

Northumbria Research Link

Citation: García-Molina, Pablo, Munoz, Jose, Ortuño, Joaquin A., Rodríguez-López, José Neptuno, García-Ruiz, Pedro Antonio, García-Cánovas, Francisco and García-Molina, Francisco (2021) Considerations about the Continuous Assay Methods, Spectro-Photometric and Spectrofluorometric, of the Monophenolase Activity of Tyrosinase. *Biomolecules*, 11 (9). p. 1269. ISSN 2218-273X

Published by: MDPI

URL: <https://doi.org/10.3390/biom11091269> <<https://doi.org/10.3390/biom11091269>>

This version was downloaded from Northumbria Research Link:
<http://nrl.northumbria.ac.uk/id/eprint/47000/>

Northumbria University has developed Northumbria Research Link (NRL) to enable users to access the University's research output. Copyright © and moral rights for items on NRL are retained by the individual author(s) and/or other copyright owners. Single copies of full items can be reproduced, displayed or performed, and given to third parties in any format or medium for personal research or study, educational, or not-for-profit purposes without prior permission or charge, provided the authors, title and full bibliographic details are given, as well as a hyperlink and/or URL to the original metadata page. The content must not be changed in any way. Full items must not be sold commercially in any format or medium without formal permission of the copyright holder. The full policy is available online: <http://nrl.northumbria.ac.uk/policies.html>

This document may differ from the final, published version of the research and has been made available online in accordance with publisher policies. To read and/or cite from the published version of the research, please visit the publisher's website (a subscription may be required.)

Article

Considerations about the Continuous Assay Methods, Spectro-Photometric and Spectrofluorometric, of the Monophenolase Activity of Tyrosinase

Pablo García-Molina ¹, José Luis Muñoz-Munoz ^{2,*}, Joaquín A. Ortuño ³, José Neptuno Rodríguez-López ¹, Pedro Antonio García-Ruiz ⁴, Francisco García-Cánovas ¹ and Francisco García-Molina ^{5,*}

Citation: García-Molina, P.; Muñoz-Munoz, J.L.; Ortuño, J.A.; Rodríguez-López, J.N.; García-Ruiz, P.A.; García-Cánovas, F.; García-Molina, F. Considerations about the Continuous Assay Methods, Spectro-Photometric and Spectrofluorometric, of the Monophenolase Activity of Tyrosinase. *Biomolecules* **2021**, *11*, 1269. <https://doi.org/10.3390/biom11091269>

Academic Editor:
Eugene A. Permyakov

Received: 12 August 2021
Accepted: 22 August 2021
Published: 25 August 2021

Publisher's Note: MDPI stays neutral with regard to jurisdictional claims in published maps and institutional affiliations.



Copyright: © 2021 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (<http://creativecommons.org/licenses/by/4.0/>).

- ¹ GENZ-Group of Research on Enzymology, Department of Biochemistry and Molecular Biology-A, Regional Campus of International Excellence "Campus Mare Nostrum", University of Murcia, 30100 Murcia, Spain; Pablo.garcia14@um.es (P.G.-M.); neptuno@um.es (J.N.R.-L.); canovasf@um.es (F.G.-C.)
 - ² Microbial Enzymology Group, Department of Applied Sciences, Ellison Building A, University of Northumbria, Newcastle Upon Tyne NE1 8ST, UK
 - ³ Department of Analytical Chemistry, Faculty of Chemistry, University of Murcia, 30100 Murcia, Spain; jortuno@um.es
 - ⁴ Group of Chemistry of Carbohydrates, Industrial Polymers and Additives, Department of Organic Chemistry, Regional Campus of International Excellence "Campus Mare Nostrum", University of Murcia, 30100 Murcia, Spain; pagr@um.es
 - ⁵ Department of Anatomía Patológica, Hospital General Universitario Reina Sofía, Av. Intendente Jorge Palacios, 1, 30003 Murcia, Spain
- * Correspondence: jose.munoz@northumbria.ac.uk (J.L.M.-M.); pacogm@um.es (F.G.-M.)

Abstract: With the purpose to obtain the more useful tyrosinase assay for the monophenolase activity of tyrosinase between the spectrofluorometric and spectrophotometric continuous assays, simulated assays were made by means of numerical integration of the equations that characterize the mechanism of monophenolase activity. These assays showed that the rate of disappearance of monophenol ($V_{ss}^{M,M}$) is equal to the rate of accumulation of dopachrome ($V_{ss}^{M,DC}$) or to the rate of accumulation of its oxidized adduct, originated by the nucleophilic attack on *o*-quinone by a nucleophile such as 3-methyl-2-benzothiazolinone (MBTH), (V_{ss}^{MA-ox}), despite the existence of coupled reactions. It is shown that the spectrophotometric methods that use MBTH are more useful, as they do not have the restrictions of the L-tyrosine disappearance measurement method, of working at pH = 8 and not having a linear response from 100 μ M of L-tyrosine. It is possible to obtain low LOD^M (limit of detection of the monophenolase activity) values with spectrophotometric methods. The spectrofluorimetric methods had a lower LOD^M than spectrophotometric methods. In the case of 4-hydroxyphenyl-propionic acid, the LOD^M obtained by us was 0.25 U/mL. Considering the relative sensitivities of 4-hydroxyanisole, compared with 4-hydroxyphenyl-propionic acid, LOD^M values like those obtained by fluorescent methods would be expected.

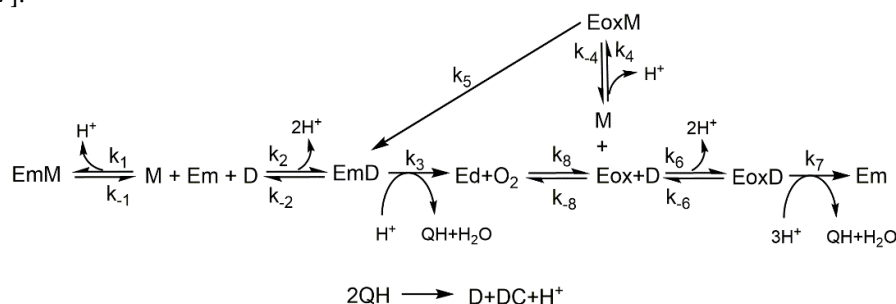
Keywords: Tyrosinase; polyphenol oxidase; monophenolase activity; fluorimetric method; spectrophotometric method; LOD^M

1. Introduction

Tyrosinase (EC 1.14.18.1) is a cuproprotein that catalyses the hydroxylation of monophenols to *o*-diphenols (monophenolase activity) and the oxidation of *o*-diphenols to *o*-quinones (diphenolase activity), with the help of molecular oxygen [1]. This enzyme is distributed throughout the phylogenetic scale [2].

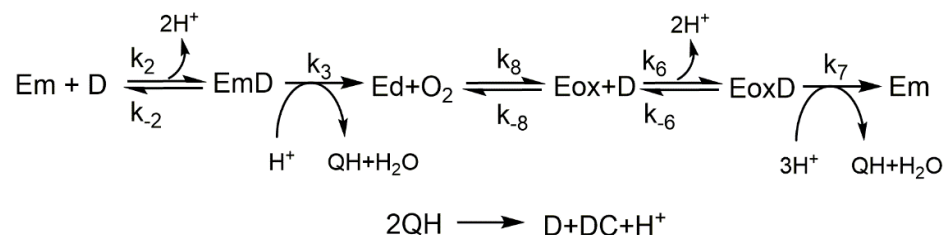
The enzyme tyrosinase, in the catalytic cycle, is found in three forms—meta, deoxy and oxy, which are differentiated by the degree of oxidation of copper: Em: metatyrosinase (Cu²⁺ Cu²⁺); Ed: deoxytyrosinase (Cu¹⁺ Cu¹⁺); Eox: oxytyrosinase (Cu²⁺ Cu²⁺ O₂⁻²) [3]. In

the case of mushroom tyrosinase, at the oxygen concentration existing in the solutions, the enzyme is saturated [4]. The two enzymatic forms Em and Eox are active on *o*-diphenols, oxidizing them to *o*-quinones, Em without oxygen and Eox with the participation of oxygen. However, with respect to monophenols, the Em form is inactive, forming an EmM dead-path complex. The Eox form is active on monophenols, carrying out its hydroxylation to *o*-diphenols through an electrophilic aromatic substitution [5–7]. This difference in activities of the Em and Eox forms is the origin of the kinetic complexity of the tyrosinase monophenolase activity. The EmM complex is inactive and therefore for the enzyme to be active on monophenols it is necessary for the Em form to be reduced to originate the Ed form and its binding to the oxygen to form Eox, closing the catalytic cycle (Scheme 1) [5–7].



Scheme 1. Monophenolase and diphenolase activities of tyrosinase. M is monophenol, D is *o*-diphenol, QH is *o*-dopaquinone protonated and DC is dopachrome. The enzymatic forms are: Em (metatyrosinase), Ed (deoxytyrosinase) and Eox (oxytyrosinase).

There is no such problem in diphenolase activity as the two forms Em and Eox are active on *o*-diphenol (D) (Scheme 2) [5]. In nature, this problem has been solved (between the Em and Eox forms), as the product of the enzyme when it acts on its physiological substrates (L-tyrosine (M), L-dopa (D)) is *o*-Q that evolves to go through chemical reactions generating D in the medium, this D, L-dopa reduces Em to Ed and thus Eox (Scheme 1) is formed [8].



Scheme 2. Diphenolase activity of tyrosinase. D is *o*-diphenol, QH is *o*-dopaquinone protonated and DC is dopachrome.

When tyrosinase acts on M, it forms D, and this becomes *o*-Q, this through its chemical evolution originates dopachrome (DC) and accumulates D in the medium (Scheme 1). That is, when the tyrosinase acts on M (monophenolase activity), the enzymatic reaction belonging to the diphenolase activity will occur but also the chemical reactions that involve evolution from *o*-Q [5].

This overlap between the activities (monophenolase and diphenolase) has led some authors to try to separate the monophenolase activity from the rest, in order to obtain the direct kinetic information of this enzymatic activity [9]. The first work that tried to separate the monophenolase activity used hydroxylamine to transform Em to Ed, and borate

at pH = 8 to block the D originated by the enzyme from M. These authors make the experimental measurements of rates at the steady-state, measuring oxygen consumption (O_2) [9].

Recently, based on the mechanism described above [9], it has been proposed as a measure of tyrosinase monophenolase activity to follow the disappearance of M directly [10–12]. According to these authors, thus, interference from the other processes (diphenolase activity and chemical reactions of evolution from *o*-dopaquinone) would be avoided. The methods proposed under different approaches are spectrofluorimetric [10–12] and, therefore, very sensitive. However, they have some limitations as detailed below.

Previously, reliable and sensitive continuous spectrophotometric methods have been developed [13,14]. The first contribution to establish a spectrophotometric measurement method for monophenolase activity, measuring the oxidation of the adducts formed between *o*-quinones and MBTH, based on the method described by Winder and Harris for diphenolase activity [13], was developed with mushroom tyrosinase and tyramine [14].

This method showed some advantages over those used up to that time:

- Radiometric methods that are more sensitive but are discontinuous and require about 30 min per activity assay [13,15].
- The spectrophotometric method normally used measuring DC [16]. This method is 8 times more sensitive and 6.5 times more sensitive than the proline-coupled assay [17].
- Adducts and oxidized adducts caused by the coupling of MBTH with *o*-quinones show high molar absorptivity coefficients (see Table S1).
- The method has no interferences since the absorbance measurement occurs in the visible area, where neither the substrates nor the enzyme absorbed.
- The adducts are stable at acidic pH and at pH > 5.5 they evolve presenting an isobestic point, this particularity makes it possible to measure different enzymes of fruits and vegetables that have an optimal pH in the range between 4.0 and 5.0 [18].

Subsequently, this continuous spectrophotometric method was improved by demonstrating that *p*-hydroxyphenylpropionic acid was a better substrate for the enzyme and had a k_{cat} 2.6 higher than tyramine [19]. Furthermore, because it is an acid, it does not show solubility problems.

Further optimization of the method was achieved using 4-hydroxyanisole as a substrate [18,20], making this substrate the best for a whole series of tyrosinases or polyphenol oxidases of fruits and vegetables such as apple, pear, strawberry, avocado, medlar and artichoke. The origin of the improvement of the continuous spectrophotometric method could be due to the higher catalytic constant that this substrate shows, corresponding to the lower value of chemical displacement of the C-4 carbon of this compound $\delta_4 = 152.35$ (p.p.m.). Subsequently, an extensive study on the substrate specificity of tyrosinase from mushroom, confirmed the results obtained with the rest of tyrosinases previously mentioned [21]. Note in Table 1 how the nature of the substrate affects the limit of detection (LOD^M) values. Thus, the importance of the value of the chemical shift (δ_4), obtained by C-4 ^{13}C NMR, which in turn is related to the nucleophilicity of the oxygen of the phenolic OH- group, is shown to predict the sensitivity of the method of measure.

Table 1. Comparison of different methods to determine monophenolase activity in mushroom tyrosinase.

Analysis Method	Analysis Manner	Substrate	LOD^M (U/mL)	S_R	Reference
Fluorescence	Continuous	L-tyrosine	0.0952		[10]
Fluorescence	Real-time	L-tyrosine	0.0851		[11]
Fluorescence	Continuous synchronous	L-tyrosine	0.0721		[12]

Coupled MBTH	Continuous	4-hydroxyphenyl propionic acid	0.25		[19]
Coupled MBTH	Continuous	Tyramine	–	0.40	[14]
Coupled MBTH	Continuous	4-hydroxyanisole	–	2.64	[20,21]
Coupled MBTH	Continuous	L-tyrosine	–	0.13	[21]

S_R = Relative sensitivity comparing with our reference method with 4-hydroxyphenyl propionic acid. $S_R = k_{cat}^M \epsilon_i^M / k_{cat}^{M(r)} \epsilon_i^{M(r)}$.

The pH value used in the fluorometric methods was high (pH = 8) [10–12], to trap the *o*-diphenol generated in the reaction through the use of borate, thus it deviates from the optimum pH of the mushroom enzyme, pH = 6.8. For the same reason, spectrofluorimetric methods could not be applied to measure monophenolase activities of enzymes of fruits and vegetables that have their optimum pH between 4.0 and 5.0 (Apple (4.5), Avocado (4.6), pear (5), medlar (4.4) and strawberry (4.3)), the spectrophotometric method does not have any pH problem [18].

The sensitivity of the spectrofluorimetric method is shown in Table 1, with the low value of the LOD^M (0.0721 U/mL). With spectrophotometric methods, sensitivities in a range like spectrofluorimetrics could be achieved by varying the nature of the substrate (from L-tyrosine to 4-hydroxyanisole: see Table 1). Relationships are also obtained between the LOD values of different fruit and vegetable enzymes, depending on the type of substrate used, like those described in Table 1. These relationships ($R = LOD^{PHPPA}/LOD^{4HA}$) are: Apple (3.78), artichoke (4.2), pear (3.91), medlar (3.60), avocado (3.76) and strawberry (3.82) [18].

Recently, related to the study on oculocutaneous albinism I, Isothermal titration calorimetry (ITC) has also been successfully used for the evaluation of kinetics of tyrosinase [22], and it is possible to extend the method in the presence of MBTH as shown for assaying manganese peroxidase [23].

The main purpose of this work is demonstrated, through simulated assays by numeric integration of the differential equations of the monophenolase activity of tyrosinase, that despite the coupled reactions that occur in the action of tyrosinase on L-tyrosine, it is true that $V_{SS}^{M,M} = V_{SS}^{M,DC}$ and with respect to oxygen consumption it is true that $V_{SS}^{M,O_2} = 1.5V_{SS}^{M,M} = 1.5V_{SS}^{M,DC} = 1.5V_{SS}^{M,A-Ox}$, proving the usefulness of spectrophotometric versus fluorometric methods. Moreover, the fluorometric assays (10–12), are compared with the spectrophotometric published by our group [14,18–21,24,25].

2. Material and Methods

Experimental results of both, fluorometric [10–12], and spectrophotometric methods [14,18–21,24,25], have been taken from the bibliography.

2.1. Materials

Tyrosine, dopa, tyramine hydrochloride, dopamine hydrochloride, 3,4-dihydroxyphenylpropionic acid (DHPPA), *p*-hydroxyphenylpropionic acid (PHPPA), 4-hydroxyanisole (HA) and 3-methyl-2-benzothiazolinone hydrazone (MBTH) were purchased from Sigma (Madrid). Stock solutions of the phenolic substrate were prepared in 0.15 mM phosphoric acid to prevent autooxidation. The acid characteristic of MBTH requires the use of 50 mM buffer in the assay medium [13].

2.2. Enzyme Source

Mushroom (*Agaricus bisporus*) tyrosinase (3900 units/mg) was purchased from Sigma and purified [26]. Protein content was determined by a modified Lowry method [27].

2.3. Spectrophotometric Assays

Absorption spectra were recorded in a Perkin-Elmer Lambda-2 spectrophotometer; online interfered with a PC laptop. Temperature was controlled at 25°C using a Hacked DIG circulating water bath with a heater/cooler and checked with a precision of $\pm 0.1^\circ\text{C}$. We did not add substrate to the reference cuvettes with a final volume of 1 mL. To dissolve the MBTH-quinone adducts, 2% (by volume) N,N-dimethylformamide (DMF) was added to assay medium [14].

2.4. Kinetic Data Analysis

The methods used to monitor the monophenolase and diphenolase activities use MBTH to generate an adduct, which when is oxidized and generates a compound with high extinction coefficient (Table S1). The precision of the method was checked by repeating the estimation of V_{SS}^M (rate of action on monophenol) 10 times for three levels of $[E]_0$, 5.1×10^{-4} , 2.6×10^{-3} and 5.2×10^{-3} unit/mL, the corresponding coefficients of variation being 5, 1.5 and 0.7%. The sensitivity of the method [28] was characterized by determining the detection limit ($\text{LOD}^M = 3 \times 10^{-5}$ unit/mL) and the quantification limit ($\text{LOQ}^M = 7.2 \times 10^{-5}$ unit/mL). In this work, the enzyme unit was defined as the amount that produced 1 μmol of the adduct/min when DHPPA was assayed [19]. The next step was the transformation of the monophenolase activity expressed in the activity units described above in the action of the enzyme on DHPPA, in the unit described in [10], which correspond to the amount of enzyme that consume 1 nmol of monophenol by minute or that produce 1 nmol of product by minute. Therefore, $\text{LOD}^M = 3 \times 10^{-5}$ unit/mL, which is equivalent for propionic p-hydroxyphenol to 3×10^{-2} U/mL, where U is the amount of enzyme that produces one nmol per minute. The relationships between catalytic constants [29] of the diphenol DHPAA and its monophenol PHPPA is 8.3, which means that one unit on DHPPA is equivalent to 8.3 of monophenolase units, so LOD^M , expressed in monophenolase units, is $\text{LOD}^M = 0.25$ U/mL. Table 1 shows the relative sensitivities of different substrates (L-tyrosine, tyramine and 4-hydroxyanisole) with respect to our reference substrate (PHPPA), it is noteworthy that for 4-hydroxyanisole, values of LOD^M similar to those obtained with fluorescent methods, would be expected [18,20,21]. Thus, the sensitivity is related to the nucleophilic potency of the oxygen of the hydroxyl group attached at C-4, related to the value of the chemical shift (δ_4) [21].

The initial rates were obtained by linear regression fitting portions of each experimental recording. These values were represented and fitted to the Michaelis–Menten equation using the Sigma Plot 9.0 program for Windows [30], providing the corresponding maximum rate (V_{\max}) and the Michaelis constant (K_M) in each case.

2.5. Simulation Assays

The simulation through numerical integration of the differential equations that define the tyrosinase mechanism, helps to understand the kinetic behaviour of the enzyme and this help to compare different assay methods, both the fluorometric [10–12], and the spectrophotometric published by our group [14,18–21,24,25]. The simulated progress curves were obtained by numerical solution of the nonlinear set of differential equations corresponding to each kinetic scheme (see Supplementary Material), using arbitrary, but reasonable, sets of rate constants and initial concentration values and setting in the data entered, according to the different cases. The simulation assays showed the kinetic behaviour of the ligand and enzymatic species involved in the reaction mechanisms described here for tyrosinase. The numerical integration used a fourth order Runge–Kutta method

and the predictor corrector Adams–Moulton algorithm [31], implemented on a PC-compatible computer programme (WES) [32].

3. Results

The enzyme under study, tyrosinase, shows two activities: monophenolase and diphenolase.

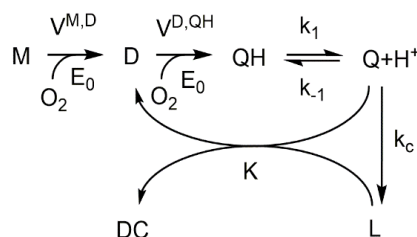
3.1. Kinetic Analysis. Diphenolase Activity

The mechanism of diphenolase activity on D has been shown in (Scheme 2), and in its structural form in Scheme S1. In the steady state, it is shown by means of the deduction of the corresponding analytical expression. Rate of product formation ($V_{SS}^{D,DC}$) and substrate disappearance V_{SS}^{D,O_2} , $V_{SS}^{D,D}$ are the same (see Supplementary Material).

3.2. Kinetic Analysis. Monophenolase Activity

In this case, tyrosinase acts on M (enzymatic stage), generating D, this can be oxidized to o-Q (enzymatic stage), or it is released into the medium, subsequently, the o-Q evolves towards DC, regenerating D in the medium (enzymatic-enzymatic-chemical mechanism EzEzC) [16].

The detailed structural mechanism is shown in the Supplementary Material Scheme S2, in a simpler way it is described in Scheme 1 and we can express it as in Scheme 3.



Scheme 3. Monophenolase and diphenolase activities of tyrosinase. M is monophenol, D is o-diphenol, QH is o-dopaquinone protonated, Q is o-dopaquinone deprotonated, L is leucodopachrome and DC is dopachrome. $V^{M,D}$ is the rate for the D formation from M. This rate is in the steady-state as $V_{SS}^{M,D}$. $V^{D,QH}$ is the rate for the QH formation from D. This rate in the steady-state is $V_{SS}^{D,QH}$.

This scheme shows the enzymatic stage of action of tyrosinase on M, its transformation into D, its oxidation to o-QH and the evolution of o-QH.

In the steady state, the quantity of matter entering the system of reactions will be shared between product and the sum the intermediates.

Considering that the rate of consumption of M in the steady-state is $V_{SS}^{M,M}$ and that the rate of accumulation of DC in the steady state is $V_{SS}^{M,DC}$, it can be established if the system has reached steady state:

$$V_{SS}^{M,M} = V_{SS}^{M,DC} \quad (1)$$

Moreover, the following material balance is fulfilled:

$$V_{SS}^{M,M}t = V_{SS}^{M,DC}t = [DC] + [D]_{SS} + [QH]_{SS} \quad (2)$$

That is, the matter that enters the system is distributed among the different intermediaries and the final product. The intermediaries are D and o-QH, which reach steady-state with the levels $[D]_{SS}$ and $[QH]_{SS}$. An analytical expression can be established for the rate of M consumption and for the accumulation of DC.

Applying the steady-state approximation for EoxD and EoxM in Scheme 1, and taking into account the relationship of the velocities between the catalytic stages, as discussed above [8], it is fulfilled:

$$[DC] = V_{SS}^{M,DC} \left(t - \frac{R[M]_0}{2k_{app}V_{SS}^{M,DC}} - \frac{1}{2k_{app}} \right) \quad (3)$$

with

$$\frac{[D]_{SS}}{[M]_{SS}} = \frac{k_5k_4(k_{-6}+k_7)}{2k_7k_6(k_{-4}+k_5)} = R \quad (4)$$

DC accumulates in the medium according to the equation of a line Equation (3), the slope of this line is the steady-state rate $V_{SS}^{M,DC} = V_{SS}^{M,M}$, the cut with the axis of time at a point $t = \tau$, in which $[DC] = 0$, resulting in

$$\tau = \frac{R[M]_0}{V_{SS}^{M,DC}} + \frac{1}{2k_{app}} \quad (5)$$

After the lag period described by Equation (5) has finished, which is when the fulfilment of Equation (4) occurs, the rate has the analytical expression indicated by Equation (S9). Therefore, it is not necessary to measure substrate disappearance to obtain a correct measure of enzyme activity.

3.3. Simulation under Different Experimental Approximations of the Monophenolase Activity Mechanism

The systems of differential equations for monophenolase activity are shown in the Supporting Information. Although the objective of this work is the measurement of tyrosinase monophenolase activity, it is convenient to consider the two activities of the enzyme (for the diphenolase activity, see Supplementary Material).

Monophenolase Activity

The monophenolase activity of tyrosinase consists of the hydroxylation of monophenols to *o*-diphenols and the oxidation of these to *o*-quinones. There are several types of monophenols [24], and depending on the chemical evolution of the *o*-quinones generated, a certain kinetic behaviour of tyrosinase is obtained. In this work, we will focus on monophenols such as L-tyrosine, tyramine, 4-hydroxyphenylpropionic, 4-hydroxyanisole whose *o*-quinones either evolve generating D in the medium (L-tyrosine, tyramine) or they can be attacked by a nucleophilic reagent as MBTH and achieve the same objective of accumulation of D in the medium and in this way carry out the passage from Em to Eox, the first inactive on monophenols while the second is.

Through different simulation conditions, it is shown that in the steady state, the rate of consumption of M (L-tyrosine) ($V_{SS}^{M,M}$) is equal to the rate of formation of DC ($V_{SS}^{M,DC}$), or in general a colored derivative that is generated by a nucleophilic attack by a reagent such as MBTH, followed by an oxidation reduction reaction, A-ox ($V_{SS}^{M,A-ox}$), in addition, V_{SS}^{M,O_2} is related to the other two rates by $V_{SS}^{M,O_2} = 1.5V_{SS}^{M,M} = 1.5V_{SS}^{M,DC} = 1.5V_{SS}^{M,A-ox}$ in such way that the correct measurement of monophenolase activity can be done in theory by measuring the consumption of M or O_2 , or the formation of DC or A-ox. The simulation of the kinetic behaviour of the mechanism of monophenolase activity under different approaches helps us to confirm and establish with adequate experimental design.

(1) Action of tyrosinase on L-tyrosine. No *o*-diphenol (L-dopa) is added to the medium.

The numerical integration of the set of differential equations that describe the mechanism (Scheme 1) (see Supplementary Material), was carried out obtaining the following results.

(a) Accumulation of dopachrome, *o*-dopaquinone and *o*-diphenol in the medium. In Figure 1A, the DC, *o*-Q and D accumulation curves are shown. At short times, it is shown in Figure S1A.

- (b) Rates of monophenol consumption ($V^{M,M}$), oxygen consumption (V^{M,O_2}) and dopachrome formation ($V^{M,DC}$). In Figure 1B, the rates of consumption of M and O_2 and the formation of DC are shown. Figure S1B shows the curves of the velocities obtained in a short time.
- (c) Evolution over time of the different enzyme forms: Eox, Em and EmM. Figure S1C shows the evolution of these enzymatic forms with time.
- (d) Variation of the concentration and rate of accumulation of o-diphenol with time. Figure S1D shows the accumulation of D over time, until it reaches a steady-state with a constant level. Thus, Figure S1E shows the accumulation of D at short times, continuously increasing towards the steady state. Figure S1F shows the rate of D accumulation, it increases at the beginning of the reaction (step of Eox \rightarrow Em), to decrease at longer times, until a value of the rate is equal to 0, in the steady state. At short times, in Figure S1G you can see a burst in the rate of D accumulation, followed by a lag and then another burst to decrease going towards the steady state.

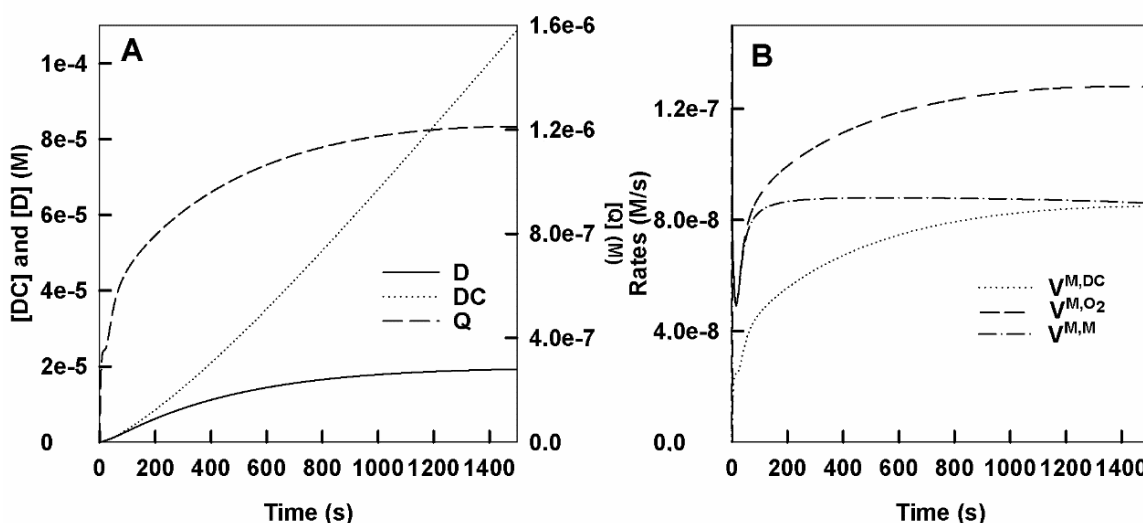


Figure 1. Simulated curves of tyrosinase action on L-tyrosine, o-diphenol is not added to the medium. (A) Accumulation curves of DC, o-Q and D in the reaction medium. (B) Evolution of O_2 consumption rates (V^{M,O_2}), monophenol ($V^{M,M}$) and DC formation ($V^{M,DC}$). Conditions: $[E]_0 = 50 \times 10^{-9} \text{ M}$, $[E_{ox}]_0 = 0.3x[E]_0$, $[E_m]_0 = 0.7 \times [E]_0$; $[M]_0 = 2 \times 10^{-3} \text{ M}$, $[O_2]_0 = 0.26 \times 10^{-3} \text{ M}$; $[D]_0 = 0$ and the rate constants were: $k_1 = 5 \times 10^5 \text{ M}^{-1}\text{s}^{-1}$, $k_{-1} = 17 \text{ s}^{-1}$, $k_2 = 3.82 \times 10^6 \text{ M}^{-1}\text{s}^{-1}$, $k_{-2} = 10 \text{ s}^{-1}$, $k_3 = 1074 \text{ s}^{-1}$, $k_4 = 3.16 \times 10^4 \text{ M}^{-1}\text{s}^{-1}$, $k_{-4} = 10 \text{ s}^{-1}$, $k_5 = 7.5 \text{ s}^{-1}$, $k_6 = 7 \times 10^5 \text{ M}^{-1}\text{s}^{-1}$, $k_{-6} = 10 \text{ s}^{-1}$, $k_7 = 107.3 \text{ s}^{-1}$, $k_8 = 2.3 \times 10^7 \text{ M}^{-1}\text{s}^{-1}$, $k_{-8} = 1.03 \times 10^3 \text{ s}^{-1}$, $k_{10} = 0.14 \text{ s}^{-1}$.

- (2) Action of tyrosinase on L-tyrosine adding enough o-diphenol to the medium to reach the steady-state.
- (a) Accumulation of dopachrome, o-dopaquinone and o-diphenol in the medium. The curves shown in Figure 2A show the concentrations of o-Q, D and DC over time. In Figure S2A the short-time curves are shown, a small lag can be seen in the accumulation of DC and a burst in the accumulation of o-Q.
- (b) Rates of consumption of monophenol and oxygen and rate of dopachrome formation. In Figure 2B the consumption rates of O_2 , M and the rate of DC formation are shown and in Figure S2B, we show the same parameters but at short times.
- (c) Evolution over time of the different enzyme species. Figure S2C shows the evolution of the different species enzymes.
- (d) Variation of the concentration and rate of accumulation of o-diphenol with time. It is shown in the Figure S2D.

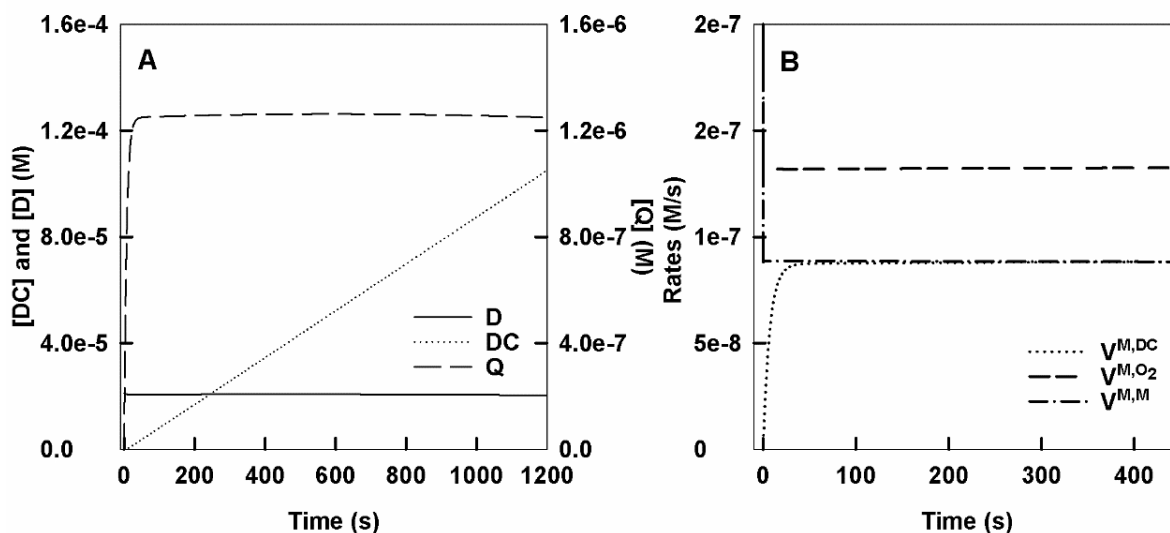


Figure 2. Simulated curves of the action of tyrosinase on L-tyrosine adding to the reaction medium the necessary concentration of *o*-diphenol to reach steady-state. (A) Accumulation of DC, *o*-Q and D in the medium. (B) Relationship between the rates of M consumption ($V^{M,M}$), O_2 (V^{M,O_2}) and DC formation rate ($V^{M,DC}$). The experimental conditions are the same than in Figure 1, but in this case, $[D]_0 = 2.114 \times 10^{-5}$ M.

- (3) Action of tyrosinase on L-tyrosine, adding before starting the reaction an amount of *o*-diphenol, less than that necessary to reach steady state.
 - (a) Accumulation of dopachrome, *o*-dopaquinone and *o*-diphenol in the reaction medium. Figures S3A and S6B show the accumulation of DC, *o*-Q and D. In Figure S6B, you can see the detail in short time.
 - (b) Rates of consumption of monophenol and oxygen and rate of dopachrome formation. Rates are shown in Figure S3C and at short times in Figure S3D.
 - (c) Evolution over time of the different enzyme species. A graph like that of Figures S1C and S2C is obtained. The enzyme accumulates mostly as EmM.
 - (d) Variation of *o*-diphenol concentration with time. Figure S3E shows D variation.
- (4) Action of tyrosinase on L-tyrosine, adding before starting the reaction an amount of *o*-diphenol greater than that necessary to reach steady state. The speeds at long and short times are shown in Figure S4A,B, respectively.
- (5) Action of tyrosinase on L-tyrosine in the presence of a nucleophile N.
 - (a) The curves obtained in this section are like those obtained in Section 1, the only difference is that the *o*-Q evolution constant becomes much larger as the nucleophilic attack of N is more powerful than the cyclization of *o*-Q. In the action of tyrosinase on M in the presence of N, Figure 3A shows the accumulation of *o*-Q, D and A-ox with time. It shows a burst for *o*-Q and D and a very small lag for A-ox.
 - (b) Rates of consumption of M, O_2 and the formation of A-ox. In Figure 3B, the curves obtained for the rate of consumption of M and O_2 and that of accumulation of A-ox are shown.
 - (c) Enzyme species evolve in a similar way to the other cases and the enzyme accumulates fundamentally as EmM, Figure 3C.
 - (d) Variation of the concentration and rate of accumulation of D over time. It is shown in Figure 3D.

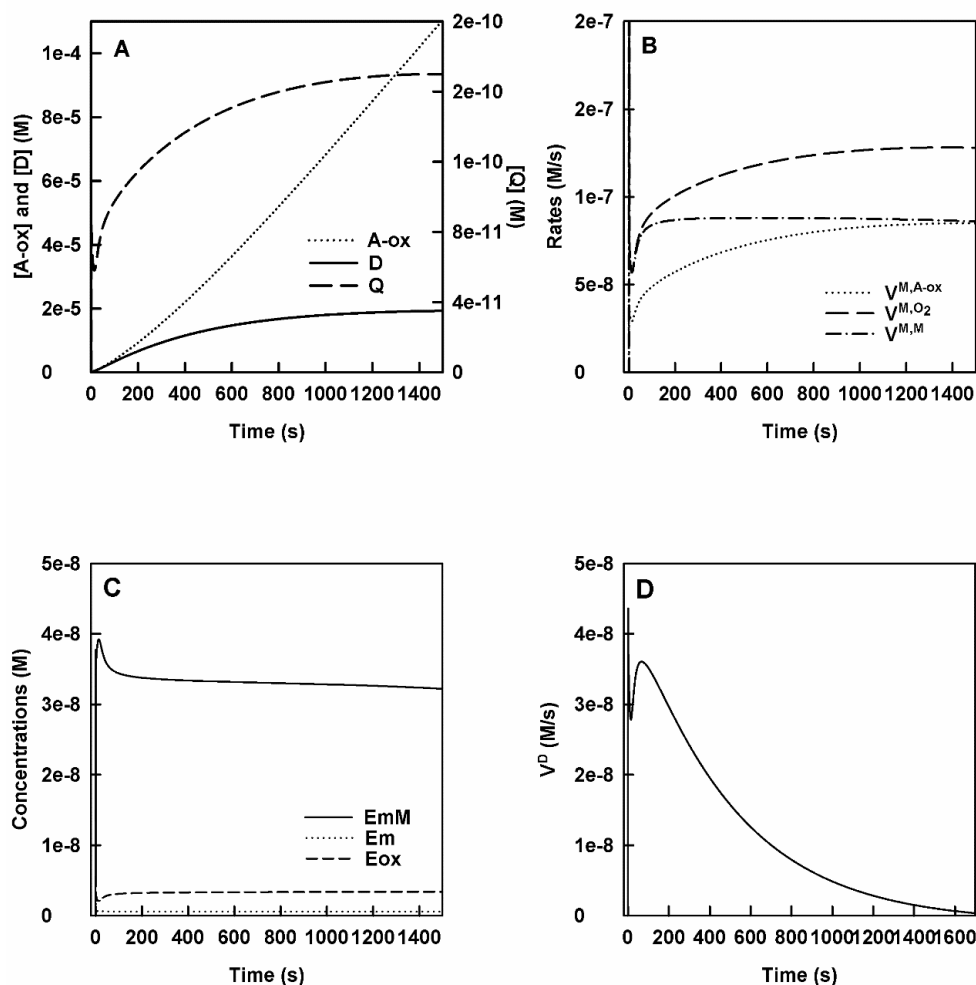


Figure 3. Simulated curves of tyrosinase action on L-tyrosine in the presence of a nucleophile (N). (A) Accumulation of o-Q, D and A-ox over time. (B) Relationship between consumption rates of M ($V^{M,M}$), O_2 (V^{M,O_2}) and DC formation ($V^{M,DC}$). (C) Evolution of enzyme species over time. (D) Variation of the rate of D accumulation with time. The experimental conditions are the same than in Figure 1, but in this case, $[D]_0 = 0$, $k_{11} = 5 \times 10^5 \text{ M}^{-1} \text{ s}^{-1}$ y $[\text{MBTH}] = 2 \times 10^{-3} \text{ M}$.

4. Discussion

The analysis of the mechanism of monophenolase activity of tyrosinase under different initial conditions was carried out by simulation by numerical integration of each particular case.

4.1. Action of Tyrosinase on L-Tyrosine. No o-Diphenol (L-Dopa) Is Added to the Medium

Figure 1A shows that DC accumulates with a lag (τ), while the o-Q and D accumulate with a burst. When the system reaches the steady state ($t_{99} \cong 4.6 \tau$), the slope of the DC accumulation line corresponds to the steady-state rate ($V_{SS}^{M,DC}$). Figure S1A shows the accumulation of these species at short times increasing to reach the steady state.

Rates of consumption of O_2 and M are higher at the beginning of the reaction (Figure 1B), because in the native enzyme there is approximately 30% in the Eox form and this species consumes M and O_2 . The rate of DC accumulation increases with time, in the steady state: $V_{SS}^{M,M} = V_{SS}^{M,DC}$. However, the rate of O_2 consumption (V_{SS}^{M,O_2}) in the steady-state is 1.5 times $V_{SS}^{M,M}$ or $V_{SS}^{M,DC}$. The explanation for these curves is that at steady-state the enzyme molecule performs two cycles in the hydroxylase pathway (see Scheme 1) consuming 2M and 2 O_2 and one cycle in the oxidase pathway consuming 2D and 1 O_2 . The

two molecules of M originate 2Q and those of 2D other 2Q, which regenerate between 4Q, 2 of D and 2 of DC, for this reason, as shown in Figure 1B ($V_{SS}^{M,O_2} = 1.5V_{SS}^{M,M} = 1.5V_{SS}^{M,DC}$) [8,33]. Note a less abrupt drop in the rate of consumption of M and O₂ and an increase in the rate of formation of DC (Figure S1B), because at $t = 0$ there is approximately a 30% enzyme in Eox form that reacts rapidly by consuming M and O₂ and practically all the enzyme is found as EmM, that is, inactive, as will be seen later. The appearance of lag in the rate of DC formation can be observed.

The assays shown in Figure S1C are explained at the beginning of the reaction and the enzyme is mostly found as: 70% Em and 30% Eox, when adding M, the Eox form reacts rapidly generating o-Q and passing to Em (inactive on M) (Figure S1C). The Em form reacts with M to form the inactive complex EmM, the simulated curves show how the enzyme is mostly as EmM.

When tyrosinase begins its action on M, a lag period, τ , is originated in the accumulation of DC and at the same time the system accumulates D in the medium, the lag period is related to the time in which the system reaches the stationary phase and that assumes a constant value of D. D variation with time is shown in Figure S1D–G.

4.2. Action of Tyrosinase on L-Tyrosine Adding Enough o-Diphenol to the Medium to Reach the Steady State

This situation assumes that the system does not have to accumulate D in the medium and thus the steady state is reached earlier at $t \rightarrow 0$. D quantity is given by Equation (S11).

Note that now the DC accumulates without a delay period, the steady state is reached at $t \rightarrow 0$ (Figure 2A and Figure S2A). The steady state is reached quickly, M and O₂ arrive before DC, due to the evolution of the o-Q. In the steady state, $V_{SS}^{M,M} = V_{SS}^{M,DC}$ and $V_{SS}^{M,O_2} = 1.5V_{SS}^{M,M} = 1.5V_{SS}^{M,DC}$ (Figure 2B). A similar effect is seen in short-time records (Figure S2B). The concentration of the enzymatic species EmM, Em and Eox shows that most of the enzyme is in the complex of dead-path EmM (Figure S2C). The variation of D with time is very small because practically the system is in the steady-state (Figure S2D).

4.3. Action of Tyrosinase on L-Tyrosine, Adding before Starting the Reaction an Amount of o-Diphenol, Less than that Necessary to Reach the Steady State

The system will accumulate the D resulting from the difference between the D of the steady state and the added at the beginning. Thus, the lag in DC accumulation will be less than in case (4.1) and the steady-state will be reached earlier.

DC is accumulated with a lag that is lower than in the absence of added D and the o-Q and D accumulate with a burst, until reaching the steady state (Figure S3A). The burst of the o-Q moves in time because it must be generated and evolve towards DC (Figure S6B). In long time, the system reaches the steady state and then it is satisfied that $V_{SS}^{M,M} = V_{SS}^{M,DC}$ and with respect to O₂: $V_{SS}^{M,O_2} = 1.5V_{SS}^{M,M} = 1.5V_{SS}^{M,DC}$ (Figure S3C). At short times, no rate has reached the steady state and appreciable differences are seen between $V^{M,M}$ and $V^{M,DC}$ (Figure S3D). D concentration reaches the steady state before τ_{an} in the case 4.1 (Figure S3E).

These conditions should be of choice to work experimentally with monophenolase activity. In order to reduce lag, a small amount of o-diphenol should be added before starting the reaction. When the substrate concentration is varied, the o-diphenol concentration must be varied such that the ratio is constant.

4.4. Action of Tyrosinase on L-Tyrosine, Adding before Starting the Reaction an Amount of o-Diphenol Greater than that Necessary to Reach Steady-State

The system advances towards the steady state decreasing the concentration of D until reaching that of the steady-state, and then fulfilling the relationships between the rate: $V_{SS}^{M,M} = V_{SS}^{M,DC}$ and $V_{SS}^{M,O_2} = 1.5V_{SS}^{M,M} = 1.5V_{SS}^{M,DC}$ (Figure S4A), and at short times (Figure S4B). The rest of the parameters follow a behaviour like cases 4.1–4.3.

4.5. Action of Tyrosinase on L-Tyrosine in the Presence of a Nucleophile N

The measurement of the activity of the enzyme on M in the presence of MBTH has led to the design of very sensitive measurement methods of the monophenolase and diphenolase activities, and this has allowed to characterize especially the monophenolase activity in fruits and vegetables that was described that many polyphenol oxidases had no monophenolase activity [13,14,25].

In this case, the arrival to the steady state is advanced and again it is true that the rate of formation of the coloured adduct, the oxidized form of the adduct (A-ox) is related to the rate of consumption of M and O₂ by the relationship $V_{SS}^{M,M} = V_{SS}^{M,A-ox}$ and $V_{SS}^{M,O_2} = 1.5V_{SS}^{M,M} = 1.5V_{SS}^{M,A-ox}$ (Figure 3A,B). Evolution of D and the enzymatic species is shown in Figure 3C,D. Figure 3D shows the variation in the rate of accumulation of D over time. Initially there is a burst in the rate of formation of D, and then at long times decrease the rate until it vanishes in the steady state.

In all the cases described, it is shown that the measurement of monophenolase activity in tyrosinase acting on M can be done either by measuring the disappearance of M or O₂, or by measuring the formation of DC or A-ox, when the system is in steady-state.

4.6. Comparison of the Measurement of Monophenolase Activity of Tyrosinase Obtained by Continuous Spectrophotometric Methods That Use MBTH as a Nucleophilic Reagent with Continuous Fluorometric Methods that Measure the Disappearance of L-Tyrosine through Quenching L-Dopa Fluorescence by Borate

Fluorometric methods are normally more sensitive than spectrophotometric ones, but when it comes to measuring the activity of an enzyme, in this case tyrosinase, one must consider several variables such as pH, substrate concentration and measurement interferences. Proposed continuous spectrophotometric methods to measure tyrosinase monophenolase and diphenolase activity may have some advantages over spectrofluorimetric:

1. In spectrophotometric methods, product formation is measured in an area where substrates do not interfere and it is more sensitive to measure product formation than substrate disappearance [24]. In the proposed spectrofluorimetric methods, the disappearance of substrate L-tyrosine, is measured.
2. In these spectrophotometric methods, product formation is measured. Moreover, there are no phototube saturation problems. Thus, the substrate concentration can be increased because it does not interfere. In the spectrofluorimetric methods proposed, when measuring the disappearance of substrate (L-tyrosine), by increasing its concentration, linearity can be lost, whereas in reality, it occurs in such a way that in the proposed methods [10–12], which follow fluorescence, only the behavior is linear up to a concentration of 100 µM of L-tyrosine.
3. What is indicated in Section 2 is very important when characterizing the kinetics of tyrosinase monophenolase activity, as the concentration cannot be increased more than 100 µM. Indeed, if the initial velocity values are adjusted, with respect to the L-tyrosine concentration, erroneous kinetic parameters are obtained, because a true hyperbolic dependence of the $V_{SS}^{M,M}$ vs. $[M]_0$ values has not been obtained. Thus, the authors working with mushroom tyrosinase, obtain a value of $K_M = 19.51$ µM [10–12], when the value found for this enzyme is between 0.21 mM [21] and 0.31 mM [34]. At high pH values, K_M values are constants [35]. In general, this problem is encountered by most of the methods that attempt to measure the disappearance of substrate [24].
4. The lack of linearity between the fluorescence measurement and the concentration of L-tyrosine above 100 µM make kinetic studies difficult and especially in the presence of inhibitors. Thus, when zinc ion is studied as a tyrosinase inhibitor [10], data are obtained that indicate that it is a competitive inhibitor, so in order to determine the K_i value, the concentration of L-tyrosine must be increased by more than 100 µM, like

this not possible, abnormal results are obtained. On the other hand, the IC₅₀ value described is 14.64 μM and the K_i value = 67.60 μM [10], but in the case of a competitive inhibitor, it can be shown that IC₅₀ > K_i, specifically IC₅₀ = (n+1)K_i, where n is the ratio of substrate concentration to K_m [36]. This relationship between IC₅₀ and K_i is well established in the data shown in Tables 1 and 2 in [37] and in Table 1 in [38].

5. The relationship of rates of consumption of M, O₂ and formation of DC described in Table S5 of the Supporting Information [10] shows in the monophenolase activity the equality of rates of consumption of O₂ and consumption of M, however the DC formation rate is not comparative; this aspect confirms the validity of the measure measuring O₂ consumption that was described previously [9]. The stoichiometry of diphenolase activity predicts that the O₂ consumption rate should be equal to the formation rate of DC, as demonstrated previously [8]. In the first column (substrate + TYR, diphenolase) of Table S5 [10], the described relationship is not fulfilled; however, it is achieved in the last column of Table S5 [10] (Substrate + TYR + hydroxylamine).
6. A feature of spectrofluorimetric methods that measure the disappearance of M could be the one that reaches the steady-state earlier, as D does not accumulate in the medium under these test conditions. This aspect can be corrected in spectrophotometric methods by adding a small concentration of D to the medium ($[D]_{\text{added}} < [D]_{\text{SS}}$) (Figure 2A) to decrease the lag period. Furthermore, it is thus achieved that the substrate concentration varies very little and therefore measured initial rates will correspond to the steady-state rate.

5. Conclusions

The numerical integrations carried out in this work revealed that to correctly measure the monophenolase activity of tyrosinase, we can measure the disappearance of substrate M (fluorimetrically) or O₂ (oximetrically) in a medium at pH = 8 with borate, or either by spectrophotometrically measuring the appearance of DC or A-ox product, always considering that the system is in a steady state. Thus, if the system is in a steady state, the rate of substrate disappearance is equal to the rate of product formation, if they are initial steady-state rates. In this sense, the proposed spectrophotometric methods measuring the formation of oxidized adducts between *o*-quinones and MBTH, can be considered more suitable to follow the monophenolase activity of tyrosinase, taking into account the following aspects: (a) There are no interferences from substrates. (b) There is no interference from the enzyme. (c) The substrate concentration can be varied. (d) It is possible to work at different pH values and thus the enzymes of fruits and vegetables can be characterized. (e) The addition of a small amount of *o*-diphenol is convenient to decrease the delay period (τ), but not to suppress it.

Supplementary Materials: The following are available online at www.mdpi.com/article/10.3390/biom11091269/s1. Scheme S1. Diphenolase pathway. Structural mechanism. Scheme S2. Monophenolase pathway. Structural mechanism. Scheme S3. Diphenolase activity of tyrosinase in which the product, *o*-quinone, is attacked by a nucleophilic reagent such as MBTH. Figure S1. Simulated curves of the action of tyrosinase on Ltyrosine, *o*-diphenol is not added to the medium. Figure S2. Simulated curves of the action of tyrosinase on Ltyrosine, the *o*-diphenol necessary to reach the steady-state is added to the medium. Figure S3. Simulated curves of the action of tyrosinase on Ltyrosine, a concentration of *o*-diphenol is added to the medium lower than that necessary to reach steady-state. Figure S4. Simulated curves of the action of tyrosinase on Ltyrosine, a concentration of *o*-diphenol is added to the medium greater than that necessary to reach the steady-state. Table S1. Kinetic characteristics of the action of tyrosinase on different monophenols and spectroscopic properties of the oxidized adducts formed with MBTH.

Author Contributions: J.L.M.-M., P.G.-M., F.G.-M. and F.G.-C. designed and directed the project and experiments. P.G.-M. performed all simulations experiments. J.A.O. and F.G.-C. analysed and interpreted the results. P.G.-M., F.G.-M., J.A.O., J.N.R.-L., P.A.G.-R., F.G.-C. and J.L.M.-M., drafted

the manuscript and figures, provided commentary and edits to the manuscript and figures and prepared the final version of the article. All authors contributed to the article and approved the submitted version.

Funding: This research received no external funding.

Institutional Review Board Statement: Not applicable.

Informed Consent Statement: Not applicable.

Acknowledgments: Fundacion Séneca, the Region de Murcia (FS-RM) (20809/PI/18). J.M.-M. receive funding from internal grants in Northumbria University.

Conflicts of Interest: The authors declare no conflict of interest.

References

1. Bubacco, L.; Salgado, J.; Tepper, A.W.J.W.; Vijgenboom, E.; Canters, G.W. 1H NMR spectroscopy of the binuclear Cu(II) active site of *Streptomyces antibioticus* tyrosinase. *FEBS Lett.* **1999**, *442*, 215–220, [https://doi.org/10.1016/S0014-5793\(98\)01662-7](https://doi.org/10.1016/S0014-5793(98)01662-7).
2. Kanteev, M.; Goldfeder, M.; Fishman, A. Structure–function correlations in tyrosinases. *Protein Sci.* **2015**, *24*, 1360–1369, <https://doi.org/10.1002/pro.2734>.
3. Solomon, E.I.; Heppner, D.E.; Johnston, E.M.; Ginsbach, J.W.; Cirera, J.; Qayyum, M.; Kieber-Emmons, M.T.; Kjaergaard, C.H.; Hadt, R.G.; Tian, L. Copper Active Sites in Biology. *Chem. Rev.* **2014**, *114*, 3659–3853, [doi:10.1021/cr400327t](https://doi.org/10.1021/cr400327t).
4. Rodríguez-López, J.N.; Ros, J.R.; Varón, R.; García-Cánovas, F. Oxygen Michaelis constants for tyrosinase. *Biochem. J.* **1993**, *293*, 859–866, [doi:10.1042/bj2930859](https://doi.org/10.1042/bj2930859).
5. Fenoll, L.G.; Rodríguez-López, J.N.; García-Sevilla, F.; García-Ruiz, P.A.; Varón, R.; García-Cánovas, F.; Tudela, J. Analysis and interpretation of the action mechanism of mushroom tyrosinase on monophenols and diphenols generating highly unstable o-quinones. *Biochim. Biophys. Acta Protein Struct. Mol. Enzymol.* **2001**, *1548*, 1–22, [https://doi.org/10.1016/S0167-4838\(01\)00207-2](https://doi.org/10.1016/S0167-4838(01)00207-2).
6. Kampatsikas, I.; Rompel, A. Similar but Still Different: Which Amino Acid Residues Are Responsible for Varying Activities in Type-III Copper Enzymes? *ChemBioChem* **2020**, *21*, 1–16, <https://doi.org/10.1002/cbic.202000647>.
7. Goldfeder, M.; Kanteev, M.; Isaschar-Ovdat, S.; Adir, N.; Fishman, A. Determination of tyrosinase substrate-binding modes reveals mechanistic differences between type-3 copper proteins. *Nat. Commun.* **2014**, *5*, 4505, [doi:10.1038/ncomms5505](https://doi.org/10.1038/ncomms5505).
8. Rodríguez-López, J.N.; Tudela, J.; Varón, R.; García-Carmona, F.; García-Cánovas, F. Analysis of a kinetic model for melanin biosynthesis pathway. *J. Biol. Chem.* **1992**, *267*, 3801–3810, [https://doi.org/10.1016/S0021-9258\(19\)50597-X](https://doi.org/10.1016/S0021-9258(19)50597-X).
9. Yamazaki, S.; Itoh, S. Kinetic Evaluation of Phenolase Activity of Tyrosinase Using Simplified Catalytic Reaction System. *J. Am. Chem. Soc.* **2003**, *125*, 13034–13035, [doi:10.1021/ja036425d](https://doi.org/10.1021/ja036425d).
10. Guo, N.; You, X.; Wu, Y.; Du, D.; Zhang, L.; Shang, Q.; Liu, W. Continuous Fluorometric Method for Determining the Monophenolase Activity of Tyrosinase on L-Tyrosine, through Quenching L-DOPA Fluorescence by Borate. *Anal. Chem.* **2020**, *92*, 5780–5786, [doi:10.1021/acs.analchem.9b05127](https://doi.org/10.1021/acs.analchem.9b05127).
11. Du, D.; Guo, N.; Zhang, L.; Wu, Y.; Shang, Q.; Liu, W. Real-time fluorometric monitoring of monophenolase activity using a matrix-matched calibration curve. *Anal. Bioanal. Chem.* **2021**, *413*, 635–647, [doi:10.1007/s00216-020-03034-4](https://doi.org/10.1007/s00216-020-03034-4).
12. Zhang, L.; Shang, Q.; Chen, C.; Tang, W.; Xu, Y.; Liu, W. Synchronous fluorometric method for continuous assay of monophenolase activity. *Spectrochim. Acta Part. A Mol. Biomol. Spectrosc.* **2021**, *252*, 119486, <https://doi.org/10.1016/j.saa.2021.119486>.
13. Winder, A.J.; Harris, H. New assays for the tyrosine hydroxylase and dopa oxidase activities of tyrosinase. *Eur. J. Biochem.* **1991**, *198*, 317–326, <https://doi.org/10.1111/j.1432-1033.1991.tb16018.x>.
14. Rodríguez-lopez, J.N.; Escribano, J.; Garcia-cánovas, F. A Continuous Spectrophotometric Method for the Determination of Monophenolase Activity of Tyrosinase Using 3-Methyl-2-benzothiazolinone Hydrazone. *Anal. Biochem.* **1994**, *216*, 205–212, <https://doi.org/10.1006/abio.1994.1026>.
15. Pomerantz, S.H. The Tyrosine Hydroxylase Activity of Mammalian Tyrosinase. *J. Biol. Chem.* **1966**, *241*, 161–168, [https://doi.org/10.1016/S0021-9258\(18\)96973-5](https://doi.org/10.1016/S0021-9258(18)96973-5).
16. Cabanes, J.; García-Cánovas, F.; Lozano, J.; García-Carmona, F. A kinetic study of the melanization pathway between L-tyrosine and dopachrome. *Biochim. Biophys. Acta—Gen. Subj.* **1987**, *923*, 187–195, [https://doi.org/10.1016/0304-4165\(87\)90003-1](https://doi.org/10.1016/0304-4165(87)90003-1).
17. Rzepecki, L.M.; Waite, J.H. A chromogenic assay for catecholoxidases based on the addition of l-proline to quinones. *Anal. Biochem.* **1989**, *179*, 375–381, [https://doi.org/10.1016/0003-2697\(89\)90148-6](https://doi.org/10.1016/0003-2697(89)90148-6).
18. Espín, J.C.; Tudela, J.; García-Cánovas, F. 4-Hydroxyanisole: The Most Suitable Monophenolic Substrate for Determining Spectrophotometrically the Monophenolase Activity of Polyphenol Oxidase from Fruits and Vegetables. *Anal. Biochem.* **1998**, *259*, 118–126, [doi:https://doi.org/10.1006/abio.1998.2598](https://doi.org/10.1006/abio.1998.2598).
19. Espín, J.C.; Morales, M.; García-Ruiz, P.A.; Tudela, J.; García-Cánovas, F. Improvement of a Continuous Spectrophotometric Method for Determining the Monophenolase and Diphenolase Activities of Mushroom Polyphenol Oxidase. *J. Agric. Food Chem.* **1997**, *45*, 1084–1090, [doi:10.1021/jf960428a](https://doi.org/10.1021/jf960428a).
20. Espín, J.C.; Varón, R.; Tudela, J.; García-Cánovas, F. Kinetic study of the oxidation of 4-hydroxyanisole catalyzed by tyrosinase. *IUBMB Life* **1997**, *41*, 1265–1276, <https://doi.org/10.1080/15216549700202361>.

21. Espín, J.C.; Varón, R.; Fenoll, L.G.; Gilabert, M.A.; García-Ruiz, P.A.; Tudela, J.; García-Cánovas, F. Kinetic characterization of the substrate specificity and mechanism of mushroom tyrosinase. *Eur. J. Biochem.* **2000**, *267*, 1270–1279, <https://doi.org/10.1046/j.1432-1327.2000.01013.x>.
22. Young, K.L.II.; Kassouf, C.; Dolinska, M.B.; Anderson, D.E.; Sergeev, YV. Human Tyrosinase: Temperature-Dependent Kinetics of Oxidase Activity. *Int. J. Mol. Sci.* **2020**, *21*, 895–906, doi:10.3390/ijms21030895.
23. Ertan, H.; Siddiqui, K.S.; Muenchhoff, J.; Charlton, T.; Cavicchioli, R. Kinetic and thermodynamic characterization of the functional properties of a hybrid versatile peroxidase using isothermal titration calorimetry: Insight into manganese peroxidase activation and lignin peroxidase inhibition. *Biochimie* **2012**, *94*, 221–231, doi:10.1016/j.biochi.2012.02.012.
24. García-Molina, F.; Muñoz, J.L.; Varón, R.; Rodríguez-López, J.N.; García-Cánovas, F.; Tudela, J. A Review on Spectrophotometric Methods for Measuring the Monophenolase and Diphenolase Activities of Tyrosinase. *J. Agric. Food Chem.* **2007**, *55*, 9739–9749, doi:10.1021/jf0712301.
25. Espin, J.C.; Morales, M.; Varon, R.; Tudela, J.; Garcia-canovas, F. A Continuous Spectrophotometric Method for Determining the Monophenolase and Diphenolase Activities of Apple Polyphenol Oxidase. *Anal. Biochem.* **1995**, *231*, 237–246, <https://doi.org/10.1006/abio.1995.1526>.
26. Rodríguez-López, J.N.; Fenoll, L.G.; García-Ruiz, P.A.; Varón, R.; Tudela, J.; Thorneley, R.N.F.; García-Cánovas, F. Stopped-Flow and Steady-State Study of the Diphenolase Activity of Mushroom Tyrosinase. *Biochemistry* **2000**, *39*, 10497–10506, doi:10.1021/bi000539+.
27. Bradford, M.M. A rapid and sensitive method for the quantitation of microgram quantities of protein utilizing the principle of protein-dye binding. *Anal. Biochem.* **1976**, *72*, 248–254, [https://doi.org/10.1016/0003-2697\(76\)90527-3](https://doi.org/10.1016/0003-2697(76)90527-3).
28. FDA. Q2B Validation of Analytical Procedures: Methodology. Available online: <https://www.fda.gov/regulatory-information/search-fda-guidance-documents/q2b-validation-analytical-procedures-methodology> (accessed on 24 August 2018).
29. Fenoll, L.G.; Rodríguez-López, J.N.; García-Molina, F.; García-Cánovas, F.; Tudela, J. Unification for the Expression of the Monophenolase and Diphenolase Activities of Tyrosinase. *IUBMB Life* **2002**, *54*, 137–141, <https://doi.org/10.1080/15216540214537>.
30. *Systat Software, Version 9.0, Sigma Plot for Windows*; Systat software: San Jose, CA, USA, 2006.
31. Gerald, C.F. *Applied Numerical Analysis*; Addison-Wesley: Reading, UK, 1978.
32. García-Sevilla, F.; Garrido-del Solo, C.; Duggleby, R.G.; García-Cánovas, F.; Peyró, R.; Varón, R. Use of a windows program for simulation of the progress curves of reactants and intermediates involved in enzyme-catalyzed reactions. *Biosystems* **2000**, *54*, 151–164, [https://doi.org/10.1016/S0303-2647\(99\)00071-4](https://doi.org/10.1016/S0303-2647(99)00071-4).
33. Sánchez-Ferrer, Á.; Neptuno Rodríguez-López, J.; García-Cánovas, F.; García-Carmona, F. Tyrosinase: A comprehensive review of its mechanism. *Biochim. Biophys. Acta Protein Struct. Mol. Enzymol.* **1995**, *1247*, 1–11, [https://doi.org/10.1016/0167-4838\(94\)00204-T](https://doi.org/10.1016/0167-4838(94)00204-T).
34. Fernandez-Julia, P.J.; Tudela-Serrano, J.; Garcia-Molina, F.; Garcia-Canovas, F.; Garcia-Jimenez, A.; Munoz-Munoz, J.L. Study of tyrosine and dopa enantiomers as tyrosinase substrates initiating l- and d-melanogenesis pathways. *Biotechnol. Appl. Biochem.* **2020**, <https://doi.org/10.1002/bab.1998>.
35. Maria-Solano, M.A.; Ortiz-Ruiz, C.V.; Muñoz-Muñoz, J.L.; Teruel-Puche, J.A.; Berna, J.; Garcia-Ruiz, P.A.; Garcia-Canovas, F. Further insight into the pH effect on the catalysis of mushroom tyrosinase. *J. Mol. Catal. B Enzym.* **2016**, *125*, 6–15, <https://doi.org/10.1016/j.molcatb.2015.12.008>.
36. Ortiz-Ruiz, C.V.; Garcia-Molina, M. del M.; Serrano, J.T.; Tomas-Martinez, V.; Garcia-Canovas, F. Discrimination between Alternative Substrates and Inhibitors of Tyrosinase. *J. Agric. Food Chem.* **2015**, *63*, 2162–2171, doi:10.1021/jf5051816.
37. Buitrago, E.; Faure, C.; Challali, L.; Bergantino, E.; Boumendjel, A.; Bubacco, L.; Carotti, M.; Hardré, R.; Maresca, M.; Philouze, C.; et al. Ditopic Chelators of Diccopper Centers for Enhanced Tyrosinases Inhibition. *Chem. Eur. J.* **2021**, *27*, 4384–4393, <https://doi.org/10.1002/chem.202004695>.
38. Kim, J.H.; Leem, H.H.; Lee, G.Y. The Guanidine Pseudoalkaloids 10-Methoxy-Leonurine and Leonurine Act as Competitive Inhibitors of Tyrosinase. *Biomolecules* **2020**, *10*, 174.