DATA NOTE



The genome sequence of the European turtle dove,

Streptopelia turtur Linnaeus 1758 [version 1; peer review:

awaiting peer review]

Jenny C. Dunn¹, Keith C. Hamer², Antony J. Morris³, Philip V. Grice⁴, Michelle Smith⁵, Craig Corton⁵, Karen Oliver⁵, Jason Skelton⁵, Emma Betteridge⁵, Jale Dolucan^{5,6}, Michael A. Quail⁵, Shane A. McCarthy^{5,7}, Marcela Uliano-Silva⁵, Kerstin Howe¹, James Torrance¹, William Chow⁵, Sarah Pelan⁵, Ying Sims⁵, Richard Challis¹, Jonathan Threlfall¹, Daniel Mead¹, Mark Blaxter¹,

¹University of Lincoln, Lincoln, LN6 7TS, UK

³Centre for Conservation Science, Royal Society for the Protection of Birds, Sandy, Bedfordshire, SG19 2DL, UK

⁴Natural England, Peterborough, PE1 1NG, UK

⁵Wellcome Sanger Institute, Hinxton, Cambridgeshire, CB10 1SA, UK

⁶Achilles Therapeutics plc, London, W6 8PW, UK

⁷Department of Genetics, University of Cambrudge, Cambridge, CB2 3EH, UK

⁸Owlstone Medical, Cambridge, CB4 0GJ, UK

First published: 27 Jul 2021, 6:191 https://doi.org/10.12688/wellcomeopenres.17060.1 Latest published: 27 Jul 2021, 6:191 https://doi.org/10.12688/wellcomeopenres.17060.1

Abstract

We present a genome assembly from an individual female *Streptopelia turtur* (the European turtle dove; Chordata; Aves; Columbidae). The genome sequence is 1.18 gigabases in span. The majority of the assembly is scaffolded into 35 chromosomal pseudomolecules, with the W and Z sex chromosomes assembled.

Keywords

Streptopelia turtur, European turtle dove, genome sequence, chromosomal



This article is included in the Tree of Life

gateway.

Open Peer Review

Reviewer Status AWAITING PEER REVIEW

Any reports and responses or comments on the article can be found at the end of the article.

²School of Biology, University of Leeds, Leeds, LS2 9JT, UK

Corresponding author: Mark Blaxter (mark.blaxter@sanger.ac.uk)

Author roles: Dunn JC: Data Curation, Formal Analysis, Investigation, Resources, Writing - Review & Editing; Hamer KC: Data Curation, Formal Analysis, Investigation, Resources, Writing - Review & Editing; Morris AJ: Data Curation, Formal Analysis, Investigation, Resources, Writing - Review & Editing; Grice PV: Data Curation, Funding Acquisition, Investigation, Resources, Writing - Review & Editing; Smith M: Formal Analysis, Investigation, Methodology, Writing - Review & Editing; Corton C: Formal Analysis, Investigation, Methodology, Writing - Review & Editing; Oliver K: Formal Analysis, Investigation, Methodology, Writing - Review & Editing; Skelton J: Formal Analysis, Investigation, Methodology, Writing - Review & Editing; Betteridge E: Formal Analysis, Investigation, Methodology, Writing - Review & Editing; Dolucan J: Formal Analysis, Investigation, Methodology, Writing - Review & Editing; Quail MA: Formal Analysis, Investigation, Methodology, Writing - Review & Editing; McCarthy SA: Formal Analysis, Investigation, Methodology, Software, Validation, Writing - Review & Editing; Uliano-Silva M: Formal Analysis, Investigation, Methodology, Software, Validation, Writing -Review & Editing; Howe K: Formal Analysis, Investigation, Methodology, Software, Supervision, Validation, Writing – Review & Editing; Torrance J: Formal Analysis, Investigation, Methodology, Software, Validation, Writing – Review & Editing; Chow W: Formal Analysis, Investigation, Methodology, Software, Validation, Writing – Review & Editing; Pelan S: Formal Analysis, Investigation, Methodology, Software, Validation, Writing - Review & Editing; Sims Y: Formal Analysis, Investigation, Methodology, Software, Validation, Writing -Review & Editing; Challis R: Formal Analysis, Methodology, Software, Validation, Visualization, Writing – Review & Editing; Threlfall J: Project Administration, Writing – Original Draft Preparation, Writing – Review & Editing; Mead D: Conceptualization, Investigation, Project Administration, Writing - Review & Editing; Blaxter M: Conceptualization, Data Curation, Funding Acquisition, Supervision, Writina - Review & Editina

Competing interests: Jonathan Threlfall was employed by F1000 Research Limited until January 2021.

Grant information: This work was supported by Wellcome through core funding to the Wellcome Sanger Institute (206194) and the Darwin Tree of Life Discretionary Award (218328). SMcC is supported by Wellcome (207492). Sample collection was jointly funded by the Royal Society for the Protection of Birds and Natural England under the Action for Birds in England (AfBiE) scheme. *The funders had no role in study design, data collection and analysis, decision to publish, or preparation of the manuscript.*

Copyright: © 2021 Dunn JC *et al.* This is an open access article distributed under the terms of the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

How to cite this article: Dunn JC, Hamer KC, Morris AJ *et al.* The genome sequence of the European turtle dove, *Streptopelia turtur* Linnaeus 1758 [version 1; peer review: awaiting peer review] Wellcome Open Research 2021, 6:191 https://doi.org/10.12688/wellcomeopenres.17060.1

First published: 27 Jul 2021, 6:191 https://doi.org/10.12688/wellcomeopenres.17060.1

Species taxonomy

Eukaryota; Metazoa; Chordata; Aves; Columbiformes; Columbidae; Streptopelia; *Streptopelia turtur* Linnaeus 1758 (NCBI:txid177155).

Introduction

The European turtle dove, Streptopelia turtur, breeds throughout Europe, Central Asia, the Middle East and North Africa, overwintering in north Sub-Saharan Africa. Populations in the Atlantic archipelago of Britain and Ireland are primarily located in southern and eastern England. S. turtur populations are in rapid decline in the UK, having fallen by 98% between 1970 and 2018, making them critically endangered; they are also vulnerable to global extinction (Burns et al., 2020). Several causes have been put forward for this collapse in population. Changes in farming practices and agricultural intensification in the UK have reduced the availability of wild plant seeds, increasing the reliance of S. turtur on anthropogenic seed sources (Browne & Aebischer, 2003); a negative association between nestling condition and consumption of seeds from anthropogenic sources has been reported, although this association was positive for adult birds (Dunn et al., 2018). Additionally, infection with the protozoan parasite Trichomonas gallinae has been identified as a cause of death in adults and nestlings (Stockdale et al., 2015). The length of breeding seasons and the number of breeding attempts of S. turtur have markedly reduced, meaning that fewer young are hatched each year (Browne & Aebischer, 2004). Large populations of migrating birds are also hunted in Mediterranean countries, such as France, Spain and Morocco, compounding this decline in numbers. The genome sequence described here

will be of utility to researchers assessing the vulnerability of *S. turtur* to parasitic infections, and to those interested in population genomics and supporting the numbers of this declining species.

Genome sequence report

The genome was sequenced from a blood sample collected from a single live female S. turtur during routine population health checks. A total of 34-fold coverage in Pacific Biosciences single-molecule long reads (N50 22 kb) and 45-fold coverage in 10X Genomics read clouds (from molecules with an estimated N50 of 34 kb) were generated. Primary assembly contigs were scaffolded with chromosome conformation Hi-C data. The Hi-C scaffolds were validated using BioNano Genomics long range restriction maps (106-fold effective coverage). Manual assembly curation corrected 54 missings/misjoins and removed 1 haplotypic duplication, reducing the scaffold number by 23.59%, increasing the scaffold N50 by 19.08% and decreasing the assembly length by 0.01%. The final assembly has a total length of 1.18 Gb in 357 sequence scaffolds with a scaffold N50 of 81.4 Mb (Table 1). The majority, 98.3%, of the assembly sequence was assigned to 35 chromosomal-level scaffolds representing 33 autosomes (numbered by synteny to the chicken, Gallus gallus domesticus: GCA_000002315.5), and the W and Z sex chromosomes (Figure 1-Figure 4; Table 2). The assembly has a BUSCO v5.1.2 (Simão et al., 2015) completeness of 95.7% using the aves_odb10 reference set. While not fully phased, the assembly deposited is of one haplotype. Contigs corresponding to the second haplotype have also been deposited.

Project accession data			
Assembly identifier	bStrTur1.2		
Species	Streptopelia turtur		
Specimen	bStrTur1		
NCBI taxonomy ID	NCBItxid:177155		
BioProject	PRJEB32724		
BioSample ID	SAMEA994735		
Isolate information	Female, blood sample		
Raw data accessions			
PacificBiosciences SEQUEL I	ERR3041797-ERR3041814		
10X Genomics Illumina	ERR3229775, ERR3316037-ERR3316040		
Hi-C Illumina	ERR4179379-ERR4179383		
BioNano	ERZ1392829		
Genome assembly			
Assembly accession	GCA_901699155.2		
Accession of alternate haplotype	GCA_901699165.2		

able 1. Genome (data for <i>Strept</i>	opelia turtur	bStrTur1.2

Genome assembly			
Span (Mb)	1,179		
Number of contigs	1,246		
Contig N50 length (Mb)	3.79		
Number of scaffolds	311		
Scaffold N50 length (Mb)	81.4		
Longest scaffold (Mb)	222		
BUSCO* genome score	C:95.7%[S:94.9%,D:0.8%],F:0.8%,M:3.5%, n:8338		

*BUSCO scores based on the aves_odb10 BUSCO set using v5.1.2. C= complete [S= single copy, D=duplicated], F=fragmented, M=missing, n=number of orthologues in comparison. A full set of BUSCO scores is available at https://blobtoolkit.genomehubs.org/view/Streptopelia%20turtur/dataset/CABFKC02/busco.



Figure 1. Genome assembly of *Streptopelia turtur*, bStrTur1.2: metrics. The BlobToolKit Snailplot shows N50 metrics and BUSCO gene completeness. An interactive version of this figure is available at https://blobtoolkit.genomehubs.org/view/Streptopelia%20turtur/dataset/ CABFKC02/snail.

Methods

The European turtle dove specimen was taken from blood collected from a live bird during routine health checks of populations in Marks Tey, Essex, UK (latitude 51.874N, longitude 0.729E; grid reference TL8823). The sample was taken under Home Office (Animals Scientific Procedures Act, ASPA) licence number PPL 7007641); the bird was caught and handled under a British Trust for Ornithology ringing license.

DNA was extracted using an agarose plug extraction from a blood sample following the BioNano Genomics Prep Blood and Cell Culture DNA Isolation Protocol. Pacific Biosciences



Figure 2. Genome assembly of *Streptopelia turtur*, bStrTur1.2: GC coverage. BlobToolKit GC-coverage plot. An interactive version of this figure is available at https://blobtoolkit.genomehubs.org/view/Streptopelia%20turtur/dataset/CABFKC02/blob.

(PacBio) CLR long read and 10X Genomics read cloud sequencing libraries were constructed according to manufacturers' instructions. Sequencing was performed by the Scientific Operations core at the Wellcome Sanger Institute on Pacific Biosciences SEQUEL I and Illumina HiSeq X instruments. Ultra-high molecular weight DNA was extracted using the BioNano Genomics Prep Animal Tissue DNA Isolation Soft Tissue Protocol and assessed by pulsed field gel and Qubit 2 fluorimetry. DNA was labeled for BioNano Genomics optical mapping following the BioNano Genomics Prep Direct Label and Stain (DLS) Protocol, and run on one Saphyr Optical Instrument chip flowcell (BioNano Genomics). Hi-C data were generated using the Arima Hi-C kit v1 by Arima Genomics, San Diego, USA, using the Illumina HiSeqX sequencing instrument.

Assembly was carried out following the Vertebrate Genome Project pipeline v1.6 (Rhie *et al.*, 2020) with Falcon-unzip (Chin *et al.*, 2016); haplotypic duplication was identified and removed with purge_haplotigs (Roach et al., 2018) and a first round of scaffolding carried out with 10X Genomics read clouds using scaff10x. Hybrid scaffolding was performed using the BioNano Genomics DLE-1 data and BioNano Solve. Scaffolding with Hi-C data (Rao et al., 2014) was carried out with SALSA2 (Ghurye et al., 2019). The Hi-C scaffolded assembly was polished with arrow using the PacBio data, then polished with the 10X Genomics Illumina data by aligning to the assembly with longranger align, calling variants with freebayes (Garrison & Marth, 2012) and applying homozygous nonreference edits using bcftools consensus. Two rounds of the Illumina polishing were applied. The assembly was checked for contamination and corrected using the gEVAL system (Chow et al., 2016) as described previously (Howe et al., 2021). Manual curation was performed using evidence from BioNano Genomics (using the BioNano Access viewer), using HiGlass and Pretext. Figure 1-Figure 3 and BUSCO v5.1.2 scores were generated using BlobToolKit (Challis et al., 2020). Table 3 gives version numbers of the software tools used in this work.



Figure 3. Genome assembly of *Streptopelia turtur,* **bStrTur1.2: cumulative sequence.** BlobToolKit cumulative sequence plot. An interactive version of this figure is available at https://blobtoolkit.genomehubs.org/view/Streptopelia%20turtur/dataset/CABFKC02/ cumulative.



Figure 4. Genome assembly of Streptopelia turtur, bStrTur1.2: Hi-C contact map. Hi-C contact map of the bStrTur1 assembly, visualized in HiGlass (Kerpedjiev et al., 2018).

INSDC accession	Chromosome	Size (Mb)	GC%
LR594551.2	1	222.12	0.40
LR594552.1	2	169.96	0.40
LR594553.2	3	126.22	0.40
LR594554.2	4	81.37	0.40
LR594556.1	5	70.85	0.41
LR594558.1	6	40.34	0.42
LR594557.1	7	42.93	0.41
LR594559.1	8	34.57	0.42
LR594560.2	9	27.80	0.43
LR594562.1	10	23.07	0.44
LR594563.1	11	22.26	0.43
LR594561.1	12	23.32	0.43
LR594564.1	13	20.81	0.45
LR594565.1	14	18.48	0.46
LR594567.2	15	15.32	0.47
LR594569.2	17	11.44	0.48

Table 2. Chr	omosomal	pseudomo	lecules i	n the
genome ass	sembly of S	treptopelia	turtur, k	StrTur1.2.

INSDC accession	Chromosome	Size (Mb)	GC%
LR594570.2	18	12.59	0.47
LR594568.1	19	10.86	0.48
LR594566.1	20	16.38	0.47
LR594571.2	21	7.81	0.49
LR594577.1	22	4.50	0.50
LR594572.1	23	5.94	0.52
LR594573.1	24	5.82	0.48
LR594579.1	25	1.00	0.58
LR594575.2	26	6.47	0.52
LR594576.2	27	6.49	0.50
LR594574.2	28	5.37	0.53
OU015479.1	29	22.41	0.44
LR594580.2	30	0.71	0.52
LR594581.1	32	0.02	0.53
LR594578.1	33	1.28	0.55
LR594555.2	Z	77.54	0.39
OU015480.1	W	8.84	0.43

Table 3. Software tools used.

Software tool	Version	Source
Falcon-unzip	falcon-kit 1.1.1	(Chin <i>et al.,</i> 2016)
purge_haplotigs	1.0.2	(Roach <i>et al.,</i> 2018)
SALSA2	2.2	(Ghurye <i>et al.,</i> 2019)
Bionano Solve	3.3_10252018	https://bionanogenomics.com/support/software-downloads/
scaff10x	3.0	https://github.com/wtsi-hpag/Scaff10X
arrow	2.2.2	https://github.com/PacificBiosciences/GenomicConsensus
longranger align	2.2.2	https://support.10xgenomics.com/genome-exome/software/ pipelines/latest/advanced/other-pipelines
freebayes	1.1.0-3-g961e5f3	(Garrison & Marth, 2012)
bcftools consensus	1.9	http://samtools.github.io/bcftools/bcftools.html
HiGlass	1.11.6	(Kerpedjiev <i>et al.,</i> 2018)
PretextView	0.0.4	https://github.com/wtsi-hpag/PretextMap
gEVAL	N/A	(Chow <i>et al.</i> , 2016)
BlobToolKit	2.6.1	(Challis <i>et al.,</i> 2020)

Data availability

European Nucleotide Archive: Streptopelia turtur (European turtle dove) genome assembly, bStrTur1. Accession number PRJEB32724.

The genome sequence is released openly for reuse. The S. turtur genome sequencing initiative is part of the Wellcome Sanger Institute's "25 genomes for 25 years" project. It is also part of the Vertebrate Genome Project (VGP) ordinal references programme and the Darwin Tree of Life (DToL) project.

All raw data and the assembly have been deposited in the ENA. The genome will be annotated and presented through the Ensembl pipeline at the European Bioinformatics Institute. Raw data and assembly accession identifiers are reported in Table 1.

Acknowledgements

We thank Mike Stratton and Julia Wilson for their continuing support for the 25 genomes for 25 years project.

References

Burns F. Eaton MA. Balmer DE. et al.: The state of the UK's birds 2020. The RSPB, BTO, WWT, DAERA, JNCC, NatureScot, NE and NRW, Sandy, Bedfordshire. 2020

Reference Source

Browne SJ, Aebischer NJ: Habitat Use, Foraging Ecology and Diet of Turtle Doves Streptopelia Turtur in Britain. Ibis. 2003; 145(4): 572-82 **Publisher Full Text**

Browne SJ, Aebischer NJ: Temporal Changes in the Breeding Ecology of European Turtle Doves *Streptopelia Turtur* in Britain, and Implications for Conservation. *Ibis.* 2004; **146**(1): 125–37. **Publisher Full Text**

Challis R, Richards E, Rajan J, et al.: BlobToolKit - Interactive Quality Assessment of Genome Assemblies. G3 (Bethesda). 2020; 10(4): 1361-74. PubMed Abstract | Publisher Full Text | Free Full Text

Chin CS, Peluso P, Sedlazeck FJ, et al.: Phased Diploid Genome Assembly with Single-Molecule Real-Time Sequencing. Nat Methods. 2016; 13(12): 1050-54. PubMed Abstract | Publisher Full Text | Free Full Text

Chow W, Brugger K, Caccamo M, et al.: gEVAL — a Web-Based Browser for Evaluating Genome Assemblies. Bioinformatics. 2016; 32(16): 2508-10. PubMed Abstract | Publisher Full Text | Free Full Text

Dunn JC, Stockdale JE, Moorhouse-Gann RJ, et al.: The Decline of the Turtle Dove: Dietary Associations with Body Condition and Competition with Other Columbids Analysed Using High-Throughput Sequencing. Mol Ecol. 2018; 27(16): 3386-3407.

PubMed Abstract | Publisher Full Text

Garrison E, Marth G: Haplotype-Based Variant Detection from Short-Read Sequencing. arXiv: 1207.3907. 2012. **Reference Source**

Ghurye RR, Sundaram K, Smith F, et al.: Novel ADA2 Mutation Presenting with Neutropenia, Lymphopenia and Bone Marrow Failure in Patients with

Deficiency in Adenosine Deaminase 2 (DADA2). Br J Haematol. 2019; 186(3): e60-64.

PubMed Abstract | Publisher Full Text

Howe K, Chow W, Collins J, et al.: Significantly Improving the Quality of Genome Assemblies through Curation. Gigascience. 2021; 10(1): giaa153. PubMed Abstract | Publisher Full Text | Free Full Text

Kerpedjiev P, Abdennur N, Lekschas F, et al.: HiGlass: Web-Based Visual Exploration and Analysis of Genome Interaction Maps. Genome Biol. 2018; **19**(1): 125.

PubMed Abstract | Publisher Full Text | Free Full Text

Rao SSP, Huntley MH, Durand NC, et al.: A 3D Map of the Human Genome at Kilobase Resolution Reveals Principles of Chromatin Looping. Cell. 2014; 159(7): 1665-80.

PubMed Abstract | Publisher Full Text | Free Full Text

Rhie A, McCarthy SA, Fedrigo O, et al.: Towards Complete and Error-Free Genome Assemblies of All Vertebrate Species. bioRxiv. 2020; 2020.05.22.110833

Publisher Full Text

Roach MJ, Schmidt SA, Borneman AR: Purge Haplotigs: Allelic Contig Reassignment for Third-Gen Diploid Genome Assemblies. BMC Bioinformatics. 2018; 19(1): 460.

```
PubMed Abstract | Publisher Full Text | Free Full Text
```

Simão FA, Waterhouse RM, Ioannidis P, et al.: BUSCO: Assessing Genome Assembly and Annotation Completeness with Single-Copy Orthologs. Bioinformatics. 2015; **31**(19): 3210–12. PubMed Abstract | Publisher Full Text

Stockdale JE, Dunn JC, Goodman SJ, et al.: The Protozoan Parasite Trichomonas Gallinae Causes Adult and Nestling Mortality in a Declining Population of European Turtle Doves, *Streptopelia Turtur*. Parasitology. 2015; **142**(3): 490–98. PubMed Abstract | Publisher Full Text