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Recommended Citation

De Vuyst, H., Mugo, N. R., Franceschi, S., McKenzie, K., Tenet, V., Njoroge, J., Rana, F. S., Sakr, S. R., Snijders, P. J., Chung, M. H. (2014). Residual disease and HPV persistence after cryotherapy for cervical intraepithelial neoplasia grade 2/3 in HIV positive women in Kenya. *PLOS ONE*, 9(10).

Available at: http://ecommons.aku.edu/eastafrica_fhs_mc_pathol/121

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Residual Disease and HPV Persistence after Cryotherapy for Cervical Intraepithelial Neoplasia Grade 2/3 in HIV-Positive Women in Kenya

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Abstract

Objective: To assess residual cervical intraepithelial neoplasia (CIN) 2/3 disease and clearance of high-risk (hr) human papillomavirus (HPV) infections at 6 months after cryotherapy among HIV-positive women.

Design: Follow-up study.

Methods: 79 HIV-positive women received cryotherapy for CIN2/3 in Nairobi, Kenya, and underwent conventional cytology 6 months later. Biopsies were performed on high grade cytological lesions and hrHPV was assessed before (cervical cells and biopsy) and after cryotherapy (cells).

Results: At 6 months after cryotherapy CIN2/3 had been eliminated in 61 women (77.2%; 95% Confidence Interval, (CI): 66.4–85.9). 18 women (22.8%) had residual CIN2/3, and all these women had hrHPV at baseline. CD4 count and duration of combination antiretroviral therapy (cART) were not associated with residual CIN2/3. CIN3 instead of CIN2 was the only significant risk factor for residual disease (odds ratio, OR vs CIN2 = 4.3; 95% CI: 1.2–15.0) among hrHPV-positive women after adjustment for age and HPV16 infection. Persistence of hrHPV types previously detected in biopsies was found in 77.5% of women and was associated with residual CIN2/3 (OR = 8.1, 95% CI: 0.9–70). The sensitivity, specificity, and negative predictive value of hrHPV test in detecting residual CIN2/3 were 0.94, 0.36, and 0.96 respectively.

Conclusions: Nearly one quarter of HIV-positive women had residual CIN2/3 disease at 6 months after cryotherapy, and the majority had persistent hrHPV. CD4 count and cART use were not associated with residual disease or hrHPV persistence. The value of hrHPV testing in the detection of residual CIN2/3 was hampered by a low specificity.

Citation: De Vuyst H, Mugo NR, Franceschi S, McKenzie K, Tenet V, et al. (2014) Residual Disease and HPV Persistence after Cryotherapy for Cervical Intraepithelial Neoplasia Grade 2/3 in HIV-Positive Women in Kenya. PLoS ONE 9(10): e111037. doi:10.1371/journal.pone.0111037

Editor: Craig Meyers, Penn State University School of Medicine, United States of America

Received: January 30, 2014; **Accepted:** August 1, 2014; **Published:** October 24, 2014

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Funding: This work was funded by the Washington Global Health Alliance, the National Institutes of Health (grant number 5K23AI065222-04), the Fondation de France (grant number 00016673), and the Bill & Melinda Gates Foundation (grant number 35537). The funders had no role in study design, data collection and analysis, decision to publish, or preparation of the manuscript.

Competing Interests: The authors have declared that no competing interests exist.

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Introduction

Women living with HIV are at increased risk for infection with human papillomavirus (HPV), cervical intraepithelial neoplasia grade 2 or 3 (CIN2/3) and invasive cervical cancer [1,2]. Residual or recurrent disease after CIN2/3 treatment is also more frequent among HIV-positive women [3] than HIV-negative women [4,5].

Most studies of CIN2/3 in HIV-positive women reported outcomes after excisional treatment, e.g. loop electrosurgical excision procedure (LEEP), or cold knife conization (CKC) [3,6–15]. However, cryotherapy is more feasible and affordable than excisional treatment in low- and middle-income countries [16].

Little information is available on the efficacy of cryotherapy for the treatment of CIN2/3 in HIV-positive women [6] and none on the impact of cryotherapy on HPV persistence, i.e., a strong risk factor for residual/recurrent disease in HIV-negative women [4,5].

The aim of this study was to assess: 1) the frequency of residual CIN2/3 disease and persistent infection of high-risk (hr) HPV types at 6 months after cryotherapy for CIN2/3, and 2) the performance of hrHPV testing after cryotherapy in the detection of residual disease in HIV-positive women in Kenya.

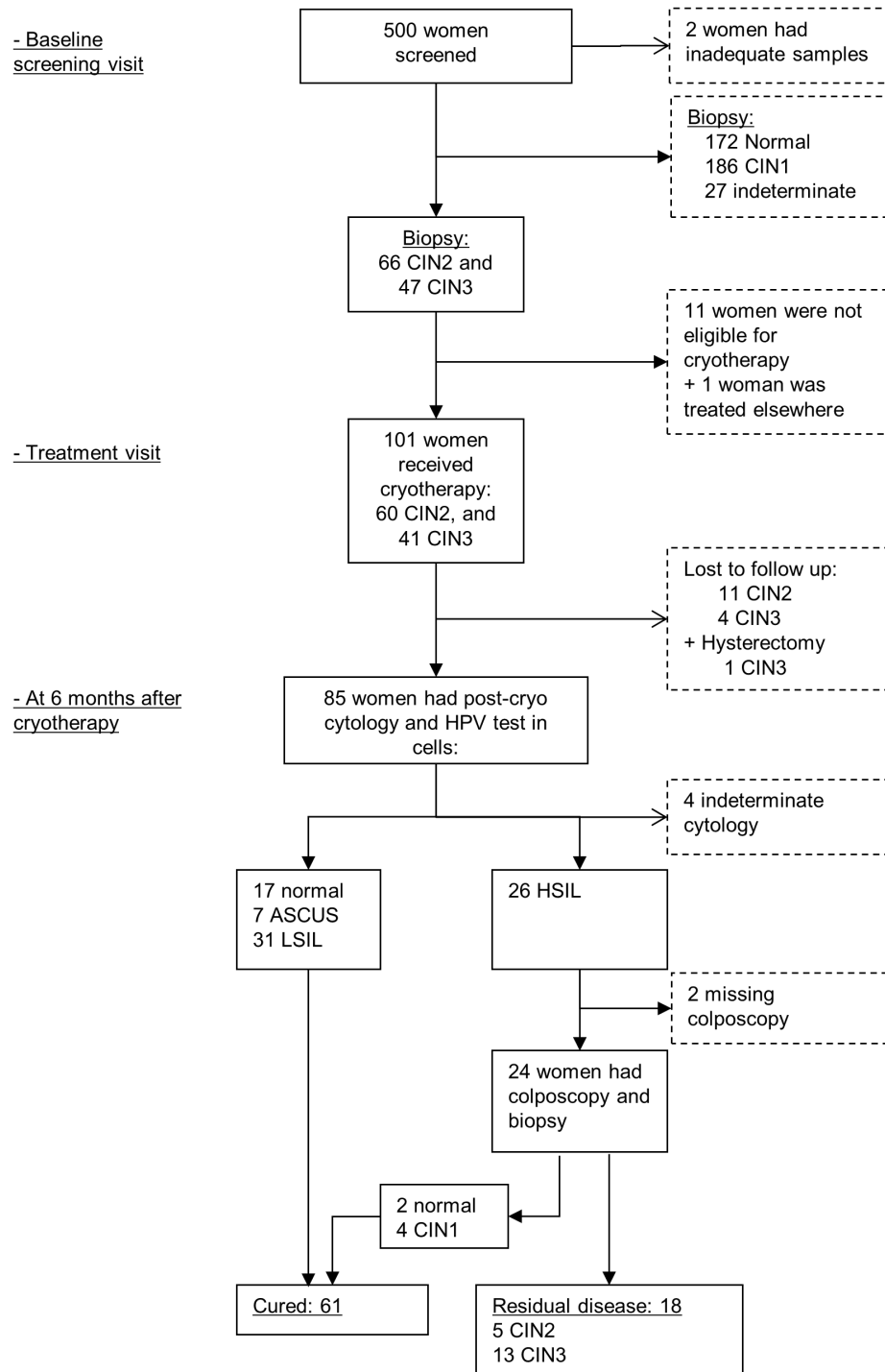


Figure 1. Flowchart study population. HPV, human papillomavirus; CIN, cervical intraepithelial neoplasia; cryo, cryotherapy; ASCUS, atypical squamous cells of undetermined significance; LSIL, low-grade squamous intraepithelial lesion; HSIL, high-grade squamous intraepithelial lesion. doi:10.1371/journal.pone.0111037.g001

Methods

Participants and study procedures

In 2009, 500 HIV-positive women in Nairobi, Kenya were enrolled in a study that compared cervical cancer screening with conventional cytology, visual inspection with acetic acid (VIA) and HPV testing, as described elsewhere [17–19]. Briefly, women who attended the Coptic Hope Center for Infectious Diseases for HIV-

related conditions were invited to participate and were eligible if they: 1) were between 18 and 55 years of age; 2) had not undergone cervical screening in the last year; and 3) had never been treated for cervical cancer or pre-cancerous lesions. After obtaining a written informed consent, information on women's characteristics and the use of combined antiretroviral treatment (cART) as well as blood samples to measure CD4 count were collected. Cervical exfoliated cells (further on referred to as "cells")

Table 1. Odds ratio (OR) and 95% confidence intervals (CI) for residual disease six months after cryotherapy (cryo) for CIN2/3 by selected pre-treatment characteristics.¹Kenya, 79 HIV-positive women.

Pre-cryo characteristics	Total N	Cured (≤LSIL/CIN1) n (%)	Residual disease (CIN2/3) n (%)	OR ² (95% CI)	OR ³ (95% CI)
Total	79	61 (77.2)	18 (22.8)		
Age					
<35	19	17 (89.5)	2 (10.5)	1	1
35–44	39	28 (71.8)	11 (28.2)	3.3 (0.7–16.9)	3.3 (0.6–19.4)
≥45	21	16 (76.2)	5 (23.8)	2.7 (0.5–15.7)	2.2 (0.3–14.7)
χ^2_1 for trend				$p = 0.340$	$p = 0.543$
Parity					
0–1	21	19 (90.5)	2 (9.5)	1	
2	23	18 (78.3)	5 (21.7)	2.2 (0.4–13.6)	
≥3	35	24 (68.6)	11 (31.4)	3.5 (0.6–19.8)	
χ^2_1 for trend				$p = 0.153$	
CD4 cells/μL at baseline					
>500	16	12 (75.0)	4 (25.0)	1	
200–500	45	35 (77.8)	10 (22.2)	1.0 (0.3–4.0)	
<200	18	14 (77.8)	4 (22.2)	1.1 (0.2–6.0)	
χ^2_1 for trend				$p = 0.891$	
CD4 count/μL at 6-month follow-up					
≥500	22	16 (72.7)	6 (27.3)	1.0	
200–500	47	37 (78.7)	10 (21.3)	0.7 (0.2–2.6)	
<200	10	8 (80.0)	2 (20.0)	0.8 (0.1–5.9)	
χ^2_1 for trend				$p = 0.750$	
cART					
No	18	16 (88.9)	2 (11.1)	1	
<2 years	37	28 (75.7)	9 (24.3)	2.1 (0.4–11.7)	
≥2 years	24	17 (70.8)	7 (29.2)	2.5 (0.4–15.3)	
χ^2_1 for trend				$p = 0.343$	
VIA					
Neg	33	28 (84.9)	5 (15.2)	1	
Pos	46	33 (71.7)	13 (28.3)	2.6 (0.8–8.6)	
Cytology⁴					
Normal/ASCUS/LSIL	25	23 (92.0)	2 (8.0)	1	
HSIL or worse	52	36 (69.2)	16 (30.8)	5.2 (1.1–25.0)	
Histology					
CIN2	45	40 (88.9)	5 (11.1)	1	1
CIN3	34	21 (61.8)	13 (38.2)	5.3 (1.6–17.4)	4.3 (1.2–15.0)
hrHPV (cells)					
Neg	18	18 (100)	0	1	
Pos	61	43 (70.5)	18 (29.5)	∞ (1.9 - ∞)	
HPV16 (cells)					
Neg	64	53 (82.8)	11 (17.2)	1	1
Pos	15	8 (53.3)	7 (46.7)	3.9 (1.1–13.1)	2.2 (0.6–8.1)

Table 1. Cont.

Pre-cryo characteristics	Total N	Cured (≤LSIL/CIN1) n (%)	Residual disease (CIN2/3) n (%)	OR ² (95% CI)	OR ³ (95% CI)
Multiple hrHPV⁵ (cells)					
No	38	25 (65.8)	13 (34.2)	1	
Yes	23	18 (78.3)	5 (21.7)	0.6 (0.2–1.9)	

¹Unless otherwise specified;

²Adjusted for age as appropriate;

³Among 61 hrHPV-positive women only. Adjusted for age, histology and HPV16 as appropriate;

⁴Do not add up to the total because of 2 indeterminate cytology results;

⁵Among hrHPV-positive women only.

cART, combination antiretroviral therapy; VIA, visual inspection with acetic acid; hrHPV, high-risk human papillomavirus; ASCUS, atypical squamous cells of undetermined significance; LSIL, low-grade squamous intraepithelial lesion; CIN, cervical intraepithelial neoplasia.

doi:10.1371/journal.pone.0111037.t001

were obtained using a Cervex-Brush (Rovers Medical Devices, Oss, The Netherlands) and placed in PreservCyt media (Hologic, Marlborough, MA, USA) for HPV testing. A medical doctor performed a colposcopic examination and took a biopsy from all women, either from the most abnormal area on the cervix identified by the colposcopic examination, or at 12 o'clock if no lesion was visualized. Biopsy tissues were immediately immersed in 10% buffered formalin and transported to the local pathology laboratory, where they were embedded in paraffin. Biopsy tissues and PreservCyt media were stored at ambient temperature and shipped to the Department of Pathology at Vrije University Medical Center (Amsterdam, the Netherlands) for HPV DNA testing.

Women who were diagnosed with CIN2/3 disease by biopsy were offered cryotherapy if the lesion was: 1) <75% of the cervix surface and not larger than the cryoprobe tip; and 2) entirely visible and not extending by more than 2 to 3 mm into the endocervical canal [20]. A follow-up visit was planned at 6 months after cryotherapy. It included the collection of cells for conventional cytology and HPV testing and a blood sample for CD4 count. Women with high-grade squamous intraepithelial lesions (HSIL) at cytology were offered a colposcopic examination including a biopsy of the residual lesion. All cytological slides and biopsies were processed under the supervision of the study pathologist (FSR) at the Aga Khan University (Nairobi), who also

read all of the cytological and histological slides. Cytology was reported according to the Bethesda 1991 revised classification [21].

Ethics Statement

The study protocol was approved by the Ethical Review Committees of the Kenyatta National Hospital, Kenya, the University of Washington, the United States, and the International Agency for Research on Cancer, France. The study conformed to the Helsinki Declaration of 1975, as revised in 2000. Each participant gave their written consent.

HPV DNA testing

HPV DNA testing was done on pre-cryotherapy cells and tissue biopsies and exclusively on cells for the post-cryotherapy visit. Biopsies were sectioned using a 'sandwich' approach, whereby inner sections were used for HPV testing and outer sections for histological examination. Testing methods have been described elsewhere [17,22]. Briefly, detection of HPV DNA was done by GP5+/6+-PCR followed by enzyme immunoassay detection with a cocktail of oligonucleotide probes [23]. Subsequent HPV typing was performed by reverse-line blot hybridization of PCR products [24]. One pre-cryotherapy sample was negative for beta-globin, but positive for hrHPV DNA, and it was included. HPV types 16,

Table 2. Presence of hrHPV after cryotherapy (cryo) by type of pre-cryo sample in women who were hrHPV-positive before cryotherapy.

Post-cryo hrHPV status	Pre-cryo hrHPV-positive women	
	Cells n (%)	Biopsy n (%)
Cells		
Persistent, HPV16	9 (14.8)	6 (12.2)
Persistent, other hr types¹	40 (65.6)	32 (65.3)
New type(s) only²	5 (8.2)	6 (12.2)
Negative	7 (11.5)	5 (10.2)
Total	61	49 ³

Kenya, 61 HIV-positive women.

¹Persistence of at least one hr type but not HPV16.

²In the absence of persistent hr types.

³Does not include 12 women who were hrHPV-positive in cells but negative in biopsy.

hrHPV, high-risk human papillomavirus.

doi:10.1371/journal.pone.0111037.t002

Table 3. Crude odds ratio (OR) and 95% confidence intervals (CI) for residual disease six months after cryotherapy (cryo) for CIN2/3 by persistence of hrHPV infections detected in pre-cryo cells or biopsies.

Persistence of hrHPV	Total n	Cured (\leq LSIL/CIN1) n (%)	Residual disease (CIN2/3) n (%)	OR (95% CI)
Pre-cryo cells				
No	12	11 (25.6)	1 (5.6)	1
Yes	49	32 (74.4)	17 (94.4)	5.8 (0.7–49.2)
Pre-cryo biopsies¹				
No	11	10 (32.3)	1 (5.6)	1
Yes	38	21 (67.7)	17 (94.4)	8.1 (0.9–69.7)

Kenya, 61 HIV-positive women.

¹Does not include 12 women who were hrHPV-positive in cells but negative in biopsy.

hrHPV, high-risk human papillomavirus; LSIL, low-grade squamous intraepithelial lesion; CIN, cervical intraepithelial neoplasia.

doi:10.1371/journal.pone.0111037.t003

18, 31, 33, 35, 39, 45, 51, 52, 56, 58, 59, and 68 were considered hr types [25].

Statistical analysis

Women with either histologically confirmed CIN2 or CIN3 were classified as having residual disease; women with either cytology \leq low-grade squamous intraepithelial lesion (LSIL) or biopsy \leq CIN1 were classified as cured. Women's CD4 counts at baseline and at 6-month follow-up were compared using the Wilcoxon signed-rank test.

Age-adjusted odds ratios (ORs) for the presence of residual disease and the corresponding 95% confidence intervals (CIs) were computed. None of the hrHPV-negative women had residual disease. A multiple logistic regression model including age and significant residual disease risk factors was therefore restricted to hrHPV-positive women. In addition, cytology and histology at baseline were closely correlated and we chose to include only histology. Tests for linear trend of ORs were computed giving increasing scores to each level of the categorized variable and fitting them into the model as continuous variables.

The persistence of HPV16, hrHPV types other than HPV16, and the appearance of new hrHPV types were assessed by comparing hrHPV findings from cells or biopsies collected at the pre-cryotherapy visit with those in cells at the post-cryotherapy visit. Crude ORs for residual disease by hrHPV persistence after cryotherapy were computed. Finally, we evaluated the performance of hrHPV testing at 6 months after cryotherapy (sensitivity, specificity, positive predictive value, PPV, and negative predictive value, NPV) for the detection of residual CIN2/3.

Results

Of the 500 women who were enrolled in our study, 498 had an adequate cell sample (Figure 1). CIN2 and CIN3 were detected in 66 and 47 women, respectively, and 101 received cryotherapy 30 days on average after inclusion in the study. A follow-up visit was scheduled at 6 months after cryotherapy (median: 182 days, Interquartile range, IQR: 165–197). Fifteen women (11 CIN2 and 4 CIN3) did not attend the follow-up visit and one woman with CIN3 underwent hysterectomy. Out of the remaining 85 women, 81 had a valid cytology and 26 had HSIL. All but two women with HSIL underwent colposcopic examination and biopsy collection. Among 18 women with residual disease, 5 had CIN2 and 13 CIN3. Only 17 women had negative cytology at the follow-up

visit. Sixty-one women who had either cytology \leq LSIL or biopsy \leq CIN1 were considered cured (Figure 1).

The median age of 79 women eventually included in the present report was 41 years (IQR: 35–45). At baseline, 61 (77.2%) of them were using cART, and 6 had started cART between the baseline and follow-up visit. Median CD4 count was lower at baseline (322 cells/ μ L, IQR: 205–477) than at the follow-up visit (386, IQR: 272–558) ($p = 0.007$).

Table 1 shows OR for residual CIN2/3 by selected baseline characteristics. Cryotherapy eliminated CIN2/3 in 77.2% (95% CI: 66.4–85.9) of all women (Table 1). The presence of HSIL cytology (OR *vs* \leq LSIL = 5.2; 95% CI: 1.1–25.0); CIN3 in biopsy (OR *vs* CIN2 = 5.3; 95% CI: 1.6–17.4); infection with hrHPV (OR = ∞ ; 95% CI: 1.9– ∞), and HPV16 (OR = 3.9; 95% CI: 1.1–13.1) in cells at baseline were significantly associated with residual disease in the age-adjusted models. Parity, CD4 cell count, duration of cART use, diagnosis at VIA, and multiple hrHPV infections at baseline (37.7% of hrHPV-positive women) were unrelated to residual disease. Residual disease was only found among 61 women who were hrHPV-positive at baseline and, among them, 70.5% (95% CI: 57.4–81.5%) were cured. The presence of CIN3 instead of CIN2 at baseline was significantly associated with residual disease (OR *vs* CIN2 = 4.3; 95% CI: 1.2–15.0) after adjustment for age and HPV16 infection (Table 1).

After cryotherapy, hrHPV infection in cells was detected in 56 (70.9%) women, of whom 22 (39.3%) had multiple hrHPV types, and 2 were new infections in women who were previously hrHPV-negative. A comparison of hrHPV findings at baseline and 6 months after cryotherapy among women who were initially hrHPV-positive in cells ($n = 61$) or biopsies ($n = 49$) is shown in Table 2. Persistence of at least one hrHPV type in cells was detected in 80.4% of women, including persistence of HPV16 in 14.8%. New hrHPV types, in the lack of persistent types, were detected in 5 women whereas 7 (11.5%) had become hrHPV-negative. Similar proportions of persistence were found when the comparison was based on hrHPV types detected in pre-treatment CIN2/3 biopsies (Table 2). The persistence of HPV16 or other hrHPV types was not influenced by CD4 count or duration of cART use (data not shown).

Table 3 shows OR for residual disease according to the presence of persistent hrHPV infection after cryotherapy. Women with persistent infection showed an increased risk of residual disease. ORs were 5.8 (95% CI: 0.7–49.2); and 8.1 (95% CI: 0.9–69.7) for persistence of types previously detected in baseline cells or biopsies, respectively. The level of persistence of hrHPV infections

Table 4. Performance of hrHPV testing for the detection of CIN2/3 six months after cryotherapy.

Post-treatment	TP	FP	FN	TN	Sensitivity	Specificity	PPV	NPV
hrHPV	17	39	1	22	0.94 (0.73–1.00)	0.36 (0.24–0.49)	0.30 (0.19–0.44)	0.96 (0.78–1.00)
HPV16	7	6	11	55	0.39 (0.17–0.64)	0.90 (0.80–0.96)	0.54 (0.25–0.81)	0.83 (0.72–0.91)
HPV18	4	8	14	53	0.22 (0.06–0.48)	0.87 (0.76–0.94)	0.33 (0.10–0.65)	0.79 (0.67–0.88)
HPV16/18	11	14	7	47	0.61 (0.36–0.83)	0.77 (0.65–0.87)	0.44 (0.24–0.65)	0.87 (0.75–0.95)
Persistent hrHPV ¹	17	32	1	29	0.94 (0.73–1.00)	0.48 (0.35–0.61)	0.35 (0.22–0.50)	0.97 (0.83–1.00)

Kenya, 79 HIV-positive women.

¹At least 1 same hrHPV type present before and 6 months after cryotherapy.

TP, true positive; FP, false positive; FN, false negative; TN, true negative; PPV, positive predictive value, hrHPV, high-risk human papillomavirus; NPV, negative predictive value, hrHPV, high-risk human papillomavirus. doi:10.1371/journal.pone.0111037.t004

was unrelated to the level of CD4 count or the use of cART (data not shown). None of the five women with new hrHPV types in the lack of persistent infection showed residual disease (data not shown).

The performance of post-treatment hrHPV testing to detect residual disease is shown in Table 4. The sensitivity and specificity of hrHPV testing at 6 months after cryotherapy were 0.94 and 0.36, respectively; PPV and NPV were 0.30 and 0.96, respectively. Testing for either or both HPV16 and 18 showed lower sensitivity and NPV than hrHPV testing, but better specificity and PPV (except for HPV18 only). Persistent hrHPV infection showed sensitivity, specificity, PPV and NPV of 0.94, 0.48, 0.35 and 0.97, respectively.

Discussion

Our study on the efficacy of cryotherapy for the treatment of CIN2/3 in HIV-positive women is one of the few that included cytological and histological evaluation prior to and after treatment and information on treatment outcome by persistence of individual hrHPV types. Cryotherapy eliminated CIN2/3 in 77% (95% CI: 66–86) of all women and 71% (95% CI: 57–82%) of women who had hrHPV infection before treatment, but only a small minority of them became hrHPV-negative. Contrary to others [8,11,13,26], we chose to define as cure the elimination of CIN2/3 rather than the disappearance of any cytological abnormalities. If we had used normal cytology as threshold, only 19 (24.1%) of our study women would have been considered cured. However, on account of the difficulty to eliminate HPV infection and its morphological correlate (low-grade lesions) in HIV-positive women, we reasoned that the cure of CIN2/3 had to be considered the most important outcome.

An initial diagnosis of CIN3, instead of CIN2, was the most important predictor of residual disease. HPV16-positivity at baseline was not significantly associated with residual disease after adjustment for the presence of CIN3. Infections with multiple hrHPV types were relatively frequent (38%) in study women but they did not increase the probability of residual disease as expected since cancer progression-prone CIN2/3 are assumed to be monoclonal [27,28]. No residual disease was found among 18 women who were initially hrHPV-negative.

Some [8,11,13,26], but not all studies [3,9,12], reported that cART treatment or high CD4 counts were favourable prognostic factors for treatment outcome. In our cross-sectional study, neither cART use nor CD4 counts prior to or after cryotherapy were associated with residual disease. Unfortunately, we did not have systematic information on CD4 count prior to cART. However, cART tended to be started relatively late in Kenya in 2009 (i.e., CD4 count <250 cells/ μ L or presence of WHO clinical III/IV stage disease) [29], when CIN2/3 may have already become refractory to immune reconstitution [17].

Our study confirms that residual disease after excisional treatment or cryotherapy of CIN2/3 may be more frequent among HIV-positive women than HIV-negative women. A study from Zimbabwe [6] reported residual CIN2/3 at 6 months after cryotherapy or LEEP in 3.8% and 1.8% of 109 HIV-positive women, respectively, but in none of 38 HIV-negative women. A few additional reports mainly from high-income countries included only HIV-positive women and consistently showed the difficulty in achieving cure of CIN2/3 and avoiding long-term recurrences in HIV-positive women. Tebeu et al [30] reviewed early studies on the outcome of conization for any degree of CIN in HIV-positive women at 6-to-74 months after treatment. Recurrences ranged between 20% and 75% but residual CIN1

was not separated from CIN2/3. Heard et al [11] reported that 54% of 75 HIV-positive women treated with LEEP or conization in France had a CIN recurrence at 36 months, and approximately one third had CIN2/3. Massad et al [26] reported residual CIN2/3 disease in 12% of 115 mostly HIV-positive women mainly treated with excisional methods in the United States. Reimers et al [3] detected residual CIN2/3 in 42% of 75 HIV-positive women at 6 months after LEEP or CKC.

Rates of residual disease and recurrences in HIV-negative women after CIN2/3 treatment were reported in many studies [31,32]. One study of 610 women from the United States showed residual CIN2/3 at 6 months after LEEP in 3% of women, and 7% after 2 years [4]. In other studies in which excisional methods were used for CIN2/3 treatment, residual disease ranged between 1% to 8% at 6 months [33,34], and between 2% and 20% at 2 years [33,35,36] or more [37]. A few studies from India [38,39] and Africa [6] showed persistent or recurrent diseases in 13% or less of HIV-negative women at 2 years or more after cryotherapy.

The difference in hrHPV clearance between HIV-positive and HIV-negative women after CIN2/3 treatment is much larger than the difference in the frequency of residual disease. Positivity for hrHPV after LEEP was shown in 57% of HIV-positive women with any degree of CIN by Gingelmaier et al [10] in Germany. The proportion of HIV-negative women who were hrHPV-positive at 6 months after treatment in PCR-based studies was 10 to 37% [4,37,40–42], except in studies that used ultra-sensitive HPV tests [33,36]. Type-specific hrHPV persistence in our study (80%) was also larger than that reported after cryotherapy or excisional treatment among HIV-negative women (7-to-37%) [4,40,43]. The association between persistent hrHPV infection and residual disease in our study was especially strong (8-fold increase, of borderline statistical significance) when the persistent type was the one previously detected in the CIN2/CIN3 biopsy.

A positive association between persistent HPV infection and residual or recurrent CIN2/3 was shown in HIV-negative women [4,37,41,44] lending support to the use of HPV testing to detect treatment failure [31]. In fact, HPV testing was shown to be more sensitive and not significantly less specific than cytology screening in a large meta-analysis that excluded verification bias [31]. The sensitivity of hrHPV testing in our study (94%) was similar to the corresponding sensitivity in HIV-negative women (92%) [31] but specificity (36%) was much lower than among HIV-negative women (76%) [31]. Sensitivity did not change but specificity improved to 48% when we used persistent hrHPV infection instead of hrHPV-positivity after cryotherapy. Testing for HPV16 and/or 18 showed a specificity of 77% but a sensitivity of only 61%. An equally valid evaluation of the performance of cytology

testing in HIV-positive women was impossible in our study due to verification bias, as only women with HSIL cytology received colposcopic examination.

Limitations of our study include the lack of random biopsies at follow-up visit and the short follow-up. A 6-month follow-up prevented the evaluation of long-term recurrences after cryotherapy in HIV-positive women and lends support to the interpretation of treatment failures as residual disease. Infections with new hrHPV types were seldom detected. We were also unable to compare the efficacy of cryotherapy to excisional methods as they were used only exceptionally. However, a meta-analysis [32] did not show systematic differences across seven different techniques of CIN treatment, including cryotherapy, in HIV-negative women. Strengths of our study include the use of high-quality cytological and histological examination and a clinically validated HPV testing method [45] on pre- and post-treatment cells and pre-treatment CIN2/3 biopsies.

In conclusion, we demonstrated that cryotherapy eliminated three quarters of CIN2/3, but only one fifth of hrHPV infections in HIV-positive women. HIV-positive women should be, therefore, carefully monitored after cryotherapy as they often remain cytologically abnormal and hrHPV-positive. In fact, the value of testing for hrHPV infection to detect residual CIN2/3 after cryotherapy was hampered by the lower specificity of the test than in HIV-negative women. The clinical implications of low hrHPV clearance will require further study and so will the assessment of the possible influence of long-term history of cART use and CD4 count on recurrence rates.

Acknowledgments

We thank the research personnel, clinic and laboratory staff, and data management teams in Nairobi, Kenya; Seattle, USA; Amsterdam, The Netherlands; and Lyon, France for their work. We are especially grateful to Dr. Evans Nyongesa-Malava for his commitment as study physician. We recognize the Coptic Hope Center for Infectious Diseases, Nairobi, Kenya, for their cooperation and our patients for their participation and support.

Dr Farzana S. Rana, our colleague and co-author, sadly passed away before the publication of this manuscript. We dedicate this manuscript to her memory. Dr Farzana S. Rana was a great scientist, collaborator and mentor on this project.

Author Contributions

Conceived and designed the experiments: MHC HDV NRM SF. Performed the experiments: FSR PJFS. Analyzed the data: HDV VT. Contributed reagents/materials/analysis tools: VT. Wrote the paper: HDV MHC SF. Data collection: MHC NRM KMJN SRS.

References

1. Clifford GM, Goncalves MA, Franceschi S, for the HPV and HIV Study Group (2006) Human papillomavirus types among women infected with HIV: a meta-analysis. *AIDS* 20: 2337–2344.
2. De Vuyst H, Lillo F, Broutet N, Smith JS (2008) HIV, human papillomavirus, and cervical neoplasia and cancer in the era of highly active antiretroviral therapy. *Eur J Cancer Prev* 17: 545–554.
3. Reimers LL, Sotardi S, Daniel D, Chiu LG, Van AA, et al. (2010) Outcomes after an excisional procedure for cervical intraepithelial neoplasia in HIV-infected women. *Gynecol Oncol* 119: 92–97.
4. Kreimer AR, Guido RS, Solomon D, Schiffman M, Wacholder S, et al. (2006) Human papillomavirus testing following loop electrosurgical excision procedure identifies women at risk for posttreatment cervical intraepithelial neoplasia grade 2 or 3 disease. *Cancer Epidemiol Biomarkers Prev* 15: 908–914.
5. Kocken M, Berkhof J, van Kemenade FJ, Louwers JA, Zaal A, et al. (2012) Long-term CIN3+ risk in women with abnormal cytology; role of hrHPV testing. *Br J Cancer* 106: 817–825.
6. Chirenje ZM, Rusakaniko S, Akino V, Munjoma M, Mlingo M (2003) Effect of HIV Disease in Treatment Outcome of Cervical Squamous Intraepithelial Lesions Among Zimbabwean Women. *J Low Genit Tract Dis* 7: 16–21.
7. Foulot H, Heard I, Potard V, Costagliola D, Chapron C (2008) Surgical management of cervical intraepithelial neoplasia in HIV-infected women. *Eur J Obstet Gynecol Reprod Biol* 141: 153–157.
8. Fruchter RG, Maiman M, Sedlis A, Bartley L, Camilien L, et al. (1996) Multiple recurrences of cervical intraepithelial neoplasia in women with the human immunodeficiency virus. *Obstet Gynecol* 87: 338–344.
9. Gilles C, Manigart Y, Konopnicki D, Barlow P, Rozenberg S (2005) Management and outcome of cervical intraepithelial neoplasia lesions: a study of matched cases according to HIV status. *Gynecol Oncol* 96: 112–118.
10. Gingelmaier A, Grubert T, Kaestner R, Mylonas I, Weissenbacher T, et al. (2007) High recurrence rate of cervical dysplasia and persistence of HPV infection in HIV-1-infected women. *Anticancer Res* 27: 1795–1798.
11. Heard I, Potard V, Foulot H, Chapron C, Costagliola D, et al. (2005) High rate of recurrence of cervical intraepithelial neoplasia after surgery in HIV-positive women. *J Acquir Immune Defic Syndr* 39: 412–418.
12. Holcomb K, Matthews RP, Chapman JE, Abulafia O, Lee YC, et al. (1999) The efficacy of cervical conization in the treatment of cervical intraepithelial neoplasia in HIV-positive women. *Gynecol Oncol* 74: 428–431.

13. Tate DR, Anderson RJ (2002) Recrudescence of cervical dysplasia among women who are infected with the human immunodeficiency virus: a case-control analysis. *Am J Obstet Gynecol* 186: 880–882.
14. Cejtin HE, Malapati R, Chaparala S (2011) A comparison of loop electrosurgical excision procedures between human immunodeficiency virus-seropositive and -seronegative women. *J Low Genit Tract Dis* 15: 37–41.
15. Lehtovirta P, Paavonen J, Heikinheimo O (2008) Risk factors, diagnosis and prognosis of cervical intraepithelial neoplasia among HIV-infected women. *Int J STD AIDS* 19: 37–41.
16. Santesso N, Schunemann H, Blumenthal P, De Vuyst H, Gage J, et al. (2012) World Health Organization Guidelines: Use of cryotherapy for cervical intraepithelial neoplasia. *Int J Gynaecol Obstet* 118: 97–102.
17. De Vuyst H, Mugo NR, Chung MH, McKenzie KP, Nyongesa-Malava E, et al. (2012) Prevalence and determinants of human papillomavirus infection and cervical lesions in HIV-positive women in Kenya. *Br J Cancer* 107: 1624–1630.
18. Chung MH, McKenzie KP, Richardson BA, John-Stewart GC, Coombs RW, et al. (2011) Cervical HIV-1 RNA shedding after cryotherapy among HIV-positive women with cervical intraepithelial neoplasia stage 2 or 3. *AIDS* 25: 1915–1919.
19. Chung MH, McKenzie KP, De Vuyst H, Richardson BA, Rana F, et al. (2013) Comparing Papanicolaou smear, visual inspection with acetic acid and human papillomavirus cervical cancer screening methods among HIV-positive women by immune status and antiretroviral therapy. *AIDS* 27: 2909–19.
20. Sellors, J W. and Sankaranarayanan R. (2003) Colposcopy and treatment of cervical intraepithelial neoplasia : a beginner's manual. Lyon: International Agency for Research on Cancer.
21. Luff RD (1992) The Bethesda System for reporting cervical/vaginal cytologic diagnoses. Report of the 1991 Bethesda workshop. *Am J Clin Pathol* 98: 152–154.
22. De Vuyst H, Chung MH, Baussano I, Mugo NR, Tenet V, et al. (2013) Comparison of HPV DNA testing in cervical exfoliated cells and tissue biopsies among HIV-positive women in Kenya. *Int J Cancer* 133: 1441–1446.
23. Jacobs MV, Walboomers JM, Snijders PJ, Voorhorst FJ, Verheijen RH, et al. (2000) Distribution of 37 mucosotropic HPV types in women with cytologically normal cervical smears: the age-related patterns for high-risk and low-risk types. *Int J Cancer* 87: 221–227.
24. van den Brule AJ, Pol R, Fransen-Daalmeijer N, Schouls LM, Meijer CJ, et al. (2002) GP5+/6+ PCR followed by reverse line blot analysis enables rapid and high-throughput identification of human papillomavirus genotypes. *J Clin Microbiol* 40: 779–787.
25. Schiffman M, Clifford G, Buonaguro FM (2009) Classification of weakly carcinogenic human papillomavirus types: addressing the limits of epidemiology at the borderline. *Infect Agent Cancer* 4: 8.
26. Massad LS, Fazzari MJ, Anastos K, Klein RS, Minkoff H, et al. (2007) Outcomes after treatment of cervical intraepithelial neoplasia among women with HIV. *J Low Genit Tract Dis* 11: 90–97.
27. Quint W, Jenkins D, Molijn A, Struijk L, van de SM, et al. (2012) One virus, one lesion—individual components of CIN lesions contain a specific HPV type. *J Pathol* 227: 62–71.
28. van der MJ, Quint WG, Schiffman M, van de Sandt MM, Zuna RE, et al. (2012) Molecular mapping of high-grade cervical intraepithelial neoplasia shows etiological dominance of HPV16. *Int J Cancer* 131: E946–E953.
29. Ojoo S (2007) Kenya national clinical manual for ART providers: a concise and practical guide to ART provision. Nairobi, Kenya: National AIDS and STI Control Program (NASCOP).
30. Tebeu PM, Major AL, Mhawech P, Rapiiti E (2006) The recurrence of cervical intraepithelial neoplasia in HIV-positive women: a review of the literature. *Int J STD AIDS* 17: 507–511.
31. Kocken M, Uijterwaal MH, de Vries AL, Berkhof J, Ket JC, et al. (2012) High-risk human papillomavirus testing versus cytology in predicting post-treatment disease in women treated for high-grade cervical disease: a systematic review and meta-analysis. *Gynecol Oncol* 125: 500–507.
32. Martin-Hirsch PP, Paraskevaidis E, Bryant A, Dickinson HO, Keep SL (2010) Surgery for cervical intraepithelial neoplasia. *Cochrane Database Syst Rev* CD001318.
33. Jones J, Saleem A, Rai N, Shylasree TS, Ashman S, et al. (2011) Human Papillomavirus genotype testing combined with cytology as a 'test of cure' post treatment: the importance of a persistent viral infection. *J Clin Virol* 52: 88–92.
34. Nobbenhuis MA, Meijer CJ, van den Brule AJ, Rozendaal L, Voorhorst FJ, et al. (2001) Addition of high-risk HPV testing improves the current guidelines on follow-up after treatment for cervical intraepithelial neoplasia. *Br J Cancer* 84: 796–801.
35. Alonso I, Torne A, Puig-Tintore LM, Esteve R, Quinto L, et al. (2006) Pre- and post-conization high-risk HPV testing predicts residual/recurrent disease in patients treated for CIN 2–3. *Gynecol Oncol* 103: 631–636.
36. Valasoulis G, Koliopoulos G, Founta C, Kyrgiou M, Tsoumpou I, et al. (2011) Alterations in human papillomavirus-related biomarkers after treatment of cervical intraepithelial neoplasia. *Gynecol Oncol* 121: 43–48.
37. Kocken M, Helmerhorst TJ, Berkhof J, Louwers JA, Nobbenhuis MA, et al. (2011) Risk of recurrent high-grade cervical intraepithelial neoplasia after successful treatment: a long-term multi-cohort study. *Lancet Oncol* 12: 441–450.
38. Nene BM, Hiremath PS, Kane S, Fayette JM, Shastri SS, et al. (2008) Effectiveness, safety, and acceptability of cryotherapy by midwives for cervical intraepithelial neoplasia in Maharashtra, India. *Int J Gynaecol Obstet* 103: 232–236.
39. Sankaranarayanan R, Rajkumar R, Esmay PO, Fayette JM, Shanthakumary S, et al. (2007) Effectiveness, safety and acceptability of 'see and treat' with cryotherapy by nurses in a cervical screening study in India. *Br J Cancer* 96: 738–743.
40. Bais AG, Eijkemans MJ, Rebolj M, Snijders PJ, Verheijen RH, et al. (2009) Post-treatment CIN: randomised clinical trial using hrHPV testing for prediction of residual/recurrent disease. *Int J Cancer* 124: 889–895.
41. Kitchener HC, Walker PG, Nelson L, Hadwin R, Patnick J, et al. (2008) HPV testing as an adjunct to cytology in the follow up of women treated for cervical intraepithelial neoplasia. *BJOG* 115: 1001–1007.
42. Strander B, Ryd W, Wallin KL, Warleby B, Zheng B, et al. (2007) Does HPV-status 6–12 months after treatment of high grade dysplasia in the uterine cervix predict long term recurrence? *Eur J Cancer* 43: 1849–1855.
43. Elfgrén K, Jacobs M, Walboomers JM, Meijer CJ, Dillner J (2002) Rate of human papillomavirus clearance after treatment of cervical intraepithelial neoplasia. *Obstet Gynecol* 100: 965–971.
44. Zielinski GD, Rozendaal L, Voorhorst FJ, Berkhof J, Snijders PJ, et al. (2003) HPV testing can reduce the number of follow-up visits in women treated for cervical intraepithelial neoplasia grade 3. *Gynecol Oncol* 91: 67–73.
45. Rijkaart DC, Berkhof J, Rozendaal L, van Kemenade FJ, Bulkman NW, et al. (2012) Human papillomavirus testing for the detection of high-grade cervical intraepithelial neoplasia and cancer: final results of the POBASCAM randomised controlled trial. *Lancet Oncol* 13: 78–88.