

Puckered, a *Drosophila* MAPK phosphatase, ensures cell viability by antagonizing JNK-induced apoptosis

Donald G. McEwen^{1,*†} and Mark Peifer^{1,2}

¹Lineberger Comprehensive Cancer Center, The University of North Carolina at Chapel Hill, Chapel Hill, NC 27599-3280, USA

²Department of Biology, The University of North Carolina at Chapel Hill, Chapel Hill, NC 27599-3280, USA

*Present address: Children's Cancer Research Institute, University of Texas Health Science Center at San Antonio, San Antonio, TX 78229, USA

†Author for correspondence (e-mail: mcewen@uthscsa.edu)

Accepted 21 June 2005

Development 132, 3935-3946

Published by The Company of Biologists 2005

doi:10.1242/dev.01949

Summary

MAPK phosphatases (MKPs) are important negative regulators of MAPKs *in vivo*, but ascertaining the role of specific MKPs is hindered by functional redundancy in vertebrates. Thus, we characterized MKP function by examining the function of Puckered (Puc), the sole *Drosophila* Jun N-terminal kinase (JNK)-specific MKP, during embryonic and imaginal disc development. We demonstrate that Puc is a key anti-apoptotic factor that prevents apoptosis in epithelial cells by restraining basal JNK signaling. Furthermore, we demonstrate that JNK signaling plays an important role in γ -irradiation-induced apoptosis, and examine how JNK signaling fits into the circuitry regulating this process. Radiation upregulates

both JNK activity and *puc* expression in a p53-dependent manner, and apoptosis induced by loss of Puc can be suppressed by p53 inactivation. JNK signaling acts upstream of both Reaper and effector caspases. Finally, we demonstrate that JNK signaling directs normal developmentally regulated apoptotic events. However, if cell death is prevented, JNK activation can trigger tissue overgrowth. Thus, MKPs are key regulators of the delicate balance between proliferation, differentiation and apoptosis during development.

Key words: Apoptosis, JNK, Phosphatase, *Drosophila*

Introduction

The balance between cell proliferation, differentiation and death is strictly regulated during development. Genetic or epigenetic disruption of this balance has severe consequences, including both degenerative and hyper-proliferative diseases. To understand the mechanistic basis by which cells balance proliferation, differentiation and apoptosis and correct imbalances triggered by outside stress, we used *Drosophila* as a model system to examine the role of mitogen-activated protein kinase (MAPK) signaling in this process.

Three evolutionarily conserved MAPK pathways have been described. These cascades relay information about the environment to the nucleus via the sequential phosphorylation of a series of kinases, with terminal activation of the MAPKs ERK, Jun N-terminal kinase (JNK) or p38 eliciting changes in gene transcription (reviewed by Schaeffer and Weber, 1999).

MAPKs are key modulators of cell proliferation, differentiation and apoptosis during oncogenesis. The Ras-Raf-ERK pathway has received the most attention for its role in promoting transformation. By contrast, the JNK pathway has both oncogenic and tumor-suppressing roles depending on the cellular context. For example, transformation of leukemia cell lines by Bcr-Abl is dependent on JNK signaling (Dickens et al., 1997; Raitano et al., 1995). By contrast, chemotherapeutic agents promote JNK-dependent apoptosis (Osborn and Chambers, 1996; Seimiya et al., 1997; Stadheim and Kucera, 2002). Thus, JNK signaling can both support and antagonize

transformation; however, the molecular mechanisms underlying this choice are not well defined.

JNK signaling is initiated by dual phosphorylation of threonine and tyrosine residues in its activation loop, while removal of either phosphate inactivates MAPKs (reviewed by Davis, 2000). Thus, steady-state levels of MAPK activity depend upon the equilibrium between MEK kinases and protein phosphatases. MAPK phosphatases (MKPs) are one class of proteins tempering MAPK activity (reviewed by Keyse, 1998). At least nine vertebrate MKPs, all very similar in their phosphatase domains, have been identified. One class, typified by MKP-1/CL100, are immediate-early genes: their transcription is activated by JNK signaling and they limit the peak level and duration of MAPK activity by completing a negative-feedback loop. As differences in peak amplitude and duration of MAPK signaling can elicit distinct cellular responses, MKP activity may play an important role in regulating biological responses to stimuli.

Functional redundancy among the three JNK isoforms complicates analysis of JNK signaling in mammals (reviewed by Davis, 2000). By contrast, a single JNK [*basket* (*bsk*)] exists in *Drosophila* and plays an important role in embryonic development (Riesgo-Escovar et al., 1996; Sluss et al., 1996). Likewise, little is known about the *in vivo* function of vertebrate MKPs, as functional redundancy among MKPs (Dorfman et al., 1996) and overlapping substrate specificity of distinct MKPs (Franklin and Kraft, 1997; Sun et al., 1993) hinder the assessment of phenotypes *in vivo*. Again, a single

MKP, Puckered (Puc), antagonizes JNK signaling in *Drosophila* (Martin-Blanco et al., 1998; Rintalen et al., 2003). Thus, *Drosophila* has emerged as an excellent model system for characterizing both the function and regulation of JNK signaling in vivo.

Drosophila JNK signaling coordinates dorsal closure, a morphogenetic event that unites the two lateral epidermal sheets. Mutations in any component of the JNK pathway, like *hemipterous* (the JNK kinase), *bsk*, *kayak* (*kay*; *Drosophila fos*) and *Djun* (*Jra* – FlyBase) disrupt dorsal closure (reviewed by Noselli and Agnes, 1999; Stronach and Perrimon, 1999). This may be an evolutionarily conserved mechanism, as JNK signaling regulates vertebrate cell migration and epithelial wound healing (reviewed by Martin and Parkhurst, 2004). *puc* zygotic mutants also exhibit defects in dorsal closure; increased JNK signaling in *puc* mutants disrupts alignment of the lateral epidermal sheets (Martin-Blanco et al., 1998). Thus, the tempering of JNK signaling by Puc is crucial during dorsal closure.

In addition to regulating morphogenesis, JNK signaling is also activated by other signaling pathways; in this context, JNK activation triggers apoptosis. In *Drosophila*, these include TNF α (Igaki et al., 2002; Moreno et al., 2002b), TGF β (Adachi-Yamada et al., 1999; Adachi-Yamada and O'Connor, 2002; Adachi-Yamada and O'Connor, 2004) and Myc (de la Cova et al., 2004; Moreno and Basler, 2004). However, the function of Puc during the induction of apoptosis is less clear.

Previous analysis of Puc focused on zygotic mutants retaining maternally contributed Puc. Here, we extend this analysis by completely removing both maternal and zygotic Puc from embryos, and by eliminating Puc function in imaginal disc cells. We find that Puc is required continuously to antagonize JNK-dependent apoptosis in a cell-autonomous manner, suggesting that basal JNK signaling is poised just below a threshold required to eliminate epithelial cells. Second, we find that JNK signaling also plays a key role in promoting apoptosis in response to a diverse array of signals, including both the p53-dependent response to DNA damage and developmentally regulated apoptosis. Finally, we demonstrate that when apoptosis is blocked, Puc prevents tissue overgrowth by preventing unbridled JNK activation. Thus, JNK signaling can promote apoptosis or proliferation in different cellular contexts, and Puc plays an important role in controlling this balance.

Materials and methods

Genetics

Wild-type was CantonS or *y w* (for immunofluorescence). Sources of stocks: *FRT^{82B} puc^{A251.1}* (L. Dobbens); *puc^{R10}* (A. Martinez-Arias – it was recombined onto *FRT^{82B}*); and *rpr-11-lacZ* (Abrams et al., 1993). *UAS-p53Gus 3.1* (Brodsky et al., 2000), *p53^{5A-1-4}* (a null allele) and other stocks were from the Bloomington Stock Center (flybase.bio.indiana.edu). Myc-Puc carries six tandem copies of the human MYC epitope in frame with the N terminus of Puc, and is cloned in p[UASp]. *UAS-myc-Puc^{Dead}* is identical except for a ¹⁸⁵D>A site-directed mutation, targeting the catalytic D conserved in all tyrosine phosphatases. It has previously been shown to inactivate PTP1B (Flint et al., 1997). Overexpression of *myc-Puc^{Dead}* has no effect on viability. Transgenics were generated by P-element transformation. Maternal-zygotic *puc* mutants were generated as described by Xu and Rubin (Xu and Rubin, 1993). *hsflp^{1/w}; FRT^{82B} puc/FRT^{82B} ovoD¹⁻¹⁸* larvae were heat-shocked for 3 hours at 37°C,

~72 hours after egg laying, and females mated to wild-type or *puc/+* males. MARCM (Lee and Luo, 1999) was used to generate GFP-marked clones. *hsflp¹; FRT^{82B} puc^{A251.1}/TM6b* males were crossed to *Act^{5C}>GAL4²⁵ UAS-GFP^{T2}; FRT^{82B} tubP-GAL80^{LL3}* females, larvae heat-shocked 1 hour at 37°C, 3–4 days after egg laying, and wing discs dissected from non-Tb wandering 3rd-instar female larvae. Mutant eye discs were derived as described by Stowers and Schwarz (Stowers and Schwarz, 1999). *FRT^{82B} w^{+90E}; FRT^{82B} puc^{A251.1}/TM3*, *FRT^{82B} puc^{R10}/TM3*, *bsk^{2/+}; FRT^{82B} puc^{A251.1}/TM3*, or *FRT^{82B} puc^{A251.1} p53^{5A-1-4}/TM3* males were mated to *y w; eyeless-GAL4^{DH1} UAS-FLP^{1(JD1)}; FRT^{82B} GMR-hid^{SS4}, l(3)/TM2* females.

Irradiation

Wandering 3rd-instar larvae were given 4000 rads of γ -radiation from a ¹³⁷Cs source, and allowed to recover for 4 hours at 25°C before dissection.

Immunofluorescence

Dechorionated embryos were fixed for 20 minutes in 1:1 3.7% formaldehyde/PBS:heptane and methanol-devitellinized. Discs were fixed with 3.7% formaldehyde/PBS+0.5% TritonX-100 (PBT5) for 20 minutes on ice. Tissues were blocked for 3 hours at room temperature in PBS+0.1% TritonX-100 (PBT) (embryos) or PBT5 (discs) + 10% BSA, incubated with primary antibody overnight at 4°C, washed in PBT or PBT5, incubated for 1 hour at room temperature with fluorescent secondary antibodies (1:1000; Molecular Probes) and washed.

Antibodies

Rat- α -DE-cad2 (1:250; DSHB), mouse- α -phosphotyrosine (4G10 1:1000; Upstate), α -Engrailed (4D9 1:50; DSHB), α - β -galactosidase (Promega; 1:1000) and α -Myc (9E10; 1:5) were used. Alexa⁶⁴⁷- or Alexa⁵⁶⁸-phalloidin (Molecular Probes) and Hoechst (Sigma) were used at 1:1000.

TUNEL labeling

Samples were post-fixed for 15 minutes in 3.7% formaldehyde in PBT5, permeabilized in 100 mM sodium citrate/0.1% TritonX-100 at 65°C for 30 minutes, rinsed in PBT5, and free DNA ends fluorescein-labeled using the In Situ Cell Death Detection Kit (Roche). Samples were mounted in Aqua Poly/mount (Polysciences).

Image collection and processing

Images of embryos, wings and legs were captured on a Nikon OPTIPHOT-2 with Kodak Technical Pan film and digitized with a Polaroid SprintScan35. Eyes, thorax and genitalia were photographed with a stereomicroscope-mounted Nikon Coolpix4500 digital camera after immersion in 70% ethanol. z-series were collected with a Zeiss 510 confocal microscope, and brightest-point projections generated with ZeissLSM software. Brightness, contrast and levels were adjusted in Photoshop 7.0.

Results

Complete loss of Puc activity promotes cell-autonomous apoptosis

puc zygotic mutants die with defects in dorsal closure and inappropriate cell fate choices in the ventral epidermis, which reflect inappropriate activation of JNK signaling (Martin-Blanco et al., 1998) and may also reflect effects on other signaling cascades, such as the Wnt pathway (McEwen et al., 2000).

Maternally contributed mRNA and protein often mask requirements for genes during early *Drosophila* development; thus, we used the *hs-flp*/DFS technique (Chou and Perrimon, 1992) to remove both maternal and zygotic Puc function.

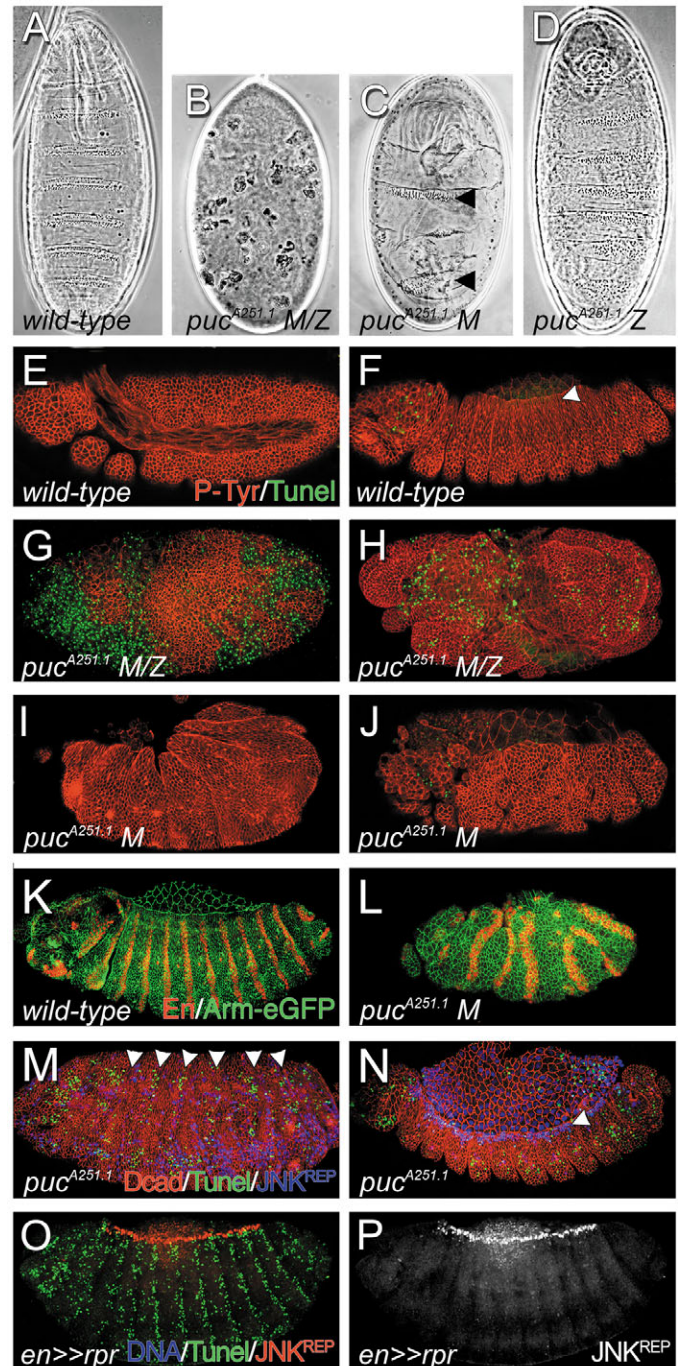
Fig. 1. Loss of Puc function triggers apoptosis in embryonic epithelia. (A-D) Cuticles, ventral view, anterior upwards. (A) Wild-type; (B) *puc*^{A251.1} M/Z; (C) *puc*^{A251.1} maternal-only (arrowheads indicate denticle belts); (D) *puc*^{A251.1} zygotic mutant. *puc*^{A251.1} M/Z mutants secrete fragmented cuticle; this is suppressed by paternal Puc. (E-N) Embryos, anterior leftwards. Dorsal is upwards unless indicated otherwise. (E-J) Lateral views, TUNEL-label (green). Anti-phosphotyrosine (red) outlines cells. (E,F) Wild type stage 10 (E) and stage 13 (F). Arrowhead in F indicates leading edge cells. (G,H) Stage 10 (G) and stage 13 (H) presumptive *puc*^{A251.1} M/Z mutants undergoing massive apoptosis. (I,J) Stage 10 (I) and stage 13 (J) presumptive *puc*^{A251.1} maternal-only mutants, with fewer TUNEL-positive cells. (K,L) Wild type (K) and *puc*^{A251.1} maternal mutants (L). Engrailed is in red. Arm-eGFP (green) indicates cell boundaries. (M,N) TUNEL-labeled (green) *puc*^{A251.1} zygotic mutants. Mutants are identified by ectopic JNK^{REP} activity (blue). Cell boundaries are indicated by DE-cad (red). (M) Ventrolateral view, stage 14. Arrowheads indicate TUNEL-positive cells. (N) Lateral view, stage 13. Arrowhead-leading edge cells with elevated JNK^{REP} activity. (O) *UAS-rpr;en-Gal4;puc*^{A251.1/+}, TUNEL-labeled (green). (P) JNK^{REP} activity from O (red). *puc* heterozygotes identified by JNK^{REP} activity at leading edge. Apoptosis is induced in segmental stripes, without induction of JNK^{REP}.

Mothers with germlines homozygous for the strongest allele, *puc*^{R10}, failed to produce eggs, but females with germlines homozygous for *puc*^{A251.1}, a strong allele, lay reduced numbers of small eggs, allowing us to assess the effect of complete loss of Puc function.

puc^{A251.1} maternal/zygotic (M/Z) mutant embryos have severe defects in embryonic development, secreting only fragmented scraps of cuticle (compare Fig. 1B with 1A; henceforth, unless noted, *puc* refers to *puc*^{A251.1}). The same phenotype was noted using different paternally contributed *puc* alleles, suggesting it is due to *puc* and not another second-site mutation (see Table S1 in the supplementary material). Similar ‘scraps’-like phenotypes are observed when epithelial cells initiate apoptosis (e.g. *crb* mutants) (Tepass et al., 1990). Thus, we used TUNEL labeling to assess whether the *puc*^{M/Z} phenotype resulted from widespread apoptosis. Late-stage *puc*^{M/Z} mutant embryos have disorganized epithelia with highly elevated numbers of TUNEL-positive cells (Fig. 1G,H) relative to similarly staged wild-type controls (Fig. 1E,F). A similar increase in apoptosis was observed in living *puc*^{M/Z} mutants stained with Acridine Orange (data not shown). Thus, Puc normally prevents the induction of apoptosis throughout the embryonic epidermis.

The cuticle defects and widespread apoptosis of *puc*^{M/Z} mutants are suppressed if the embryo receives paternal wild-type Puc (Fig. 1C,I,J), but this fails to rescue embryo viability. *puc* maternal-only mutants have patterning defects distinct from those of zygotic mutants, exhibiting segmental deletions and fusions, as evidenced by loss of Engrailed stripes (compare Fig. 1L with 1K) and the corresponding loss of denticle belts (compare Fig. 1C with 1D). Thus, maternal Puc plays a distinct role in establishing segmentation, but we have not pursued this role further.

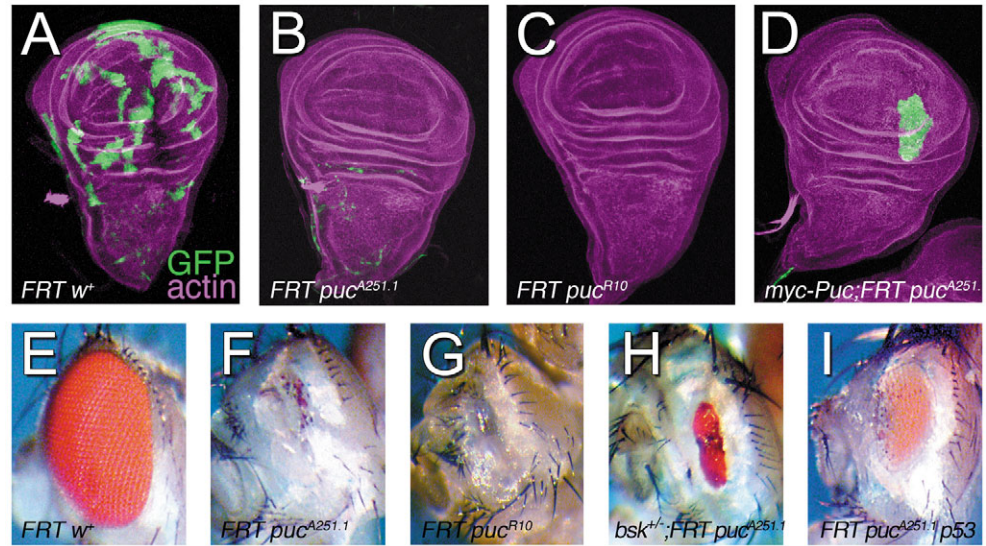
Given the clear role for Puc in preventing apoptosis in *puc*^{M/Z} mutants, we re-examined the ventral segment-polarity phenotype of *puc* zygotic mutants (McEwen et al., 2000) to see if this might also be, in part, the result of apoptosis. Increased numbers of apoptotic cells were observed in a segmentally



repeated pattern in both the ventral and lateral epidermis (compare Fig. 1M with 1F); JNK signaling is upregulated in a similar segmental pattern (Fig. 1M). Thus, the *puc* zygotic mutant phenotype also reflects an increase in JNK-dependent apoptosis.

To further characterize the role of Puc during epithelial development, we examined its function in the imaginal discs, precursors of the adult epidermis. Clones of *puc* mutant tissue were generated in the wing disc, with mutant cells positively marked by GFP (Fig. 2A-D). Although clones were readily derived from wild-type controls (Fig. 2A), neither *puc*^{A251.1} (Fig. 2B) nor *puc*^{R10} (Fig. 2C) mutant clones were recovered

Fig. 2. JNK signaling promotes the loss of *puc* mutant imaginal tissues. (A–D) Third-instar wing discs. GFP-expressing (green) cells are clonal patches of cells homozygous for: (A) wild-type control, (B) *puc*^{A251.1}, (C) *puc*^{R10}, (D) *UAS-myc-Puc*; *puc*^{A251.1}. Actin (purple). *puc* mutant clones are not recovered unless *Puc*⁺ function is transgenically restored (D). The GFP-signal in B is from clones outside the disc proper in the parapodial membrane. (E–I) Adult eyes. All eye cells homozygous for: (E) wild-type control, (F) *puc*^{A251.1}, (G) *puc*^{R10}, (H) *bsk*^{2/+}; *puc*^{A251.1} and (I) *puc*^{A251.1} *p53*^{5A-1-4J}. Eye tissue reappears in *bsk* heterozygotes or *p53* homozygotes.



anywhere throughout the wing disc. We did not observe *puc* mutant clones even when we looked as soon as 8 hours after clone induction, when wild-type clones were only a few cells in size. We cannot rule the possibility that loss of *Puc* may reduce proliferation, but given the clear induction of apoptosis in the embryonic epidermis following *Puc* loss, we think apoptosis is the most likely explanation for clone loss in the wing.

To confirm that loss of epithelial cells resulted from loss of *Puc*, a Myc-epitope tagged *puc* transgene (*myc-puc*) was used to rescue *puc* clones. Addition of *Puc*⁺ activity back to *puc* clones at the time of clone induction, using this transgene, rescued cell viability (e.g. Fig. 2D). Rescued *puc* mutant clones were recovered throughout the wing disc, though at lower frequencies than wild-type controls; we suspect that the delay in *myc-puc* transgene expression after clone induction (Gal4-mediated Myc-*Puc* accumulation requires degradation of Gal80) means that many *puc* cells are committed to die before rescue can occur.

Next, we assessed whether *Puc* plays a similar role in eye discs. Removal of *Puc* function from the entire eye imaginal disc results in nearly complete loss of eye tissue (*puc*^{A251.1}, Fig. 2F; *puc*^{R10}, Fig. 2G). To assess whether loss of *puc* mutant eye tissue resulted from inappropriate JNK activation, *puc* mutant eyes were generated in flies with reduced levels of JNK. Heterozygosity for *bsk*² partially rescued *puc* mutant eye tissue (Fig. 2H).

Together, these data demonstrate a crucial role for *Puc* in preventing apoptosis in embryonic and larval epithelial cells. Apoptosis occurred in the absence of any outside stress (other than the loss of *Puc*) that might stimulate JNK activity, suggesting that *Puc* prevents apoptosis by buffering basal levels of JNK signaling.

JNK and MKP activities regulate radiation-induced apoptosis in a p53-dependent manner

Next, we explored whether *Puc* plays a broader role in regulating JNK-induced apoptosis by examining γ -radiation-induced cell death. To assess whether JNK activity is upregulated in response to γ -irradiation, β -galactosidase (β -gal) expression from an enhancer trap in *puc* (Martin-Blanco

et al., 1998) was monitored. *puc* enhancer traps have been widely used to monitor JNK activity in vivo (e.g. Adachi-Yamada et al., 1999; Igaki et al., 2002; Tatenos et al., 2000). For simplicity we subsequently refer to this readout as JNK reporter (JNK^{REP}) activity. In the wing disc, JNK^{REP} activity is normally limited to the peripodial membrane overlying the notal region of the wing disc (Fig. 3B). However, γ -irradiation strongly induced both JNK^{REP} activity (Fig. 3C) and apoptosis (Ollmann et al., 2000) throughout the wing disc within 4 hours.

To confirm that JNK^{REP} activation reflects JNK activation in situ, we used Myc-tagged *Puc* (Myc-*Puc*) to antagonize JNK activity. Expression of Myc-*Puc* in the posterior compartment of the wing disc (Fig. 3A) prevented radiation-induced JNK^{REP} activation (Fig. 3D). Interestingly, it also substantially reduced radiation-induced apoptosis (Fig. 3G,G'). By contrast, expression of a catalytically-inactive form of *Puc* (Myc-*Puc*^{DEAD}) failed to block either JNK^{REP} activity (Fig. 3E) or apoptosis (Fig. 3H,H'). Thus, in *Drosophila*, radiation-induced apoptosis depends, at least in part, on JNK signaling.

Discontinuities in morphogen gradients induce JNK-dependent apoptosis, perhaps by promoting discontinuities in a JNK activation gradient (Adachi-Yamada et al., 1999; Adachi-Yamada and O'Connor, 2002). However, lowering JNK activity by expressing Myc-*Puc* in the posterior compartment did not trigger apoptosis along the anteroposterior (AP) compartment border (Fig. 3F). Furthermore, expression of Myc-*Puc* in a stripe anterior to the A-P compartment boundary using *patched-Gal4* or in clonal patches throughout the wing disc by *flp*-out mediated gene induction did not induce autonomous or non-autonomous apoptosis, but it blocked radiation-induced apoptosis (data not shown).

p53 is a key regulator of cell cycle progression and apoptosis in mammals, and is mutated in more than half of all tumors. Like its ortholog, *Drosophila p53* is required for radiation-induced apoptosis and can induce apoptosis when overexpressed (Brodsky et al., 2000; Brodsky et al., 2004; Ollmann et al., 2000; Jassim et al., 2003; Lee et al., 2003;

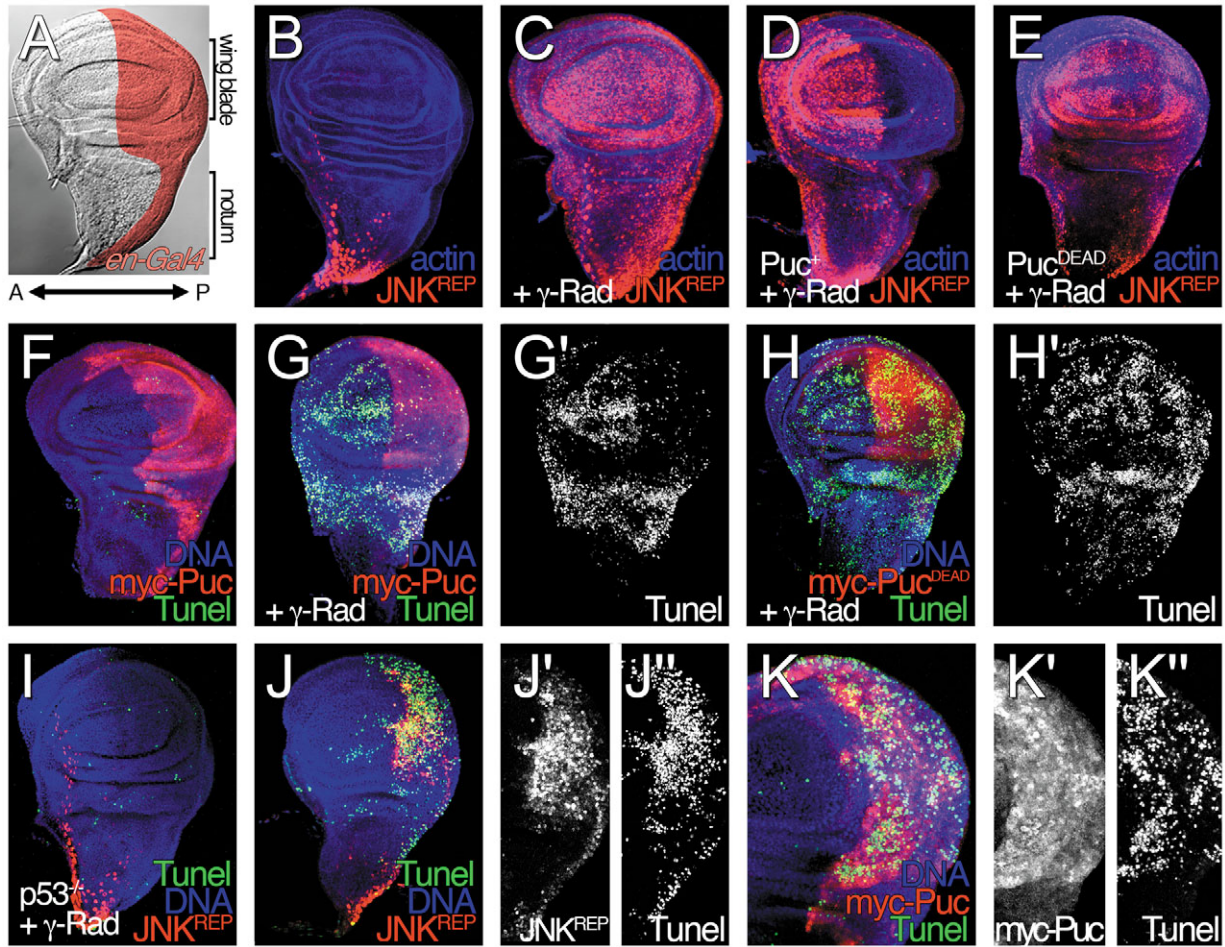


Fig. 3. p53-dependent activation of the JNK signaling pathway is required for γ -radiation-induced apoptosis. Third-instar wing discs. (A) DIC. *en-Gal4* (schematically overlaid in red) is expressed in the posterior compartment. (B-E) Wing discs oriented as in A. JNK^{REP} (red), actin (blue). (B) Untreated *puc*^{A251.1/+}. (C-E) Four hours after γ -radiation (4000 rads here and below). (C) *puc*^{A251.1/+}, (D) *en-Gal4/UAS-myc-Puc*; *puc*^{A251.1/+} and (E) *en-Gal4/UAS-myc-Puc*^{DEAD}; *puc*^{A251.1/+}. (F-H) TUNEL (green), Myc-Puc (red) and DNA (blue). Untreated (F) or γ -irradiated (G,H) discs expressing Myc-Puc (G) or Myc-Puc^{DEAD} (H) in the posterior compartment. (G',H') TUNEL alone. (I,J) TUNEL (green), JNK^{REP} (red) and DNA (blue). (I) γ -irradiated *puc*^{A251.1} *p53*^{[5A-1-4]/+} *p53*^{[5A-1-4]/+} disc. (J) Untreated *en-Gal4/UAS-p53*^{GUS2.1}; *puc*^{A251.1/+} disc. (J',J'') Posterior compartment of J: JNK^{REP} (J') and TUNEL (J''). (K) *en-Gal4 UAS-myc-Puc/UAS-p53*^{GUS2.1} mutant. TUNEL (green, K'), Myc-Puc (red, K') and DNA (blue, K'').

Sogame et al., 2003). However, unlike mammalian p53, overexpression of fly p53 does not induce G₁ arrest (Ollmann et al., 2000) and it is not required for G₂/M arrest in response to radiation (Brodsky et al., 2000).

To assess how JNK signaling and p53 function are integrated in the hierarchy of signaling pathways regulating radiation-induced apoptosis, we asked if p53 is required for JNK^{REP} induction. Radiation-induced JNK^{REP} expression in wing discs requires p53 (compare Fig. 3I with 3C), whereas developmentally regulated JNK^{REP} expression in the peripodial membrane does not. Furthermore, p53 mis-expression in the posterior compartment of the wing disc is sufficient to induce both JNK activation, as evidenced by induction of the JNK^{REP} (Fig. 3J,J'), and apoptosis (Fig. 3J,J''). Thus, p53 is both necessary for radiation-induced JNK^{REP} activity and sufficient to promote JNK activation and apoptosis. However, co-expression of Myc-Puc failed to block apoptosis induced by sustained expression of p53 (Fig. 3K,K',K'').

Although p53 is not required for cell viability, it is thought to monitor genomic integrity during normal development. We thus asked whether the p53 and JNK pathways interact in the developing eye disc in the absence of stress. Although inactivation of *puc* in the eye disc results in ablation of the adult eye (Fig. 2F,G), concomitant loss of p53 suppressed cell death to some extent (Fig. 2I). As loss of p53 alone has no effect on normal eye development, this may suggest that p53 plays a more general role in cell viability by regulating basal JNK activation.

Reaper functions downstream of JNK signaling

Reaper (Rpr), Head Involution Defective (Hid) and Grim (collectively referred to as RHG proteins) are key regulators of apoptosis that promote caspase activation via IAP degradation. Transcriptional upregulation of RHG genes precedes developmental and stress-induced apoptosis, whereas mis-expression of any these genes induces apoptosis. In developing

embryos, *rpr-11-lacZ* has been used to assess *rpr* induction in response to both developmental defects as well as radiation (Nordstrom et al., 1996).

To test whether JNK signaling can regulate *rpr* reporter expression in wing discs, we examined the effects of JNK activation on *rpr-11-lacZ* expression. In un-irradiated *rpr-11-lacZ* wing discs, basal levels of reporter are seen, with β -gal expression highest at the wing margin and along the anterior side of the AP compartment boundary (Fig. 4A). γ -Radiation triggers a substantial increase in β -gal expression throughout the wing pouch (Fig. 4B), consistent with previous work. To determine whether this induction by radiation is JNK dependent, we overexpressed Myc-Puc throughout the posterior compartment of *rpr-11-lacZ* larvae. Inactivation of JNK signaling by Myc-Puc overexpression in the posterior compartment blocked induction of the reporter by irradiation (Fig. 4D; it also blocked β -gal expression along the wing margin in un-irradiated discs; Fig. 4C). Together, these results suggest that JNK signaling may promote apoptosis by transcriptionally upregulating *rpr* expression; this is subject to the caveat that the *rpr-11-lacZ* reporter does not perfectly reflect endogenous *rpr* expression (e.g. Nordstrom et al., 1996). However, JNK-induced upregulation of *hid* was observed in eye imaginal discs (Moreno et al., 2002b).

Previous work suggested that caspase activation might regulate JNK signaling in *Drosophila* (Kuranaga et al., 2002; Ryoo et al., 2004). To test directly whether RHG protein-mediated induction of apoptosis triggers JNK activation, we examined JNK^{REP} activity in response to *rpr* and *hid* mis-expression. When we used *en-Gal4* to direct *rpr* expression in embryos, cell death was observed in *en* stripes, without concomitant JNK activation (Fig. 10,P). As *en-GAL4*-driven *rpr* is embryonic lethal, we could not use this to examine wing discs. We thus used heat-shock to express *hid* in wing discs. A one-hour heat-shock of *puc*^{A251.1}/*hs-hid* larvae induced apoptosis throughout the wing disc (data not shown), but did

not elevate JNK^{REP} activity (Fig. 4G,H; a slight increase in JNK^{REP} was sometimes seen in the overlying peripodial membrane, as determined by the superficial position and size of the nuclei in H). This contrasts with the effect of γ -irradiation (Fig. 3C). Heat-shock of control *puc*^{A251.1}/+ wing discs did not induce JNK^{REP} (compare Fig. 3B to Fig. 4E,F) or apoptosis (data not shown). Thus, RHG-induced apoptosis can occur without JNK activation.

Caspase-mediated cleavage of cellular substrates can promote activation of mammalian JNK and p38 (Graves et al., 1998). To examine whether caspase activity is required for JNK activation, we blocked caspase activation in the posterior compartment. *en-Gal4/UAS-p35*; *puc*/+ larvae were irradiated and JNK^{REP} activity assessed 4 hours later when apoptosis is readily detectable in the anterior but not the posterior compartment (Fig. 6N). JNK^{REP} activity was observed throughout the developing disc (Fig. 6M), suggesting that radiation-induced JNK activation is independent of effector caspase activity.

JNK signaling regulates programmed cell death during normal development

In *Drosophila*, apoptosis plays crucial roles in establishing tissue architecture and removing excess cells during normal development. For example, inactivation of *hid* leads to mis-orientation of the external male genitalia (Abbott and Lengyel, 1991), and inactivation of either *dark* (*Drosophila* Apaf) or *Traf1* (*Drosophila* tumor-necrosis factor receptor-associated factor1) leads to extra scutellar bristles (Kanuka et al., 1999; Kuranaga et al., 2002; Rodriguez et al., 1999). To test whether JNK signaling is required for developmentally regulated apoptosis, we blocked JNK signaling in the genital and wing discs.

Inhibition of apoptosis by expressing the baculovirus caspase inhibitor p35 resulted in genital mis-orientation (Fig. 5B; quantified in 5D), phenocopying reduction in Hid function.

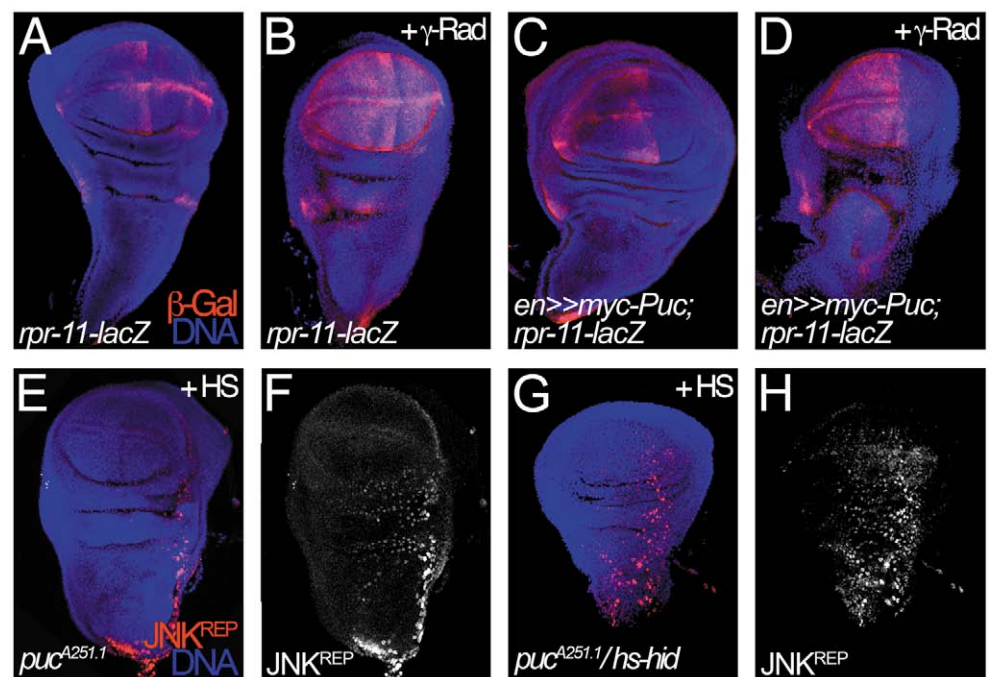


Fig. 4. JNK signaling functions upstream of Rpr and Hid. (A-D) Control (A,C) or irradiated (B,D) wing discs from *rpr-11-lacZ* (A,B) or *en-Gal4 UAS-myc-Puc*; *rpr-11-lacZ* (C,D) larvae. β -Gal (red) and DNA (blue). *rpr*-reporter is blocked by Myc-Puc expression. (E-H) Discs, *puc*^{A251.1}/+ (E,F) or *puc*^{A251.1}/*hs-hid* (G,H) four hours after heat shock. (E,G) JNK^{REP} (red) and DNA (blue). (F,H) JNK^{REP} alone from E,G, respectively. Neither heat shock alone nor heat shock-*hid* induction strongly induces JNK^{REP}.

Mis-expression of Myc-Puc had a similar effect (Fig. 5C, quantified in 5D), while control constructs (GFP or Myc-Puc^{DEAD}) failed to affect genital orientation (Fig. 5D). Extra scutellar bristles were produced when we prevented apoptosis in the scutellar region of the notum by expressing p35 with *ptc-Gal4* (Fig. 5F; mean=5.4 bristles/fly, range=4-10, compared with four bristles in wild-type). *ptc-Gal4*-mediated expression of Myc-Puc also increased bristle number (Fig. 5G; mean=6.71 bristles/fly; range=4-10), whereas neither GFP nor Myc-Puc^{DEAD} altered the number of scutellar bristles (Fig. 5H; means=4.07 and 4.15 bristles/fly, respectively). While our manuscript was in preparation, Macias et al. (Macias et al., 2004) also reported that Puc or p35 mis-expression both block genital rotation. Together, these results suggest that JNK signaling is required for developmentally regulated, caspase-dependent apoptosis.

When cell death is blocked, inappropriate activation of JNK signaling can promote overgrowth

Growth of different compartments of the wing disc is strictly coordinated so that the adult wing has a defined shape and precisely matched surfaces. In normal wings, proliferation and apoptosis are coordinated to correct experimental perturbations, and presumably to correct defects arising during normal development (de la Cova et al., 2004; Neufeld et al., 1998). However, apoptosis does not appear to play an essential role in patterning of the wild-type wing, as over-expression of the caspase inhibitor p35 in the posterior compartment has little or no effect on wing pattern (over 96% of the wings of *en-Gal4/UAS-p35* flies appear normal; Fig. 6B) (see also Pérez-Garijo et al., 2004), although altering apoptosis may affect wing size (de la Cova et al., 2004).

By contrast, when apoptosis is blocked in the posterior compartment of *puc* heterozygotes (*en-Gal4/UAS-p35; puc/+*), we observed tissue overgrowths at random positions

throughout the posterior compartment in >90% of the flies (Fig. 6C-G). Overgrowths arising in the interior of the wing blade resemble wing blisters; however, there appear to be excess cells in only one compartment, as outgrowths project from the dorsal or ventral surfaces, but not both (Fig. 6C,E). When overgrowths arise along the wing margin, excess tissue is readily apparent as an extension of the wing (Fig. 6D,F). Cells within overgrowths respond to developmental cues, adopting appropriate cell fates (Fig. 6E-G). Similar but less frequent overgrowths were observed in posterior regions of adult legs (Fig. 6H). Thus, when cell death is blocked and restraints on JNK activation are relaxed by *puc* heterozygosity, small groups of cells escape regulation by the machinery that normally synchronizes growth of different cell populations in wing discs.

Overgrowths could already be observed in wing discs from *en-Gal4/UAS-p35;puc/+* larvae (Fig. 6I-L). Multiple, randomly-positioned focal disruptions of disc architecture were observed in the posterior compartment where apoptosis was selectively blocked (Fig. 6I,L). These foci contain excess cells, resulting in abnormal folding of the disc epithelium. Furthermore, cells associated with the foci invariably exhibited elevated JNK^{REP} activity (Fig. 6J-L). Most cells within the foci were not apoptotic (Fig. 6J,K); however, occasional TUNEL-positive cells were observed. While JNK^{REP} activity and disc overgrowth were sometimes confined to small foci (Fig. 6I-K), at other times activation of the JNK^{REP} and areas of overgrowth were more extensive (Fig. 6L).

While our manuscript was in preparation, three groups reported that when they induced apoptosis by various stimuli and blocked caspase activity, it also triggered overgrowth (Ryoo et al., 2004; Pérez-Garijo et al., 2004; Huh et al., 2004). Each group found that some of the 'undead' cells expressed the morphogen and disc growth factor Wg. We examined whether this was also the case in our situation. In wing discs

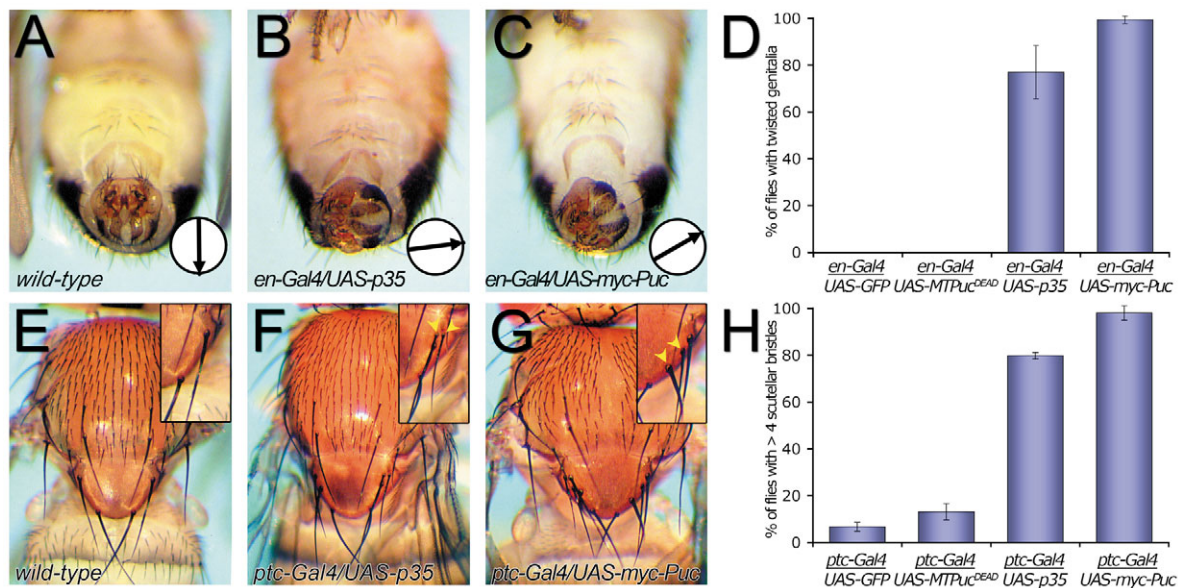


Fig. 5. JNK-dependent cell death is required for normal development. (A-C) Male external genitalia. (A) Wild type, (B) *en-Gal4/UAS-p35* and (C) *en-Gal4/UAS-myc-Puc*. Arrow in lower right-hand corner, penis-to-anus orientation. Mis-expression of either p35 or Myc-Puc disrupts genital orientation. (D) Quantitation. (E-G) Adult thorax. (E) Wild type, (F) *ptc-Gal4/UAS-p35* and (G) *ptc-Gal4/UAS-myc-Puc*. (Insets) Higher magnification of the scutellar bristles; arrowheads indicate extra macrochaetae. (H) Quantitation.

from *en-Gal4/UAS-p35;puc/+* larvae, a subset of cells in JNK^{REP}-positive foci in the wing pouch ectopically expressed Wg (Fig. 6P); however, in control discs Wg is only expressed in a stripe at the dorsal/ventral compartment boundary (Fig. 6O). Ectopic Wg expression, like JNK^{REP} activation, was confined to the posterior compartment where p35 was expressed.

The correlation between foci with activated JNK signaling in discs and outgrowths in adults suggested that JNK activation promotes overgrowth when apoptosis is blocked. To test this, we examined whether reducing the dose of *kayak* (previously *D-fos*) affected outgrowth frequency. Consistent with a role for JNK signaling in promoting overgrowth, reducing *kayak* by 50% resulted in a 25% decrease in outgrowth frequency ($P < 0.001$, Student's *t*-test). Thus, when apoptosis is blocked, JNK signaling promotes tissue overgrowth.

Discussion

We characterized the role of MAPK/MKP interactions in *Drosophila* epithelial tissues by examining the function of Puc, a MKP that selectively antagonizes JNK signaling. Puc plays a key role in regulating cell viability in response to a diverse array of normal and abnormal developmental signals.

Puc restrains apoptosis in the absence of stress

Mammalian JNK signaling promotes apoptosis in response to both extrinsic (e.g. TNF α) and intrinsic (e.g. DNA damage) cues (Davis, 2000). *Drosophila* JNK signaling plays a similar role in apoptosis triggered by extrinsic cues (Adachi-Yamada et al., 1999; Adachi-Yamada and O'Connor, 2002; Burke and Basler, 1996; Igaki et al., 2002; Kuranaga et al., 2002; Martin et al., 2004; Moreno et al., 2002a; Moreno et al., 2002b), but its role in response to intrinsic cues remained to be established. The role of Puc in regulating responses to either sort of signal was also unclear.

We found that removal of Puc triggers apoptosis in epithelia, even in the absence of stress. Thus, basal JNK signaling exists in the absence of stress in embryonic and larval epithelia, and if Puc is not present to restrain this intrinsic JNK activity, it exceeds a threshold and triggers apoptosis. This suggests cells may regulate apoptosis without exogenous JNK stimulation, by regulating MKP levels. Biochemical studies suggest a possible mechanism. Several MKPs are labile proteins, whose half-lives can be shortened or lengthened by post-translational modification (Brondello et al., 1999; Lin et al., 2003). Thus, cell signals may influence apoptosis by modulating MKP accumulation.

Although Puc is crucial to restrain JNK activity and prevent apoptosis in most epithelia, high-level JNK signaling does not

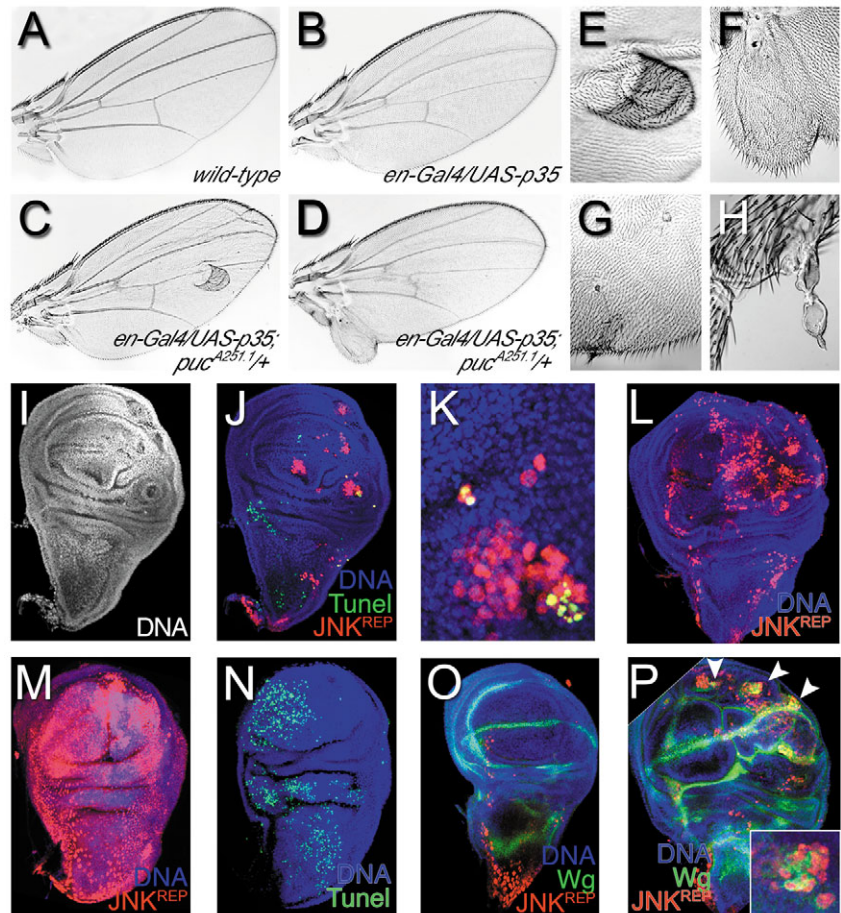


Fig. 6. Elevating JNK signaling while blocking apoptosis promotes ectopic overgrowth of imaginal tissues. Adult wings (A-G). Foreleg (H). (A) Wild type, (B) *en-Gal4/UAS-p35*, (C-H) *en-Gal4/UAS-p35;puc^{A251.1/+}*. (E,F) Higher magnification of the overgrowths from C,D, respectively. Overgrowths occur only in the posterior compartment. Cells adopt appropriate fates. (G) Additional wing defects (disruption of polarity, extra wing vein material). (I-L) *en-Gal4/UAS-p35;puc^{A251.1/+}*. (I) DNA. (J,L) JNK^{REP} (red) and DNA (blue). (J) TUNEL label (green). Ectopic JNK^{REP} activity is associated with regions of overgrowth. (J) Less severe example. (L) More severe example. (K) Higher magnification of J. (M,N) Irradiated *en-Gal4/UAS-p35;puc^{A251.1/+}*. (M) JNK^{REP} (red) and DNA (blue). p35 does not inhibit JNK^{REP} activation in the posterior compartment. (N) DNA (blue). TUNEL label (green). p35 blocks radiation-induced apoptosis. (O,P) JNK^{REP} (red), Wg (green) and DNA (blue). (O) *puc^{A251.1/+}* control. Normal pattern of Wg. (P) *en-Gal4/UAS-p35;puc^{A251.1/+}*. Wg is ectopically expressed by a subset of cells that activate the JNK^{REP} (arrowheads). (Inset) Higher magnification of P.

always induce apoptosis. JNK signaling is required for embryonic dorsal closure. High JNK activity is normally restricted to the dorsal-most epithelial cells, though in *puc* mutants it extends into adjacent cells (Martin-Blanco et al., 1998). However, this JNK signaling does not trigger apoptosis (Fig. 1N). By contrast, ectopic JNK signaling in ventral and lateral epithelial cells in *puc* mutants does trigger apoptosis. Thus, cells where JNK activity is normally high are somehow refractory to JNK-induced apoptosis. JNK signaling normally activates Dpp in dorsal epithelial cells, with ectopic Dpp activation in more lateral cells in *puc* mutants – perhaps this is anti-apoptotic, as Dpp is a survival factor in wing discs (e.g. Moreno et al., 2002a). Interplay between death and survival

signals may also explain the strongly synergistic stimulation of apoptosis in the embryonic epidermis we previously observed when we simultaneously eliminated zygotic Puc (lowering the threshold for JNK signaling) and removed the survival signal Wg (McEwen et al., 2000).

Fitting JNK into the circuitry regulating radiation-induced apoptosis

Studies of JNK signaling in mammalian cells suggest that it plays a role in radiation-induced apoptosis (Davis, 2000). We tested this hypothesis. When we block JNK activation by expressing Myc-Puc using a JNK-independent promoter, radiation-induced apoptosis is attenuated. Thus, JNK signaling plays a crucial role in promoting apoptosis in response to radiation in *Drosophila*, paralleling its essential role in mammalian UV-induced apoptosis (Tournier et al., 2000).

In mammalian cells, prolonged JNK activity correlates with apoptosis (Chen et al., 1996), while transient activation induces cell proliferation (Sabapathy et al., 1999). Thus, differences in both signal amplitude and duration are key determinants of the biological outcome. One mechanism modulating signal amplitude is a negative feedback loop; JNK signaling induces expression of MKPs such as Puc. Our data provide an instance of how this may regulate the DNA damage response. *puc* is upregulated by γ -irradiation in a JNK-dependent manner, and artificially prolonged Myc-Puc expression prevents radiation-induced apoptosis. Thus, the initial increase in Puc expression following irradiation may create a 'grace period' during which Puc elevates the threshold of JNK activation required to induce apoptosis. During this time, cells could attempt to repair radiation-induced damage. However, if damage persists and JNK stimulation continues, the Puc-defined threshold may be exceeded. Indeed, artificially prolonged p53 overexpression overcomes the anti-apoptotic effects of Puc overexpression, perhaps overwhelming the ability of Puc to restrain JNK.

p53 is a key regulator of decisions between life and death in response to DNA damage. Thus, we investigated how JNK signaling and p53 are integrated. Expression of p53 activated JNK, as revealed by its effect on the JNK^{REP}, while loss of p53 prevented radiation-induced JNK activation. These results suggest that p53 acts upstream of JNK signaling in response to cellular stress. Several mechanisms are possible; e.g. p53 might upregulate transcription of JNK pathway components, whose overexpression can induce apoptosis (Adachi-Yamada et al., 1999; McEwen et al., 2000).

p53 normally monitors genome integrity. Loss of p53 significantly, although not completely, suppresses apoptosis induced by Puc inactivation. One model to explain this suggests that basal levels of DNA damage or replication errors act through p53 to regulate basal JNK activity during normal development. This basal activity is kept below the apoptotic threshold by Puc. In *p53* mutants, basal JNK activity would be lowered enough that it could not exceed the apoptotic threshold, even in the absence of Puc. Alternatively, p53 may also function downstream of JNK. Consistent with this, p38 can activate p53 in response to UV (Bulavin et al., 1999; Huang et al., 1999), and JNK can regulate p53 stability/activity via direct phosphorylation (Buschmann et al., 2001). This would set up a positive-feedback loop: DNA damage-triggered activation of p53 would induce JNK

activation, which would further elevate p53 activity. Additional experiments are required to test these alternate hypotheses.

Signal transduction pathways regulate apoptosis, at least in part, by regulating transcription of *rpr*, *hid* and *grim* (the RHG proteins), the key developmental effectors of apoptosis in *Drosophila* (reviewed by Martin, 2002). The relationship between JNK signaling, RHG proteins and caspase activation are not well understood. Caspase 3 cleavage of Mst1, an upstream regulator of JNK and p38, has been suggested to amplify apoptotic responses (Graves et al., 1998), while other data suggest that Mst1 activates caspases via a JNK-dependent pathway (Ura et al., 2001). Likewise, in *Drosophila*, initiation of an apoptotic response by inactivation of DIAP1 may lead to caspase-independent induction of a JNK^{REP} (Kuranaga et al., 2002; Ryoo et al., 2004).

We directly assessed regulatory relationships between JNK activation, RHG proteins and caspase activation. Our data suggest that JNK signaling acts through RHG proteins and caspases to induce apoptosis. JNK signaling is required for *rpr*-reporter induction in response to radiation. Thus, RHG proteins may be JNK-responsive target genes upregulated to elicit apoptosis. Consistent with this hypothesis, mis-expression of *eiger*, a known JNK activator, promotes *hid* expression and apoptosis in eye discs (Moreno et al., 2002b).

To assess whether JNK signaling can be triggered by caspase activation, we examined JNK^{REP} activity in response to Rpr or Hid expression. Both induced apoptosis without concomitant JNK activation. Furthermore, our data suggest that, in at least some contexts, caspases act downstream of JNK: p35-mediated caspase blockade allows cells with elevated JNK signaling to survive in imaginal discs, but does not prevent JNK activation in response to irradiation. Thus, RHG proteins elicit caspase-mediated apoptosis downstream of JNK activation.

JNK-induced apoptosis during normal development

Work on JNK-induced apoptosis in *Drosophila* has focused on its role in the complex processes shaping organ size. Wing discs cells make decisions about whether to die or proliferate by integrating levels of different developmental signals they receive, and comparing their status with that of their neighbors. This occurs in part by competition for survival signals like the TGF β family member Dpp (Burke and Basler, 1996; Martin et al., 2004; Moreno et al., 2002a). Complex crosstalk among this and other signaling pathways precisely regulates the size and pattern of the wing, in a process that is very resistant to tissue damage or developmental errors. Discontinuities in smooth morphogen gradients, which may arise from errors in patterning or tissue injury, are corrected in part by JNK-dependent apoptosis (e.g. Adachi-Yamada and O'Connor, 2002).

Our data clarifies roles for JNK signaling in adult development (Agnes et al., 1999). Our failure to recover *puc* clones anywhere in the wing disc suggests that basal JNK activity is sufficient to promote cell-autonomous apoptosis independent of other signals activating JNK. However, JNK signaling does not play a crucial role in wing patterning, as JNK inactivation in the posterior compartment (by Myc-Puc mis-expression) did not have drastic consequences. We did identify roles for JNK signaling in genitalia and thoracic bristles, where it regulates developmentally programmed apoptosis; this was also reported by Macias et al. (Macias et al., 2004).

Can MKPs act as tumor suppressors?

JNK signaling plays complex roles during oncogenesis. It can prevent tumorigenesis by promoting apoptosis, and it can promote tumorigenesis by supporting Ras- (Behrens et al., 2000; Smeal et al., 1991) or BCR-Abl-mediated (Dickens et al., 1997; Raitano et al., 1995) transformation. As each tumor type has a unique set of mutations in oncogenes and/or tumor suppressors, the phenotypic effects of JNK activation probably differ depending on the activity of other pathways.

Inhibition of apoptosis is one prerequisite for tumorigenesis. We thus examined the consequences of JNK activation when apoptosis was blocked. When cell death was blocked in the posterior compartment of the wing and the restraints on JNK activation were relaxed by *puc* heterozygosity, tissue overgrowth occurred. Groups of posterior cells, presumably of clonal origin, exhibited elevated levels of JNK activation, and formed small overgrowths both in developing imaginal discs and in the resulting adult wings and legs. Thus, when apoptosis is suppressed, JNK activation can lead to tissue over-growth.

Ryoo et al. (Ryoo et al., 2004), Pérez-Garijo et al. (Pérez-Garijo et al., 2004), and Huh et al. (Huh et al., 2004) recently reported related results, inducing apoptosis by *diap* inactivation, irradiation or Hid expression, while simultaneously blocking caspase activation. This triggered non-autonomous proliferation of neighboring cells, presumably to compensate for cell lost by apoptosis. Furthermore, some of the 'undead' cells produced by these treatments show JNK-dependent upregulation of Wg or Dpp. Ryoo et al. show that Wg signaling is required for compensatory proliferation. Our results extend theirs, as our experiment differed in one significant way: we did not actively induce apoptosis, but simply blocked caspase activation in *puc* heterozygotes. Thus, when apoptosis is inhibited and Puc repression is reduced, cells become susceptible to runaway JNK activation. It is likely that analogous focal JNK activation occurs in cells in which apoptosis is not blocked (e.g. anterior compartment cells in our experiment), but JNK-induced apoptosis rapidly eliminates them. We also found that at least a subset of the undead cells activated expression of Wg, which may promote excess growth. Interestingly, however, this ectopic Wg expression does not alter cell fates, at least in those situations where overgrowths survive to be observed in the adult wing.

Together, these data have interesting implications. In tumor cells in which apoptosis is prevented, JNK signaling might switch from promoting apoptosis to promoting proliferation by inducing Wnt or TGF β , suggesting how JNK signaling can be both pro- and anti-tumorigenic. Future experiments should address mechanisms by which JNK activation triggers Wnt and TGF β signaling. Our data also suggest that runaway JNK activation can promote cell-autonomous proliferation, as groups of cells with elevated JNK activity are associated with overgrowths. Thus, JNK activation or loss of MKP-mediated JNK repression may play multiple roles in tumors whose cells have lost the ability to die.

Ectopic JNK activation occurred in small groups of cells in random positions throughout the posterior compartment. The event(s) that initiate unrestrained JNK activation in these cells remain to be defined. Perhaps there are stochastic variations in JNK signaling that are normally below the threshold triggering

apoptosis. However, when Puc activity is reduced, some cells may exceed this threshold, triggering runaway JNK activation in the initial cell and its descendants. Variations in JNK activity may also be induced by spontaneous DNA damage. Normally, such damage would trigger JNK-dependent apoptosis; however, if apoptosis is blocked, the cells proliferate. Finally, loss-of-heterozygosity at the *puc* locus could create cells lacking restraints on JNK signaling. As mitotic recombination in somatic tissue can occur (Baker et al., 1978), Puc might act in a manner analogous to classic tumor suppressor genes. Additional studies will be required to distinguish between these possibilities.

In summary, our work establishes that Puc is a key negative regulator of apoptosis throughout *Drosophila* development. In its absence, basal JNK activity is poised to eliminate cells from the developing epithelium. Our results position JNK signaling in the hierarchy of events regulating radiation-induced apoptosis. Finally, our data support the possibility previously suggested by cytogenetics (Armes et al., 2004; Furukawa et al., 2003) that MKPs may act as tumor suppressor genes. These data prompt many new mechanistic questions regarding the role of JNK signaling in apoptosis and oncogenesis.

We thank the Bloomington *Drosophila* Stock Center, Developmental Studies Hybridoma Bank, A. Martinez-Arias, T. Uemura and L. Dobens for reagents, and J. Gates, V. Bautch, T. Harris, A. Fanning and B. Duronio for comments. This work was supported by NIH GM47857 (M.P.). M.P. was supported in part by the Army Breast Cancer Research Program and D.G.M. was supported by NIH 1F32GM19824.

Supplementary material

Supplementary material for this article is available at <http://dev.biologists.org/cgi/content/full/132/17/3935/DC1>

References

- Abbott, M. K. and Lengyel, J. A. (1991). Embryonic head involution and rotation of male terminalia require the *Drosophila* locus head involution defective. *Genetics* **129**, 783-789.
- Abrams, J. M., White, K., Fessler, L. I. and Steller, H. (1993). Programmed cell death during *Drosophila* embryogenesis. *Development* **117**, 29-43.
- Adachi-Yamada, T., Fujimura-Kamada, K., Nishida, Y. and Matsumoto, K. (1999). Distortion of proximodistal information causes JNK-dependent apoptosis in *Drosophila* wing. *Nature* **400**, 166-169.
- Adachi-Yamada, T. and O'Connor, M. B. (2002). Morphogenetic apoptosis: a mechanism for correcting discontinuities in morphogen gradients. *Dev. Biol.* **251**, 74-90.
- Adachi-Yamada, T. and O'Connor, M. B. (2004). Mechanisms for removal of developmentally abnormal cells: cell competition and morphogenetic apoptosis. *J. Biochem. (Tokyo)* **136**, 13-17.
- Agnes, F., Suzanne, M. and Noselli, S. (1999). The *Drosophila* JNK pathway controls the morphogenesis of imaginal discs during metamorphosis. *Development* **126**, 5453-5462.
- Armes, J. E., Hammet, F., de Silva, M., Ciciulla, J., Ramus, S. J., Soo, W. K., Mahoney, A., Yarovaya, N., Henderson, M. A., Gish, K. et al. (2004). Candidate tumor-suppressor genes on chromosome arm 8p in early-onset and high-grade breast cancers. *Oncogene* **23**, 5697-5702.
- Baker, B. S., Carpenter, A. T. C. and Ripoll, P. (1978). The utilization during mitotic cell division of loci controlling Meiotic recombination and disjunction in *Drosophila melanogaster*. *Genetics* **90**, 531-578.
- Behrens, A., Jochum, W., Sibilio, M. and Wagner, E. F. (2000). Oncogenic transformation by ras and fos is mediated by c-Jun N-terminal phosphorylation. *Oncogene* **19**, 2657-2663.
- Brodsky, M. H., Nordstrom, W., Tsang, G., Kwan, E., Rubin, G. M. and Abrams, J. M. (2000). *Drosophila* p53 binds a damage response element at the reaper locus. *Cell* **101**, 103-113.

- Brodsky, M. H., Weinert, B. T., Tsang, G., Rong, Y. S., McGinnis, N. M., Golic, K. G., Rio, D. C. and Rubin, G. M. (2004). Drosophila melanogaster MNK/Chk2 and p53 regulate multiple DNA repair and apoptotic pathways following DNA damage. *Mol. Cell. Biol.* **24**, 1219-1231.
- Brondello, J. M., Pouyssegur, J. and McKenzie, F. R. (1999). Reduced MAP kinase phosphatase-1 degradation after p42/p44MAPK-dependent phosphorylation. *Science* **286**, 2514-2517.
- Bulavin, D. V., Saito, S., Hollander, M. C., Sakaguchi, K., Anderson, C. W., Appella, E. and Fornace, A. J., Jr (1999). Phosphorylation of human p53 by p38 kinase coordinates N-terminal phosphorylation and apoptosis in response to UV radiation. *EMBO J.* **18**, 6845-6854.
- Burke, R. and Basler, K. (1996). Dpp receptors are autonomously required for cell proliferation in the entire developing Drosophila wing. *Development* **122**, 2261-2269.
- Buschmann, T., Potapova, O., Bar-Shira, A., Ivanov, V. N., Fuchs, S. Y., Henderson, S., Fried, V. A., Minamoto, T., Alarcon-Vargas, D., Pincus, M. R. et al. (2001). Jun NH2-terminal kinase phosphorylation of p53 on Thr-81 is important for p53 stabilization and transcriptional activities in response to stress. *Mol. Cell. Biol.* **21**, 2743-2754.
- Chen, Y. R., Meyer, C. F. and Tan, T. H. (1996). Persistent activation of c-Jun N-terminal kinase 1 (JNK1) in gamma radiation-induced apoptosis. *J. Biol. Chem.* **271**, 631-634.
- Chou, T. B. and Perrimon, N. (1992). Use of a yeast site-specific recombinase to produce female germline chimeras in Drosophila. *Genetics* **131**, 643-653.
- Davis, R. J. (2000). Signal transduction by the JNK group of MAP kinases. *Cell* **103**, 239-252.
- de la Cova, C., Abril, M., Bellosta, P., Gallant, P. and Johnston, L. A. (2004). Drosophila myc regulates organ size by inducing cell competition. *Cell* **117**, 107-116.
- Dickens, M., Rogers, J. S., Cavanagh, J., Raitano, A., Xia, Z., Halpern, J. R., Greenberg, M. E., Sawyers, C. L. and Davis, R. J. (1997). A cytoplasmic inhibitor of the JNK signal transduction pathway. *Science* **277**, 693-696.
- Dorfman, K., Carrasco, D., Gruda, M., Ryan, C., Lira, S. A. and Bravo, R. (1996). Disruption of the erp/mkp-1 gene does not affect mouse development: normal MAP kinase activity in ERP/MKP-1-deficient fibroblasts. *Oncogene* **13**, 925-931.
- Flint, A. J., Tiganis, T., Barford, D. and Tonks, N. K. (1997). Development of 'substrate-trapping' mutant to identify physiological substrates of protein tyrosine phosphatases. *Proc. Natl. Acad. Sci. USA* **94**, 1680-1685.
- Franklin, C. C. and Kraft, A. S. (1997). Conditional expression of the mitogen-activated protein kinase (MAPK) phosphatase MKP-1 preferentially inhibits p38 MAPK and stress-activated protein kinase in U937 cells. *J. Biol. Chem.* **272**, 16917-16923.
- Furukawa, T., Sunamura, M., Motoi, F., Matsuno, S. and Horii, A. (2003). Potential tumor suppressive pathway involving DUSP6/MKP-3 in pancreatic cancer. *Am. J. Pathol.* **162**, 1807-1815.
- Graves, J. D., Gotoh, Y., Draves, K. E., Ambrose, D., Han, D. K., Wright, M., Chernoff, J., Clark, E. A. and Krebs, E. G. (1998). Caspase-mediated activation and induction of apoptosis by the mammalian Ste20-like kinase Mst1. *EMBO J.* **17**, 2224-2234.
- Huang, C., Ma, W. Y., Maxiner, A., Sun, Y. and Dong, Z. (1999). p38 kinase mediates UV-induced phosphorylation of p53 protein at serine 389. *J. Biol. Chem.* **274**, 12229-12235.
- Huh, J. R., Guo, M. and Hay, B. A. (2004). Compensatory proliferation induced by cell death in the Drosophila wing disc requires activity of the apical cell death caspase Dronc in a nonapoptotic role. *Curr. Biol.* **14**, 1262-1266.
- Igaki, T., Kanda, H., Yamamoto-Goto, Y., Kanuka, H., Kuranaga, E., Aigaki, T. and Miura, M. (2002). Eiger, a TNF superfamily ligand that triggers the Drosophila JNK pathway. *EMBO J.* **21**, 3009-3018.
- Jassim, O. W., Fink, J. L. and Cagan, R. L. (2003). Dmp53 protects the Drosophila retina during a developmentally regulated DNA damage response. *EMBO J.* **22**, 5622-5632.
- Kanuka, H., Sawamoto, K., Inohara, N., Matsuno, K., Okano, H. and Miura, M. (1999). Control of the cell death pathway by Dapaf-1, a Drosophila Apaf-1/CED-4-related caspase activator. *Mol. Cell* **4**, 757-769.
- Keyse, S. M. (1998). Protein phosphatases and the regulation of MAP kinase activity. *Semin. Cell Dev. Biol.* **9**, 143-152.
- Kuranaga, E., Kanuka, H., Igaki, T., Sawamoto, K., Ichijo, H., Okano, H. and Miura, M. (2002). Reaper-mediated inhibition of DIAP1-induced DTRAF1 degradation results in activation of JNK in Drosophila. *Nat. Cell Biol.* **4**, 705-710.
- Lee, J. H., Lee, E., Park, J., Kim, E., Kim, J. and Chung, J. (2003). In vivo p53 function is indispensable for DNA damage-induced apoptotic signaling in Drosophila. *FEBS Lett.* **550**, 5-10.
- Lee, T. and Luo, L. (1999). Mosaic analysis with a repressible cell marker for studies of gene function in neuronal morphogenesis. *Neuron* **22**, 451-461.
- Lin, Y. W., Chuang, S. M. and Yang, J. L. (2003). ERK1/2 achieves sustained activation by stimulating MAPK phosphatase-1 degradation via the ubiquitin-proteasome pathway. *J. Biol. Chem.* **278**, 21534-21541.
- Macias, A., Romero, N. M., Martin, F., Suarez, L., Rosa, A. L. and Morata, G. (2004). PVF1/PVR signaling and apoptosis promotes the rotation and dorsal closure of the Drosophila male terminalia. *Int. J. Dev. Biol.* **48**, 1087-1094.
- Martin, F. A., Pérez-Garijo, A., Moreno, E. and Morata, G. (2004). The brinker gradient controls wing growth in Drosophila. *Development* **131**, 4921-4930.
- Martin, P. and Parkhurst, S. M. (2004). Parallels between tissue repair and embryo morphogenesis. *Development* **131**, 3021-3034.
- Martin, S. J. (2002). Destabilizing influences in apoptosis: sowing the seeds of IAP destruction. *Cell* **109**, 793-796.
- Martin-Blanco, E., Gampel, A., Ring, J., Virdee, K., Kirov, N., Tolkovsky, A. M. and Martinez-Arias, A. (1998). puckered encodes a phosphatase that mediates a feedback loop regulating JNK activity during dorsal closure in Drosophila. *Genes Dev.* **12**, 557-570.
- Martin-Blanco, E., Pastor-Pareja, J. C. and Garcia-Bellido, A. (2000). JNK and decapentaplegic signaling control adhesiveness and cytoskeleton dynamics during thorax closure in Drosophila. *Proc. Natl. Acad. Sci. USA* **97**, 7888-7893.
- McEwen, D. G., Cox, R. T. and Peifer, M. (2000). The canonical Wg and JNK signaling cascades collaborate to promote both dorsal closure and ventral patterning. *Development* **127**, 3607-3617.
- Moreno, E. and Basler, K. (2004). dMyc transforms cells into supercompetitors. *Cell* **117**, 117-129.
- Moreno, E., Basler, K. and Morata, G. (2002a). Cells compete for decapentaplegic survival factor to prevent apoptosis in Drosophila wing development. *Nature* **416**, 755-759.
- Moreno, E., Yan, M. and Basler, K. (2002b). Evolution of TNF signaling mechanisms. JNK-dependent apoptosis triggered by Eiger, the Drosophila homolog of the TNF superfamily. *Curr. Biol.* **12**, 1263.
- Neufeld, T. P., de la Cruz, A. F., Johnston, L. A. and Edgar, B. A. (1998). Coordination of growth and cell division in the Drosophila wing. *Cell* **93**, 1183-1193.
- Nordstrom, W., Chen, P., Steller, H. and Abrams, J. M. (1996). Activation of the reaper gene during ectopic cell killing in Drosophila. *Dev. Biol.* **180**, 213-226.
- Noselli, S. and Agnes, F. (1999). Roles of the JNK signaling pathway in Drosophila morphogenesis. *Curr. Opin. Genet. Dev.* **9**, 466-472.
- Ollmann, M., Young, L. M., Di Como, C. J., Karim, F., Belvin, M., Robertson, S., Whittaker, K., Demsky, M., Fisher, W. W., Buchman, A. et al. (2000). Drosophila p53 is a structural and functional homolog of the tumor suppressor p53. *Cell* **101**, 91-101.
- Osborn, M. T. and Chambers, T. C. (1996). Role of the stress-activated/c-Jun NH2-terminal protein kinase pathway in the cellular response to adriamycin and other chemotherapeutic drugs. *J. Biol. Chem.* **271**, 30950-30955.
- Pérez-Garijo, A., Martin, F. A. and Morata, G. (2004). Caspase inhibition during apoptosis causes abnormal signalling and developmental aberrations in Drosophila. *Development* **131**, 5591-5598.
- Raitano, A. B., Halpern, J. R., Hambuch, T. M. and Sawyers, C. L. (1995). The Bcr-Abl leukemia oncogene activates Jun kinase and requires Jun for transformation. *Proc. Natl. Acad. Sci. USA* **2**, 11746-11750.
- Riesgo-Escovar, J. R., Jenni, M., Fritz, A. and Hafen, E. (1996). The Drosophila Jun-N-terminal kinase is required for cell morphogenesis but not for DJun-dependent cell fate specification in the eye. *Genes Dev.* **10**, 2759-2768.
- Rintelen, F., Hafen, E. and Nairz, K. (2003). The Drosophila dual-specificity ERK phosphatase DMKP3 cooperates with the ERK tyrosine phosphatase PTP-ER. *Development* **130**, 3479-3490.
- Rodriguez, A., Oliver, H., Zou, H., Chen, P., Wang, X. and Abrams, J. M. (1999). Dark is a Drosophila homologue of Apaf-1/CED-4 and functions in an evolutionarily conserved death pathway. *Nat. Cell Biol.* **1**, 272-279.
- Ryoo, H. D., Gorenc, T. and Steller, H. (2004). Apoptotic cells can induce compensatory cell proliferation through the JNK and the Wingless signaling pathways. *Dev. Cell* **7**, 491-501.
- Sabapathy, K., Hu, Y., Kallunki, T., Schreiber, M., David, J. P., Jochum,

- W., Wagner, E. F. and Karin, M. (1999). JNK2 is required for efficient T-cell activation and apoptosis but not for normal lymphocyte development. *Curr. Biol.* **9**, 116-125.
- Schaeffer, H. J. and Weber, M. J. (1999). Mitogen-activated protein kinases: specific messages from ubiquitous messengers. *Mol. Cell. Biol.* **19**, 2435-2444.
- Seimiya, H., Mashima, T., Toho, M. and Tsuruo, T. (1997). c-Jun NH2-terminal kinase-mediated activation of interleukin-1beta converting enzyme/CED-3-like protease during anticancer drug-induced apoptosis. *J. Biol. Chem.* **272**, 4631-4636.
- Sluss, H. K., Han, Z., Barrett, T., Davis, R. J. and Ip, Y. T. (1996). A JNK signal transduction pathway that mediates morphogenesis and an immune response in *Drosophila*. *Genes Dev.* **10**, 2745-2758.
- Smeal, T., Binetruy, B., Mercola, D. A., Birrer, M. and Karin, M. (1991). Oncogenic and transcriptional cooperation with Ha-Ras requires phosphorylation of c-Jun on serines 63 and 73. *Nature* **354**, 494-496.
- Sogame, N., Kim, M. and Abrams, J. M. (2003). *Drosophila* p53 preserves genomic stability by regulating cell death. *Proc. Natl. Acad. Sci. USA* **100**, 4696-4701.
- Stadheim, T. A. and Kucera, G. L. (2002). c-Jun N-terminal kinase/stress-activated protein kinase (JNK/SAPK) is required for mitoxantrone- and anisomycin-induced apoptosis in HL-60 cells. *Leuk. Res.* **26**, 55-65.
- Stowers, R. S. and Schwarz, T. L. (1999). A genetic method for generating *Drosophila* eyes composed exclusively of mitotic clones of a single genotype. *Genetics* **152**, 1631-1639.
- Stronach, B. E. and Perrimon, N. (1999). Stress signaling in *Drosophila*. *Oncogene* **18**, 6172-6182.
- Sun, H., Charles, C. H., Lau, L. F. and Tonks, N. K. (1993). MKP-1 (3CH134), an immediate early gene product, is a dual specificity phosphatase that dephosphorylates MAP kinase in vivo. *Cell* **75**, 487-493.
- Tateno, M., Nishida, Y. and Adachi-Yamada, T. (2000). Regulation of JNK by Src during *Drosophila* development. *Science* **287**, 324-327.
- Tepass, U., Theres, C. and Knust, E. (1990). crumbs encodes an EGF-like protein expressed on apical membranes of *Drosophila* epithelial cells and required for organization of epithelia. *Cell* **61**, 787-799.
- Tournier, C., Hess, P., Yang, D. D., Xu, J., Turner, T. K., Nimnual, A., Barsagi, D., Jones, S. N., Flavell, R. A. and Davis, R. J. (2000). Requirement of JNK for stress-induced activation of the cytochrome c-mediated death pathway. *Science* **288**, 870-874.
- Ura, S., Masuyama, N., Graves, J. D. and Gotoh, Y. (2001). MST1-JNK promotes apoptosis via caspase-dependent and independent pathways. *Genes Cells* **6**, 519-350.
- Xu, T. and Rubin, G. M. (1993). Analysis of genetic mosaics in developing and adult *Drosophila* tissues. *Development* **117**, 1223-1237.