

17:07:00

OCA PAD INITIATION - PROJECT HEADER INFORMATION

10/04/88

Active

Project #: E-25-614  
Center # : R6595-0A0

Cost share #:  
Center shr #:

Rev #: 0  
OCA file #:  
Work type : RES  
Document : OTH  
Contract entity: GTRC

Contract#: LTR DTD 880725  
Prime #: 1 R01 HL41175

Mod #:

Subprojects ? : N  
Main project #:

Project unit: ME  
Project director(s):  
NEREM R M ME

Unit code: 02.010.126

Sponsor/division names: UNIVERSITY OF TEXAS  
Sponsor/division codes: 400

/ SAN ANTONIO, TX  
/ 045

Award period: 880701 to 890430 (performance) 890430 (reports)

Sponsor amount	New this change	Total to date
Contract value	74,620.00	74,620.00
Funded	74,620.00	74,620.00
Cost sharing amount		0.00

Does subcontracting plan apply ? : N

Title: VASCULAR HEALING: CELL BIOLOGY AND RHEOLOGIC FACTORS

PROJECT ADMINISTRATION DATA

OCA contact: Ina R. Lashley

894-4820

Sponsor technical contact

Sponsor issuing office

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Security class (U,C,S,TS) : U  
Defense priority rating : N/A  
Equipment title vests with: Sponsor  
NONE PROPOSED.

ONR resident rep. is ACO (Y/N): N  
N/A supplemental sheet  
GIT

Administrative comments -

CONSORTIUM AGREEMENT ESTABLISHED UNDER AN NIH GRANT. NIH GRANT GUIDELINES GOVERNS ITS ADMINISTRATION.



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SR-525  
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GEORGIA INSTITUTE OF TECHNOLOGY  
OFFICE OF CONTRACT ADMINISTRATION

NOTICE OF PROJECT CLOSEOUT

Date 7/3/89

Project No. E-25-614

Center No. R6595-OA0

Project Director R. M. Nerem

School/Lab ME

Sponsor University of Texas Health Science Center

Contract/Grant No. Agreement dtd 7/27/88

GTRC XX GIT     

Prime Contract No. 1 R01 HL41175

Title Vascular Healing: Cell Biology and Rheologic Factors

Effective Completion Date 4/30/89

(Performance) 4/30/89

(Reports)

Closeout Actions Required:

- None
- Final Invoice or Copy of Last Invoice - Submitted
- Final Report of Inventions and/or Subcontracts - Patent questionnaire sent to PI.
- Government Property Inventory & Related Certificate
- Classified Material Certificate
- Release and Assignment
- Other \_\_\_\_\_

Includes Subproject No(s). \_\_\_\_\_

Subproject Under Main Project No. \_\_\_\_\_

Continues Project No. \_\_\_\_\_

Continued by Project No. E-25-M80

Distribution:

- |  |   |
|--|---|
| <input checked="" type="checkbox"/> Project Director                 | <input checked="" type="checkbox"/> Reports Coordinator (OCA)       |
| <input checked="" type="checkbox"/> Administrative Network           | <input checked="" type="checkbox"/> GTRC                            |
| <input checked="" type="checkbox"/> Accounting                       | <input checked="" type="checkbox"/> Project File                    |
| <input checked="" type="checkbox"/> Procurement/GTRI Supply Services | <input checked="" type="checkbox"/> Contract Support Division (OCA) |
| <input checked="" type="checkbox"/> Research Property Management     | <input type="checkbox"/> Other _____                                |
| <input checked="" type="checkbox"/> Research Security Services       | _____   |

SECTION IV  
PROGRESS REPORT SUMMARY

1 ROI H141175-02

PRINCIPAL INVESTIGATOR OR PROGRAM DIRECTOR	PERIOD COVERED BY THIS REPORT	
<b>Schwartz, Colin J.</b>	FROM	THROUGH
APPLICANT ORGANIZATION	4/1/89	3/31/90
<b>The University of Texas Health Science Center</b>		
TITLE OF PROJECT (Repeat title shown in item 1 on first page)		
<b>Vascular Healing -- Cell Biology and Rheologic Factors</b>		

(SEE INSTRUCTIONS)

1. Brief Summary of Plans:

Based on preliminary data obtained in growth studies, no significant changes in our research plan are anticipated. Specific Aim #1 will be completed within the first 6 months of this next year (-02) and the experiments within Specific Aim 2; to determine the influence of hemodynamic preconditioning on maintenance of endothelial integrity in response to shear stress; and Specific Aim 3; to examine platelet and monocyte adherence on hemodynamically pre-conditioned endothelial cells; will be initiated in year -02 both at UTHSCSA and GIT as outlined in the original proposal.

2. Current Studies:

The initial seven months of this study have concentrated on characterization of the growth of bovine aortic endothelial cells (BAEC) and porcine aortic endothelial cells (PAEC) on several of the proposed substrates. These substrates include 1 micron pore polyester mesh, 1 micron pore dacron, a nonporous polyester film, and nonporous polystyrene (culture ware plastic). Additionally, endothelial cell replication on these surfaces was examined using either the native surface or subsequent to collagen and/or fibronectin coating.

Endothelial cells were seeded at a density of  $10^4$  cell/cm<sup>2</sup> on the respective substrates. Cell replication was monitored on days 1,3,5,7 after seeding by cell counting, <sup>3</sup>H-thymidine incorporation, and <sup>3</sup>H-thymidine autoradiography.

The results indicate that endothelial cells seeded upon noncoated polyester mesh, dacron mesh, or nonporous polyester exhibit a slower growth rate and attain a lower maximum number and density relative to cells plated on standard polystyrene culture ware. Either collagen and/or

fibronectin coating of these surfaces allowed endothelial cells to achieve full confluence by day 5 at a number and density similar to that observed on standard culture surfaces. In experiments utilizing coated surfaces, <sup>3</sup>H-thymidine incorporation and autoradiographic labeling were maximum at days 1 and 3 and sharply decreased to a minimum plateau at days 5 and 7.

Current efforts involve examining endothelial cell growth on uncoated vs collagen and/or fibronectin coated mesh under either steady state or pulsatile shear stress conditions. Secondly, current studies are now extending to evaluate PAEC and BAEC growth on expanded polytetrafluoroethylene (ePTFE). This studies were delayed due to difficulty in obtaining the desired material from the original proposed source (W.C. Gore and Associates). We now have ePTFE provided to us by Shiley, Inc., Irvine, CA.

The most significant achievements of the current year involve characterization of endothelial cell growth on several potential vascular graft materials and the attainment of a confluent endothelial cell monolayer on the porous polyester mesh within 5 days postseeding. These basic accomplishments allow us to proceed with the major objectives of this proposal. Specifically, studies examining the influence of hemodynamic shear stress on cell replication, cell to cell and cell to substrate interactions, and platelet and monocyte adherence to endothelium cultured on these potential graft surfaces can now proceed. Finally, these results indicate the feasibility of attaining a totally preendothelialized vascular graft which, potentially, may exhibit superior nonthrombogenicity and patency to currently available small diameter vascular grafts.

3. Human Subjects: No change

4. Vertebrate Animals: No change

5. Publications:

1. Schwartz CJ, Valente AJ, Kelley JL, Sprague EA and Edwards EH. Thrombosis and the Development of Atherosclerosis: Rokitansky Revisited. Seminars in Thrombosis and Hemostasis. 14:189-195, 1988

2. Valente AJ, Delgado R, Metter JD, Cho C, Sprague EA, Schwartz CJ and Graves DT. Cultured Primate Aortic Smooth Muscle Cells Express Both the PDGF-A and PDGF-B Genes, But Do Not Secrete a Mitogenic Platelet-Derived Growth Factor-Like Protein. *J Cell Physiol.* 136:479-485, 1988
3. Edwards EH, Sprague EA, Schwartz CJ. Low Density Lipoprotein (LDL) Endocytosis. I. Influence of the Multivalent Ligand Cationized Ferritin on Normal and Receptor-Negative Human Fibroblasts. *Exp Molec Path.* 48:353-372, 1988
4. Sprague EA, Edwards EH, Valente AJ, Suenram CA, Kerbacher JJ and Schwartz CJ. Modified Low Density Lipoprotein (Acetyl-LDL) Endocytosis II: Influence of the Multivalent Ligand, Cationized Ferritin, on Cultured Cells. *Exp and Molec Path.* 48:373-390, 1988
5. Kelley JL, Rozek MM, Suenram CA and Schwartz CJ. Activation of Human Peripheral Blood Monocytes by Lipoproteins. *Am J Path.* 130:223-231, 1988
6. Valente AJ, Graves DT, Vialle-Valentin CE, Delgado R and Schwartz CJ. Purification of a Monocyte Chemotactic Factor (SMC-CF) Secreted by Non-Human Primate Vascular Smooth Muscle Cells in Culture. *Biochemistry,* 27:4162-4168, 1988
7. Kelley JL, Suenram CA, Rozek MM and Schwartz CJ. Influence of hypercholesterolemia and cholesterol accumulation on rabbit carrageenan granuloma macrophage activation. *Am J Pathol,* 131:539, 1988
8. Sprague EA, Moser M, Edwards EH and Schwartz CJ. Stimulation of Receptor-Mediated Low Density Lipoprotein Endocytosis in Neuraminidase-Treated Cultured Bovine Aortic Endothelial Cells. *J. Cell Physiol.* 137:251-262, 1988
9. Kelley JL, Suenram CA, Rozek MM, Schaffer SA and Schwartz CJ. Influence of the Acyl-CoA: Cholesterol O-acyltransferase Inhibitor, CL 277082 on Cholesteryl Ester Accumulation in Rabbit Macrophage-Rich Granulomas and Hepatic Tissue. *Biochim et Biophys Acta* 960:83-90, 1988

10. Gwyne JT and Schwartz CJ. (eds). A Symposium: Second International Conference on hypercholesterolemia. Examining New Data on Probucol After a Decade of Use. Amer J Cardiol, 62:1B-81B, 1988
11. Schwartz CJ. Introduction - The Probucol Experience: A Review of the Past and a Look at the Future. In a Symposium: Second International Conference on Hypercholesterolemia. Examining New Data on Probucol After a Decade of Use. (JT Gwyne, CJ Schwartz, eds). Am J Cordiol, 62:1B-5B, 1988
12. Schwartz CJ. Advances in Plasma Cholesterol: Key Questions for Clinical Use and Research. Consultant, 28:48-49, 1988
13. Sprague EA, Steinbach BL, Logan SA, Nerem RM and Schwartz CJ. Influence of Shear Stress on Lipoprotein Endocytosis. Proceedings: Biology of the Arterial Wall. "Interaction in the Arterial Wall and Atherosclerosis". CIC Edizioni Internazionale: 183-188, 1988
14. Schwartz CJ, Sprague EA, Valente AJ, Kelley JL, Edwards EH and Suenram CA. Inflammatory Components of the Human Atherosclerotic Plaque. Workshop on the Evolution of the Human Atherosclerotic Plaque. Am Heart Assoc, Rockville, Maryland, September 20-23, Springer-Verlag. In press, 1989
16. Schwartz CJ, Sprague EA, Valente AJ, Kelley JL and Edwards, EH. Cellular Mechanisms in the Response of the Arterial Wall to Injury and Repair. Toxic Path. 17: , In press, 1989
17. Edwards EH, Sprague EA, Kelley JL, Kerbacher JJ, Schwartz CJ and Elbein AD. Castanospermine Inhibits the Function of the Low-Density Lipoprotein Receptor. Submitted, 1989

#### Abstracts:

1. Sprague EA, Edwards EH, Logan S, Schwartz CJ and Nerem RM. Enhanced LDL Receptor Expression in Cultured Arterial Endothelial Cells Exposed to Elevated Fluid-Imposed Wall Shear Stress. Symposium on Engineering Approaches to Atherosclerosis, 1988

2. Schwartz CJ, Sprague EA, Nerem RM and Grover FL. Vascular Healing: Cell Biology and Rheologic Factors. NIH Devices and Technology Branch, Program:33, 1988
3. Prasad ARS, Schwartz CJ. and Sprague EA. Phosphoinositide Metabolism and Low Density Lipoprotein Receptor-Mediated Endocytosis. *Circulation* 78:484a, 1988
4. Kelley JL, Kerbacher JJ and Schwartz CJ. Purification to Homogeneity of the Acetyl-LDL "Scavenger" Receptor from Rabbit Carrageenan Granulomas. *Circulation* 78:13a, 1988
5. Sprague EA, Prasad ARS and Schwartz CJ. Role of Signal Transduction in Low Density Lipoprotein (LDL) Receptor-Mediated Endocytosis. *J Cell Biol*, 107:810a, 1989