Original Article

ANTIMICROBIAL AND ANTI-INFLAMMATORY STUDIES ON SARGASSUM WIGHTII EXTRACTS

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Received: 08 June 2014 Revised and Accepted: 20 Jul 2014

ABSTRACT

Objective: To evaluate the antimicrobial and anti-inflammatory effect of hexane, chloroform and ethanolic extract of Sargassum wightii.

Methods: Hexane, chlorofom and ethanolic extractions of these algae were done and evaluated for biological activity. Assay of antimicrobial activity was performed by disc diffusion method against various microorganisms. Assay of anti-inflammatory activity was performed in albino rats with paw edema, induced by Carrageenan.

Results: Chloroform and ethanol extracts exhibited antimicrobial activity against various microorganisms. All the extracts revealed antiinflammatory activity, in which chloroform extract showed maximum activity in dose dependent manner.

Conclusions: Among the three different extracts tested, chloroform extract of *S. wightii* possess significant antimicrobial and anti-inflammatory activities. Further in-depth studies, could result in the development of a good antimicrobial and anti-inflammatory agent from this chloroform extract of *S.wightii*.

Keywords: Antimicrobial, Anti-inflammatory, Sargassum wightii.

INTRODUCTION

Algae are a group of organisms with little tissue differentiation, no true vascular tissues, no roots, stems or leaves and no flowers. Based on the pigments present and morphological and anatomical features, they are classified as Blue green algae (Cyanophyceae/ Myxophyceae), Green algae (Chlorophyceae), Brown algae (Phaeophyceae), and Red algae (Rhodophyceae).

The brown algae belong to a large group, the heterolontophyta, a eukaryotic group of organism distinguished most prominently by having chloroplasts surrounded by four membranes. There are about 2000 multicellular marine algae species which include many seaweed of colder northern hemisphere waters. They play an important role in the marine environment as both food and for the habitats they form [1]. *Sargassum wightii* is marine algae that grows 20-30 cm in length with a well marked holdfast, upper portion richly branched. They are dark-brown in color. They grow in intertidal and subtidal habitats [2]. Since they contain 8-10% mannitol, can be used as a substitute for sugar, fertilizer and medicine [3]. The remarkable medicinal properties include antibacterial, antifungal, antiprotozoal, antifertility, antiviral, anticancer, and a scope of pharmacological, hypoglycemic and antimalarial activities [4] [5] [6].

MATERIALS AND METHODS

Collection of seaweed material

The fresh seaweed of the *Sargassum wightii* was collected around the Mandapam region, Rameswaram, Tamilnadu, India. The seaweed was taxonomically identified and authenticated by Dr V.T. Sridharan, Professor (Rtd.), Department of Botany, National College, Trichy. The whole seaweed material was shade dried and coarsely powdered.

Preparation of extracts

Nearly 1000g of shade dried coarse material was subsequently extracted with sufficient volume of various organic solvents in the order of increasing polarity and various extracts (Hexane, Chloroform and ethanol) were prepared. Then the extract was filtered and evaporated using a vacuum rotary evaporator at 40°C.

Antimicrobial Activity

Microorganisms

Pure bacterial cultures were purchased from the microbial type culture collection (MTCC), Chandigarh. The various extracts of *S.wightii* were tested against various microorganisms such as *E. coli* (MTCC 118), *Bacillus cereus* (MTCC 2389), *Bacillus subtilis* (MTCC 1305), *Klebsiella pneumoniae* (MTCC 1305), *Staphylococcus aureus* (MTCC 737), *Streptococcus pyogenes* (MTCC 1924), *Salmonella typhimurium* (MTCC 1253) and *Proteus vulgaris* (MTCC 1771).

Method

Assay of antimicrobial activity was performed by disc diffusion method. The Muller-Hinton agar plates were prepared and the organism was swabbed over it using a sterile cotton swab. The antimicrobial discs were placed on the surface of the agar plates and then, the plates were incubated at 37°C for 24 hrs. After incubation, the zone of inhibition was measured [7].

Anti-inflammatory activity

Animals used

Albino rats of either sex weighing 250-300g were used. The animals were maintained under suitable conditions (Temperature $25\pm2^{\circ}c$) with dark and light cycle and fed with standard dry pellets and water ad libitum throughout the experiment. The experiment was carried out after getting the necessary approval from the institutional animal ethical committee (Reg.No:790/03/ac/CPCSEA).

Carrageenin Induced Paw Edema

All rats were induced for inflammation by using 0.1ml of 1% carrageenin (w/v) solution in normal saline injected into the right hind paw of the albino rats for creating inflammation or edema [8]. Rats weighing between 250-300g were selected and divided into eleven groups of six each.

Group 1: Control group

Group 2: Sargassum wightii 100 mg/kg, p.o (Hexane extract) Group 3: Sargassum wightii 200 mg/kg, p.o (Hexane extract) Group 4: Sargassum wightii 300 mg/kg, p.o (Hexane extract) Group 5: Sargassum wightii 100 mg/kg, p.o (Chloroform extract) Group 6: Sargassum wightii 200 mg/kg, p.o (Chloroform extract) Group 7: Sargassum wightii 300 mg/kg, p.o (Chloroform extract) Group 8: Sargassum wightii 100 mg/kg, p.o (Ethanol extract) Group 9: Sargassum wightii 200 mg/kg, p.o (Ethanol extract) Group 10: Sargassum wightii 300 mg/kg, p.o (Ethanol extract)

Group 11: Standard - Indomethacin 10 mg/kg, p.o Extracts and standard drug, Indomethacin [9] were administrated orally in the albino rats. 0th hour and 3rd hour treatment i.e., right hind paw was measured (mm) and the values were statistically analyzed. The values were used to identify the percentage inhibition in paw volume by using the formula, = Control (% increase in paw volume in third hour) – Test (% increase in paw volume in third hour) X 100 / Control (% increase in paw volume in third hour)

RESULTS

4.1 Antimicrobial activity

The various extracts of *S. wightii* were tested against various microorganisms such as *E. coli, Bacillus cereus, Bacillus subtilis, Klebsiella pneumoniae, Staphylococcus aureus, Streptococcus*

 $pyogenes,\ Salmonella\ typhimurium\ and\ Proteus\ vulgaris\ using\ different\ concentrations\ viz.,\ 0.8mg,\ 1.2mg,\ 1.6mg,\ 2mg\ /\ disc\ and\ the\ results\ were\ tabulated.$

Hexane extract

The Hexane extract of the *Sargassum wightii* exhibited no zone of inhibition and hence hexane extract possess no significant antimicrobial activity against tested microorganisms.

Chloroform extract

The chloroform extract of the *Sargassum wightii* tested against various microorganism and the results were presented in table 1. It showed no zone of inhibition against two microorganisms viz., *Streptococcus pyogenes* and *Bacillus subtilis* and exhibited maximum antimicrobial activity against *Klebsiella pneumoniae* (Fig 1).

Ethanol extract

Ethanol extract of *S. wightii* was tested against various microorganisms and found that it showed no activity against four microorganisms viz, *E. coli, Bacillus cereus, Staphylococcus aureus, Proteus vulgaris* and maximum antimicrobial activity against *Salmonella typhimurium* (Table 2 and Fig 2).

Table 1: Antimicrobial activity	of chloroform extract of the Sara	assum wiahtii

S. No.	Microorganism	Chloroform extract of S.wightii					
		0.8 mg	1.2 mg	1.6 mg	2 mg	Negative control	Positive control
1.	Bacillus cereus	6 mm	7 mm	10 mm	11mm	-	39 mm
2.	Klebsiella pneumonia	6mm	6 mm	6 mm	13 mm	-	10 mm
3.	Proteus vulgaris	10 mm	11 mm	11 mm	12 mm	-	8 mm
4.	Salmonella typhimurium	9 mm	12mm	13mm	-	-	8 mm
5.	E. coli	6 mm	6 mm	9 mm	11 mm	-	7 mm
6.	Staphylococcus	9 mm	9 mm	10 mm	10 mm	-	8 mm
	Aureus						

Antimicrobial activity of chloroform extract of the Sargassum wightii



A) Bacillus cereus



C) Proteus vulgaris



B)Klebstella pneumontae



D)Salmonella typhimurium

Fig. 1: Antimicrobial activity of chloroform extract of the Sargassum wightii

Antimicrobial activity of chloroform extract of the Sargassum wightii







F)Staphylococcus aureus

Antimicrobial activity of Ethanol extract of the Sargassum wightii





A)Salmonella typhimurium

Antimicrobial activity of Ethanol extract of the Sargassum wightii





C) Bacillus subtillis

D) Streptococcus pyogenes

Fig. 2: Antimicrobial activity of Ethanol extract of the Sargassum wightii

In-vivo studies

Anti-inflammatory studies

Carrageenan induced hind paw edema: Treatment with three different extracts viz hexane, chloroform and ethanol extract of *S.wightii* (100, 200, and 300 mg/kg.) showed a significant inhibition of paw volume. It was observed that the extracts exhibited dose dependant anti-inflammatory activity and maximum was observed at a dose of 300 mg/kg of the chloroform extract against Carrageenan induced hind paw edema (Table 3, 4 and 5).

Table 2: Antimicrobial activity of Ethanol extract of the Sargassum wightii

S. No.	Microorganism	Ethanol					
		0.8 mg	1.2 mg	1.6 mg	2 mg	Positive control	Negative control
1.	Salmonella typhimurium	6 mm	6 mm	7mm	12 mm	-	7 mm
2.	Klebsiella pneumoniaee	6 mm	6 mm	8mm	10mm	-	8 mm
3.	Bacillus subtillis	6 mm	6 mm	6 mm	11mm	-	10 mm
4.	Streptococcus pyogenes	6 mm	6 mm	8mm	9mm	-	10 mm

The antimicrobial properties in *Sargassum wightii* may be due to multiple inhibitory properties found in algae. The results were encouraging for further in depth studies, could result in a good antibacterial agent from this marine algal source.

Table 2. Anti-inflammatory activit	w of Hovano ovtract of Saraassu	ım wiahtii əqəinct Cərrəqoon	an induced new Edema in Albino rate
Table 5. Anti-minaminatory activit	ly of fiexalle extract of Surgussu	ini wiyinni agamsi carrageen	an muuceu paw Euema in Albino rats

	% increase in paw vo	olume Mean ± S.E (n = 6)	
Treatment	In 0 min	In 3 hrs	% inhibition in paw volume
Control (0.5 ml/kg)	37.25 ± 3.21	126.42 ± 9.52	-
100 mg of Hexane extract	32.25 ± 2.8	70.70 ± 6.3	44.07
200 mg of Hexane extract	30.64 ± 2.7	63.5±4.2	49.77
300 mg of Hexane extract	33.64 ± 2.7	51.54 ± 5.1	59.23
Indomethacin	31.64 ± 2.7	34.32 ± 3.1	72.85
(10 mg/kg)			

Table 4: Anti-inflammatory activity of Chloroform extract of Sargassum wightii against Carrageenan induced paw Edema in Albino rats

	% increase in paw v	olume Mean ± S.E (n = 6)	
Treatment	In 0 min	In 3 hrs	% inhibition in paw volume
Control (0.5 ml/kg)	37.25 ± 3.21	126.42 ± 9.52	-
100 mg of Chloroform extract	33.25 ± 2.8	56.32 ± 6.3	55.45
200 mg of Chloroform extract	31.25 ± 1.12	48.32±1.3	61.77
300 mg of Chloroform extract	28.25 ± 1.12	40.5 ± 2.2	67.95
Indomethacin	31.64 ± 2.7	34.32 ± 3.1	72.85
(10 mg/kg)			

Table 5: Anti-inflammatory activity of Ethanol extract of Sargassum wightii against Carrageenan induced paw Edema in Albino rats

	% increase in paw vo	olume Mean ± S.E (n = 6)	
Treatment	In 0 min	In 3 hrs	% inhibition in paw volume
Control (0.5 ml/kg)	37.25 ± 3.21	126.42 ± 9.52	-
100 mg of Ethanol extract	32.64 ± 2.7	79.72 ± 5.1	36.94
200 mg of Ethanol extract	33.52 ± 2.7	65.3 ±4.6	48.34
300 mg of Ethanol extract	34.23 ± 1.12	54.25 ± 2.2	57.08
Indomethacin	31.64 ± 2.7	34.32 ± 3.1	72.85
(10 mg/kg)			

DISCUSSION

Antimicrobial activity of various extracts of *S.wightii* revealed that the hexane extract has no significant antimicrobial activity against the tested microorganism. Chloroform and ethanol extracts exhibited significant antimicrobial activity, whereas the chloroform extract showed activity against six tested microorganism viz., *Klebsiella pneumoniaee, Bacillus cereus, Staphylococcus aureus, Salmonella typhimurium, Proteus vulgaris, and E.coli.*

Carrageenan induced hind paw edema study is a model used to analyze the acute inflammation mechanism. Carrageenan induced inflammatory response involves three phases of chemical mediators released in succession.[10] In the first phase (1 h) histamine and serotonin are released and is characterized by an increase in vascular permeability.

The second phase (2 h) is mediated by the release of bradykinin, an important chemical mediator of both pain and inflammation. Release of prostaglandins and cyclooxygenases products takes place in the third and final phase (3 h). It was observed that there was inhibition of edema formation after 3 hours, which indicated the acute antiinflammatory activity. The chloroform extract of *S. wightii* showed an inhibition of 67.95% at a dose of 300 mg/kg, which was significant as the standard drug, which exhibited an inhibition of 72.85%. The drug showed inhibition at 3 hours which means the drug exerts its anti-inflammatory activity by preventing prostaglandin biosynthesis through inhibition of the COX enzyme.

CONCLUSION

Among the three different extracts tested for antimicrobial and antiinflammatory activities, chloroform extract of *S. wightii* possess significant antimicrobial and anti-inflammatory activities. Further in-depth studies, could result in the development of a good antimicrobial and anti-inflammatory agent from this chloroform extract of marine source.

CONFLICT OF INTERESTS

Declared None

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