## **Supporting Information**

### Imaging of Neurotransmitters and Small Molecules in Brain Tissues using Laser Desorption/Ionization Mass Spectrometry Assisted with Zinc Oxide Nanoparticles

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### **Conditions of Supporting Experiments with LC-MS**

#### **Sample Preparations for LC-MS**

A frozen rat brain of 112 mg was homogenized in 400  $\mu$ L of prechilled MeOH/H<sub>2</sub>O (1:1) using tissue grinder pestle in a 1.5 mL microtube, followed by centrifugation at 13000 g for 20 min at 4 °C. Aliquots of 100  $\mu$ L of the supernatant were transferred into microtubes and then spun in a vacuum concentrator for 3 h at 43 °C until dry. The aqueous extracts of the tissue samples were finally reconstituted in 200  $\mu$ L of solvent mixture of MeOH/H<sub>2</sub>O (1:1), after centrifugation for 20 min at 13000 g and 4 °C. Following aqueous extraction, the residual pellet was homogenized in 400  $\mu$ L of prechilled CHCl<sub>3</sub>/MeOH (3:1) using tissue grinder pestle in 1.5 mL microtube, followed by centrifugation at 13000 g for 20 min at 4 °C. Aliquots of 150  $\mu$ L of the supernatant were transferred into another microtube and were allowed to evaporate at room temperature in fume hood overnight. The organic extracts of the tissue samples were reconstituted in 25  $\mu$ L of the solvent mixture of H<sub>2</sub>O /ACN/isopropanol (ISP) (1:1:2), after centrifugation for 10 min at 5000 g and 4 °C.

#### LC-MS analysis of rat brain

LC/MS/MS analysis was performed with an Agilent 1290 HPLC system coupled to a Q-Exactive Quadrupole Orbitrap mass spectrometer (Thermo Scientific). Chromatographic conditions consisted of an Eclipse Plus C18 RRHD column  $(2.1 \times 100 \text{ mm}, 1.8 \mu\text{m}; \text{Agilent Technologies})$  maintained at 35 °C using an established gradient program. The mobile phase consisted of water with 0.1% formic acid (mobile phase A) and acetonitrile with 0.1% formic acid (mobile phase B) (Optima grade, Fisher Scientific). Mobile phase B was held at 0% B for 0.5 min before increasing to 100% over 3.5 min, held at 100% for 2.5 min, before returning to 0% B over 0.5 min. Brain metabolite aqueous and organic extracts were injected with a volume of 5  $\mu$ L each and separated at a flow rate of 0.3 mL/min.

Heated electrospray ionization was used in both positive and negative ion mode with the following settings: capillary voltage, 3.9 kV and 3.5 kV respectively in positive and negative ion mode; capillary temperature, 400 °C; sheath gas, 17 units; auxiliary gas, 8 units; probe heater temperature, 450 °C; S-Lens RF level, 50%. MS data were acquired using untargeted DDA that included a full MS scan at 35 000 resolution, with a scan range of 70–1000 *m/z*; automatic gain control target,  $1 \times 10^6$ ; and a maximum injection time of 128 ms. The five highest intensity ions were selected from each full scan for MS/MS analysis using a 1.2 Da isolation window and were analyzed using the following conditions: resolution, 17 500; automatic gain control target,  $1 \times 10^6$ ; max IT, 64 ms; normalized stepped collision energy, 20/40; intensity threshold,  $2 \times 10^5$ ; dynamic exclusion, 7 s. Raw data files were imported into MZmine. The *m/z* peak list was matched against the theoretical *m/z* of tentatively identified molecules from MALDI MSI using 5 ppm.

analyte	detection mode	derivatizaion	matrix	tissue type	spatial resolution	reference
glycine	+	yes	DHB	mouse brain	125 µm	[28]
		no	ZnO TiOn	mouse brain	70 µm	This work
aspartate	+	yes	DHB	mouse brain	125 µm	[28]
		no	TiO <sub>2</sub>	mouse brain	70 µm	[55]
		no	ZnO TiO <sub>2</sub>	mouse brain	70 µm	This work
	_	no	9-AA	mouse brain	50 µm	[18]
		no	NEDC	mouse brain	100 µm	[20]
		no	PNA	rat brain	200 µm	[21]
		no	BNDM	rat brain	100 µm	[23]
serine	+	yes	DHB	mouse brain	125 µm	[28]
		no	TiO <sub>2</sub>	mouse brain	70 µm	[55]
		no	TiO <sub>2</sub> ZnO	mouse brain mouse brain Bat brain	70 μm 70 μm 80 μm	This work
GABA	+	yes	DHB	mouse brain	125 μm	[28]
		yes	CHCA	rat brain	100 µm	[29]
		yes	CHCA	rat brain pig adrenal gland	50 μm 200 μm	[30]
		yes	TPP	rat brain	120 µm	[33]
		no	TiO <sub>2</sub>	mouse brain	70 µm	[55]
		no	TiO <sub>2</sub> ZnO	mouse brain mouse brain rat brain	70 μm 70 μm 80 μm	This work
		no	modified-TiO2	mouse brain	30 um	[58]
glutamate	+	yes	DHB	mouse brain	125 μm	[28]
		yes	CHCA	brain	-	[29]
		yes	CHCA	rat brain	50 µm	[30]
		no	TiO <sub>2</sub>	mouse brain	70 µm	[55]
		no	TiO <sub>2</sub> ZnO	mouse brain mouse brain rat brain	70 μm 70 μm 80 μm	This work
		no	nodified-TiO <sub>2</sub>	mouse brain	30 µm	[58]
	-	no	9-AA	mouse brain	50 µm	[18]
		no	NEDC	mouse brain	100 µm	[20]
		no	PNA	rat brain	200 µm	[21]
		no	BNDM	rat brain	100 µm	[23]
alanine	+	yes	DHB	mouse brain	125 µm	[28]
		no	TiO <sub>2</sub> ZnO	mouse brain mouse brain rat brain	70 μm 70 μm 80 μm	This work
taurine	+	yes	DHB	mouse brain	125 µm	[28]
		no	ZnO	mouse brain rat brain	70 μm 80 μm	This work
	-	no	NEDC	mouse kidney	50 µm	[20]
cysteine	+	no	PNA TiO2	rat brain mouse brain	200 μm 70 μm	[21]
cysteme	4	110	7.0	incuse brain	70 µm	[55]
		no	ZnO	mouse brain	70 µm	This work

# Table S1. Summary of Common Neurotransmitters Imaged by LDI MSI in the Literature.

adenosine	+	No	TiO <sub>2</sub>	mouse brain	70 µm	this work
			ZnO	mouse brain	70 µm	
				rat brain	80 µm	
acetylcholine	+	Yes	CHCA	rat brain	15 µm	[29]
				mouse brain	100 µm	
dopamine	+	Yes	DHB	mouse brain	125 µm	[28]
		Yes	CHCA	rat brain	100 µm	[29]
		Yes	CHCA	pig adrenal gland	200 µm	[30]
		Yes	TPP	rat brain	120 µm	[33]
		No	TiO <sub>2</sub>	mouse brain	70 µm	
			ZnO	mouse brain	70 µm	this work
				rat brain	80 µm	
norepinephrine	+	Yes	CHCA	pig adrenal gland	200 µm	[30]
		No	TiO <sub>2</sub>	mouse brain	70 µm	this work
			ZnO	mouse brain	70 µm	
			TiO <sub>2</sub>	rat brain	80 µm	
epinephrine	+	Yes	CHCA	pig adrenal gland	200 µm	[30]
		No	TiO <sub>2</sub>	mouse brain	70 µm	this work
			ZnO	rat brain	80 µm	
serotonin	+	Yes	CHCA	-	-	[29]
tyramine	+	Yes	CHCA	brain	-	[29]
tryptamine	+	Yes	CHCA	brain	-	[29]

**Table S2.** A List of the Small Molecules Detected in Mouse Brain Tissues by LDI MSI with  $TiO_2 NP$  or Dopamine-Modified  $TiO_2$  Monolith.

	,		1 1		1
m/z	compd	TiO2 in this	TiO <sub>2</sub> [54]	IS donamine-modified	classification
		work	1102[54]	TiO <sub>2</sub> [57]	
99.00	glycine	$[M + Na]^+$		1102[07]	amino acids
113.97	67	$[M + K]^{+}$			
122.03	cysteine		$[M + H]^{+}$		amino acids
127.98	alanine	$[M + K]^{+}$			amino acids
133.08	ornithine		$[M + H]^{+}$		amino acids
134.05	aspartic acid		$[M + H]^{+}$		amino acids
172.01		[M + K]			,
132.07	creatine	$M + K 1^{+}$	[M + H]		amino acids
170.03		[MI + K]		$[M + K]^+$	
143.99	serine	$[M + K]^+$		[]	amino acids
154.00	proline	$[M + K]^{+}$			amino acids
154.02	*			$[M + K]^{+}$	
156.02	valine	$[M + K]^{+}$			amino acids
120.06	threonine		$[M + H]^{+}$		amino acids
158.01	1 11 1	$[M + K]^+$	$D ( + III^{\ddagger})$		,
166.08	phenylalanine	$10.4 \pm 12.1^{+}$	[M + H]		amino acids
167.01	2-amino-4-cyano-	[M + K]		$[M + K]^+$	amino acids
168.00	pyroglutamic acid	$[M + K]^+$			amino acids
168.01	pyrogramme uera	[]		$[M + K]^{+}$	unnio uolus
169.04	glutamine	$[M + Na]^+$			amino acids
169.06	-			$[M + Na]^{+}$	
185.02		$[M + K]^{+}$			
148.06	glutamic acid		$[M + H]^{+}$		amino acids
186.00		[M + K]		DM   121 <sup>+</sup>	
150.02	mathianina		$IM \pm HI^+$	[IVI+K]	amino agida
188.00	methonne	$[M + K]^{+}$	[WI + II]		ammo acius
194.02	histidine	$[M + K]^+$			amino acids
196.00	amino-muconic acid			$[M + K]^{+}$	amino acids
196.01		$[M + K]^{+}$			
198.09	amino-octanoic acid			$[M + K]^{+}$	amino acids
204.03	phenylalanine	$[M + K]^{+}$			amino acids
175.11	arginine		$[M + H]^{+}$		amino acids
213.06		$[M + K]^+$			
234.07	methoxytyrosine			$[M + Na]^+$	amino acids
89.10	putrescine	$IM + N_{2}1^{+}$	[M + H]		alkaloids
122.06	niaatinamida	[IVI + INA]	$IM \pm HI^+$		alkalaida
160.99	incomannue	$[M + K]^{+}$	[WI + II]		aikaioius
168.13	spermidine	$[M + Na]^+$			alkaloids
184.09	1	$[M + K]^{+}$			
180.04	arecaidine			$[M + K]^{+}$	alkaloids
241.17	spermine	$[M + K]^+$			alkaloids
136.06	adenine		$[M + H]^{+}$		purine
174.01		$[M + K]^{+}$			
137.05	hypoxanthine	$D(t + N_{t-})^{\pm}$	[M + H]		purine
159.00		[M + Na] $[M + K1^+$			
113.03	uracil	[WI + K]	$[M + H]^+$		nvrimidine
150.96	uraen	$[M + K]^{+}$	[MI + II]		pyrimaine
104.06	γ-aminobutyric acid	,	$[M + H]^{+}$		neurotransmitters
126.02	(GABA)	$[M + Na]^+$			
142.00		$[M + K]^{+}$			
142.03	1	DM + 123+		$[M + K]^+$	· · · · · · · · · ·
191.03	aopamine	[M + K]'			neurotransmitters
208.02	norepinephrine	$[M + K]^{+}$			neurotransmitters
306.06	adenosine	$[M + K]^+$			neurotransmitters
409 34	cholesterol	[IVI + K]		$[M + Na]^+$	sterol linids
409.36		$[M + Na]^{+}$		[	
425.32				$[M + K]^+$	
425.34		$[M + K]^+$		-	
429.24	hydroxy-oxo-cholan-			$[M + K]^{+}$	sterol lipids
420.20	oic acid	├		$D_{1}(x) = 123^{+}$	
439.30	OH-/- dehydrocholostorol	$[M + V]^+$		[M + K]	sterol lipids
439.31	denydrocholesterol	[1VI + K]		D.C. TOL	
465.33	cholesta-6,8(14)-dien-			$[M + K]^{T}$	sterol lipids
2/3 00	Met-Ala	$[M + N_0]^+$			dinentides
245.08	witt-Ala	[IVI INA]		$[M + K]^+$	apeptides
277.06	Gly-Tyr			$[M + Na]^+$	dipeptides
283.07	Pro-Glu	l l		$[M + K]^+$	dipeptides
		•			

(table continued on next page)

m/z	compd		detected ions		classification
	tompa	TiO <sub>2</sub> in this	TiO <sub>2</sub> [54]	dopamine-	
		work		modified	
				TiO <sub>2</sub> [57]	
413.12	Cys Gly Pro Val			$[M + K]^{+}$	peptides
453.17	Gly Glu Pro Ile			$[M + K]^{+}$	peptides
457.21	Met Leu Ala Thr			$[M + Na]^{+}$	peptides
459.22	Thr Leu Gly Phe			$[M + Na]^+$	peptides
469.15	Glu Trp Pro			[M + K] <sup>+</sup>	peptides
481.21	Asp Val Leu Pro			$[M + K]^+$	peptides
495.16	Gly Phe Ala Tyr			$[M + K]^+$	peptides
407.18	Acn Acn Bro Mot			$[M + N_0]^+$	peptides
497.18	Ash Ash Pro Met	$DM + 121^{+}$		[NI + Na]	fertines for the second s
126.99	butyric acid	[M + K]			fatty acids
168.98	5-oxohexanoic acid	M + K			fatty acids
277.18	FA(16:1)	[M + Na]		51 C + 172±	fatty acids
293.19	71460			[M + K]	
279.23	FA(16:0)			[M + Na]	fatty acids
295.20				$[M + K]^{+}$	
305.24	FA(18:1)			$[M + Na]^{+}$	fatty acids
321.23		$[M + K]^{+}$		. ,	
307.26	FA(18:0)			$[M + Na]^+$	fatty acids
323.23	( · · · /	$[M + K]^{+}$			
323.23				$[M + K]^{+}$	
327.23	FA(20:4)			$[M + Na]^+$	fatty acids
343,20				$[M + K]^{+}$	,
343.21		$[M + K]^+$		[	
337.21	hydroxy-oleic acid	,		$[M + K]^{+}$	fatty acids
341.19	FA(20:5)			$[M + K]^+$	fatty acids
351.22	FA(22:6)	[M + Nal <sup>+</sup>		[	fatty acids
351.22		[111 . 144]		$[M + N_2]^+$	intry neitos
367.20				$[M + K]^+$	
367.21		$[M + K]^+$		[IVI + IX]	
355.26	decesstatreancia said			$[M \pm N_0]^+$	fatty agida
555.20	(22:4)			[IVI + IVA]	fatty actus
262.27	(22.4) EA(21.1)			$[M + V]^+$	fatty agida
120.00	FA(21:1)	$D_{1} + N_{1} + 1^{\pm}$		[NI + K]	Tatty actus
139.00	Tumaric acid	[M + Na]			organic acids
144.98	glyceric acid	[M + K]			organic acids
153.00	citraconic acid	[M + Na]			organic acids
154.99	levulinic acid	$[M + K]^{+}$			organic acids
156.98	succinic acid	$[M + K]^{+}$			organic acids
159.00	threonic acid	$[M + Na]^{+}$			organic acids
187.01	cinnamic acid	$[M + K]^{+}$			organic acids
189.03	3-phenylpropionic	$[M + K]^{+}$			organic acids
	acid				-
212.97	aconitic acid	$[M + K]^{+}$			organic acids
217.02	glucuronic acid	$[M + Na]^+$			organic acids
104.09	choline	M <sup>+</sup>			others
110.03	hypotaurine		$[M + H]^+$		others
112.01	oxamic acid	$[M + Na]^+$			others
112.06	cytosine	[]	$[M + H1^{+}]$		others
150.00	0. 100 mile	$[M + K]^+$	[		oulors
130.08	ninecolic acid		$[M + H1^+]$		others
126.04	procedule actu	├	[IVI + IT]	$IM \perp N_{-}1^{+}$	others
150.04	creatinine	$[M + V]^+$		[IVI + INA]	others
152.01		[IVI + K]		$[M + k']^+$	
120.02	aminoponteramida	├		[101   IX] [M_1 N1-1 <sup>+</sup>	others
139.08	ammopentanamide	├	$M + 111^+$	[IVI⊤INA]	others
143.08	ectoine	D.4 - 37.24	[M + H]		otners
146.98	phosphono-	$[M + Na]^{T}$			others
162.96	acetaldehyde	$[M + K]^{+}$	. <i>c</i> ⊥		-
164.07	S-methylmethionine		M		others
165.04	lumazine		$[M + H]^{+}$		others
184.07	phosphocholine		$M^+$		others
192.98	dihydroxyacetone	$[M + Na]^{+}$			others
	phosphate				
203.04	myo-inositol	$[M + Na]^{+}$			others
219.01		$[M + K]^{+}$			
207.03	3-methoxy-4-	$[M + Na]^+$			others
223.01	hydroxyphenylglycol	$[M + K]^{+}$			
214.00	N-acetylaspartic acid	$[M + K]^+$			others
228,01	N-acetylglutamic acid	$[M + K]^+$			others
260.05	N-acetylglucosamine	$[M + K1^+]$			others
266.05	cytidine	$[M + N_2]^+$			others
307.05	guanosine	$[M + K1^+]$			others
375.20	prostaglandin E2	$[M + N_0]^+$			others
391.20	prostagianulli E2	$[M + K]^+$			otters

### A. Myo-inositol



**Figure S1.** ZnO NP-assisted LDI mass spectra of standards: 30 ng of **(A)** myo-inositol and **(B)** GABA in positive ion mode and **(C)** 300 ng of palmitic acid in negative ion mode.



**Figure S2.** Calibration curve of GABA-d6 signals measured by ZnO NP-assisted LDI MS. The GABA-d6 concentration refers to that of the standard solution spiked on rat brain tissue sections. Error bars show +/– the standard deviation from 9 replicate measurements.



**Figure S3.** Ion images of sagittal sections of mouse brain acquired with ZnO NP-assisted LDI-MSI at a raster size of 70  $\mu$ m in the positive ion mode (figure continued on next page).



(figure continued on next page)



*m*/z 423.31



Figure S4. Ion images of sagittal sections of mouse brain acquired with ZnO NP-assisted LDI-MSI at a raster size of 70  $\mu$ m in the negative ion mode.



**Figure S5**. Peak height intensity ratios of the sodiated and potassiated ions from two adjacent mouse brain tissue sections without (control) and with the application of 5 mM potassium acetate (a) or 5 mM sodium acetate (b) by automated sprayer. M represent the molecular weight. The figure inset shows the isotope patterns of  $[M + K]^+$  at m/z 156.88 without and with the application of 5 mM potassium acetate.