucosceptoside **A** (**46**) [28]. Glycoside compounds; sitosterol- β -*D*-glucopyranoside(**62**), stigmasterol- β -*D*-glucopyranoside(**63**), 3, 4-dimethylphenol- β -*D*-glucopyranoside(**64**) and 3, 4, 5-trimethylphenol- β -*D*-glucopyranoside(**65**) were isolated from leaves and stems of *Rhinacanthusnasutus* plant of family Acanthaceae [13].

Flavonoids

The preliminary phytochemical screening of *Dipteracanthuspatulus* Jacq. Nees (Acanthaceae) revealed the presence of flavonoids [38].

From the crude ethanol extract of Brillantaisiapalisatii (Acanthaceae) 3-epi-ursolic acid (66), verbascoside(44) and lespedin(67) were isolated[39]. Following compounds were reported from Andrographispaniculata (Acanthaceae); ßsitosterol(68), andrographolide(8),14-deoxy-ll,12-dedihydro andrographolide,14-deoxyandrographolide (69), 7-0methyldihydrowogonin (70), (2S)-5,7,2',3'-tetramethoxyflavanone (71), dihydroskullcapflavone I (72), 7-0-methylwogonin (73), 5hvdroxy-7,8,2',5'-tetramethoxy-flavone (74), 5-hydroxy-7,8,2',3'tetramethoxyflavone (75), 5-hydroxy-7,2,6'-trimethoxyflavone(76), 5-hydroxy-7,2,3'-trimethoxyflavone(77), skullcapflavone12'methylether (78), cinnamic acid(79), caffeic acid (80), ferulic acid (81), chlorogenic acid (82), 7-0-methylwogonin 5-glucoside (83), skullcapflavone I 2'-glucoside (84), 14-deoxy- 15-isopropylidene-11,12-didehydro-andrographolide (85), 14-deoxy-11-hydroxyandro grapholide (86), neoandrographolide(87) and andro graphoside(88) [40]. Chromatographic resolution of ethyl acetate extract of leaves of Barleriacristata furnished two flavonoid compounds, which were further identified and characterizes as luteoline(89) and 7-methoxy luteoline(90)[41]. Coumarins, (+)pracruptorin, umbelliferone(**91**) and 2, 6-dimethoxy benzoquinone (92) derived from the whole plant of Rhinacanthusnasutus plant of family Acanthaceae [13].

Phenolic compounds

Petroleum ether extract of leaves of *Barleriacristata* furnished two phenolic compounds, which were characterized as p-coumaric acid **(93)** and α -tocopherol**(94)** [41].

Naphthoquinone

Rhinacanthin-A **(12)**, rhinacanthin-B **(13)**, rhinacanthin-C **(14)**, rhinacanthin-D **(15)**, rhinacanthin-G, rhinacanthin-H, rhinacanthin-I, rhinacanthin-J, rhinacanthin-K, rhinacanthin-L, rhinacanthin-M,

rhinacanthin-N	(16) ,	rhinacanthin-O,		rhinacanthin-P		and
rhinacanthin-Q	(17)	were	isolated	aerial	parts	of
Rhinacanthusnasutus plant of family Acanthaceae [13].						

Benzonoids

The benzonoids compounds p-hydroxy-benzaldehyde(95), vanillic acid (96), syringic acid (97), 2-methoxy-propionolphenol (98), methyl valinate(99) and syringaldehyde(100) were isolated from leaves, roots and stems of *Rhinacanthusnasutus* plant of family Acanthaceae [41].

Triterpnoids

The triterpenoids compounds β -amyrin(101), glutinol(102) and lupeol(103) were derived from roots of *Rhinacanthusnasutus* plant of family Acanthaceae [13].

Pharmacological reports on genus Aphelandra

Antibacterial activity and Antifungal activity

From the roots of genus *Aphelandra*2-benzoxazolinones **(104)** and 6-methoxy-2-benzoxazolinone **(105)** were separated. These compound and their related synthetic derivatives showed antibacterial and antifungal activity [42]. Plants belonging to genus *Aphelandra* (*Acanthaceae*)showed antimicrobial activity against *E. coli, P. aeruginosa, S. aureus, and C. albicans*[43]. The methanol extracts of *Aphelandra squarrosa*showed antibacterial activity against at least one of the following test organisms, *Staphylococcus aureus, Escherichia coli, Proteus vulgaris andPseudomonas aeruginosa* using the tube dilution and plate diffusion methods [44].

Immunomodulatory activity

2-benzoxazolinone **(104)** and 2-hydroxy-1, 4-benzoxazin-3-one **(106)** were isolated from *Aphelandra tetragona*. These were bio transformed into 2-amino-3-*H*-phenoxazine-3-one **(107)** by endophytic fungi.2-amino-3-*H*-phenoxazine-3-one **(107)** has potent anti-inflammatory and immunoregulatory properties. These properties may provide a promising therapeutic strategy for the treatment of T cell-mediated inflammatory auto immune diseases as well as for bacteria induced chronicinflammatory diseases[45].

Both 2-benzoxazolinone **(104)** and 2-hydroxy-1, 4-benzoxazin-3one **(106)** are compounds calledphytoanticipins and present in healthy plants as substances toovercome microbial diseases and herbivore[46].

Phytochemical reports on genus Aphelandra

Alkaloids

Macrocyclic polyamine alkaloid aphelandarine**(108)** was isolated from the roots of *Aphelandra tetragona*, which consists of spermine and two units of 4-hydroxycinnamic acid [47].

 N^{1} ,N⁵-Di-*p*-coumaroylspermidine (109), N⁵,N[10]-di-*p*coumaroylspermidine (110), and N¹,N⁵,N[10]-tri-*p*coumaroylspermidine (111) were isolated from anthers of *Aphelandratetragona* A. .chamissoniana[48].

18-*O*-Methylchaenorpine and iso-18-*O*-methylchaenorpine two novel stereo-isomeric spermine alkaloids were separated from Aphelandra*tetragona*[49].

Macrocyclicspermine alkaloids namely 6-hydroxy-aphelandrine **(112)** and 6-acetoxy-aphelandrine **(113)** were isolatedfrom the roots of *Aphelandra fuscopunctata*[50].

Benzoxazinoids-cyclic hydroxamic acids and their correspondingglucosides

From roots of *Aphelandra squarrosa* and *Aphelandra fuscopunctata* following compounds were separated; 2,4-dihydroxy-1,4benzoxazin-3(H)-one **(114)**, 2,4-dihydroxy-7-methoxy-1,4benzoxazin-3(H)-3one **(115)**, 2-hydroxy-1,4-benzoxazin-3(2H)-one **(116)**, 2-hydroxy-7-methoxy-1,4-benzoxazin-3(2H)-one **(117)**, and their corresponding glycosides as well as bezoxazolinones[51]

In Aphelandra squarrosaglycosides are present in the roots [52].

Flavonoids and Isoflavones

Several compounds were isolated and identified from the leaves and stems of *Aphelandra aurantiaca* which consists of scopoletin(**118**), chrysin(**119**), eucalyptin(**120**), gnaphalin(**121**), nevadensin(**122**) and *p*-coumaric acid (**123**) in the form of ester with glucose [53].

Fig. 1: Structure of compounds present

CONCLUSION

Literature actually reports chemical investigations of a large family of nearly 250 genera and 2500 species, distributed mainly in the tropics and extending to Mediterranean, Australia and Southern U. .S. A. In Pakistan it is represented by 18 genera and 60 specific and infraspecific taxa; of which 44 are native. There are no published data concerning either the toxicity of the whole remedies and their isolated compounds. Further investigations on phytochemical discovery and subsequent screening are needed for opening new opportunities to develop pharmaceuticals based on Family Acanthaceae and genus Aphelandra.

CONFLICT OF INTERESTS

Declared None

REFERENCE

- Mughal TA. Ethnomedicinal studies of flora of Southern Punjab and isolation of biologically active principles: Department of Chemistry Lahore College for Women University, Lahore; 2008.
- Muthu C, Ayyanar M, Raja N, Ignacimuthu S. Medicinal plants used by traditional healers in Kancheepuram District of Tamil Nadu, India. J Ethnobiology Ethnomedicine 2006;2:1-10.
- 3. Jeruto P, Lukhoba C, Ouma G, Otieno D, Mutai C. An ethnobotanical study of medicinal plants used by the Nandi people in Kenya. J Ethnopharmacol 2008;116:370-6.
- Sandhya B, Thomas S, Isabel W, shenbagarathai R. Ethnomedicinal plants used by valaiyan community of piranmalia hills (reserved forest), tmil nadu, india. –a pilot study. Afr J Tradit Complementary Altern Med 2006;3:101-14.
- Mia MMK, Kadir MF, Hossan MS, Rahmatullah M. Medicinal plants of Garo tribe inhabiting the Madhupur forest region of Bangladesh. Am-Eur J Sustainable Agriculture 2009;3:165-71.
- Ignacimuthu S, Ayyanar M, Sankarasivaraman K. Ethnobotanical study of medicinal plants used by paliyartribals in theni district of tamil nadu, india. Fitoterapia 2008;79:562-8.
- 7. Husain SZ, Malik RN, Javaid M, Bibi S. Ethnobotanical properties and uses of medicinal plants of Morgah biodiversity park, rawalpindi. Par J Boany 2008;40:1897-911.
- Krishnaraju AV, Rao TVN, Sundararaju D, Vanisree M, Tsay HS, Subbaraju GV. Assessment of bioactivity of indian medicinal plants using brin shrimo (artemiasalina) lethality assay. Int J Appl Sci Eng 2005;3:125-34.
- 9. Singha PK, Roy S, Dey S. Antimicrobial activity of Andrographispaniculata. Fitoterapia 2003;74:692-4.
- 10. Samy RP. Antimicrobial activity of some medicinal plants from India. Fitoterapia 2005;76:697-9.

- 11. Alagesaboopathi C. Antimicrobial potential and phytochemical screening of Andrographisaffis Nees. An endemic medicinal plant from India. Int J Pharm Pharm Sci 2011;3:157-9.
- Radhika P, Lakshmi K. Antimicrobial activity of the chloroform exracts of the roots and the stem of andrographispaniculata nees. Int Res J Microbiol 2010;1:037-9.
- Bukke S, Raghu PS, Sailaja G, Kedam TR. The study on Morphological, phytochemical and pharmacological aspects of rhinacanthusnasutus. (L) Kurz (A Review). J Appl Pharm Sci 2011;1:26-32.
- 14. Rasoamiaranjanahary L, Marstan A, Guilet D, Schenk K, Randimbivololona F, Hostettmann K. Antifungal diterpenes from hypoestesserpens (Acanthaceae). Phytochem 2003;62:333-7.
- Radhika P, Lakshmi K. Antimicrobial activity of the chloroform exracts of the roots and the stem of andrographispaniculata nees. Int Res J Microbiology 2010;1:037-9.
- Navarro E, Alonso SJ, Trujillo J, Jorge E, Perez C. General Behavior, Txicity, and Cytotoxic Activity of Elenoside, a lignin from Justciahyssopifolia. J Nat Pro 2001;64:134-5.
- Susplugas S, Hung NV, Bignon J, Thoison O, Kruczynsk A, Sevenet T, Gueritte F. Cytotoxic arylnaphthalene lignans from a vietnamese acanthaceae, justiciapatentiflora. J Nat Pro 2005;68:737-8.
- Kumar RA, Seridevi K, Kumar NV, Nanduri S, Rajagopal S. Anticancer and immunstimulatory compounds from andrographispaniculata. J Ethnopharm 2004;92:291-5.
- Nanduri S, Nyavanandi VK, Thunuguntla SSR, Kasu S, Pallerla MK, Ram, et al. Synthesis and structure activity relationship of andrographolide analogues as novel cytotoxic agents. Bioorg Med Chem Lett 2004;14:4711-7.
- Patra A, Jha S, Murthy PN, Vaibhav A, Chattopadhyay P, Panigrahi G, *et al.* Anti-inflammatory and antipyretic activities of Hygrophilaspinosa T. Anders leaves (Acanthaceae). Trop J Pharm Res 2009;8:133-7.
- 21. Richard SW, Marus L, Noya S, Pierre GI, Germaine NOO. Antinflammatory, analgesic and antipyretic effects of Lepidagathisanobrya Nees (Acanthaceae). Afr J Tradit Complementary Altern Med 2011;8:420-4.
- 22. Sawadogo wR, Boly R, Lompo M, Some N, Lamien CE, *et al*. Antiinflammatory, analgesic and antipyretic activities of diclipteraverticillata. Int J Pharmacol 2006;2:435-8.
- Jothimanivannan C, Kumar RS, Subramanian N. Antiinflammatory and analgesic activity of ethanol extract of aerial parts of justiciagenarussa burn. Int J Pharmacol 2010;1:1-6.
- Khadse CD, Kakde RB. Anti-inflamatory activity of aqueous extract fractions of Barleriaprionitis L. roots. Asian J Plant Sci Res 2011;1:63-8.

- Chandrasekaran CV, Gupta A, Agarwal A. Effect of an extract of Andrographispaniculata leaves on inflammatory and allergic mediators *in vitro*. J Ethnopharmacol 2010;129:203-7.
- Jimenez GS, Aquino CR, Martinez LC, Torres KB, Monroy MR. Antioxidant activity and content of Phenolic compounds and flavonoids from Justiciaspicigera. J Biol Sci 2009;9:629-32.
- 27. Charoenchai P, Vajrodaya S, Somprasong W, Mahidol C, Ruchirawat S, Kittakoop P. Antiplasmodial, acytotoxic, radical scavenging and antioxidant activities of thai plants in the family acanthaceae. Planta Med 2010;76:1940-43.
- Ashour MAG. Isolation, HPLC/UV Characterization and antioxidant activityof phenylethnoids from Blepharisedulis (Forssk.) pers. Growing in Egypt. Bull Faculty Pharm Cairo Univer 2012;50:67-72.
- Bhragual DD, Kumar N, Garg VK, Sharma PK. Review on plants having Hepatoprotective activity. J Pharm Res 2010;3:2077-82.
- Amin E, Radwan MM, Hawary SSE, Fathy MM, Mohammad R, Becnel JJ, *et al.* Potent insecticidal secondary metabolites from medicinal plant acanthus montanus. Record Nat Pro 2012;6:301-5.
- 31. Kanchanapoom T, Kamel MS, Kasai R, Picheansoonthon G, Hiraga Y, Yamasaki K. Benzoxanoid glucosides from Acanthus ilicifolius. Phytochem 2001;58:637-40.
- 32. Kanchanapoom T, Ruchirawat S. Megastigmane glucoside from Asystasiagangetica (L.). J Nat Medicine 2007;61:430-3.
- 33. Kanchanapoom T, Noiarsa P, Ruchirawat S, Kasai R, Otsuka H. Triterpinoidal glycosides from Justiciabetonica. Phytochem 2004;65:2613-18.
- Mawla AMAAE, Ahmed AS, Ibraheim ZZ, Ernst L. Phenylethnoid glycosides from Barleriacristata L. callus cultures. Bull Pharm Sci 2005;28:199-204.
- 35. Noiarsa P, Ruchirawat S, Kanchanapoom T. Acanmontanoside, a new phenyl ethanoid diglycoside from acanthus montanus. Molecules 2010;15:8967-72.
- 36. Worawittayanon P, Ruadreo J, Disadee W, Sahakitpichan P, Sitthimonchai S, Thasana N, *et al.* Iridoid and flavone glycosides from Asystasiagangetica subsp. micrantha and their antioxidant activities. Biochem Systematics Ecol 2012;40:38-42.
- Hemalatha K, Hareeka N, Sunitha D. Chemical constituents isolated from leaves of barleriacristata linn. Int J Pharm Bio Sci 2012;3:609-15.
- Bumrela SB, Naik SR. Phytochemical and morphoanatimical studies for establishing quality standards of Dipteracanthuspatulus (Jacq.) Nees. Int J Pharm Biomed Res 2011;2:98-103.
- Berrondo LF, Gabriel FT, Fernandes SBDO, Menezes FDS. Dirhamnosyl flavonoid and other constituents from Brillantaisiapalisatii. Quim Nova 2003;26:922-3.

- 40. Rao YK, Vimalamma G, Rao CV, Tzeng YM. Flavonoids and andrographolides from Andrographispaniculata. Phytochem 2004;65:2317-21.
- Hemalatha K, Hareeka N, Sunitha D. Chemical constituents isolated from leaves of barleriacristata linn. Int J Pharma Bio Sci 2012;3:609-15.
- Bravo HR, Copaja SV, Lazo W. Antimicrobial activity of Natural 2-Benzoxazolines and related derivatives. J Agricultureand Food Chem 1997;45:3255-7.
- Grimes BM, McBeth DL, Hallihan B, Delph S. Antimicrobial activity in medicinal plants of the Scrophulariaceae and Acanthaceae. J Pharm Biol 1996;34:243-8.
- 44. Roia FC, Smith RA. The antibacterial screening of some common ornamental plants. Economic Bot 1977;31:28-37.
- Kohno k, Miyake M, Sano O, Katoka MT, Yamamota S, Miyata SK, et al. Anti-inflammatory and Immunomodulatory properties of 2-Amino-3-H-phenoxazine. Biology Pharm Bull 2008;31:1938-45.
- Zikmundova M, Drandarov K, Bigler L, Hesse M, Werner C. Biotransformation of 2-benzoxazolinone (BOA) and 2-hydroxy-1,4-benzoxazin-3-one (HBOA) by endophytic fungi isolated from Aphelandra tetragona. Appl Environ Microbiol 2002;68:4863-70.
- 47. Hedberg C, Hesse M, Werner C. Spermine and spermidine hydroxyl cinnamoyl transeferases in Aphelandra tetragona. Plant Sci 1996;113:149-56.
- Warner C, Hu W, Riatsch AL, Hesse M. Dicoumaroylspermidines and Tri-coumaroylspermidines in anthers of different species of genus Aphelandra. Phytochem 1995;40:461-5.
- Tawil BF, Zhu JP, Piantini U, Hesse M. New spermine alkaloids from aphelandra tetragona (vahl) nees. Helvetica Chimica Acta 2004;72:180-4.
- Youhnovski N, Filipov S, Linden A, Guggisberg A, Werner C, Hesse M. Two macrocyclicspermine alkaloids from Aphelandra fuscopunctata (Acanthaceae). Phytochem 1999;52:1717-23.
- Baumeler A, Hesse M, Werner C. Benzoxazinoids cyclic hydroxamic acids, lactams and their corresponding glucosides in the genus Aphelandra (Acanthaceae). Phytochem 2000;53:213-22.
- 52. Hedberg C, Hesse M, Werner C. Spermine and spermidine hydroxyl cinnamoyl transeferases in Aphelandra tetragona. Plant Sci 1996;113:149-56.
- 53. Bratoeff EA, Amador PMC. Phytochemical study of Aphelandra aurantica Scheider (Acanthaceae). Phyton-Int J Exper Bot 1994;56:27-32.