



## Using Pharmacokinetic and Viral Kinetic Modeling To Estimate the Antiviral Effectiveness of Telaprevir, Boceprevir, and Pegylated Interferon during Triple Therapy in Treatment-Experienced Hepatitis C Virus-Infected Cirrhotic Patients.

Cédric Laouénan, Patrick Marcellin, Martine Lapalus, Feryel Khelifa-Mouri, Nathalie Boyer, Fabien Zoulim, Lawrence Serfaty, Jean-Pierre Bronowicki, Michelle Martinot-Peignoux, Olivier Lada, et al.

### ► To cite this version:

Cédric Laouénan, Patrick Marcellin, Martine Lapalus, Feryel Khelifa-Mouri, Nathalie Boyer, et al.. Using Pharmacokinetic and Viral Kinetic Modeling To Estimate the Antiviral Effectiveness of Telaprevir, Boceprevir, and Pegylated Interferon during Triple Therapy in Treatment-Experienced Hepatitis C Virus-Infected Cirrhotic Patients.: Effectiveness of triple therapy in cirrhotic patients. *Antimicrobial Agents and Chemotherapy*, American Society for Microbiology, 2014, 58 (9), pp.5332-41. <10.1128/AAC.02611-14>. <inserm-01059165>

**HAL Id: inserm-01059165**

**<http://www.hal.inserm.fr/inserm-01059165>**

Submitted on 29 Aug 2014

**HAL** is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from

teaching and research institutions in France or abroad, or from public or private research centers.

L'archive ouverte pluridisciplinaire **HAL**, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d'enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.

1 **Using pharmacokinetic and viral kinetic modeling to estimate the antiviral effectiveness**  
2 **of telaprevir, boceprevir and Peg-IFN during triple therapy in treatment-experienced**  
3 **HCV infected cirrhotic patients (ANRS CO20-CUPIC)**

4

5 Cédric Laouénan,<sup>a,b,#</sup> Patrick Marcellin,<sup>c,d</sup> Martine Lapalus,<sup>c</sup> Feryel Khelifa-Mouri,<sup>d</sup> Nathalie  
6 Boyer,<sup>d</sup> Fabien Zoulim,<sup>e,f</sup> Lawrence Serfaty,<sup>g</sup> Jean-Pierre Bronowicki,<sup>h,i</sup> Michelle Martinot-  
7 Peignoux,<sup>c</sup> Olivier Lada,<sup>c</sup> Tarik Asselah,<sup>c,d</sup> Céline Dorival,<sup>j</sup> Christophe Hézode,<sup>k,l</sup> Fabrice  
8 Carrat,<sup>j,m</sup> Florence Nicot,<sup>n</sup> Gilles Peytavin,<sup>a,o</sup> France Mentré,<sup>a,b</sup> Jeremie Guedj<sup>a</sup>.

9

10 INSERM, IAME, UMR 1137, Univ Paris Diderot, Sorbonne Paris Cité, Paris, France<sup>a</sup>; AP-  
11 HP, Hôpital Bichat, Département of Biostatistic, Paris, France<sup>b</sup>; INSERM, CRI Paris  
12 Montmartre, UMR 1149, Univ Paris Diderot, Clichy, France<sup>c</sup>; AP-HP, Hôpital Beaujon,  
13 Hepatology, Physiopathology and Treatment of Viral Hepatitis, Clichy, France<sup>d</sup>; INSERM,  
14 UMR 1052, Univ Lyon, Lyon, France<sup>e</sup>; Hospices Civils de Lyon, Department of Hepatology,  
15 Lyon, France<sup>f</sup>; AP-HP, Hôpital Saint-Antoine, Department of Hepatology, Paris, France<sup>g</sup>;  
16 INSERM, UMR 954, Univ Lorraine, Vandoeuvre-les-Nancy, France<sup>h</sup>; Centre Hospitalier  
17 Universitaire de Nancy, Department of Hepatology, Vandoeuvre-les-Nancy, France<sup>i</sup>;  
18 INSERM, UMR 707, Univ Pierre et Marie Curie, Paris, France<sup>j</sup>; INSERM, UMR 955, Univ  
19 Paris-Est, Créteil, France<sup>k</sup>; AP-HP, Hôpital Henri Mondor, Department of Hepatology,  
20 Créteil, France<sup>l</sup>; AP-HP, Hôpital Saint-Antoine, Department of public health, Paris, France<sup>m</sup>;  
21 CHU Toulouse, IFB Purpan, Virology, laboratory, Toulouse, France<sup>n</sup>; AP-HP, Hôpital Bichat,  
22 Department of Clinical Pharmacokinetics, Paris, France<sup>o</sup>.

23

24 **Running Head:** Effectiveness of triple therapy in cirrhotic patients

25

26 # Address correspondence to Cédric Laouénan, [cedric.laouenan@inserm.fr](mailto:cedric.laouenan@inserm.fr)

27

28 **Word count:**

29 - Abstract: 243

30 - Article: 4685

31

32 **Abstract**

33 **Background** Triple therapy combining a protease inhibitor (PI) telaprevir or boceprevir,  
34 pegylated-interferon (Peg-IFN) and ribavirin (RBV) have dramatically increased the chance  
35 to eradicate hepatitis C virus (HCV). However the efficacy of this treatment remains  
36 suboptimal in cirrhotic experienced-patients. Here we aimed to better understand the origin of  
37 this impaired response by estimating the antiviral effectiveness of each drug.

38 **Methods** Fifteen genotype 1-patients with compensated cirrhosis, non-responders to a prior  
39 Peg-IFN/RBV therapy were enrolled in a non-randomized study. HCV-RNA and drug  
40 concentrations of PIs, Peg-IFN and RBV were frequently assessed in the first 12 weeks of  
41 treatment and were analyzed using a pharmacokinetics/viral kinetics model.

42 **Results** Both PIs achieved similar level of molar concentrations ( $P=0.5$ ), but there was a  
43 significant difference of  $EC_{50}$  ( $P=0.008$ ), leading to a larger antiviral effectiveness than  
44 boceprevir in blocking viral production (99.8% vs 99.0%, respectively,  $P=0.002$ ). In all  
45 patients the antiviral effectiveness of Peg-IFN was modest (43.4%) and there was no  
46 significant contribution of RBV exposure on the total antiviral effectiveness. The second  
47 phase of viral decline, which is attributed to the loss rate of infected cells, was slow ( $0.19 \text{ day}^{-1}$ )  
48 and was higher in patients that subsequently eradicated HCV ( $P=0.03$ ).

49 **Conclusion** Both PIs achieved a high level of antiviral effectiveness. However the suboptimal  
50 antiviral effectiveness of Peg-IFN/RBV and the low loss of infected cells suggest that longer  
51 treatment duration might be needed in cirrhotic treatment experienced-patients and that future  
52 IFN-free regimen may be particularly beneficial to these patients.

53

54 **Keywords:** Hepatitis C virus; Non-linear mixed effect models; Early viral kinetics; Protease  
55 inhibitor; Pegylated-interferon; Ribavirin; Mathematical modeling; Pharmacokinetic

56

57 **Introduction**

58 Chronic infection with hepatitis C virus (HCV) affects approximately 160 million people  
59 worldwide (1) and is the leading cause of cirrhosis, liver cancer and liver transplantation (2).  
60 The goal of treatment is to achieve a sustained virological response (SVR), marker of viral  
61 eradication, assessed by the absence of detectable HCV RNA six months after treatment  
62 discontinuation. The approval in 2011 of two protease inhibitors (PI), telaprevir and  
63 boceprevir, in combination with pegylated-interferon-alpha and ribavirin (Peg-IFN/RBV) (3),  
64 has marked an important milestone with SVR rates higher than 70% in HCV genotype 1  
65 infected patients (4, 5). Recently two new triple therapy involving sofosbuvir, a nucleoside  
66 polymerase inhibitor, and simeprevir, a new protease inhibitor, have been approved by the  
67 European and American regulatory agencies, showing in clinical trials even higher SVR rates  
68 of 90% (6). However the cost of these new treatments, about twice as much as telaprevir or  
69 boceprevir-based therapy (7), will make them out of reach for many countries. Therefore  
70 triple therapy with Peg-IFN, RBV and telaprevir/boceprevir will continue to be vastly used in  
71 the next years and will remain the only therapeutic option for many patients.

72 Although these results suggest that a functional cure might be obtained in a large majority of  
73 patients, one should keep in mind that issues remain. In particular the proportion of patients  
74 with advanced liver disease and cirrhosis and/or who had failed a previous treatment with  
75 Peg-IFN/RBV is under represented in the patient population in clinical trials (8–11). The  
76 evaluation of the triple therapy in this population was precisely the goal of the ANRS-CO20-  
77 CUPIC cohort (Compassionate Use of Protease Inhibitors in viral C Cirrhosis;  
78 ClinicalTrials.gov number: NCT01514890) (12), where 511 genotype 1 treatment-  
79 experienced cirrhotic patients were included. In this study the SVR rates 12 weeks after  
80 treatment discontinuation (SVR12) were equal to 52% and 43% in telaprevir and boceprevir  
81 treated patients, respectively (13). The origin of this impaired response might encompass a

82 variety of factors, in particular impaired drug pharmacokinetics (PK) or limited sensitivity to  
83 PI agents and/or Peg-IFN/RBV in this particular population.

84 One way to evaluate treatment antiviral effectiveness and to optimize therapy is to use PK-  
85 viral kinetic (VK) models that provide a useful tool to quantitatively describe the relationship  
86 between drug exposure and viral response (reviewed in (14)). However no such analysis has  
87 been published with boceprevir and results published for telaprevir were mostly based on  
88 treatment naive and/or non-cirrhotic patients (15–17).

89 Here, we aimed to get new insights into the determinants of the response to triple therapy by  
90 analyzing in details, within a subset of 15 patients enrolled in the ANRS-CO20-CUPIC study,  
91 the relationship between drug concentrations and early virological response. We used the  
92 techniques of PK-VK modeling in order to tease out the relative antiviral effectiveness of  
93 each of the agents involved in the triple therapy (*i.e.*, boceprevir or telaprevir, Peg-IFN and  
94 RBV) and to investigate for a possible association with long term virological response.

95

96 **Materials and methods**

97 **Patients and data**

98 MODCUPIC is a substudy of the French multicentre prospective ANRS-CO20-CUPIC  
99 cohort. In four centres, from September 2011 to September 2012, patients chronically  
100 monoinfected with HCV genotype 1, compensated cirrhosis (Child-Pugh class A), non-  
101 responders to a prior IFN-based therapy and who started triple therapy were recruited. The  
102 diagnosis of cirrhosis was made by liver biopsy or non-invasive tests, Fibrotest® or  
103 Fibroscan® or Fibrometer® or Hepascore® at the discretion of the investigator, according to  
104 the French recommendations (18). The choice between TVR- or BOC-based therapies was at  
105 the investigator's discretion without randomization. TVR-based therapy included 12 weeks of  
106 telaprevir (750 mg/8 hours) in combination with Peg-IFN- $\alpha$ 2a (180  $\mu$ g/week) and RBV (1,000  
107 or 1,200 mg/day, depending on body weight) then 36 weeks of Peg-IFN- $\alpha$ 2a/RBV (named  
108 group telaprevir in the following). BOC-based therapy included 4 weeks (lead-in phase) of  
109 Peg-IFN- $\alpha$ 2b (1.5  $\mu$ g/kg/week) or Peg-IFN- $\alpha$ 2a (180  $\mu$ g/week) and RBV (800 or 1,400  
110 mg/day, depending on body weight) then 44 weeks of Peg-IFN- $\alpha$ 2b/RBV and boceprevir (800  
111 mg/8 hours) (named group boceprevir in the following). Patients were followed up to six  
112 months after treatment discontinuation to assess SVR.

113 Written informed consent was obtained before enrolment. The protocol was conducted in  
114 accordance with the Declaration of Helsinki and was approved by the "Ile-de-France IX  
115 Ethics Committee" (Créteil, France).

116

117 **Bioanalytical methods**

118 HCV RNA and drug concentrations were measured post PIs initiation at hours 0, 8, days 1, 2,  
119 3 and weeks 1, 2, 3, 4, 8 and 12. Patients treated with boceprevir had two additional VL and  
120 concentrations measurements during the lead-in phase. Blood samples were collected early in



121 the morning before the first daily dose of PIs and RBV and therefore only trough pre-dose  
122 drug concentrations were collected. All samples were collected on SST (serum) vacutainers,  
123 kept at 4°C until centrifuged at 3,000 RPM for 10 minutes in a 4°C centrifuge, within 1 hour  
124 after collection, aliquoted and kept at -80°C until analysis.

125 PIs concentrations in serum were determined using ultra-performance liquid chromatography  
126 coupled with tandem mass spectrometry with a lower limit of quantification (LOQ) of 5 ng/ml  
127 and 10 ng/ml for boceprevir and telaprevir, respectively (19). PI concentrations were  
128 converted to  $\mu\text{mol/l}$  for analysis using molar masses of 519.68 g/mol and 679.85 g/mol for  
129 boceprevir and telaprevir, respectively. RBV concentrations in serum were determined using  
130 ultra-performance liquid chromatography coupled with UV detection with a LOQ of 100  
131 ng/ml (20). Peg-IFN- $\alpha$ 2a and - $\alpha$ 2b in serum were determined with a bioassay which was  
132 chosen because the objective was to quantify the antiviral activity of Peg-IFN- $\alpha$  and not only  
133 the concentration. Immunoassay measures the physical quantity of material but does not  
134 differentiate between active and inactive molecules while bioassay for IFN- $\alpha$  is based on the  
135 protection of cultured cells against the cytopathic effect of a challenge virus and also was  
136 suitable for assaying both Peg-IFN- $\alpha$ -2a and Peg-IFN- $\alpha$ -2b. The reference solutions  
137 contained 2.8–180 ng/ml of Peg-IFN- $\alpha$ 2a (Roche Diagnostics, Germany) (21).

138 HCV-RNA levels were measured with a real-time PCR-based assay, Cobas®  
139 Ampliprep/Cobas TaqMan® assay (Roche Diagnostics, Germany), with a lower limit of  
140 detection (LOD) of 15 IU/ml. DNA samples were genotyped for the IL28B rs12979860  
141 polymorphism (AmpliTaq gold® DNA polymerase and BigDye® terminator cycle  
142 sequencing kit, Applied Biosystems, UK).

143

144 **Drug pharmacokinetic modeling**

145 All drug concentrations were fitted separately in telaprevir and boceprevir treatment groups.  
 146 For both Peg-IFN and RBV, the trough serum concentrations, noted  $C^{Peg-IFN}(t)$  and  $C^{RBV}(t)$ ,  
 147 respectively were fitted using an exponential model to reflect the progressive increase in  
 148 trough drug concentrations over time:

$$149 \quad C^{Peg-IFN}(t) = C_{ss}^{Peg-IFN} \times (1 - e^{-kt}) \quad \text{Eq. (1)}$$

$$150 \quad C^{RBV}(t) = C_{ss}^{RBV} \times (1 - e^{-kt}) \quad \text{Eq. (2)}$$

151 where  $C_{ss}$  is the trough concentration at steady state and  $k$  the rate constant of elimination  
 152 which reflects the progressive increase in  $C(t)$  over time.

153 For both PI drugs, consistent with the fact that they have a short elimination half-life (22), no  
 154 significant increase of trough concentrations over time was observed. Therefore  
 155 concentrations for both telaprevir and boceprevir were fitted using a constant model, where  
 156  $C_{ss}$  is the trough concentration:

$$157 \quad C^{PI}(t) = C_{ss}^{PI} \quad \text{Eq. (3)}$$

158

### 159 **Viral kinetic modeling**

160 The following model of HCV viral kinetics (VK) was used to fit the changes in HCV RNA  
 161 (23):

$$162 \quad \frac{dI}{dt} = bVT - \delta I$$

$$163 \quad \frac{dV}{dt} = p(1 - \varepsilon(t))I - cV \quad \text{Eq. (4)}$$

164 where  $T$  represent the target cells that can be infected by virus,  $V$ , with rate  $b$ . Infected cells,  $I$ ,  
 165 are lost with rate  $\delta$  and produce  $p$  virions per day, which are cleared from serum with rate  $c$ .  
 166 The target cell level is assumed constant throughout the study period (12 weeks) and remains  
 167 at its pre-treatment value  $T_0 = c\delta/p\beta$ . Treatment is assumed to reduce the average rate of viral  
 168 production per cell from  $p$  to  $p(1-\varepsilon)$ , where  $\varepsilon$  represents the drug antiviral effectivenesses, *i.e.*,  
 169  $\varepsilon = 0.99$  implying the drug is 99% effective in blocking viral production. This model predicts

170 that VL will fall in a biphasic manner, with a rapid first phase lasting for a couple of days that  
 171 reduce the VL with a magnitude equal to  $\log_{10}(1-\varepsilon)$ , followed by a second slower but  
 172 persistent second phase of viral decline with rate  $\varepsilon\delta$ . Therefore a difference between  $\varepsilon =$   
 173 99.9% and  $\varepsilon = 99.0\%$  corresponds to a 10-fold difference in the viral production under  
 174 treatment and will lead to 1-log difference between the two curves of viral decline (24). We  
 175 fixed  $p$  and  $b$  to 100 IU/ml/cell/day and  $10^{-7}$  (IU/ml)<sup>-1</sup>/day, respectively, without loss of  
 176 generality (25).

177 The effectiveness of each drug in blocking viral production was described by an  $E_{\max}$  model  
 178 assuming a maximum inhibition of 100%:

$$179 \quad \varepsilon^{PI}(t) = \frac{C^{PI}(t)}{C^{PI}(t) + EC_{50}^{PI}}$$

$$180 \quad \varepsilon^{Peg-IFN}(t) = \frac{C^{Peg-IFN}(t)}{C^{Peg-IFN}(t) + EC_{50}^{Peg-IFN}} \quad \text{Eq. (5)}$$

181 where  $EC_{50}^{PI}$  (respectively  $EC_{50}^{Peg-IFN}$ ) is the PI (resp. Peg-IFN) concentration at which the PI  
 182 (resp. Peg-IFN) is 50% effective, and  $C^{PI}(t)$  (resp.  $C^{Peg-IFN}(t)$ ) are the individual predictions  
 183 (see below) given by the PK models (Eq. 1 and 3).

184 The combined effect of PIs and Peg-IFN was modeled using a Bliss independent action model  
 185 (26) and the total efficacy  $\varepsilon(t)$  was given by:

$$186 \quad (1 - \varepsilon(t)) = (1 - \varepsilon^{PI}(t))(1 - \varepsilon^{Peg-IFN}(t)) \quad \text{Eq. (6)}$$

187 Since the effect of RBV on the early virological response is expected to be modest (27–29)  
 188 we did not incorporate the effect of RBV into the reference model (Eq. 4-6). In a second step  
 189 we tested whether the effectiveness of RBV, also modeled using an  $E_{\max}$  model could enhance  
 190 the effect in blocking viral production or reduce viral infectivity, as suggested previously (30).

191

## 192 **Data analysis and parameter estimation**

193 The pharmacokinetics/viral kinetics (PK-VK) model given by Eq. 4-6 can be used only to  
194 characterize the viral kinetics of drug sensitive virus and therefore cannot fit viral rebounds  
195 due to the emergence of drug-resistant virus. Therefore only HCV RNA data until virologic  
196 rebounds (with no indication of lack of compliance) were used to estimate the viral kinetic  
197 parameters.

198 Parameters  $V_0$ ,  $c$ ,  $\delta$ ,  $EC_{50}^{PI}$  and  $EC_{50}^{Peg-IFN}$  were estimated using non-linear mixed-effect  
199 models (NLMEM). In this approach, each individual parameter  $\theta_i$  is comprised of a fixed part  
200  $\theta$ , which represents the mean value of the parameter in the population (fixed effects), and a  
201 random part  $\eta_i$  chosen from a Gaussian distribution with mean 0 and standard deviation  $\omega_i$   
202 that accounts for the inter-individual variability. Therefore, for all parameters  $\theta_i = \theta e^{\eta_i}$   
203 where  $\eta_i \sim N(0, \omega^2)$ . Both PK data and  $\text{Log}_{10}(\text{HCV RNA})$  were best described using an  
204 additive residual error with constant variance.

205 Model parameters were estimated using the Stochastic Approximation Expectation  
206 Minimization (SAEM) algorithm in MONOLIX v4.2 (available at <http://www.lixoft.eu>). Of  
207 note this approach is based on maximum likelihood estimation which take into account the  
208 information brought by data under the LOD as left-censored data (31, 32).

209 Model selection was done using the Bayesian information criteria (BIC), a fitting criterion  
210 derived for each model from the computation of likelihood that takes into account the number  
211 of estimated parameters used (the lower the better (33)). Model evaluation was performed  
212 using goodness-of-fit plots, as well as the individual weighted residuals (IWRES) and the  
213 normalized prediction distribution errors (NPDE) over time.

214

215 **Difference in PK-VK model parameters between telaprevir and boceprevir treatment**  
216 **group**

217 A Wald test on the PK-VK model parameters ( $c, \delta, EC_{50}^{PI}$ ) was used to assess the difference  
 218 in population parameters between the two groups. Because we previously showed that this  
 219 approach could lead to an inflation of the type I error in case of small sample size ( $N < 20$  per  
 220 group) (34), a permutation test was performed to confirm statistical significance when the  
 221 Wald test was significant at the level of 5%. In brief, 1,000 datasets were simulated by  
 222 randomly allocating patients to telaprevir or boceprevir group, maintaining a similar  
 223 proportion of patients allocated to each groups than in the original dataset. Then the P-value  
 224 of the Wald test was calculated for each simulated data set. Finally the corrected P-value of  
 225 the permutation test is equal to the proportion of simulated datasets having a P-value lower  
 226 than the one found one the original dataset.

227 Because the genetic barrier to resistance of PI (*i.e.*, the number of change in amino acids  
 228 needed to generate mutants with high level of resistance) depends of HCV subgenotype and  
 229 therefore lead to different SVR rate, we also estimated the effect of HCV subgenotype (1a vs  
 230 non-1a) on viral kinetic parameters. IL28B polymorphism, which is also associated with  
 231 response to IFN-based therapy, was not investigated because all these patients had failed to a  
 232 previous bitherapy.

233

#### 234 **Prediction and comparison of individual parameters**

235 Individual Empirical Bayesian Estimates (EBE) parameters for both PK and VK were  
 236 obtained by computing for each patient the Maximum A Posteriori (MAP) estimate. The  
 237 individual antiviral effectiveness at steady state,  $\epsilon_{SS}$ , of each agent was defined by:

238

$$\epsilon_{SS}^{PI} = \frac{C_{SS}^{PI}}{C_{SS}^{PI} + EC_{50}^{PI}}$$

239

$$\epsilon_{SS}^{Peg-IFN} = \frac{C_{SS}^{Peg-IFN}}{C_{SS}^{Peg-IFN} + EC_{50}^{Peg-IFN}} \quad \text{Eq. (7)}$$

240 Non-parametric two-sided tests (Wilcoxon test) were used to compare i) individual EBE PK  
241 parameters between patients who received telaprevir vs boceprevir and between patients who  
242 received Peg-IFN- $\alpha$ 2a vs - $\alpha$ 2b, and ii) individual EBE PK parameters between SVR and non-  
243 SVR patients. Because all patients were non-responder to Peg-IFN, the effect of IL28B  
244 genotype on PK and VK parameters was not tested.  
245

## 246 Results

247 Fifteen HCV genotype 1 patients were included 9 receiving telaprevir and 6 receiving  
248 boceprevir. Twelve (80%) were men, with a median [min; max] age of 55 [44; 64] years.  
249 Seven (47%) patients were infected with subgenotype 1a, 2 (22%) in telaprevir group and 5  
250 (83%) in boceprevir group. Prior treatment responses were partial response, null response,  
251 relapse and early discontinuation for adverse events in 2, 5, 6 and 2 patients, respectively.  
252 Only two patients had the most favorable IL28B CC genotype (35). Main characteristics of  
253 the patients are presented in Table 1.

254 Two patients had a viral breakthrough (at weeks 3 and 8). Eleven patients received Peg-IFN-  
255  $\alpha$ 2a (8 in telaprevir group and 3 in boceprevir group), 3 patients Peg-IFN- $\alpha$ 2b (all in  
256 boceprevir group) and one patient in telaprevir group did not receive any injection of Peg-IFN  
257 (and this patient had a viral breakthrough at week 3).

258 Fig. 1 shows the observed drug concentrations versus time and Table 2 gives the estimated  
259 steady state trough concentrations,  $C_{ss}$ , for all drugs. There was no significant difference in  
260 the molar medians steady state concentrations of telaprevir and boceprevir ( $C_{ss}^{telaprevir} = 3.77$   
261 [2.68; 5.98]  $\mu\text{mol/l}$  *i.e.* 2,563.0 ng/ml [1,822.0; 4,065.5] and  $C_{ss}^{boceprevir} = 3.92$  [3.22; 7.64]  
262  $\mu\text{mol/l}$  *i.e.* 2037.1 ng/ml [1,673.4; 3,970.4],  $P=0.5$ ). There was no significant difference in the  
263 median steady state concentrations of Peg-IFN- $\alpha$ 2a and - $\alpha$ 2b ( $C_{ss}^{Peg-IFN-2a} = 89.6$  [52.8; 110.4]  
264 ng/ml and  $C_{ss}^{Peg-IFN-2b} = 55.4$  [55.3; 57.9] ng/ml,  $P=0.2$ ). The concentrations of RBV increased  
265 over time in all patients and could be well captured by our model (Eq. 2) with a median  $k$   
266 equal to  $0.10 \text{ day}^{-1}$ , corresponding to a half-life of increase of about 7 days. At equilibrium  
267 medians  $C_{ss}^{RBV}$  were equal to 2,860 [2,428; 3,874] ng/ml.

268 After the PK parameters were estimated, the predicted individual PK time courses were  
269 plugged into the PK-VK model (see methods). Baseline VL was higher in the telaprevir group  
270 than in the boceprevir group, thus a treatment group effect was added on baseline VL

271 ( $V_0^{telaprevir} = 6.43 \log_{10}$  IU/ml vs  $V_0^{boceprevir} = 5.52 \log_{10}$  IU/ml,  $P=0.0001$ ). A greater proportion  
272 of patients that received boceprevir were genotype 1a relative to those that received telaprevir  
273 ( $P=0.04$ ). Subgenotype is an important predictor of the response to treatment, in particular  
274 with telaprevir with a lower genetic barrier to resistance with genotype 1a than 1b (only one  
275 nucleotide change in genotype 1a viral genomes is required to generate mutations V36M and  
276 R155K/T, vs two in genotype 1b) (36). This may explain why genotype 1a patients were  
277 preferentially treated with boceprevir. We did not find any significant effect of subgenotype  
278 on any of the parameters.

279 The model could well describe the kinetics of HCV decline observed both during the lead-in  
280 phase (in the boceprevir group) and after the initiation of the PIs (in both groups, see Fig. 2).  
281 There was no evidence of model misspecification as showed by the goodness-of-fit plot (Fig.  
282 3) and all parameters could be estimated with a good precision (Table 3).

283 The model predicted a mean  $EC_{50}^{Peg-IFN}$  equal to 106 ng/ml, leading to a low antiviral  
284 effectiveness at steady state of Peg-IFN at steady state of 43.4% [0.0; 52.7], consistent with  
285 the modest 0.67  $\log_{10}$  IU/ml drop observed during the four weeks lead-in phase in patients  
286 treated with boceprevir (Fig. 2).

287 After PI initiation, VL declines in a biphasic manner in all patients, where a rapid first phase  
288 was followed by a second slower phase. The rapid first phase was attributed to a clearance  
289 rate of virus,  $c$ , equal to 3.98  $\text{day}^{-1}$  and to a high level of antiviral effectivenesses for both PIs.  
290 The intrinsic potency of the two molecules, as measured by the  $EC_{50}^{PI}$ , was significantly  
291 higher for telaprevir than boceprevir ( $EC_{50}^{telaprevir} = 0.009 \mu\text{mol/l}$  vs  $EC_{50}^{boceprevir} = 0.04$   
292  $\mu\text{mol/l}$ ,  $P=0.008$ ). Importantly the statistical significance of this difference was obtained after  
293 taking into account the small sample size (see methods) and adjusted on baseline VL. Since  
294 telaprevir had a lower  $EC_{50}$  than boceprevir and that both drugs achieved similar levels of  
295 molar concentrations the model predicted that the median individual antiviral effectiveness of



296 PI agent in blocking viral production was significantly higher in patients that received  
297 telaprevir than in those who received boceprevir ( $\varepsilon_{ss}^{telaprevir} = 99.8\% [99.3; 99.9]$  and  $\varepsilon_{ss}^{boceprevir}$   
298  $= 99.0\% [98.0; 99.6]$ ,  $P=0.002$ ). Interestingly this model could well capture the relationship  
299 between the serum exposure and its antiviral effectiveness, demonstrating that the variability  
300 in drug exposure needs to be taken into account to understand the between-subject variability  
301 in PIs antiviral effectiveness (Fig. 4A). Lastly because the effectiveness of both PIs were  
302 much larger than that of Peg-IFN (Fig. 4B), the total antiviral effectiveness obtained by the  
303 combination of PI and Peg-IFN was largely similar to the one obtained with the PIs only.  
304 After the VL was rapidly reduced as a result of the strong antiviral effectiveness of both PIs,  
305 the model predicted that a second slower phase of viral decline ensued, driven by the loss rate  
306 of infected cells,  $\delta$ . We estimated  $\delta$  to be equal to  $0.18 \text{ day}^{-1}$ , corresponding to a half-life of  
307 infected cells of 3.9 days, with no significant differences between patients receiving telaprevir  
308 and boceprevir ( $P=0.5$ ).

309 Next we investigated the relationship between the PK-VK parameters and SVR. Among the 7  
310 patients (47%) who achieved SVR, 5 received telaprevir and 2 received boceprevir (56% vs  
311 33%, respectively,  $P=0.6$ ). As shown in Fig. 5, neither the antiviral effectivenesses of PIs nor  
312 that of Peg-IFN was significantly associated with the long term virological response. However  
313 the loss rate of infected cells,  $\delta$ , was significantly higher in patients that subsequently  
314 achieved SVR (median  $\delta^{SVR} = 0.27 \text{ day}^{-1}$  vs median  $\delta^{non-SVR} = 0.14 \text{ day}^{-1}$ ,  $P=0.03$ ).

315 Lastly we verified that incorporating the effect of RBV exposure in the PK-VK model, either  
316 on the block of viral production or in the decrease of viral infectivity (data not shown) did not  
317 improve the fit of the data. Furthermore there was no significant association between the  
318 predicted  $C_{ss}^{RBV}$  and long term virological response ( $P=0.5$ ).

319

320 **Discussion**

321 Here we used a PK-VK model to provide the first detailed picture of the relationship between  
322 the exposure to all drugs involved in triple therapy (Peg-IFN, RBV and telaprevir or  
323 boceprevir) and the early virological response. This novel model provides important insights  
324 into the understanding of the response to triple therapy in hard-to treat patients.

325 We predicted that both PIs achieved a high level of antiviral effectiveness in blocking viral  
326 production that was higher than 97.9% in all patients. However telaprevir had a higher  
327 intrinsic potency than boceprevir, as measured by  $EC_{50}$  ( $P=0.008$  after correcting for small  
328 sample size), leading to a significantly higher level of antiviral effectiveness than boceprevir  
329 ( $\epsilon_{ss}^{telaprevir} = 99.8\%$  vs  $\epsilon_{ss}^{boceprevir} = 99.0\%$ ,  $P=0.002$ ) *i.e.* a 5-fold difference in the viral  
330 production under treatment. Importantly the difference in  $EC_{50}$  was obtained despite the fact  
331 that the study was not randomized and that patients who received telaprevir had less favorable  
332 baseline characteristics than those who received boceprevir with higher baseline VL (6.43  
333  $\log_{10}$  IU/ml vs 5.52  $\log_{10}$  IU/ml, respectively,  $P<10^{-4}$ ) and a higher proportion of null  
334 responder to previous bitherapy (4/9 vs 1/6).

335 The comparison of drug's antiviral effectiveness should be taken with caution because of  
336 small sample size, the absence of randomization, and the fact that only trough concentrations  
337 were used to estimate the  $EC_{50}$  of PI which may lead to underestimation. Yet these results  
338 demonstrate for the first time a significant association between serum exposure to PI agents  
339 and the antiviral effectiveness achieved. To confirm the significance of this association we  
340 fitted HCV RNA data to a simplified model where drug exposure was not taken into account  
341 (37). As compared to this model, we found that the PK-VK model both improved the fitting  
342 criterion (BIC decreases from 181.3 to 176.3, *i.e.* an improvement of 5 points which is  
343 regarded as positive evidence) and reduced the between-patient parameter variability by 26%

344 ( $\omega_{EC_{50}PI}$  from 0.85 to 0.61), thus demonstrating that serum PK is an important predictor of the  
345 antiviral effectiveness of triple therapy.

346 Our estimate that telaprevir achieves an antiviral effectiveness of 99.8% is largely similar to  
347 the one found in naïve patients (15), suggesting that compensated cirrhosis does not affect the  
348 maximal antiviral effectiveness of telaprevir. Whether this is also true for boceprevir is not  
349 known as to our knowledge there is no published viral kinetic modeling study evaluating the  
350 *in vivo* antiviral effectiveness of boceprevir.

351 In contrast to the high effectiveness achieved by both PIs, Peg-IFN was found to have a  
352 modest contribution in blocking viral production, with a mean value of 43.4%. Of note  
353 including the patient who did not receive Peg-IFN in our analysis allow us to add information  
354 on telaprevir antiviral effectiveness. Further RBV exposure had no significant contribution on  
355 the early viral kinetics. Together these results indicate that Peg-IFN and RBV have a minimal  
356 contribution on the early virologic response, at least on this population of previous non-  
357 responders to a Peg-IFN/RBV therapy.

358 In order to achieve a rapid viral decline, it is important to achieve not only a high level of  
359 effectiveness but also a rapid second phase of viral decline. Here the latter was rather slow in  
360 both treatment groups compared to what had been than found in telaprevir treated patients,  
361 and this was attributed in our model to a low loss rate of infected cells,  $\delta$ , about three times  
362 smaller than in non-cirrhotic naive-patients ( $\delta$  of 0.18 day<sup>-1</sup> vs 0.60 day<sup>-1</sup>) (15, 16). Those  
363 lower values may encompass several factors, such a lower penetration of PIs into infected  
364 cells in a highly scarced liver. Because the loss rate of infected cells is strongly related to the  
365 treatment duration needed to achieve SVR (15), our results suggest that the time to achieve  
366 SVR in this population could be longer than what had been predicted from clinical trials (15).  
367 Consistent with this prediction, the relapse rate in the CUPIC trial was equal to 41% in both

368 treatment groups (13), *i.e.*, much higher than what reported in treatment experienced patients  
369 phase 3 clinical trials (12% to 27%) (9, 11, 22).

370 Regarding the use of early viral kinetic parameters for treatment prediction, we found that  $\delta$   
371 was higher in patients that subsequently achieved SVR (median  $\delta^{SVR} = 0.27 \text{ day}^{-1}$  vs median  
372  $\delta^{non-SVR} = 0.14 \text{ day}^{-1}$ ,  $P=0.03$ ) suggesting that  $\delta$  could be a relevant predictor of the outcome of  
373 triple therapy, as it was the case for Peg-IFN/RBV bitherapy (38). In contrast there was no  
374 significant relationship between antiviral effectiveness of PIs on SVR (Fig. 6A). This absence  
375 of relationship is consistent with the hypothesis that in order to achieve SVR, it is necessary  
376 not only to have a high antiviral effectiveness at treatment initiation, when the viral  
377 population is predominantly wild-type and drug-sensitive, but also at later times, when the  
378 viral population is predominantly resistant to PI agents (39, 40). The fact that neither Peg-IFN  
379 effectiveness nor RBV were associated with SVR is more surprising, as one would expect  
380 these agents to be equally active against wild-type and resistant virus. However our patient  
381 population was both treatment experienced and cirrhotic, two major causes of insensitivity to  
382 Peg-IFN/RBV.

383 Clearly the main limitation of this study was its small size. In a previous study we evaluated  
384 by simulation the power to detect a difference of antiviral effectiveness between two  
385 treatment groups for a variety of designs (34). With a design comparable to the present study,  
386 *i.e.*, 10 patients per group, 7 VL per patient and an antiviral effectiveness of 99% vs 99.9%,  
387 the power to detect this difference was 100% with the same statistical method that we used in  
388 this analysis. Yet, further studies on larger populations will still be needed to estimate more  
389 precisely the exposure-effect relationship (Fig. 4) and other kinetic parameters involved on  
390 the long-term virologic response. A second limitation is that only trough pre-dose drug  
391 concentrations were collected and modeled. Thus  $C_{ss}$  is the steady-state  $C_{trough}$ . Moreover no  
392 information was collected on treatment adherence. The data analysis did not show any signal

393 of lack of adherence such as viral oscillations, which indicates that missed doses, if they  
394 occurred, did not have a major effect on the observed kinetic of decline. Here we considered  
395 that concentrations of PIs were constant over time. Detailed pharmacokinetic analysis showed  
396 that steady state of residual concentrations is attained after two days of treatment (41). As  
397 explained in details in Guedj *et al.* (42), the fact that we neglected this initial build up may  
398 explain why our estimate of the viral clearance rate,  $c$ , was lower than previously found in  
399 treatment naïve patients (15). Further the lack of information on the time of Peg-IFN injection  
400 also precluded a precise characterization between Peg-IFN exposure and the virological  
401 response. The fact that we used rather empirical models is less problematic for RBV, whose  
402 long elimination half-life resulting in a slow increase over time could be well characterized  
403 here (27). Moreover, as mentioned previously, in order to achieve SVR, it is important for  
404 drugs to achieve a higher effectiveness against PI-resistant virus. Because no sequencing was  
405 done here, we focused only the early virological response where presumably the virus is  
406 predominantly drug-sensitive. In order to estimate PI effectiveness against resistant virus it  
407 would be needed to quantify and follow the proportion of resistant virus over time, as early as  
408 possible, for instance using pyrosequencing (43).

409 A greater proportion of patients that received boceprevir were genotype 1a relative to those  
410 that received telaprevir ( $P=0.04$ ). It has been well established that subgenotype is an important  
411 predictor of the response to treatment and for instance the fact that telaprevir has a higher  
412 genetic barrier to resistance with genotype 1b than 1a (36) may explain why genotype non-1a  
413 patients were preferentially treated with telaprevir than boceprevir. However the effect of  
414 subgenotype on the early viral kinetics, where most of the virus is drug-sensitive is unknown,  
415 and has never been investigated as far as we know. In our study no significant effect of  
416 subgenotype on any of the parameters ( $c$ ,  $\delta$ ,  $EC_{50}^{PI}$ ) was found.

417 The effect of RBV was analyzed using serum drug concentrations. Some authors preferably  
418 used erythrocyte RBV concentration (44), which was not measured in the present study.  
419 However a significant relationship was shown between erythrocyte RBV concentrations and  
420 serum concentrations (45), suggesting that serum RBV can be used for the assessment of early  
421 and sustained virological responses (46, 47).

422 To summarize this study provides the first characterization of the relationship between drug  
423 concentrations involved in triple therapy and early HCV viral kinetics treated with telaprevir  
424 or boceprevir. We found that median values of antiviral effectiveness for telaprevir was  
425 similar to what had been found in treatment naïve patients and significantly larger than in  
426 boceprevir treated patients. In all patients the second phase of viral decline was slow and may  
427 explain the high relapse rate observed in the ANRS-CO20-CUPIC cohort. This suggests that,  
428 notwithstanding safety issues, longer treatment duration could improve the treatment efficacy  
429 and lead to a higher SVR rate. Lastly the antiviral effectiveness of Peg-IFN was modest (less  
430 than 50%) suggesting that cirrhotic treatment experienced-patients may particularly benefit  
431 from upcoming IFN-free treatment. Our approach, which shows the importance of PK data to  
432 disentangle the effects of drug combination and to understand the variability in the virological  
433 response, is not specific to triple therapy and could also be used to optimize future IFN-free  
434 regimen, in particular in hard-to-treat patients.

435

436 **Acknowledgements**

437 The study was sponsored and funded by The National Agency for research on Aids and viral  
438 Hepatitis (ANRS) and in part by the Association Française pour l'Etude du Foie (AFEF).  
439 Sponsors had no role in interpretation of data, in the writing of the report, or in the decision to  
440 submit the paper for publication.

441 The authors thank Ventzislava Petrov Sanchez and Setty Allam (from unit Basic and Clinical  
442 research on viral hepatitis, French National Agency for research on Aids and viral Hepatitis,  
443 Paris, France) and Cécilie Dufour (from Inserm UMR 707, University Pierre et Marie Curie,  
444 Paris, France). The authors thank Dr Marie Anne Loriot (from Inserm UMR 1147, University  
445 Paris Descartes, Paris, France) for genotyping the IL28B rs12979860 polymorphism.

446

447 **Author Contributions:** CL, FM, and JG made the analysis and drafted the manuscript; all  
448 authors provided the data; all authors read and approved the final manuscript.

449

450 **Disclosure statement**

451 JG: has consulted with Gilead SC.

452 FZ: received speakers/consulting fees from Gilead SC, MSD, BMS, Janssen cilag, Abbvie,  
453 Boehringer Ingelheim.

454 CH: has been a clinical investigator, speaker and/or consultant for Abbvie, Boehringer  
455 Ingelheim, BMS, Gilead Sciences, Janssen, Merck Sharp & Dohme, and Roche.

456 PM: has been a clinical investigator, speaker and/or consultant for Roche, Gilead, Vertex,  
457 Novartis, Janssen - Tibotec, MSD, Boehringer, Abbott, Pfizer, Alios BioPharma.

458 GP: has received travel grants, consultancy fees, honoraria or study grants from various  
459 pharmaceutical companies, including Bristol-Myers-Squibb, Gilead SC, Janssen, Merck, ViiV  
460 Healthcare and Splicos.

461

462 **References**

- 463 1. World Health Organization. WHO Fact Sheet 164-Hepatitis C. Available at:  
464 <http://www.who.int.gate2.inist.fr/mediacentre/factsheets/fs164/en/>. Accessed Januray 31,  
465 2014.
- 466 2. **Shepard CW, Finelli L, Alter, MJ.** 2005. Global epidemiology of hepatitis C virus  
467 infection. *Lancet Infect. Dis.* **5**:558–567.
- 468 3. **Pearlman BL.** 2012. Protease inhibitors for the treatment of chronic hepatitis C  
469 genotype-1 infection: the new standard of care. *Lancet Infect. Dis.* **12**:717-728.
- 470 4. **Poordad F, McCone J Jr, Bacon BR, Bruno S, Manns MP, Sulkowski MS,**  
471 **Jacobson IM, Reddy KR, Goodman ZD, Boparai N, DiNubile MJ, Sniukiene V,**  
472 **Brass CA, Albrecht JK, Bronowicki J-P.** 2011. Boceprevir for untreated chronic HCV  
473 genotype 1 infection. *N. Engl. J. Med.* **364**:1195–1206.
- 474 5. **McHutchison JG, Everson GT, Gordon SC, Jacobson IM, Sulkowski M, Kauffman**  
475 **R, McNair L, Alam J, Muir AJ.** 2009. Telaprevir with peginterferon and ribavirin for  
476 chronic HCV genotype 1 infection. *N. Engl. J. Med.* **360**:1827–38.
- 477 6. **Asselah T, Marcellin P.** 2014. Second-wave IFN-based triple therapy for HCV  
478 genotype 1 infection: simeprevir, faldaprevir and sofosbuvir. *Liver Int.* **34**:60–68.
- 479 7. **Deuffic-Burban S, Schwarzingler M, Obach D, Mallet V, Pol S, Pageaux GP, Canva**  
480 **V, Deltenre P, Roudot-Thoraval F, Larrey D, Dhumeaux D, Mathurin P,**  
481 **Yazdanpanah Y.** 2014. Should we await IFN-free regimens to treat HCV genotype 1  
482 treatment-naive patients? A cost-effectiveness analysis (ANRS 12188). *J. Hepatol.*, in  
483 press.
- 484 8. **Flamm SL, Lawitz E, Jacobson I, Bourlière M, Hezode C, Vierling JM, Bacon BR,**  
485 **Niederau C, Sherman M, Goteti V, Sings HL, Barnard RO, Howe JA, Pedicone LD,**  
486 **Burroughs MH, Brass CA, Albrecht JK, Poordad F.** 2013. Boceprevir with  
487 peginterferon alfa-2a-ribavirin is effective for previously treated chronic hepatitis C  
488 genotype 1 infection. *Clin. Gastroenterol. Hepatol.* **11**:81–87.
- 489 9. **Zeuzem S, Andreone P, Pol S, Lawitz E, Diago M, Roberts S, Focaccia R, Younossi**  
490 **Z, Foster GR, Horban A, Ferenci P, Nevens F, Müllhaupt B, Pockros P, Terg R,**  
491 **Shouval D, van Hoek B, Weiland O, Van Heeswijk R, De Meyer S, Luo D,**  
492 **Boogaerts G, Polo R, Picchio G, Beumont M, REALIZE Study Team.** 2011.  
493 Telaprevir for retreatment of HCV infection. *N. Engl. J. Med.* **364**:2417–2428.



- 494 10. **McHutchison JG, Manns MP, Muir AJ, Terrault NA, Jacobson IM, Afdhal NH,**  
495 **Heathcote EJ, Zeuzem S, Reesink HW, Garg J, Bsharat M, George S, Kauffman**  
496 **RS, Adda N, Di Bisceglie AM.** 2010. Telaprevir for previously treated chronic HCV  
497 infection. *N. Engl. J. Med.* **362**:1292–303.
- 498 11. **Bacon BR, Gordon SC, Lawitz E, Marcellin P, Vierling JM, Zeuzem S, Poordad F,**  
499 **Goodman ZD, Sings HL, Boparai N, Burroughs M, Brass CA, Albrecht JK,**  
500 **Esteban R.** 2011. Boceprevir for previously treated chronic HCV genotype 1 infection.  
501 *N. Engl. J. Med.* **364**:1207–1217.
- 502 12. **Hézode C, Fontaine H, Dorival C, Larrey D, Zoulim F, Canva V, de Ledinghen V,**  
503 **Poynard T, Samuel D, Bourlière M, Zarski JP, Raabe JJ, Alric L, Marcellin P,**  
504 **Riachi G, Bernard PH, Loustaud-Ratti V, Métivier S, Tran A, Serfaty L, Abergel**  
505 **A, Causse X, Di Martino V, Guyader D, Lucidarme D, Grando-Lemaire V, Hillon**  
506 **P, Feray C, Dao T, Cacoub P, Rosa I, Attali P, Petrov-Sanchez V, Barthe Y,**  
507 **Pawlotsky JM, Pol S, Carrat F, Bronowicki JP; CUPIC Study Group.** 2013. Triple  
508 therapy in treatment-experienced patients with HCV-cirrhosis in a multicentre cohort of  
509 the French Early Access Programme (ANRS CO20-CUPIC). *J. Hepatol.* **59**:434-441.
- 510 13. **Hézode C, Fontaine H, Dorival C, Zoulim F, Larrey D, Canva V, de Ledinghen V,**  
511 **Poynard T, Samuel D, Bourlière M, Alric L, Raabe J-J, Zarski J-P, Marcellin P,**  
512 **Riachi G, Bernard P-H, Loustaud-Ratti V, Chazouilleres O, Abergel A, Guyader D,**  
513 **Metivier S, Tran A, di Martino V, Causse X, Dao T, Lucidarme D, Portal I, Cacoub**  
514 **P, Gournay J, Grando-Lemaire V, Hillon P, Attali P, Fontanges T, Rosa I, Petrov-**  
515 **Sanchez V, Barthe Y, Pawlotsky J-M, Pol S, Carrat F, Bronowicki J-P, the cupic**  
516 **study group.** 2014. Effectiveness of Telaprevir or Boceprevir in Treatment-experienced  
517 Patients with HCV Genotype 1 Infection and Cirrhosis. *Gastroenterology*, in press.
- 518 14. **Chatterjee A, Guedj J, Perelson AS.** 2012. Mathematical modelling of HCV infection:  
519 what can it teach us in the era of direct-acting antiviral agents? *Antivir. Ther.* **17**:1171–  
520 1182.
- 521 15. **Guedj J, Perelson AS.** 2011. Second-phase hepatitis C virus RNA decline during  
522 telaprevir-based therapy increases with drug effectiveness: implications for treatment  
523 duration. *Hepatology* **53**:1801-1808.
- 524 16. **Adiwijaya BS, Kieffer TL, Henshaw J, Eisenhauer K, Kimko H, Alam JJ,**  
525 **Kauffman RS, Garg V.** 2012. A viral dynamic model for treatment regimens with  
526 direct-acting antivirals for chronic hepatitis C infection. *PLoS Comput. Biol.*  
527 **8**(1):e1002339.

- 528 17. **Adiwijaya BS, Herrmann E, Hare B, Kieffer T, Lin C, Kwong AD, Garg V, Randle**  
529 **JC, Sarrazin C, Zeuzem S, Caron PR.** 2010. A multi-variant, viral dynamic model of  
530 genotype 1 HCV to assess the in vivo evolution of protease-inhibitor resistant variants.  
531 PLoS Comput. Biol. **6(4):e1000745.**
- 532 18. **Fontaine H, Petitprez K, Roudot-Thoraval F, Trinchet J-C.** 2007. Guidelines for the  
533 diagnosis of uncomplicated cirrhosis. *Gastroenterol. Clin. Biol.* **31:504–509.**
- 534 19. **Farnik H, El-Duweik J, Welsch C, Sarrazin C, Lötsch J, Zeuzem S, Geisslinger G,**  
535 **Schmidt H.** 2009. Highly sensitive determination of HCV protease inhibitors boceprevir  
536 (SCH 503034) and telaprevir (VX 950) in human plasma by LC-MS/MS. *J. Chromatogr.*  
537 *B Analyt. Technol. Biomed. Life. Sci.* **877:4001–4006.**
- 538 20. **Homma M, Jayewardene AL, Gambertoglio J, Aweeka F.** 1999. High-performance  
539 liquid chromatographic determination of ribavirin in whole blood to assess disposition in  
540 erythrocytes. *Antimicrob. Agents Chemother.* **43:2716–2719.**
- 541 21. **Boulestin A, Kamar N, Legrand-Abravanel F, Sandres-Saune K, Alric L, Vinel J-P,**  
542 **Rostaing L, Izopet J.** 2004. Convenient biological assay for polyethylene glycol-  
543 interferons in patients with hepatitis C. *Antimicrob. Agents Chemother.* **48:3610–3612.**
- 544 22. **Kieran J, Schmitz S, O’Leary A, Walsh C, Bergin C, Norris S, Barry M.** 2013. The  
545 relative efficacy of boceprevir and telaprevir in the treatment of hepatitis C virus  
546 genotype 1. *Clin. Infect. Dis.* **56:228–235.**
- 547 23. **Neumann AU, Lam NP, Dahari H, Gretch DR, Wiley TE, Layden TJ, Perelson AS.**  
548 1998. Hepatitis C viral dynamics in vivo and the antiviral efficacy of interferon-alpha  
549 therapy. *Science* **282:103–7.**
- 550 24. **Guedj J, Rong L, Dahari H, Perelson AS.** 2010. A perspective on modelling hepatitis  
551 C virus infection. *J. Viral Hepat.* **17:825–833.**
- 552 25. **Shudo E, Ribeiro RM, Talal AH, Perelson AS.** 2008. A hepatitis C viral kinetic model  
553 that allows for time-varying drug effectiveness. *Antivir. Ther.* **13:919–926.**
- 554 26. **Rong L, Dahari H, Ribeiro RM, Perelson AS.** 2010. Rapid emergence of protease  
555 inhibitor resistance in hepatitis C virus. *Sci. Transl. Med.* **2:30ra32.**
- 556 27. **Pawlotsky J-M, Dahari H, Neumann AU, Hezode C, Germanidis G, Lonjon I,**  
557 **Castera L, Dhumeaux D.** 2004. Antiviral action of ribavirin in chronic hepatitis C.  
558 *Gastroenterology* **126:703–714.**
- 559 28. **Rotman Y, Nouredin M, Feld JJ, Guedj J, Witthaus M, Han H, Park YJ, Park S-**  
560 **H, Heller T, Ghany MG, Doo E, Koh C, Abdalla A, Gara N, Sarkar S, Thomas E,**  
561 **Ahlenstiel G, Edlich B, Titerence R, Hogdal L, Rehmann B, Dahari H, Perelson**

- 562 AS, Hoofnagle JH, Liang TJ. 2014. Effect of ribavirin on viral kinetics and liver gene  
563 expression in chronic hepatitis C. *Gut* **63**:161–169.
- 564 29. Mihm U, Welker M-W, Teuber G, Wedemeyer H, Berg T, Sarrazin C, Böhm S,  
565 Alshuth U, Herrmann E, Zeuzem S. 2014. Impact of ribavirin priming on viral kinetics  
566 and treatment response in chronic hepatitis C genotype 1 infection. *J. Viral Hepat.*  
567 **21**:42–52.
- 568 30. Dixit NM, Layden-Almer JE, Layden TJ, Perelson AS. 2014. Modelling how  
569 ribavirin improves interferon response rates in hepatitis C virus infection. *Nature*  
570 **432**:922–924.
- 571 31. Samson A, Lavielle M, Mentre F. 2006. Extension of the SAEM algorithm to left  
572 censored data in nonlinear mixed-effects model: Application to HIV dynamics model.  
573 *Comput. Stat. Data Anal.* **51**:1562–1574.
- 574 32. Thiébaud R, Guedj J, Jacqmin-Gadda H, Chêne G, Trimoulet P, Neau D,  
575 Commenges D. 2006. Estimation of dynamical model parameters taking into account  
576 undetectable marker values. *BMC Med. Res. Methodol.* **6**:38.
- 577 33. Guedj J, Pang PS, Denning J, Rodriguez-Torres M, Lawitz E, Symonds W,  
578 Perelson AS. 2014. Analysis of the hepatitis C viral kinetics during administration of  
579 two nucleotide analogues: sofosbuvir (GS-7977) and GS-0938. *Antivir. Ther.* **19**:211-  
580 220.
- 581 34. Laouénan C, Guedj J, Mentré F. 2013. Clinical trial simulation to evaluate power to  
582 compare the antiviral effectiveness of two hepatitis C protease inhibitors using nonlinear  
583 mixed effect models: a viral kinetic approach. *BMC Med. Res. Methodol.* **13**:60.
- 584 35. Holmes JA, Desmond PV, Thompson AJ. 2012. Does IL28B genotyping still have a  
585 role in the era of direct-acting antiviral therapy for chronic hepatitis C infection? *J. Viral*  
586 *Hepat.* **19**:677–684.
- 587 36. Cunningham M, Foster GR. 2012. Efficacy and safety of telaprevir in patients with  
588 genotype 1 hepatitis C infection. *Ther. Adv. Gastroenterol.* **5**:139–151.
- 589 37. Laouénan C, Guedj J, Lapalus M, Khelifa-Mouri F, Martinot-Peignoux M, Boyer  
590 N, Serfaty L, Bronowicki JP, Zoulim F, Mentré F, Marcellin P. 2013.  
591 Characterization of the early viral kinetics in compensated cirrhotic treatment-  
592 experienced patients treated with boceprevir and telaprevir, abstr 08c. 48th annual  
593 meeting of the European Association for the Study of the Liver (EASL), Amsterdam,  
594 The Netherlands.

- 595 38. **Talal AH, Ribeiro RM, Powers KA, Grace M, Cullen C, Hussain M, Markatou M,**  
596 **Perelson AS.** 2006. Pharmacodynamics of PEG-IFN alpha differentiate HIV/HCV  
597 coinfectd sustained virological responders from nonresponders. *Hepatology* **43**:943–  
598 953.
- 599 39. **Halfon P, Locarnini S.** 2011. Hepatitis C virus resistance to protease inhibitors. *J.*  
600 *Hepatol.* **55**:192–206.
- 601 40. **Sarrazin C, Zeuzem S.** 2010. Resistance to direct antiviral agents in patients with  
602 hepatitis C virus infection. *Gastroenterology* **138**:447–462.
- 603 41. **Yamada I, Suzuki F, Kamiya N, Aoki K, Sakurai Y, Kano M, Matsui H, Kumada**  
604 **H.** 2012. Safety, pharmacokinetics and resistant variants of telaprevir alone for 12 weeks  
605 in hepatitis C virus genotype 1b infection. *J. Viral Hepat.* **19**:e112–119.
- 606 42. **Guedj J, Dahari H, Shudo E, Smith P, Perelson AS.** 2012. Hepatitis C viral kinetics  
607 with the nucleoside polymerase inhibitor mericitabine (RG7128). *Hepatology* **55**:1030–  
608 1037.
- 609 43. **Chevaliez S, Rodriguez C, Pawlotsky J-M.** 2012. New virologic tools for management  
610 of chronic hepatitis B and C. *Gastroenterology* **142**:1303–1313.
- 611 44. **Inoue Y, Homma M, Matsuzaki Y, Shibata M, Matsumura T, Ito T, Kohda Y.**  
612 Erythrocyte ribavirin concentration for assessing hemoglobin reduction in interferon and  
613 ribavirin combination therapy. 2006. *Hepatol. Res.* **34**:23–27.
- 614 45. **Dominguez S, Ghosn J, Cassard B, Melica G, Poizot-Martin I, Solas C, Lascaux**  
615 **AS, Bouvier-Alias M, Katlama C, Lévy Y, Peytavin G.** 2012. Erythrocyte and plasma  
616 ribavirin concentrations in the assessment of early and sustained virological responses to  
617 pegylated interferon-alpha 2a and ribavirin in patients coinfectd with hepatitis C virus  
618 and HIV. *J. Antimicrob. Chemother.* **67**:1449–1452.
- 619 46. **Breilh D, Djabarouti S, Trimoulet P, Le Bail B, Dupon M, Ragnaud JM, Fleury H,**  
620 **Saux MC, Thiébaud R, Chêne G, Neau D.** 2009. Ribavirin plasma concentration  
621 predicts sustained virological response to peginterferon Alfa 2a plus ribavirin in  
622 previously treated HCV-HIV-coinfectd patients. *J. Acquir. Immune Defic. Syndr.*  
623 **52**:428–430.
- 624 47. **Morello J, Rodríguez-Novoa S, Jiménez-Nácher I, Soriano V.** 2008. Usefulness of  
625 monitoring ribavirin plasma concentrations to improve treatment response in patients  
626 with chronic hepatitis C. *J. Antimicrob. Chemother.* **62**:1174–1180.
- 627

628 **Figure legends**

629

630 **Fig. 1: Observed concentrations over time.**

631 (a) telaprevir in 9 patients (black,  $\mu\text{mol/ml}$ ) and boceprevir in 6 six patients (grey,  $\mu\text{mol/ml}$ );  
632 (b) Peg-IFN in telaprevir group (black,  $\text{ng/ml}$ ) and in boceprevir group (grey,  $\text{ng/ml}$ ); (c) RBV  
633 in telaprevir group (black,  $\text{ng/ml}$ ) and in boceprevir group (grey,  $\text{ng/ml}$ ). Patients who  
634 received a boceprevir-based therapy had only two blood samples during the lead-in phase at  
635 baseline and week 2.

636

637 **Fig. 2: Individual fits of the viral decline ( $\log_{10}$  IU/ml).**

638 Nine patients in telaprevir group (black curve) and 6 patients in boceprevir group (grey  
639 curve). Black crosses represent the observed viral load and grey stars represent the viral load  
640 under the limit of detection.

641

642 **Fig. 3. Goodness-of-fit of the viral kinetic-pharmacokinetic model**

643 Residuals (weighted residuals calculated using individual predictions: IWRES and normalized  
644 prediction distribution errors: NPDE) versus time and versus predictions plots. Residuals  
645 seem to distribute homogenously around 0.

646 Observed viral load are plotted as black crosses and viral load under the limit of detection as  
647 grey stars.

648

649 **Fig. 4. Relationship between predicted trough concentration at steady state ( $C_{ss}$ ) and**  
650 **predicted antiviral effectivenesses ( $\epsilon_{ss}$ ).**

651 (a) for the protease inhibitor (telaprevir in black and boceprevir in grey,  $\mu\text{mol/l}$ ); (b) for Peg-  
652 IFN (Peg-IFN- $\alpha$ 2a in black and Peg-IFN- $\alpha$ 2b in grey,  $\text{ng/ml}$ ). The lines denote the predictions

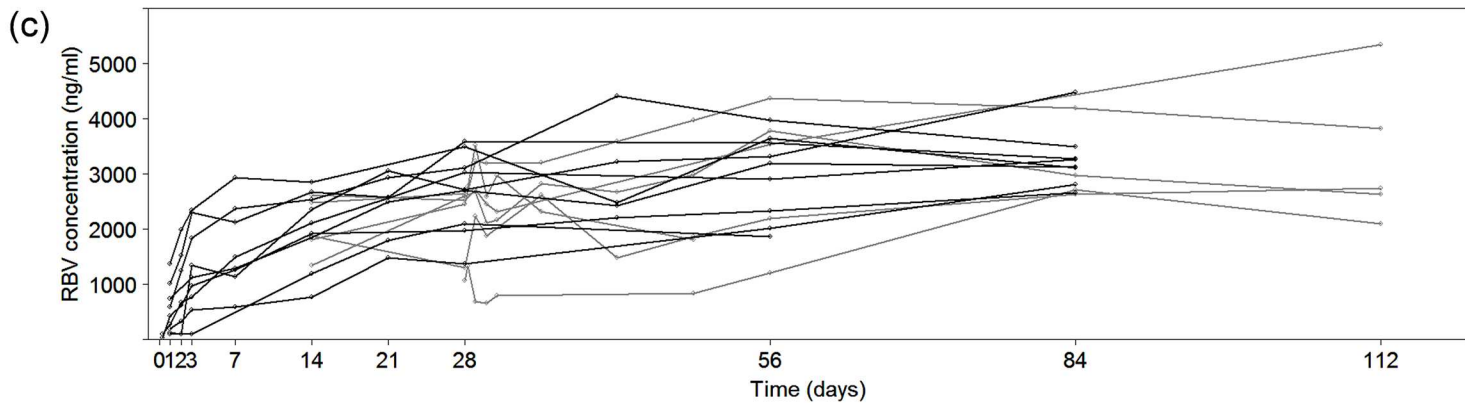
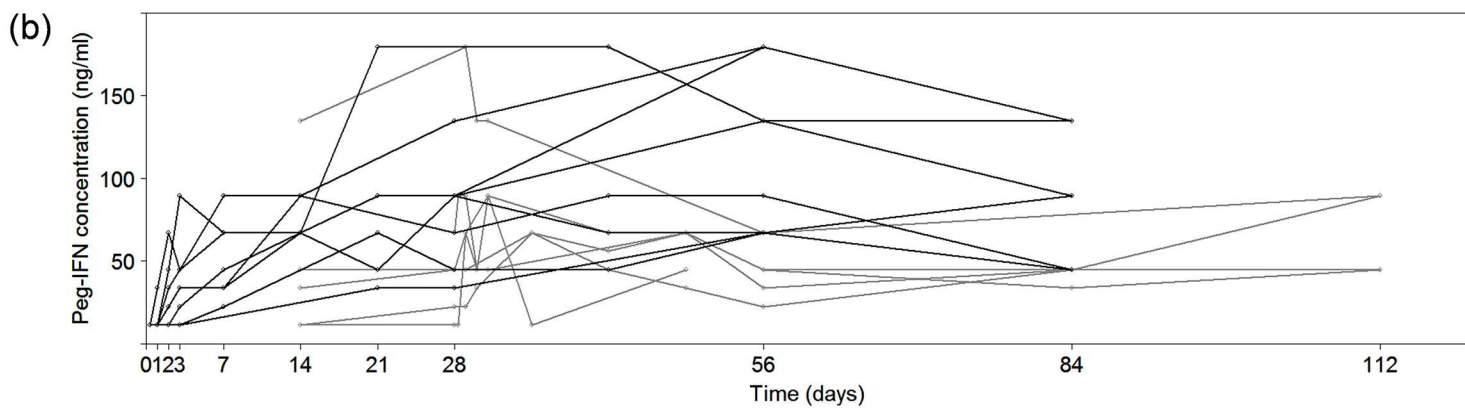
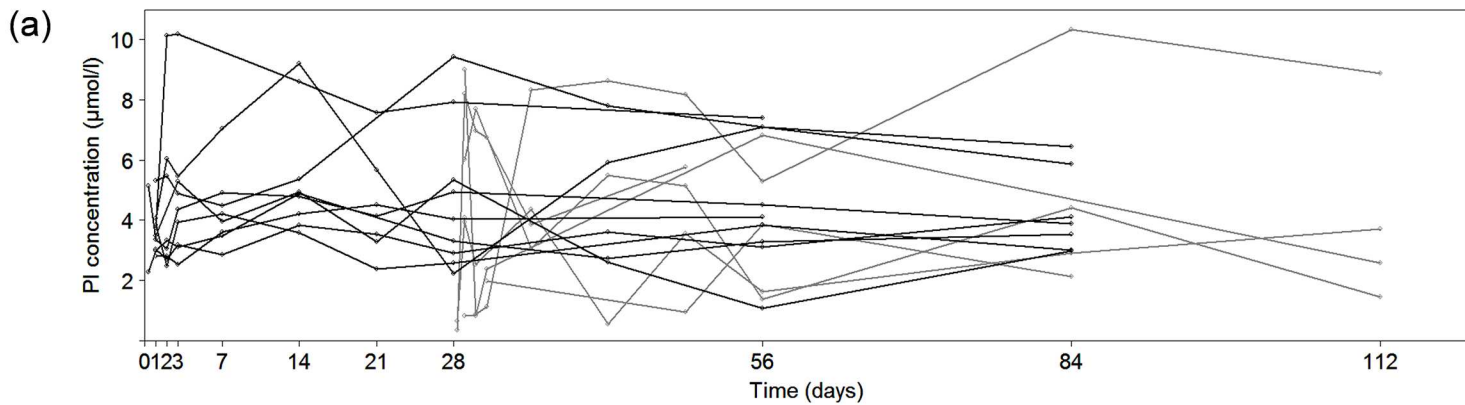
653 with the mean antiviral effectiveness and the dotted lines denote 95% confidence interval  
654 computed with the standard errors predicted by the Fisher Information Matrix.

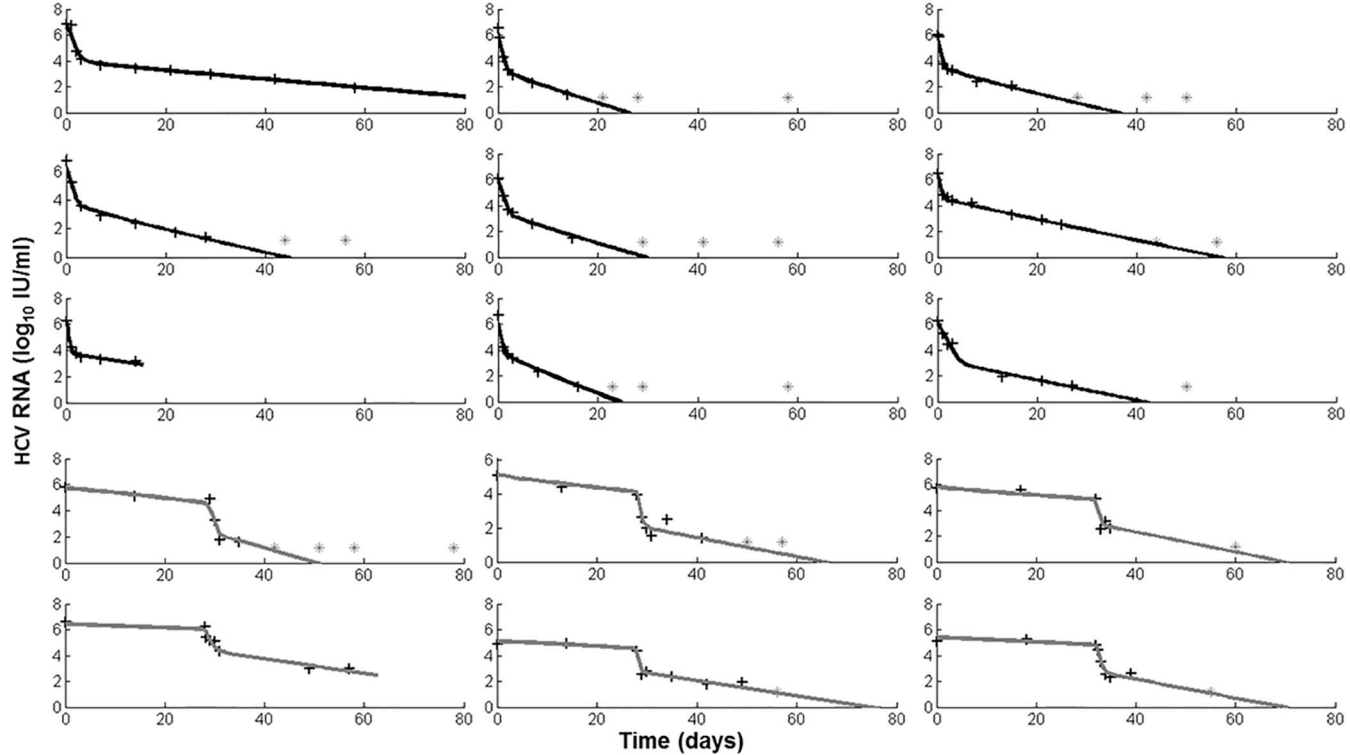
655

656 **Fig. 5: Relationship between long term virological response (SVR) and parameters**  
657 **estimated by the viral kinetic-pharmacokinetic model.**

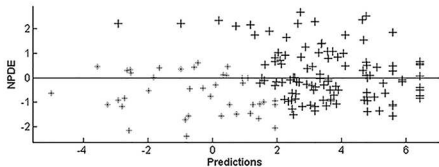
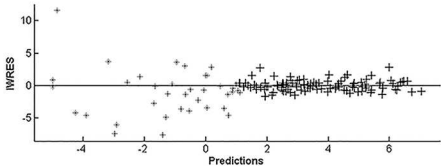
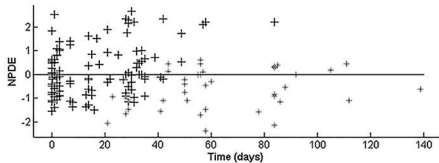
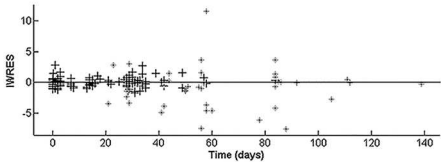
658 (a) predicted antiviral effectivenesses ( $\epsilon_{ss}$ ) of PIs; (b) predicted antiviral effectivenesses ( $\epsilon_{ss}$ )

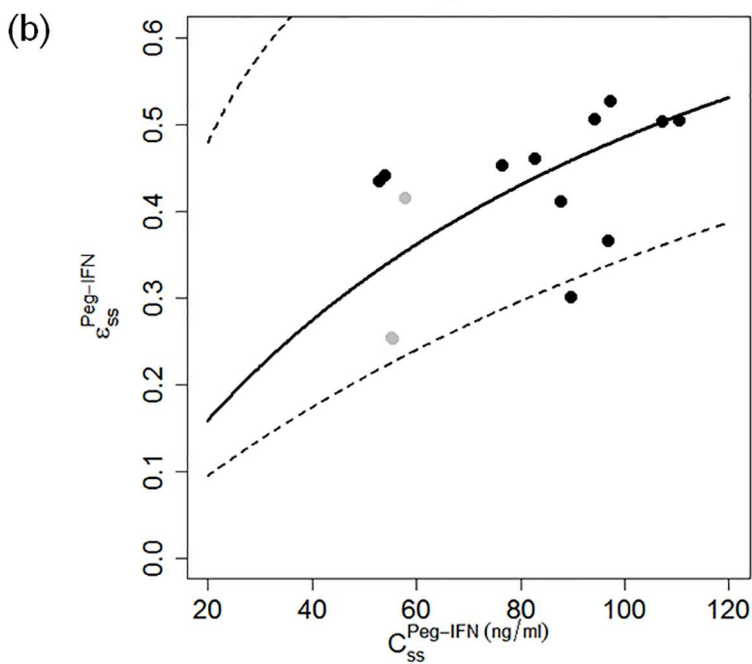
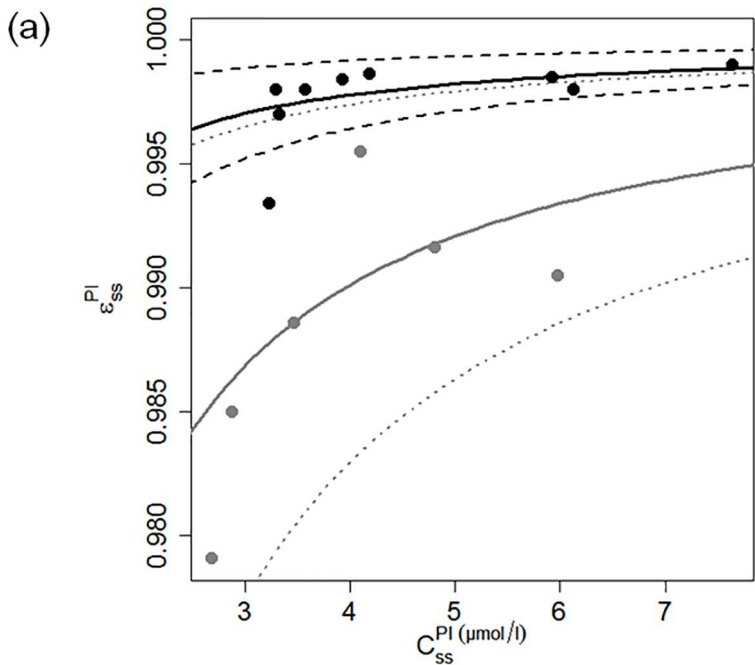
659 of Peg-IFN; (c)  $\delta$  delta parameter (loss rate of infected cells). P-value from Wilcoxon tests.

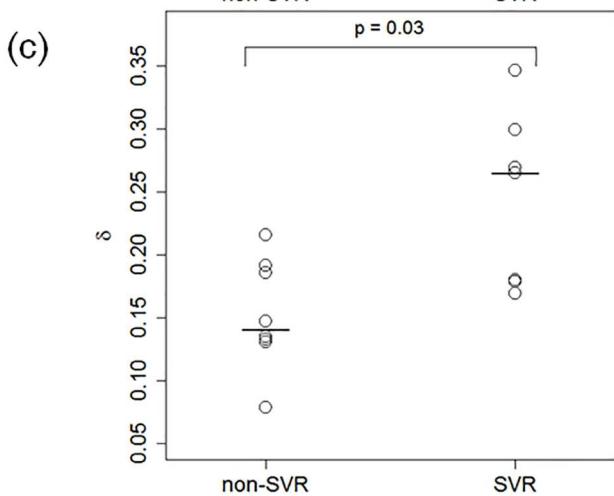
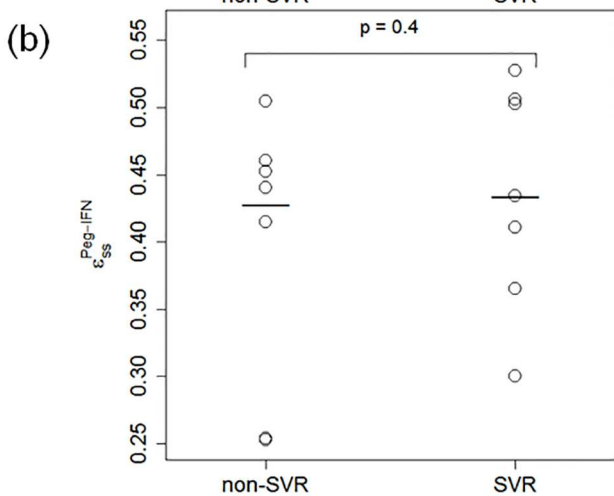
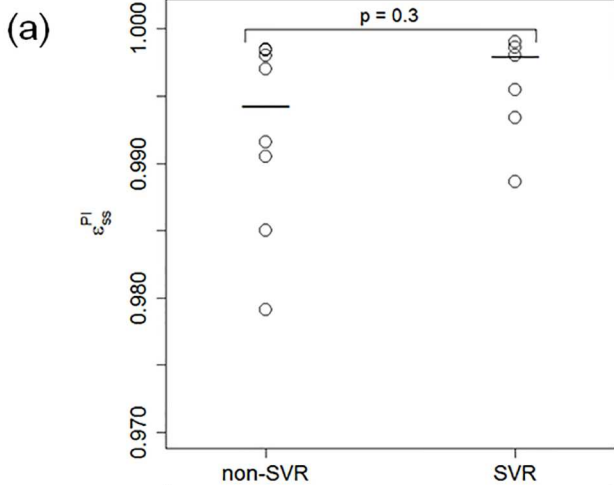












1 **Table 1. Main patient characteristics**

	<b>Peg-IFN/RBV + telaprevir</b>	<b>Peg-IFN/RBV + boceprevir</b>	<b>Total</b>
	<b>n=9</b>	<b>n=6</b>	<b>n=15</b>
Age (years), median [min-max]	55 [49-59]	53 [44-64]	55 [44-64]
Males, n (%)	8 (89)	4 (67)	12 (80)
HCV RNA (log <sub>10</sub> IU/ml), median [min-max]	6.5 [6.0-6.8]	5.4 [4.9-6.6]	6.2 [4.9-6.8]
HCV genotype, n (%):			
1a	2 (22)	5 (83)	7 (47)
Non 1a	7 (78)	1 (17)	8 (53)
IL28B genotype (rs12979860), n (%):			
C/C	2 (22)	-	2 (13)
C/T	6 (67)	6 (100)	12 (80)
T/T	1 (11)	-	1 (7)
Response to previous bitherapy, n (%):			
Partial responder	-	2 (33)	2 (13)
Null responder	4 (44)	1 (17)	5 (33)
Relapser	3 (33)	3 (50)	6 (40)
Early discontinuation for adverse event	2 (22)	-	2 (13)

2

3

4 **Table 2. Individual predicted trough concentrations at steady state ( $C_{ss}$ )**

	n	median [min; max]
$C_{ss}^{\text{telaprevir}}$ ( $\mu\text{mol/l}$ )	9	3.77 [2.68; 5.98]
$C_{ss}^{\text{boceprevir}}$ ( $\mu\text{mol/l}$ )	6	3.92 [3.22; 7.64]
$C_{ss}^{\text{Peg-IFN-}\alpha 2a}$ (ng/ml)	11	89.6 [52.8; 110.4]
$C_{ss}^{\text{Peg-IFN-}\alpha 2b}$ (ng/ml)	3	55.4 [55.3; 57.9]
$C_{ss}^{\text{RBV}}$ (ng/ml)	15	2,860 [2,428; 3,874]

5

6 **Table 3. Parameter estimates and relative standard errors (RSE)**

	Estimate	RSE (%)
$V_0^{\text{telaprevir}}$ (log <sub>10</sub> IU/ml)	6.43	2
$V_0^{\text{boceprevir}}$ (log <sub>10</sub> IU/ml)	5.52	3
$c$ (day <sup>-1</sup> )	3.98	12
$\delta$ (day <sup>-1</sup> )	0.18	11
$EC_{50}^{\text{Peg-IFN}}$ (ng/ml)	106	40
$EC_{50}^{\text{telaprevir}}$ (μmol/l)	0.009	30
$EC_{50}^{\text{boceprevir}}$ (μmol/l)	0.04	43
$\omega_{V_0}$	0.07	20
$\omega_c$	0.47	19
$\omega_\delta$	0.42	16
$\omega_{EC_{50}^{\text{Peg-IFN}}}$	0.67	30
$\omega_{EC_{50}^{\text{PI}}}$	0.61	32
$\sigma$	0.27	7

7  $V_0$ : baseline viral load;  $c$ : clearance rate of virus from serum;  $\delta$ : loss rate of  
8 infected cells;  $EC_{50}$ : half maximal effective concentration;  $\omega$ : inter-  
9 individual variability;  $\sigma$ : standard deviation of residual error; RSE: relative  
10 standard errors of parameter estimates, PI: protease inhibitor.