



# Complete Genome Sequences of 12 Quinolone-Resistant *Escherichia coli* Strains Containing *qnrS1* Based on Hybrid Assemblies

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**ABSTRACT** In total, 12 quinolone-resistant *Escherichia coli* (QREC) strains containing *qnrS1* were submitted to long-read sequencing using a FLO-MIN106 flow cell on a MinION device. The long reads were assembled with short reads (Illumina) and analyzed using the MOB-suite pipeline. Six of these QREC genome sequences were closed after hybrid assembly.

The presence of quinolone-resistant *Escherichia coli* (QREC) in the animal reservoir is a potential public health concern, especially related to plasmid-mediated quinolone resistance genes, as they might spread to more pathogenic bacteria. The *qnrS1* gene is known to be situated on plasmids with different incompatibility (Inc) groups (1, 2). Here, we aimed to select QREC strains encoding *qnrS1* on plasmids with different Inc groups to complete circular plasmid contigs.

We previously sequenced 280 QREC isolates from broilers, pigs, red foxes, and wild birds, collected through the NORM-VET program from 2006 to 2017, using short-read sequencing (Illumina, San Diego, CA) (3). The samples were either selectively isolated on MacConkey agar containing 0.06 mg/liter ciprofloxacin or randomly collected from *E. coli* isolated on MacConkey agar. In total, 12 QREC isolates encoding *qnrS1* from these four animal species were selected for long-read sequencing. Here, we report the hybrid assembly of these isolates, including six closed genome sequences. The hybrid assemblies were further analyzed using MOB-suite (4).

Extraction of genomic DNA was performed using the Genomic-tip 100/G kit (Qiagen, Hilden, Germany). Bacteria were enriched overnight at 37°C in 2 to 3 ml heart infusion broth (Difco, Omagh, UK). The DNA concentration was determined using the Qubit double-stranded DNA (dsDNA) broad-range (BR) assay kit (Thermo Fisher Scientific, Waltham, MA, USA), and the DNA was quality assessed using a NanoDrop One spectrophotometer (Thermo Fisher Scientific). Approximately 400 ng of high-quality DNA was subjected to library preparation using a rapid barcoding kit (SQK-RBK004; Oxford Nanopore Sequencing [ONT], Oxford, UK). Four samples were run with smaller amounts (104, 154, 324, and 369 ng), as only a maximum volume of 7.5  $\mu$ l of template was allowed into the library preparation reaction. The constructed libraries were indexed using barcodes RB1 to RB12, loaded onto a FLO-MIN106 flow cell on a MinION device (Oxford Nanopore Sequencing), and run for 40 h. The raw sequence data were base called separately after the run using Guppy v.3.4.5 (5) and demultiplexed using qcat v.1.1.0 (ONT, <https://github.com/nanoporetech/qcat>). The sequence quality of the demultiplexed data sets was checked with NanoPlot v.1.30.0 (6). Default parameters were used for all software unless otherwise specified.

Canu v.1.9 (7) was used to improve the accuracy of the long reads, followed by Filtlong v.0.2.0 (<https://github.com/rrwick/Filtlong>) to remove reads of <1,000 bp from the corrected long reads. Hybrid assemblies were generated using Unicycler v.0.4.8 (8),

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**TABLE 1** Characteristics and accession numbers of the quinolone-resistant *Escherichia coli* qnrS1 strains

Strain	Plasmid Inc type (pMLST)	ST <sup>a</sup>	No. of Illumina reads for:			Data for Nanopore reads:			Total size (Mbp)	Replicon size (bp)	GC content (%)	No. of genes	Coverage (x)	ENA accession no. for:	
			Read 1	Read 2	No. of reads	Avg length (bp)	No. of contigs	No. of reads						Raw reads	Assembly
2015-01-2097	IncX1 <sup>b</sup>	1421	818,798	865,881	331,312	5,292.7	2 <sup>c</sup>	4.68	21,374	50.8	4,507	275.1	ERR4592247	LR881940.1	
2015-01-466	IncF (F-A1:B1)	10	761,941	819,989	211,523	5,227.7	5 <sup>c</sup>	4.87	113,096	44.5	28	437.9	ERR4592248	LR881941.1	
	IncH								87,822	50.6	4,695	247.4		LR882052.1	
	IncF <sup>b</sup> (F2:A-B-)								50,909	52.6	137	256.3		LR882053.1	
	IncX1								46,065	47.9	96	164.3		LR882054.1	
2016-02-324	IncF <sup>b</sup> (F-A-B53)	7036	654,152	713,188	258,316	4,232.1	2 <sup>c</sup>	4.90	94,955	53.0	63	316.1	ERR4592249	LR882055.1	
2016-02-418	IncX1 <sup>b</sup>	58	596,773	650,657	174,481	2,309.8	29	4.96	46,447 <sup>d</sup>	40.7	55	294.5	ERR4592250	LR882056.1	
2016-02-522	IncY <sup>b</sup>	1011	795,118	867,426	166,584	4,176.7	4	4.94	78,634	51.0	108	225.2	ERR4592251	LR882051.1	
2016-02-620	IncX3 <sup>b</sup>	694	676,465	740,782	438,687	3,794.8	5	4.71	44,425	52.8	4,786	191.0	ERR4592252	CAJGEF01	
2016-17-164	IncF <sup>b</sup> (F89:A-B53)	7593	654,299	713,350	588,805	2,983.2	8	4.93	118,361	50.8	55	310.5	ERR4592253	CAJGEG01	
2016-17-292	IncF (F24:A-B1)	23	695,093	720,319	310,224	5,196.4	3 <sup>c</sup>	4.99	97,083	50.1	133	106.0	ERR4592254	LR882493.1	
2016-17-363	IncI2	48	761,196	825,502	404,780	2,644.6	5	4.67	59,944	48.7	99	121.3	ERR4592255	LR882494.1	
2016-17-550	IncH <sup>b</sup> (unknown)	2165	988,537	1,058,892	218,828	4,398.6	2 <sup>c</sup>	4.82	86,214	42.1	83	136.4	ERR4592256	LR882495.1	
2015-01-2838	IncY <sup>b</sup>	117	388,306	418,338	129,950	3,457.6	15	5.14	104,732	50.7	100	221.7	ERR4592257	CAJGWN01	
2014-01-7375	IncX2 <sup>b</sup>	453	472,494	482,585	209,994	4,667.7	5 <sup>c</sup>	5.27	39,630	48.0	118	128.2	ERR4592258	LR883965	
	IncI1								98,997	50.7	4,899	98.0	ERR4592259	LR883966	
	IncF (F-A-B56)								82,142	46.0	50	337.3		CAJGWP01	
	IncX1 <sup>b</sup>								47,686	50.6	5,119	34.1		LR882057.1	
	IncF (F-A-B114)								42,660	49.4	110	62.5		LR882058.1	
									42,660	47.8	89	46.4		LR882059.1	
									42,660	43.1	56	64.0		LR882060.1	
									42,660	52.5	54	88.3		LR882061.1	

<sup>a</sup> ST, sequence type.

<sup>b</sup> Plasmid with qnrS.

<sup>c</sup> Genome closed.

<sup>d</sup> Plasmid not circularized.

followed by Prokka v.1.14.5 (9) to annotate the hybrid assemblies. The GC content of each assembly was calculated using the EMBOSS v.6.6.0 (10) commands “union” and “infoseq.” MOB-suite v.1.4.9 (4) was used to predict plasmid sequences from the hybrid assemblies and identify their respective replicon types. Each plasmid FASTA file generated by MOB-suite was subjected to ResFinder v.4.0 (11), VirulenceFinder v.2.0 (12), and PlasmidFinder v.2.1 (13). Plasmids containing *qnrS1* were confirmed by genome annotation with Prokka. The Illumina reads were mapped back to the assembly using BWA v.0.7.17 (14), and the depth of coverage was calculated using SAMtools v.1.10 (15) using the depth (genome-wide) and coverage (replicon) options.

The characteristics and accession numbers are presented in Table 1. The plasmid assemblies with Inc groups that allowed further typing were run on pMLST v.2.0 (13) on the Center for Epidemiology Genomics website to further determine the respective replicon types.

**Data availability.** All data sets are deposited in ENA under accession number PRJEB40547 (Table 1).

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