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# Multitarget Stool DNA Testing for Colorectal-Cancer Screening

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#### ABSTRACT

#### **BACKGROUND**

An accurate, noninvasive test could improve the effectiveness of colorectal-cancer screening.

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#### **METHODS**

We compared a noninvasive, multitarget stool DNA test with a fecal immunochemical test (FIT) in persons at average risk for colorectal cancer. The DNA test includes quantitative molecular assays for *KRAS* mutations, aberrant *NDRG4* and *BMP3* methylation, and  $\beta$ -actin, plus a hemoglobin immunoassay. Results were generated with the use of a logistic-regression algorithm, with values of 183 or more considered to be positive. FIT values of more than 100 ng of hemoglobin per milliliter of buffer were considered to be positive. Tests were processed independently of colonoscopic findings.

# RESULTS

Of the 9989 participants who could be evaluated, 65 (0.7%) had colorectal cancer and 757 (7.6%) had advanced precancerous lesions (advanced adenomas or sessile serrated polyps measuring ≥1 cm in the greatest dimension) on colonoscopy. The sensitivity for detecting colorectal cancer was 92.3% with DNA testing and 73.8% with FIT (P=0.002). The sensitivity for detecting advanced precancerous lesions was 42.4% with DNA testing and 23.8% with FIT (P<0.001). The rate of detection of polyps with high-grade dysplasia was 69.2% with DNA testing and 46.2% with FIT (P=0.004); the rates of detection of serrated sessile polyps measuring 1 cm or more were 42.4% and 5.1%, respectively (P<0.001). Specificities with DNA testing and FIT were 86.6% and 94.9%, respectively, among participants with nonadvanced or negative findings (P<0.001) and 89.8% and 96.4%, respectively, among those with negative results on colonoscopy (P<0.001). The numbers of persons who would need to be screened to detect one cancer were 154 with colonoscopy, 166 with DNA testing, and 208 with FIT.

# CONCLUSIONS

In asymptomatic persons at average risk for colorectal cancer, multitarget stool DNA testing detected significantly more cancers than did FIT but had more false positive results. (Funded by Exact Sciences; ClinicalTrials.gov number, NCT01397747.)

ana University School of Medicine, the Regenstrief Institute, the Simon Cancer Center, and the Center for Innovation at Roudebush Veterans Affairs Medical Center — all in Indianapolis (T.F.I.); the Departments of Medicine and Epidemiology and the Lineberger Comprehensive Cancer Center, University of North Carolina at Chapel Hill, Chapel Hill (D.F.R.); the Dr. Henry D. Janowitz Division of Gastroenterology, Department of Medicine, Icahn School of Medicine at Mount Sinai, New York (S.H.I.); Kaiser Permanente Medical Center, Walnut Creek, CA (T.R.L.); Boston Biostatistics Research Foundation, Framingham MA (P.L.); Exact Sciences, Madison, WI (G.P.L., B.M.B.); and the Division of Gastroenterology and Hepatology, Mayo Clinic, Rochester, MN (D.A.A.). Address reprint requests to Dr. Imperiale at Indiana University Medical Center-Regenstrief Institute, 1050 Wishard Blvd., Indianapolis, IN 46202.

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N Engl J Med 2014;370:1287-97. DOI: 10.1056/NEJMoa1311194 Copyright © 2014 Massachusetts Medical Society. OLORECTAL CANCER IS A MAJOR CAUSE of death and disease among men and women in the United States.<sup>1</sup> The underlying neoplastic processes of colorectal carcinogenesis lend themselves to screening.<sup>2</sup> Evidence supports and guidelines endorse several tests and strategies,<sup>3-5</sup> and screening for colorectal cancer has been found to be cost-effective.<sup>5-7</sup>

Despite the supporting evidence, recommendations, and availability of several screening tests, a substantial proportion of the U.S. population is not up to date with screening.<sup>8</sup> A simple, noninvasive test with high sensitivity for both colorectal cancer and advanced precancerous lesions might increase uptake and adherence rates, which could improve clinical outcomes.

Colorectal cancer arises from accumulated genetic and epigenetic alterations, which provide a basis for the analysis of stool to identify tumorspecific changes.9 Large-scale screening studies of previously available stool-based DNA tests showed only fair sensitivity for the detection of colorectal cancer (i.e., the capacity to detect cancers, or true positive tests [see Glossary]) and low sensitivity for the detection of advanced adenomas.10,11 Important advances have since been incorporated, including the use of a stabilizing buffer,12,13 more discriminating markers,14,15 more sensitive analytic methods, 14,16,17 automation, 16 and an overall determination of results with the use of a logistic-regression algorithm, which together result in higher sensitivity for the detection of both cancer and advanced precancerous lesions.14,16 However, evaluation of the more recent tests was based largely on analyses of archived specimens, including those collected from patients after the diagnosis but before the resection of colorectal cancer or advanced precancerous polyps.

In this study, we evaluate the multitarget stool DNA test as a tool for screening. The primary aim was to determine the performance characteristics of the DNA test in the detection of colorectal cancer. The secondary aims were to determine the performance of the DNA test in the detection of advanced precancerous lesions and to compare it with a commercially available fecal immunochemical test (FIT) for human hemoglobin in the detection of both colorectal cancer and advanced precancerous lesions.

#### METHODS

### STUDY DESIGN

From June 2011 through November 2012, we enrolled participants in this cross-sectional study at 90 sites throughout the United States and Canada, including private-practice and academic settings. The study was approved by the institutional review board at each site, and all participants provided written informed consent.

The study, which was funded by Exact Sciences, was designed by the authors; Health Decisions, a contract research organization, gathered and monitored the data. The first author wrote the first draft of the manuscript, incorporating the other authors' contributions; one of the authors, who is a statistician, analyzed the data and, along

## **Glossary of Screening Terms**

Sensitivity (true positive rate): The proportion of persons with disease who have a positive test (positive test results among persons with disease).

**Specificity (true negative rate):** The proportion of persons without disease who have a negative test (negative test results among persons without disease).

False negative rate (1 minus sensitivity): The proportion of persons with disease who have a negative test (negative test results among persons with disease).

False positive rate (1 minus specificity): The proportion of persons without disease who have a positive test (positive test results among persons without disease).

**Positive predictive value:** The proportion of persons with disease among those with a positive test (disease present among those with positive test results).

**Negative predictive value:** The proportion of persons without disease among those with a negative test (disease absent among those with negative test results).

**Number needed to screen:** The number of persons who would need to be screened to identify one person with the disease.

with the last author, vouches for the data and adherence to the study protocol, which is available with the full text of this article at NEJM.org. All the authors signed confidentiality agreements with Exact Sciences.

#### STUDY POPULATION

The target population was asymptomatic persons between the ages of 50 and 84 years who were considered to be at average risk for colorectal cancer and who were scheduled to undergo screening colonoscopy. Enrollment was weighted toward persons 65 years of age or older in order to increase the prevalence of cancer. We excluded participants who had a personal history of colorectal neoplasia, digestive cancer, or inflammatory bowel disease; had undergone colonoscopy within the previous 9 years or a barium enema, computed tomographic colonography, or sigmoidoscopy within the previous 5 years; had positive results on fecal blood testing within the previous 6 months; had undergone colorectal resection for any reason other than sigmoid diverticula; had overt rectal bleeding within the previous 30 days; had a personal or family history of colorectal cancer; had participated in any interventional clinical study within the previous 30 days; or were unable or unwilling to provide written informed consent.

### CLINICAL PROCEDURES

All participants were required to provide a stool specimen and undergo screening colonoscopy within 90 days after providing informed consent. Stool was collected before routine bowel preparation. No dietary or medication restrictions were required. Colonoscopists were required to describe the extent of the examination, document cecal visualization, rate the quality of preparation (on a modified Aronchick scale),<sup>18</sup> and record the size and location of lesions.

Although colonoscopists reported the location and size of all lesions, only the most advanced colorectal epithelial lesion (the index lesion) and its location (proximal or distal) were used to categorize participants for the analysis. If two similarly advanced lesions were present, the larger of the two was designated as the index lesion. The proximal colon was considered to include the splenic flexure and all segments proximal to it, an insertion depth of more than 60 cm, or any

part described by the phrase "right colon"; the distal colon was considered to include all other segments, an insertion depth of 60 cm or less, or any part described by the phrase "left colon."

The biopsy and surgical specimens underwent histopathological analysis at the laboratory typically used by each study site. Polyps with highgrade dysplasia or 25% or more villous elements in adenomas measuring less than 1 cm, as well as sessile serrated or hyperplastic polyps measuring 1 cm or larger, were re-reviewed centrally by a gastrointestinal pathologist for confirmation, with diagnostic disagreements resolved by consensus of at least two central pathologists.

### PRIMARY AND SECONDARY OUTCOMES

The primary outcome was the ability of the DNA test to detect colorectal cancer (i.e., adenocarcinoma), with disease stage determined with the use of the American Joint Committee on Cancer (AJCC) staging system.¹9 The secondary outcome was the performance of the DNA test for the detection of advanced precancerous lesions, including advanced adenomas (high-grade dysplasia or with ≥25% villous histologic features or measuring ≥1 cm in the greatest dimension) and sessile serrated polyps measuring 1 cm or more in diameter.

#### LABORATORY PROCEDURES

A central biorepository received all stool specimens. Laboratory testing was performed without knowledge of the results of either the comparator FIT or clinical findings. (Details of stool collection and processing for DNA testing are shown in Fig. S1 in the Supplementary Appendix, available at NEJM.org.) Buffered stool samples were homogenized, separated into aliquots, and frozen at -80°C on receipt. Stool aliquots were subsequently sent in batches to one of three laboratories: Exact Sciences (Madison, WI), Mayo Medical Laboratory (Rochester, MN), and Molecular Pathology Laboratory Network (Knoxville, TN). Each laboratory received, in a blinded fashion, a similar distribution of specimens on the basis of colonoscopic findings.

The multitarget stool DNA test consists of molecular assays for aberrantly methylated *BMP3* and *NDRG4* promoter regions, mutant *KRAS*, and  $\beta$ -actin (a reference gene for human DNA quantity), as well as an immunochemical assay for

human hemoglobin. Quantitative measurements of each marker were incorporated into a validated, prespecified logistic-regression algorithm, with a value of 183 or more indicating that the test result was positive (for details, see the Supplementary Appendix). Analytic results were transferred to the study's biostatistician.

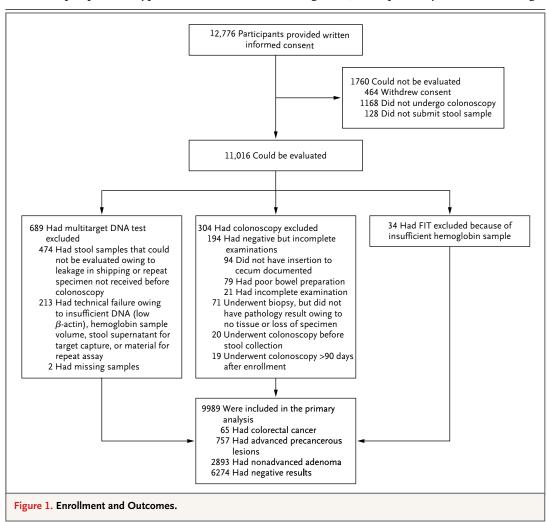
FIT (OC FIT-CHEK, Polymedco) was performed according to the manufacturer's instructions with the use of the same stool sample used for the DNA test.<sup>20</sup> Samples were refrigerated on receipt and sent in batches to a separate single laboratory for blinded analysis. Stool samples with more than 100 ng of hemoglobin per milliliter of buffer were considered to be positive.<sup>20</sup>

#### STATISTICAL ANALYSIS

The study was designed to have a power of 90% to test the prespecified hypothesis that the DNA

test would have a sensitivity of 65% or more for the detection of colorectal cancer (AJCC stages I through IV) under the null hypothesis, at a one-sided type I error rate of 0.05. A secondary hypothesis was to rule out a 5% noninferiority margin for sensitivity for the detection of colorectal cancer with the DNA test as compared with FIT, at a one-sided type I error rate of 0.05. Testing of the two hypotheses with a power of at least 80% required the diagnosis of 49 and 56 adjudicated colorectal cancers, respectively, which required the enrollment of 10,500 to 12,000 participants, under the assumption of a colorectal-cancer prevalence of 4.5 cases per 1000 population.

We conducted prespecified analyses to determine the sensitivity of the multitarget DNA test, as compared with FIT, for the detection of screening-relevant colorectal cancer (AJCC stages I through III); the specificity of the multitarget



DNA stool test (i.e., true negative rate), with advanced precancerous lesions on colonoscopy excluded and only nonadvanced adenomas and negative results included (the primary measure of specificity) and with only negative results included (the secondary measure of specificity); and the sensitivity of the multitarget stool DNA test, as compared with FIT, for the detection of advanced precancerous lesions. The analyses were based on data from all participants who had valid results on multitarget stool DNA testing, FIT, and colonoscopy; all reported subgroup analyses were prespecified.

For test characteristics, 95% lower boundaries were computed with the use of an exact binomial test. Lower 95% confidence limits for comparative analyses were computed with the use of a one-sided McNemar paired-comparisons test for the observed difference in sensitivity between the DNA test and FIT. The Hanley–McNeil method was used to calculate P values for the analysis of the receiver operating characteristic (ROC) curve.<sup>21</sup> There were no interim analyses of the data. All analyses were conducted with the use of SAS software, version 9.1, and StatXact software, version 7.

### RESULTS

#### STUDY POPULATION

A total of 12,776 participants were enrolled at 90 sites; 9989 of these participants (78.2%) had results that could be fully evaluated (Fig. 1). The participants whose results could be fully evaluated and those whose results could not be fully evaluated differed significantly with respect to mean age and race, although the magnitudes of the differences were small (Table S1 in the Supplementary Appendix).

A total of 65 participants who could be evaluated were found to have colorectal cancer on colonoscopy (prevalence, 0.7%). Of these participants, 60 had screening-relevant (stage I to III) cancers. A total of 757 participants who could be evaluated had advanced precancerous lesions (prevalence, 7.6%).

### **DNA TEST CHARACTERISTICS**

Multitarget stool DNA testing identified 60 of 65 participants with cancer, including 56 of the 60 participants with screening-relevant cancers, for respective sensitivities of 92.3% (95% confidence interval [CI], 83.0 to 97.5) and 93.3% (95% CI,

Most Advanced Finding	Colonoscopy (N = 9989)	Multitarget DNA Test (N = 9989)		FIT (N = 9989)	
		Positive Results	Sensitivity (95% CI)	Positive Results	Sensitivity (95% CI)
	no.	no.	%	no.	%
Colorectal cancer					
Any	65	60	92.3 (83.0–97.5)	48	73.8 (61.5–84.0)
Stage I to III*	60	56	93.3 (83.8–98.2)	44	73.3 (60.3–83.9)
Colorectal cancer and high-grade dysplasia	104	87	83.7 (75.1–90.2)	66	63.5 (53.5–72.7)
Advanced precancerous lesions†	757	321	42.4 (38.9–46.0)	180	23.8 (20.8–27.0)
Nonadvanced adenoma	2893	498	17.2 (15.9–18.6)	220	7.6 (6.7–8.6)
			Specificity (95% CI)		Specificity (95% CI)
All nonadvanced adenomas, non-neoplastic findings, and negative results on colonoscopy	9167	1231	86.6 (85.9–87.2)	472	94.9 (94.4–95.3)
Negative results on colonoscopy	4457	455	89.8 (88.9–90.7)	162	96.4 (95.8–96.9)

<sup>\*</sup> These stages of colorectal cancer, as defined by the system recommended by the American Joint Committee on Cancer, are associated with an increased rate of cure.

 $<sup>\</sup>dagger$  Advanced precancerous lesions include advanced adenomas and sessile serrated polyps measuring 1 cm or more.

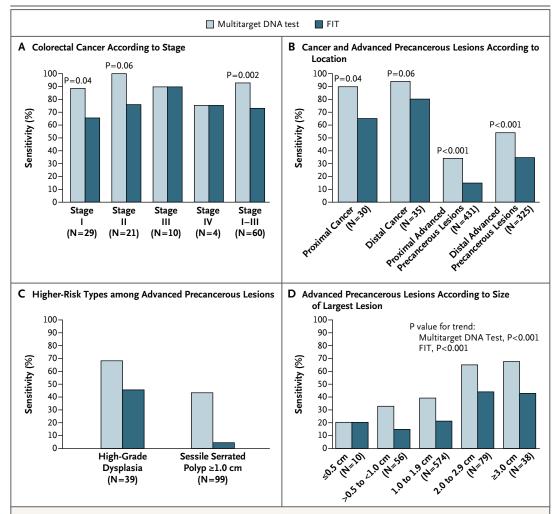


Figure 2. Sensitivity of the Multitarget Stool DNA Test and the Commercial Fecal Immunochemical Test (FIT), According to Subgroup.

Shown are the sensitivities of the DNA test and FIT for the detection of colorectal cancer according to tumor stage (Panel A), for the detection of colorectal cancer and advanced precancerous lesions according to the location in the colon (Panel B), and for the detection of higher-risk subtypes among participants with advanced precancerous lesions (Panel C) and according to lesion size (Panel D). The numbers in parentheses are the number of participants in each category. In Panel A, the stage of 1 of 65 colorectal cancers was not available. In Panel B, the location of 1 of 757 advanced precancerous lesions was not available.

83.8 to 98.2) (Table 1). Sensitivity did not vary significantly according to cancer stage (Fig. 2A) or location within the colon (Fig. 2B). Among 757 participants with advanced precancerous lesions, DNA testing detected 321 (42.4%; 95% CI, 38.9 to 46.0). A total of 69.2% (95% CI, 52.4 to 83.0) of 39 participants with high-grade dysplasia and 42.4% (95% CI, 32.6 to 52.8) of 99 par-

1 cm or larger were identified on DNA testing (Fig. 2C). The sensitivity of the DNA test was higher for distal advanced precancerous lesions (177 of 325 [54.5%; 95% CI, 48.9 to 60.0]) than for proximal lesions (143 of 431 [33.2%; 95% CI, 28.8 to 37.8]) (Fig. 2B); test sensitivity increased as the lesion size increased (Fig. 2D). The sensitivity for the detection of cancer or advanced preticipants with sessile serrated polyps measuring cancerous lesions did not differ significantly according to age or laboratory-testing site (data not shown).

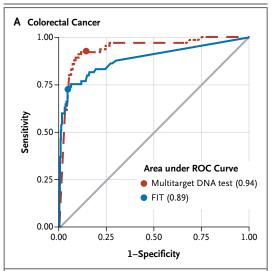
Among 9167 participants who had findings other than colorectal cancer or advanced precancerous lesions (e.g., nonadvanced adenomas or negative results), the specificity of the DNA test (true negative rate) was 86.6% (95% CI, 85.9 to 87.2). Among the 4457 participants with totally negative results on colonoscopy, the specificity was 89.8% (95% CI, 88.9 to 90.7); within this subgroup, the specificity was 94.0% among participants younger than 65 years of age and 87.1% among those 65 years of age or older (P<0.001).

#### **COMPARISON WITH FIT**

FIT detected 48 of 65 cancers (73.8%; 95% CI, 61.5 to 84.0), 44 of 60 AJCC stage I to III cancers (73.3%; 95% CI, 60.3 to 83.9), and 180 of 757 advanced precancerous lesions (23.8%; 95% CI, 20.8 to 27.0), findings that were all significantly inferior to those with DNA testing (Table 1). FIT detected 20 of 30 proximal cancers (66.7%) and 28 of 35 distal cancers (80.0%) (P=0.35 for the comparison between proximal and distal location). Comparative results for the detection of cancer according to stage and for higher-risk subsets of advanced precancerous lesions are shown in Figure 2. The DNA test was more sensitive than FIT for the detection of lesions with high-grade dysplasia (69.2% vs. 46.2%, P=0.004) or sessile serrated polyps measuring 1 cm or more (42.4% vs. 5.1%, P<0.001) (Fig. 2C) and for the detection of advanced precancerous lesions within the size ranges observed (Fig. 2D).

DNA testing detected 13 of 60 screening-relevant cancers that were undetected by FIT, whereas FIT detected 1 cancer that was undetected by DNA testing (P<0.001). DNA testing detected 170 of 757 advanced precancerous lesions (22.5%) that were undetected by FIT, whereas FIT detected 29 such lesions (3.8%) undetected by DNA testing (P<0.001).

Among 9167 participants with findings other than colorectal cancer or advanced precancerous lesions, the specificity of FIT was 94.9% (95% CI, 94.4 to 95.3). Among 4457 participants with negative results on colonoscopy, the specificity was 96.4% (95% CI, 95.8 to 96.9). In these two subgroups, the specificity values were superior to those of the DNA test (Table 1).



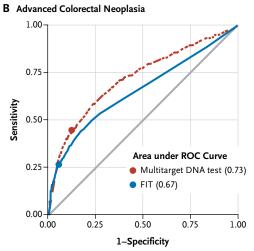


Figure 3. Receiver Operating Characteristic (ROC) Curves Comparing DNA Testing and FIT for the Detection of Colorectal Cancer and Advanced Colorectal Neoplasia.

Shown are ROC curves for the multitarget stool DNA test and FIT for the detection of colorectal cancer (Panel A) and advanced colorectal neoplasia (colorectal cancer plus advanced precancerous lesions) (Panel B). For colorectal cancer, the area under the ROC curve was 0.94 for the DNA test and 0.89 for FIT (95% confidence interval [CI] for the difference in area, 0.003 to 0.10; P=0.04). For advanced colorectal neoplasia, the area under the ROC curve was 0.73 for the DNA test and 0.67 for FIT (95% CI for the difference in area, 0.04 to 0.09; P<0.001). The respective performance thresholds were a value of 183 or more for the DNA test and more than 100 ng of hemoglobin per milliliter of buffer for FIT.

The specificity of FIT varied minimally according to age.

As measured by the area under the ROC curve (AUC), the discrimination between colorectal cancer and the combination of nonadvanced neoplasia and lesser findings was significantly higher with DNA testing than with FIT (0.94 vs. 0.89, P=0.04) (Fig. 3A); the AUC values for discrimination between advanced colorectal neoplasia (colorectal cancer plus advanced precancerous lesions) and all other findings were 0.73 and 0.67, respectively (P<0.001) (Fig. 3B). Positive and negative predictive values are shown in Table S2 in the Supplementary Appendix.

The isolated performance of the hemoglobin immunoassay component of the multitarget DNA test was similar to that of FIT, with specificities of 94.8% and 94.9%, respectively; sensitivities were 72.3% and 73.8%, respectively, for the detection of colorectal cancer and 22.7% and 23.8%, respectively, for the detection of advanced precancerous lesions.

Table 2 shows the number of persons who would need to be screened with colonoscopy, multitarget DNA testing, and FIT in order to detect one colorectal cancer (154 with colonoscopy, 166 with multitarget DNA testing, and 208 with FIT) and to detect one advanced precancerous polyp (13, 31, and 55 persons, respectively). These calculations show that multitarget DNA testing detected clinically significant lesions more efficiently than FIT.

# EXTRAPOLATION TO AN EXPANDED SCREENING POPULATION

In an extrapolation of our results to a hypothetical reference population of 10,000 participants at average risk for colorectal cancer, the various screening techniques of colonoscopy, DNA testing, and FIT would identify, respectively, 65, 60, and 48 persons with colorectal cancer; 758, 321, and 180 persons with advanced precancerous lesions;

2896, 498, and 220 persons with nonadvanced adenomas; and 6281, 732, and 248 persons with non-neoplastic findings or negative results on colonoscopy (Table 3).

The protocol specified the detection of colorectal cancer and advanced precancerous polyps as positive findings and the detection of nonadvanced adenomas as negative findings. In the hypothetical reference population of 10,000 persons, the numbers of persons who would be referred for colonoscopy on the basis of positive test results would be 1611 (16.1%) with DNA testing and 696 (7.0%) with FIT. Of the positive test results, the numbers that would be viewed as false positives would be 1230 of 1611 (76.4%) with DNA testing and 468 of 696 (67.2%) with FIT. Of 8389 negative results for DNA testing, 442 (5.3%) would be viewed as false negatives, consisting of 5 cancers and 437 advanced precancerous polyps. Of 9304 negative results for FIT, 595 (6.4%) would be viewed as false negatives, consisting of 17 cancers and 578 precancerous polyps. If nonadvanced adenomas were considered to be positive findings, then the proportions of positive tests viewed as false positives would be 732 of 1611 (45.4%) with DNA testing and 248 of 696 (35.6%) with FIT. The numbers of negative tests viewed as false negatives would be 2840 of 8389 (33.9%) with DNA testing and 3271 of 9304 (35.2%) with FIT. Most of these false negative results would be small, nonadvanced adenomas (in 2398 of 2840 participants [84.4%] with DNA testing and 2676 of 3271 participants [81.8%] with FIT), with only rare instances of colorectal cancers (5 of 2840 [0.2%] and 17 of 3271 [0.5%], respectively).

#### DISCUSSION

We compared a multitarget stool DNA test with a commercial FIT among patients at average risk for colorectal cancer. The sensitivity of the DNA

Table 2. Numbers of Persons Who Would Need to Be Screened with Colonoscopy, Multitarget DNA Test, and FIT to Detect One Colorectal Cancer and One Advanced Precancerous Lesion.

Finding	Number Needed to Screen (95% CI)				
	Colonoscopy	Multitarget DNA Test	FIT		
Any colorectal cancer	154 (120–200)	166 (130–217)	208 (156–286)		
Stage I to III colorectal cancer	166 (130–217)	178 (140–238)	227 (169–313)		
Advanced precancerous lesion	13 (12–14)	31 (28–35)	55 (48–65)		

Table 3. Extrapolation of Findings to an Expanded Population of 10,000 Persons at Average Risk for Colorectal Cancer Undergoing Screening with Colonoscopy, Multitarget Stool DNA Test, and FIT.\*

Colonoscopy Finding	Persons with Finding	Multitarget DNA Test		FIT	
		Positive Results (N=1611)	Negative Results (N=8389)	Positive Results (N = 696)	Negative Results (N=9304)
	no.	no. (%)			
Colorectal cancer	65	60 (3.7)	5 (0.06)	48 (6.9)	17 (0.18)
Advanced precancerous lesions	758	321 (19.9)	437 (5.2)	180 (25.9)	578 (6.2)
Nonadvanced adenomas	2896	498 (30.9)	2398 (28.6)	220 (31.6)	2676 (28.8)
Negative results: no colorectal cancer, advanced precancerous lesions, or nonadvanced adenomas	6281	732 (45.4)	5549 (66.1)	248 (35.6)	6033 (64.8)

<sup>\*</sup> Listed are data from the study that have been extrapolated to a theoretical population of 10,000 persons.

test for the detection of both colorectal cancer (92.3%) and advanced precancerous lesions (42.4%) exceeded that of FIT by an absolute difference of nearly 20 percentage points. This difference may be attributed to the DNA marker and algorithm components of the test, since the test performance of the hemoglobin immunoassay component of the DNA test was nearly identical to that of FIT. However, FIT was more specific for the detection of both colorectal cancer and advanced precancerous lesions, by absolute differences of 6.6 to 8.3 percentage points.

Sensitivity is the most important characteristic for screening tests because the primary role of such testing is to rule out diseases such as cancer. The sensitivity of the DNA test for the detection of advanced precancerous lesions was approximately half that for the detection of colorectal cancer; it exceeded the performance of the FIT overall and in important subsets of lesions, including adenomas measuring 2 cm or more (in which the prevalence of high-grade dysplasia is 5 to 44%22-25) and large, sessile serrated polyps (which may account for up to one third of colorectal cancers<sup>26,27</sup>). In our study, DNA testing was associated with a relative increase of 27% in the rate of detection of stage I to III colorectal cancers and a relative increase of 78% in the rate of detection of advanced precancerous lesions, as compared with FIT. A negative result on DNA testing reduced the chance of having colorectal cancer to a greater extent than did a negative result on FIT, from a baseline risk of approximately 1 in 154 (0.7%) to 1 in 1675 (0.06%) after DNA testing and 1 in 556 (0.18%) after FIT.

Although high sensitivity is the most important attribute of cancer-screening tests, specificity is also important, since it affects the number of persons who have positive test results, a majority of whom will have false positive results because of the low prevalence of cancer. The specificity of FIT (94.9 to 96.4%) was superior to that of the DNA test (86.6 to 89.8%), with false positive rates of 3.6 to 5.1% and 10.2 to 13.4%, respectively. Positive results on the DNA test increased the probability of having colorectal cancer from 0.7% to 3.7%, as compared with 6.9% for FIT, and increased the probability of having an advanced precancerous lesion from 7.3% to 19.9%, as compared with 25.9% for FIT.

Two points regarding the specificity of DNA testing deserve comment. First, analysis of the primary measure of specificity (86.6%) included participants with nonadvanced adenomas, which can cause the test to be positive, and those with a negative result on colonoscopy. Among persons with only a negative result on colonoscopy, the specificity of the DNA test was nearly 90%, although that was still inferior to the specificity of FIT, which exceeded 96%. Second, specificity correlated inversely with age. Among participants with any findings other than advanced neoplasia, specificity varied from 91.5% among participants between the ages of 50 and 64 years to 83.7% among those 65 years of age or older. Among persons 50 to 64 years of age with a negative result on colonoscopy, the specificity of the DNA test was 94%, which was similar to that of FIT. Age-related variation in specificity could be due to the presence of lesions that were missed on colonoscopy (which are more prevalent among persons older than 70 years<sup>28</sup>) or to age-related DNA methylation.<sup>29,30</sup>

The performance of FIT in our study was similar to that in previous studies in which colonoscopy was the reference standard.<sup>31,32</sup> For FIT in our study, we used the manufacturer's threshold of 100 ng of hemoglobin per milliliter of buffer.<sup>20</sup> Whether results would differ with the use of another type of FIT is not known; however, the detection rate with the FIT we used was higher than the rate with at least one other FIT.<sup>33</sup>

A discussion of the role of multitarget stool DNA testing in colorectal-cancer screening is beyond the scope of this report because it requires the assessment of several factors aside from sensitivity and specificity, which are the focus of this report. Other factors include performance characteristics of alternative tests, testing intervals, complications, costs, patient acceptance, and adherence.<sup>34,35</sup> Downstream effects of these factors on outcomes, including both cause-specific and

overall morbidity and mortality, require modeling studies to compare various screening tests and strategies. Although our study provides some of the important values for modeling, it cannot determine which test or strategy is better or preferred.

Screening rates for colorectal cancer remain low despite strong evidence of the effectiveness of several tests and strategies. The U.S. Preventive Services Task Force states that there is no preferable screening test, as supported by several cost-effectiveness analyses.<sup>5-7</sup> Offering a choice among tests may improve uptake of screening.<sup>32,36</sup> A noninvasive test with a high single-application sensitivity for curable-stage cancer may provide an option for persons who prefer noninvasive testing. Questions about testing intervals and tailoring require further consideration.

In conclusion, a stool test combining altered human DNA and fecal hemoglobin showed higher single-application sensitivity than a commercial FIT for both colorectal cancer and advanced precancerous lesions, although with lower specificity.

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Disclosure forms provided by the authors are available with the full text of this article at NEJM.org.

#### REFERENCES

- 1. Siegel R, Naishadham D, Jemal A. Cancer statistics, 2013. CA Cancer J Clin 2013:63:11-30.
- 2. Vogelstein B, Papadopoulos N, Velculescu VE, Zhou S, Diaz LA Jr, Kinzler KW. Cancer genome landscapes. Science 2013:339:1546-58.
- 3. Levin B, Lieberman DA, McFarland B, et al. Screening and surveillance for the early detection of colorectal cancer and adenomatous polyps, 2008: a joint guideline from the American Cancer Society, the US Multi-Society Task Force on Colorectal Cancer, and the American College of Radiology. Gastroenterology 2008; 134:1570-95.
- 4. Rex DK, Johnson DA, Anderson JC, Schoenfeld PS, Burke CA, Inadomi JM. American College of Gastroenterology guidelines for colorectal cancer screening 2009. Am J Gastroenterol 2009;104:739-50. [Erratum, Am J Gastroenterol 2009; 104:1613.]
- 5. Parekh M, Fendrick AM, Ladabaum U. As tests evolve and costs of cancer care rise: reappraising stool-based screening for colorectal neoplasia. Aliment Pharmacol Ther 2008;27:697-712.
- **6.** Heitman SJ, Hilsden RJ, Au F, Dowden S, Manns BJ. Colorectal cancer screening for average-risk North Americans: an economic evaluation. PLoS Med 2010;7(11): e1000370.

- 7. Sharaf RN, Ladabaum U. Comparative effectiveness and cost-effectiveness of screening colonoscopy vs. sigmoidoscopy and alternative strategies. Am J Gastroenterol 2013;108:120-32.
- **8.** Vital signs: colorectal cancer screening test use United States, 2012. MMWR Morb Mortal Wkly Rep 2013;62:881-8.
- **9.** Berger BM, Ahlquist DA. Stool DNA screening for colorectal neoplasia: biological and technical basis for high detection rates. Pathology 2012;44:80-8.
- **10.** Ahlquist DA, Sargent DJ, Loprinzi CL, et al. Stool DNA and occult blood testing for screen detection of colorectal neoplasia. Ann Intern Med 2008;149:441-50.
- 11. Imperiale TF, Ransohoff DF, Itzkowitz SH, Turnbull BA, Ross ME. Fecal DNA versus fecal occult blood for colorectal-cancer screening in an average-risk population. N Engl J Med 2004;351:2704-14.
- 12. Boynton KA, Summerhayes IC, Ahlquist DA, Shuber AP. DNA integrity as a potential marker for stool-based detection of colorectal cancer. Clin Chem 2003; 49:1058-65.
- **13.** Zou H, Harrington JJ, Klatt KK, Ahlquist DA. A sensitive method to quantify human long DNA in stool: relevance to colorectal cancer screening. Cancer Epidemiol Biomarkers Prev 2006;15:1115-9.
- **14.** Ahlquist DA, Zou H, Domanico M, et al. Next-generation stool DNA test accu-

- rately detects colorectal cancer and large adenomas. Gastroenterology 2012;142: 248-56.
- **15.** Itzkowitz S, Brand R, Jandorf L, et al. A simplified, noninvasive stool DNA test for colorectal cancer detection. Am J Gastroenterol 2008;103:2862-70.
- **16.** Lidgard GP, Domanico MJ, Bruinsma JJ, et al. Clinical performance of an automated stool DNA assay for detection of colorectal neoplasia. Clin Gastroenterol Hepatol 2013:11:1313-8.
- 17. Li M, Chen WD, Papadopoulos N, et al. Sensitive digital quantification of DNA methylation in clinical samples. Nat Biotechnol 2009;27:858-63.
- **18.** Aronchick CA, Lipshutz WH, Wright SH, Dufrayne F, Bergman G. A novel tableted purgative for colonoscopic preparation: efficacy and safety comparisons with Colyte and Fleet Phospho-Soda. Gastrointest Endosc 2000:52:346-52.
- **19.** American Joint Committee on Cancer. AJCC cancer staging manual. 7th ed. Chicago: American Joint Committee on Cancer, 2009.
- **20.** OC-Auto Micro 80 FOB Test (product insert). Cortland Manor, NY: Polymedco (http://www.fobt-tests.com).
- 21. Hanley JA, McNeil BJ. A method of comparing the areas under receiver operating characteristic curves derived from the same cases. Radiology 1983;148:839-43.

- **22.** Bardan E, Bat L, Melzer E, Shemesh E, Bar-Meir S. Colonoscopic resection of large colonic polyps a prospective study. Isr J Med Sci 1997;33:777-80.
- **23.** Christie JP. Colonoscopic excision of large sessile polyps. Am J Gastroenterol 1977;67:430-8.
- **24.** Lieberman D, Moravec M, Holub J, Michaels L, Eisen G. Polyp size and advanced histology in patients undergoing colonoscopy screening: implications for CT colonography. Gastroenterology 2008; 135:1100-5.
- **25.** Nivatvongs S, Snover DC, Fang DT. Piecemeal snare excision of large sessile colon and rectal polyps: is it adequate? Gastrointest Endosc 1984;30:18-20.
- **26.** Kahi CJ, Li X, Eckert GJ, Rex DK. High colonoscopic prevalence of proximal colon serrated polyps in average-risk men and women. Gastrointest Endosc 2012; 75:515-20.
- **27.** Rex DK, Ahnen DJ, Baron JA, et al. Serrated lesions of the colorectum: review and recommendations from an expert panel. Am J Gastroenterol 2012;107:1315-29.
- 28. Corley DA, Jensen CD, Marks AR, et

- al. Variation of adenoma prevalence by age, sex, race, and colon location in a large population: implications for screening and quality programs. Clin Gastroenterol Hepatol 2013;11:172-80.
- **29.** Ahlquist DA. Molecular detection of colorectal neoplasia. Gastroenterology 2010;138:2127-39.
- **30.** Ahlquist DA, Taylor WR, Yab TC, et al. Aberrantly methylated gene marker levels in stool: effects of demographic, exposure, body mass, and other patient characteristics. J Mol Biomark Diagn 2012; 3(5):e1000133.
- **31.** Morikawa T, Kato J, Yamaji Y, Wada R, Mitsushima T, Shiratori Y. A comparison of the immunochemical fecal occult blood test and total colonoscopy in the asymptomatic population. Gastroenterology 2005;129:422-8.
- **32.** Quintero E, Castells A, Bujanda L, et al. Colonoscopy versus fecal immunochemical testing in colorectal-cancer screening. N Engl J Med 2012;366:697-706.
- **33.** Raginel T, Puvinel J, Ferrand O, et al. A population-based comparison of immunochemical fecal occult blood tests for

- colorectal cancer screening. Gastroenterology 2013;144:918-25.
- **34.** Screening for colorectal cancer: U.S. Preventive Services Task Force recommendation statement. Ann Intern Med 2008;149:627-37.
- **35.** Zauber AG, Lansdorp-Vogelaar I, Knudsen AB, Wilschut J, van Ballegooijen M, Kuntz KM. Evaluating test strategies for colorectal cancer screening age to begin, age to stop, and timing of screening intervals: a decision analysis of colorectal cancer screening for the U.S. Preventive Services Task Force from the Cancer Intervention and Surveillance Modeling Network (CISNET). Rockville, MD: Agency for Healthcare Research and Quality, March 2009. (AHRQ publication no. 08-05124-EF-2.)
- **36.** Inadomi JM, Vijan S, Janz NK, et al. Adherence to colorectal cancer screening: a randomized clinical trial of competing strategies. Arch Intern Med 2012;172: 575.82

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