



10-1-2015

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## Recommended Citation

Xin, Yan-Le; Yu, Jie-Zhong; Yang, Xin-Wang; Liu, Chun-Yun; Li, Yan-Hua; Feng, Ling; Chai, Zhi; Yang, Wan-Fang; Wang, Qing; Jiang, Wei-Jia; Zhang, Guang-Xian; Xiao, Bao-Guo; and Ma, Cun-Gen, "FSD-C10: A more promising novel ROCK inhibitor than Fasudil for treatment of CNS autoimmunity." (2015). *Department of Neurology Faculty Papers*. Paper 86.  
<http://jdc.jefferson.edu/neurologyfp/86>

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## OPEN ACCESS

# FSD-C10: A more promising novel ROCK inhibitor than Fasudil for treatment of CNS autoimmunity

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## Synopsis

Rho-Rho kinase (Rho-ROCK) triggers an intracellular signalling cascade that regulates cell survival, death, adhesion, migration, neurite outgrowth and retraction and influences the generation and development of several neurological disorders. Although Fasudil, a ROCK inhibitor, effectively suppressed encephalomyelitis (EAE), certain side effects may limit its clinical use. A novel and efficient ROCK inhibitor, FSD-C10, has been explored. In the present study, we present chemical synthesis and structure of FSD-C10, as well as the relationship between compound concentration and ROCK inhibition. We compared the inhibitory efficiency of ROCK I and ROCK II, the cell cytotoxicity, neurite outgrowth and dendritic formation, neurotrophic factors and vasodilation between Fasudil and FSD-C10. The results demonstrated that FSD-C10, like Fasudil, induced neurite outgrowth of neurons and dendritic formation of BV-2 microglia and enhanced the production of neurotrophic factor brain-derived neurotrophic factor (BDNF), glial cell line-derived neurotrophic factor (GDNF) and neurotrophin-3 (NT-3). However, the cell cytotoxicity and vasodilation of FSD-C10 were relatively small compared with Fasudil. Although Fasudil inhibited both ROCK I and ROCK II, FSD-C10 more selectively suppressed ROCK II, but not ROCK I, which may be related to vasodilation insensitivity and animal mortality. Thus, FSD-C10 may be a safer and more promising novel ROCK inhibitor than Fasudil for the treatment of several neurological disorders.

**Key words:** Rho kinase, Rho kinase inhibitor, Fasudil, FSD-C10.

Cite this article as: Bioscience Reports (2015) 35, e00247, doi:10.1042/BSR20150032

## INTRODUCTION

Rho kinase (ROCK), a serine/threonine kinase, is activated by binding to the active GTP-bound form of the small GTPase Rho. ROCK is expressed both centrally and peripherally where it is implicated in fundamental cellular processes including migration, proliferation and survival [1]. The research in RhoA/ROCK pathway has attracted much attention for more than a decade since the discovery of ROCK in 1996. A series of studies have demonstrated that the blockade of Rho/ROCK is considered to be beneficial for inflammatory demyelination and degeneration in the central nervous system (CNS) and has proved to be effi-

acious in animal models of stroke [2], multiple sclerosis (MS) [3–6], amyotrophic lateral sclerosis (ALS) [7], Alzheimer's disease (AD) [8] and Parkinson's disease (PD) [9,10]. Therefore, Rho/ROCK pathway is a promising therapeutic target in neurodegenerative and neurotraumatic diseases and ROCK inhibitor should be a promising drug for preventing neurodegeneration and stimulating neuroregeneration in several neurological diseases [11].

Fasudil hydrochloride [hexahydro-1-(5-isoquinolinylsulfonyl)-1H-1,4-diazepine monohydrochloride (HA 1077)] is a isoquinoline sulfonamide derivative which is the most commonly used for pharmacological ROCK inhibitor, both for *in vitro* and *in vivo* studies. At present, Fasudil is only used

**Abbreviations:** AD, Alzheimer's disease; ALS, amyotrophic lateral sclerosis; Arg-1, arginase-1; BDNF, brain-derived neurotrophic factor; CNS, central nervous system; ddH<sub>2</sub>O, double-distilled water; EAE, Encephalomyelitis; GDNF, glial cell line-derived neurotrophic factor; i.p., intraperitoneally; LDH, lactate dehydrogenase; MOG, mouse myelin oligodendrocyte glycoprotein; MS, multiple sclerosis; NT-3, neurotrophin-3; p.i., post-immunization; PD, Parkinson's disease; ROCK, Rho kinase.

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in clinic as a ROCK inhibitor for preventing and improving the cerebral vasospasm after subarachnoid haemorrhage and symptoms of cerebral ischaemia. Previous studies showed that ROCK inhibitor also promotes the survival of neural stem cells, axonal regeneration and differentiation of bone marrow mesenchymal cell into neurons [12,13]. Yamashita and colleagues [14] observed that Fasudil can effect on neurons directly by reducing the activity of ROCK and protect neuronal ischaemic damage in persistent model of cerebral ischaemia.

When Fasudil displays certain beneficial effect, there are many limitations in clinical use, including short-course treatment, low oral bioavailability, cell toxicity and blood pressure fluctuation. Therefore, a considerable interest and efforts have been devoted to the development of novel ROCK inhibitors that should be taken orally for long-term use, with low cytotoxicity and blood pressure fluctuation. We have designed a novel ROCK inhibitor FSD-C10 that exhibits therapeutic potential in experimental autoimmune encephalomyelitis (EAE), an animal model of MS. In the present study, we explored and compared the cell cytotoxicity, neurite outgrowth and dendritic formation, neurotrophic factors, vasodilation and safety between Fasudil and FSD-C10.

## MATERIALS AND METHODS

### ROCK inhibition by mobility shift assay

The inhibition efficiency of Fasudil and FSD-C10 on ROCK activity was measured by mobility shift assay with ATP concentration (Sigma) at 3.6  $\mu\text{M}$  against ROCK I (Carna) and at 5.3  $\mu\text{M}$  against ROCK II (Carna) according to the manufacturer's protocol. Staurosporine (Sigma) was used as positive control and saline was used as negative control.

Fasudil and FSD-C10 were diluted to the final desired highest compound concentration (10  $\mu\text{M}$ ) by 100% DMSO and serially diluted on 96-well plate by transferring 30–60  $\mu\text{l}$  of 100% DMSO in the next well for a total of 10 concentrations in duplicate. DMSO (100  $\mu\text{l}$ ) was added to two empty wells for no compound control and no enzyme control in the same 96-well plate. Mobility shift assay was performed according to the manufacturer's protocol. Briefly, compound Fasudil and FSD-C10 (10  $\mu\text{l}$ ) were mixed with 90  $\mu\text{l}$  of kinase buffer (50 mM HEPES, pH 7.5, 0.0015% Brij-35, 10 mM  $\text{MgCl}_2$ , 2 mM DTT) in 96-well plate. The mixtures (5  $\mu\text{l}$ ) were incubated with 2.5 $\times$  enzyme solution (10  $\mu\text{l}$ ) in 384-well plate at room temperature for 10 min and control was performed by adding 5  $\mu\text{l}$  of kinase buffer. Substrate solution (10  $\mu\text{l}$ ) was added at 28°C and the enzyme reaction was stopped by adding 25  $\mu\text{l}$  of stop solution (100 mM HEPES, pH 7.5, 0.015% Brij-35, 50 mM EDTA) to all wells.

$$\text{Percent inhibition} = \left( \frac{\text{max} - \text{conversion}}{\text{max} - \text{min}} \right) \times 100$$

where max stands for DMSO control; min stands for low control.

### Primary neuron culture

Embryonic 18-day-mouse cortex (Shanghai SLAC Laboratory Animal Co. Ltd) were dissected under a microscope and then triturated into the suspension in Neurobasal-A-medium (Gibco) supplemented with 2% B27 (Gibco), 100 units/ml penicillin and 100  $\mu\text{g}/\text{ml}$  streptomycin (Gibco). The cells were then plated on cover slips coated with poly-D-lysine (0.1 mg/ml; Sigma) at a density of  $1 \times 10^6$  cells per  $\text{cm}^2$  at 37°C in a humidified cell incubator under a 95%/5% (v/v) mixture of air and  $\text{CO}_2$ , with the cell media being replaced every 48 h. After 7 days, neurons were incubated with different conditioned medium.

### BV-2 microglial culture

The BV-2 immortalized microglial cell line [15] was purchased from ShenKe Biological Technology Co. and cultured in Dulbecco's modified Eagle's medium (DMEM; Gibco), supplemented with 10% FBS (Gibco), 100 units/ml penicillin and 100  $\mu\text{g}/\text{ml}$  streptomycin (Gibco) at 37°C in a humidified cell incubator under a 95%/5% (v/v) mixture of air and  $\text{CO}_2$ . Cells were plated on a 96-well plate at a density of  $1.0 \times 10^6$  /ml (200  $\mu\text{l}/\text{well}$ ), on a 24-well plate at  $1.0 \times 10^6$ /ml (500  $\mu\text{l}/\text{well}$ ) and at  $0.2 \times 10^6$  /ml (500  $\mu\text{l}/\text{well}$ ) for different experiments.

### Evaluation of cell state

We used two widely accepted assays, lactate dehydrogenase (LDH) release and MTT reduction assay, for the measurement of cell viability and death. These assays are considered reliable and reproducible with high predictive validity and widely used in various pharmacological studies [16].

#### MTT assay

Cell viability of BV-2 microglia and primary neurons was detected by MTT assay. In brief, 100  $\mu\text{l}$  of MTT solution (0.5 mg  $\text{ml}^{-1}$ , Ducheфа) was added to cultured cells and incubated for an additional 4 h at 37°C, until the medium turned purple. Absorbance at 570 nm was measured by a microplate reader after addition of 100  $\mu\text{l}$  DMSO. Each experiment was performed in triplicate and repeated three times with separate cell preparations. Results were expressed as a percentage of the control value at 570 nm.

#### LDH assay

The death of BV-2 microglia and primary neurons was measured by LDH release in the culture medium. Levels of LDH release in supernatants of cultured cells were measured using the cytotoxicity detection kit (Promega) according to the manufacturer's protocol. Maximum LDH release ( $A_{570}$ ) was determined following the treatment of cells with lysis buffer and considered as control value (100% LDH release). Data were expressed as a percentage of the control value.

## Animals

Female C57BL/6 mice (8–10 weeks old and 18–20 g of body weight) were purchased from Vital River Laboratory Animal Technology Co. Ltd. All experiments were conducted in accordance with the guidelines of the International Council for Laboratory Animal Science. The study was approved by the Ethics Committee of Shanxi Datong University. All mice were housed under pathogen-free conditions and maintained in a reversed 12:12-h light/dark cycle in a temperature controlled room ( $25 \pm 2^\circ\text{C}$ ) for 1 week prior to experimental manipulation.

## Induction and clinical evaluation of EAE

Mouse myelin oligodendrocyte glycoprotein peptide 35–55 (MOG<sub>35–55</sub>, MEVGW YRSPFSRVVHLYRNGK) was produced in an automatic synthesizer (CL Bio-Scientific Company). The purity of the peptide was >95% as determined by HPLC. Chronic EAE was induced by subcutaneous immunization on the upper dorsal flanks with 300  $\mu\text{g}$  of MOG<sub>35–55</sub> in Freund's complete adjuvant (Sigma) supplemented with 3 mg/ml of *M. Tuberculosis* H37Ra (BD Difco; 400 mg/mice). Mice were injected intraperitoneally (i.p.) with 500 ng of pertussis toxin (Enzo Life Sciences) on days 0 and 2 post-immunization (p.i.). Animals were weighed and evaluated for clinical score every other day in a blinded fashion by at least two investigators. Clinical score of EAE was graded according to the following criteria: 0: healthy; 1: limp tail; 2: ataxia and/or paresis of hind limbs; 3: paralysis of hind limbs and/or paresis of forelimbs; 4: tetraparalysis; and 5: moribund or death. When the clinical score of EAE reaches 3, we give suitable care including softening of food with water in dish and additional nutrients such as egg, making mice easy to obtain food and nutrition.

## Administration of Fasudil and FSD-C10

Mice were divided into three groups: Fasudil-treated group ( $n = 8$ ), FSD-C10-treated group ( $n = 8$ ) and double-distilled water (ddH<sub>2</sub>O) EAE control group ( $n = 9$ ). Fasudil or FSD-C10 (from Tianjin Chase Sun Pharmaceutical Co.) was dissolved in sterile ddH<sub>2</sub>O. Mice were i.p. injected with 800  $\mu\text{g}/200 \mu\text{l}$  daily from day 3 to day 27 p.i. Mice that received the same volume of ddH<sub>2</sub>O i.p. served as control.

## Assessment of neurite outgrowth and dendritic length

Fluorescent photomicrographs of neurons with microtubule-associated protein-2 (MAP-2) staining or photomicrographs of BV-2 cells under inverted microscope were captured with a 20 $\times$  objective and a digital camera of Olympus microscope. Images were acquired from randomly selected fields for each group. The length of the longest neurite outgrown or dendritic formation in the cell body was measured by an investigator blinded to the experimental groups using a public-domain image-processing program (Image J, <http://rsb.info.nih.gov/ij>). Average lengths of neuronal neurite or dendritic formation were used for quantification.

## Western blot analysis

Protein from brains was homogenized on ice with an extraction kit and protein concentration was determined by a Bradford protein assay. Protein extracts (30  $\mu\text{g}$ ) were separated by SDS/PAGE and transferred on to a PVDF membrane (Immobilon-P; Millipore Corp.). The membranes were then incubated with anti-GDNF (glial cell line-derived neurotrophic factor; 1:1000, Epitomics), anti-BDNF (brain-derived neurotrophic factor; 1:1000, Promega), anti-NT-3 (neurotrophin-3; 1:1000, Epitomics), anti-Nogo (1:1000, Epitomics), anti-iNOS (inducible nitric oxide synthase) (1:1000, Cell Signaling Technology) Enzo Life Sciences, anti-arginase-1 (Arg-1) (1:1000, BD Bioscience) and anti- $\beta$ -actin (1:10000, Cell Signaling Technology) for overnight at 4 $^\circ\text{C}$ . Bands were visualized by horseradish peroxidase (HRP-) conjugated secondary antibodies and ECL kit under ECL system (GE Healthcare Life Sciences).

## The test of vasodilation

Both Fasudil and FSD-C10 were dissolved in sterile ddH<sub>2</sub>O and the concentration was 4 mg/ml. Naive female C57BL/6 mice were i.p. injected with 200  $\mu\text{l}$  of Fasudil or FSD-C10 and the pictures of limbs were taken at 30 and 60 min after drug injection.

## Statistical analysis

All the experiments were repeated two or three times and GraphPad Prism software was used for statistical analysis. Student's *t* test was performed to analyse the difference between any two groups. A statistically significant difference was assumed at  $P < 0.05$ .

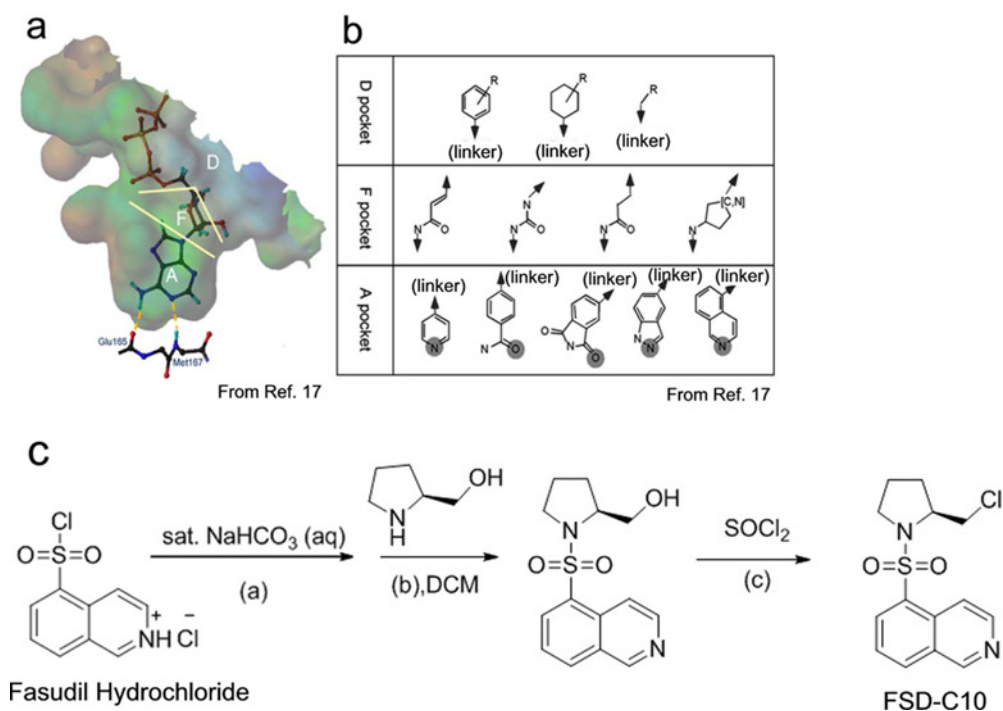
# RESULTS

## Chemical synthesis and structure of FSD-C10

In 2004, Takami et al. [17] reported a ROCK ligand-binding pocket model that is divided into three parts: region A, region F and region D (Figure 1a). It was speculated that the nitrogen atom of region D of Fasudil on the homopiperazine moiety might form a similar hydrogen bond with ATP, which could improve the bioactivity of the compound. Therefore, we designed and synthesized an isoquinoline ROCK inhibitor FSD-C10 targeting region D of Fasudil as the lead compound and on the basis of ligand-binding pocket theory (Figure 1b).

## The inhibition of ROCK by Fasudil and FSD-C10

Figure 2 shows the relationship between compound concentration and percentage of ROCK inhibition. As shown in Table 1, the inhibition of ROCK activity (IC<sub>50</sub>) was measured with ATP concentration at 3.6  $\mu\text{M}$  against ROCK I and at 5.3  $\mu\text{M}$  against ROCK II and observed in Fasudil (ROCK I = 385 nM; ROCK II = 344 nM) and FSD-C10 (ROCK I = 1141 nM; ROCK II = 711 nM) as compared with negative controls



**Figure 1** Synthesis of FSD-C10

(a) Ligand-binding pocket of ROCK homology model and (b) chemical substructures designated for each division of the ligand-binding pocket that are reproduced from [17]: Takami, A., Iwakubo, M., Okada, Y., Kawata, T., Odai, H., Takahashi, N., Shindo, K., Kimura, K., Tagami, Y., Miyake, M. et al. (2004) Design and synthesis of Rho kinase inhibitors. *Bioorg. Med. Chem.* **12**, 2115–2137. The pocket is divided into three parts: region A, F and D. The pocket D region is cleft-like in shape and a wide range of chemical fragments would fit this cleft [17]. Therefore, we designed and synthesized an isoquinoline FSD-C10 targeting region D of the Fasudil. (c) Synthetic process of FSD-C10. (a)  $\text{CH}_2\text{Cl}_2$ , saturated sodium bicarbonate solution,  $0^\circ\text{C}$ . (b)  $\text{CH}_2\text{Cl}_2$ ,  $\text{Et}_3\text{N}$ , 2 h,  $0^\circ\text{C}$ , 95% yield. (c)  $\text{CH}_2\text{Cl}_2$ , reflux, 7 h, 75% yield.

**Table 1** The inhibition efficiency of Fasudil and FSD-C10 on ROCK activity

Compound	Unit (nM)	ROCK I ATP con. (3.6 $\mu\text{M}$ )	ROCK II ATP con. (5.3 $\mu\text{M}$ )
Fasudil	$\text{IC}_{50}$	385	344
FSD-C10	$\text{IC}_{50}$	1141	711
Positive control	$\text{IC}_{50}$	1.7	0.93
Negative control	$\text{IC}_{50}$	>10000	>10000

(ROCK I > 10000; ROCK II > 10000). However, the inhibitory efficiency of FSD-C10 on ROCK activity was weaker than that of Fasudil, especially in ROCK I activity (Table 1).

### The therapeutic effect of Fasudil and FSD-C10 in EAE

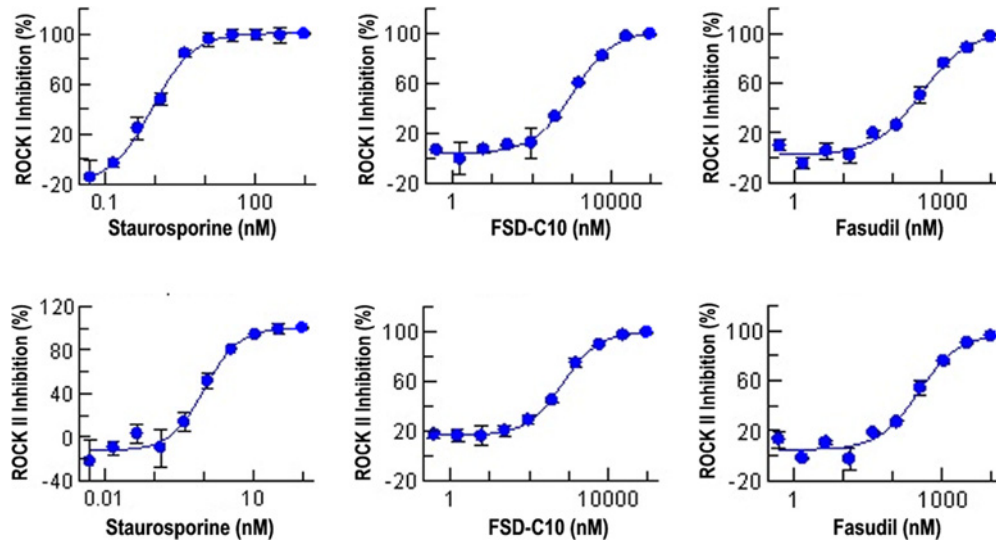
In the present study, mice were immunized with MOG<sub>35-55</sub> peptide to develop an EAE model that closely imitates many characteristics of MS. Starting on day 3 p.i., mice received 800  $\mu\text{g}/200 \mu\text{l}$  daily of Fasudil or FSD-C10 until on day 27 p.i. by i.p. injection. As shown in Figure 3, maximum clinical score in EAE control group ( $n=9$ ) was 2.78, mean clinical

score was  $1.29 \pm 1.08$  compared with mice injected with Fasudil ( $n=8$ , maximum clinical score = 0.19; mean clinical score =  $0.04 \pm 0.07$ ,  $P < 0.001$ ) and FSD-C10 ( $n=8$ , maximum clinical score = 0.50; mean clinical score =  $0.28 \pm 0.23$ ,  $P < 0.001$ ). There is a relationship between loss of body weight and severity of clinical score during the development of EAE. The treatment of Fasudil or FSD-C10 had less body weight loss as compared with that of EAE mice (Figure 3, both  $P < 0.001$ ).

### The effect of Fasudil and FSD-C10 on cell viability and death

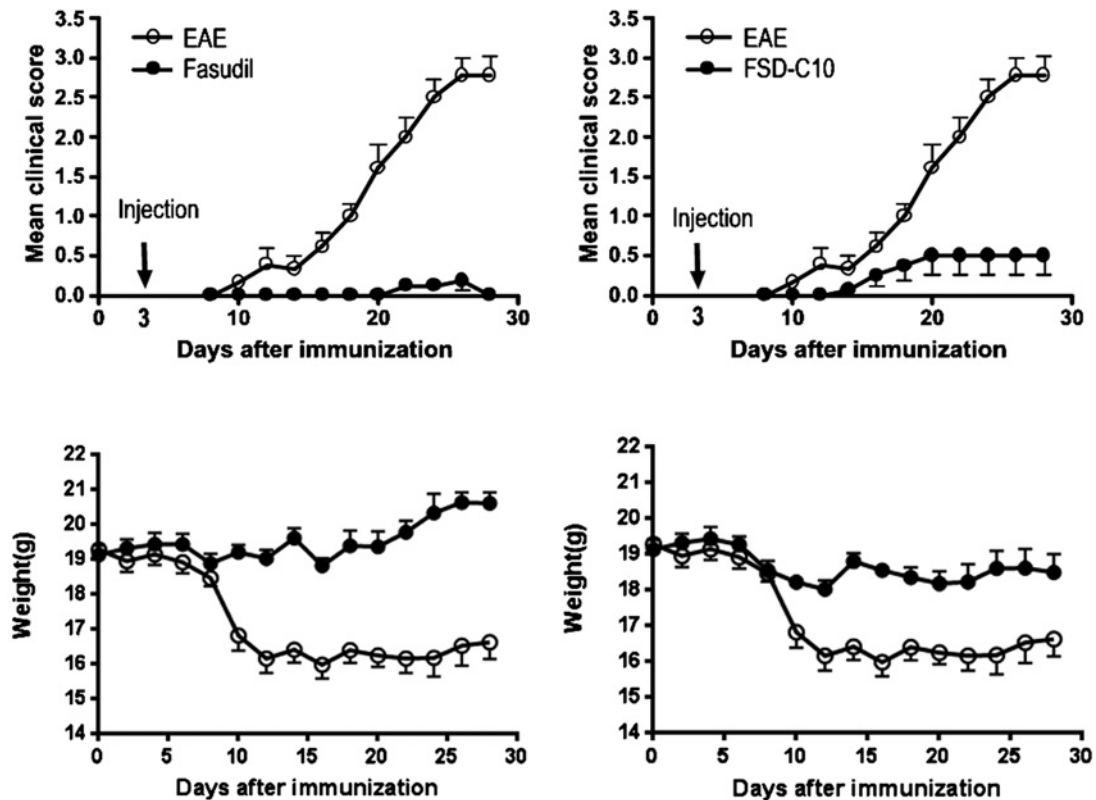
Cell viability and death after culture with Fasudil or FSD-C10 were determined by the MTT and LDH assays (Figure 4). Low/medium-dose Fasudil or FSD-C10 (ranging 0.6–15  $\mu\text{g}/\text{ml}$ ) did not influence the viability and death of cultured primary neurons, whereas high concentration of Fasudil (75  $\mu\text{g}/\text{ml}$ ) caused a significant decrease in neuron viability ( $P < 0.05$ ) and neuron death ( $P < 0.05$ ). In contrast, the same concentration of FSD-C10 did not lead to significant decrease in neuron viability and death compared with control (Figure 4a).

Similarly, low/medium-dose Fasudil or FSD-C10 (ranging 0.6–15  $\mu\text{g}/\text{ml}$ ) did not influence the viability and death of BV-2



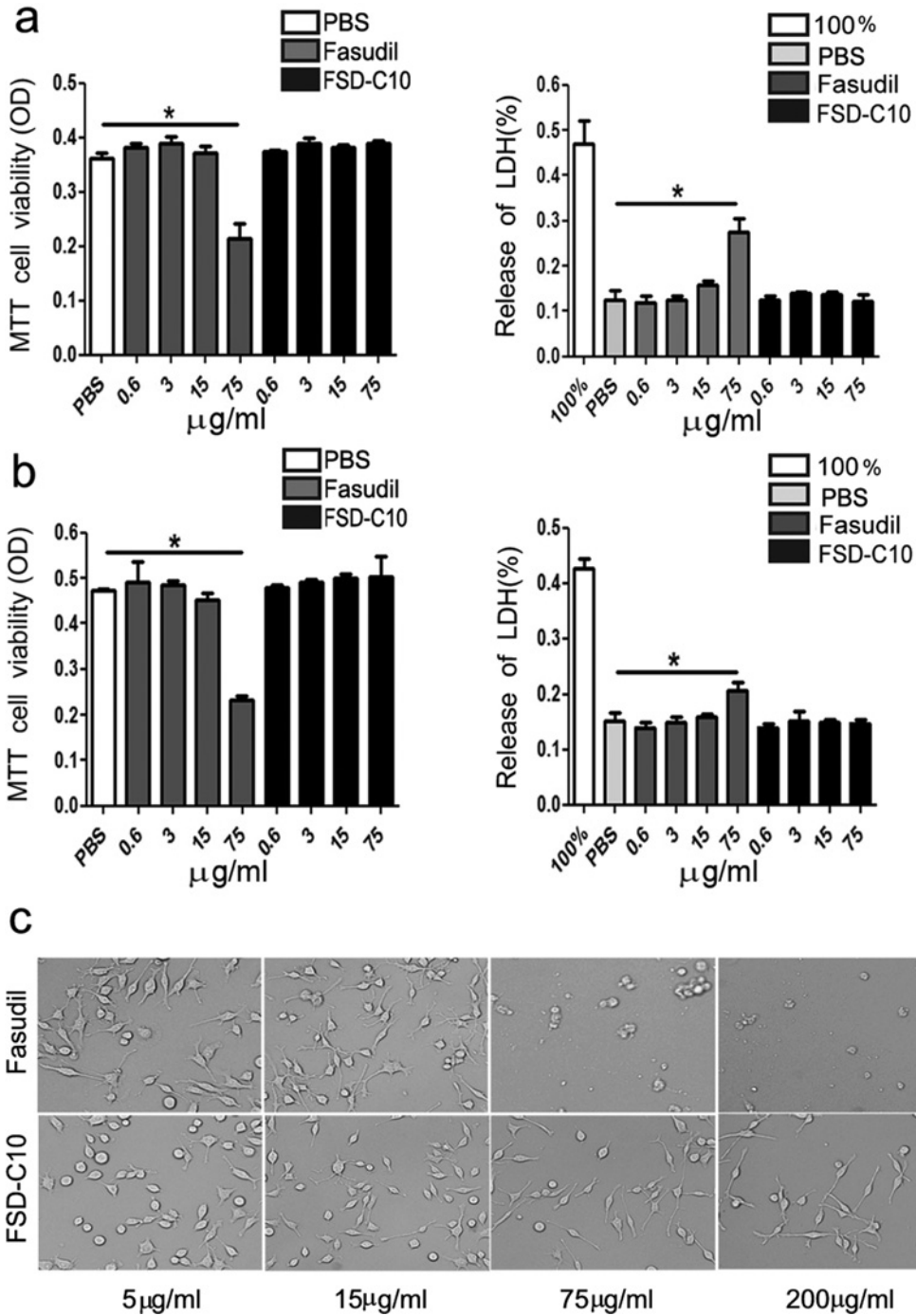
**Figure 2** The relationship between compound concentration and ROCK inhibition

Fasudil and FSD-C10 were diluted to the final desired highest compound concentration (10  $\mu$ M) by 100% DMSO and serially diluted on 96-well plate by transferring 30–60  $\mu$ l of 100% DMSO in the next well for a total of 10 concentrations in duplicate. DMSO (100  $\mu$ l) was used for no compound control and no enzyme control. Percentage of ROCK inhibition was calculated by  $\left(\frac{\text{max-conversion}}{\text{max-min}}\right) \times 100$ . 'Max' stands for DMSO control; 'min' stands for low control.



**Figure 3** Fasudil and FSD-C10 ameliorated the severity of EAE

Chronic EAE was induced in C57BL/6 mice with MOG<sub>35–55</sub> peptide. Mice received Fasudil or FSD-C10 by i.p. injection from day 3 to 27 p.i. Mean clinical score and mean body weight were recorded. The comparison in each time point was separately analysed by Mann–Whitney U test after non-parametric Kruskal–Wallis test.



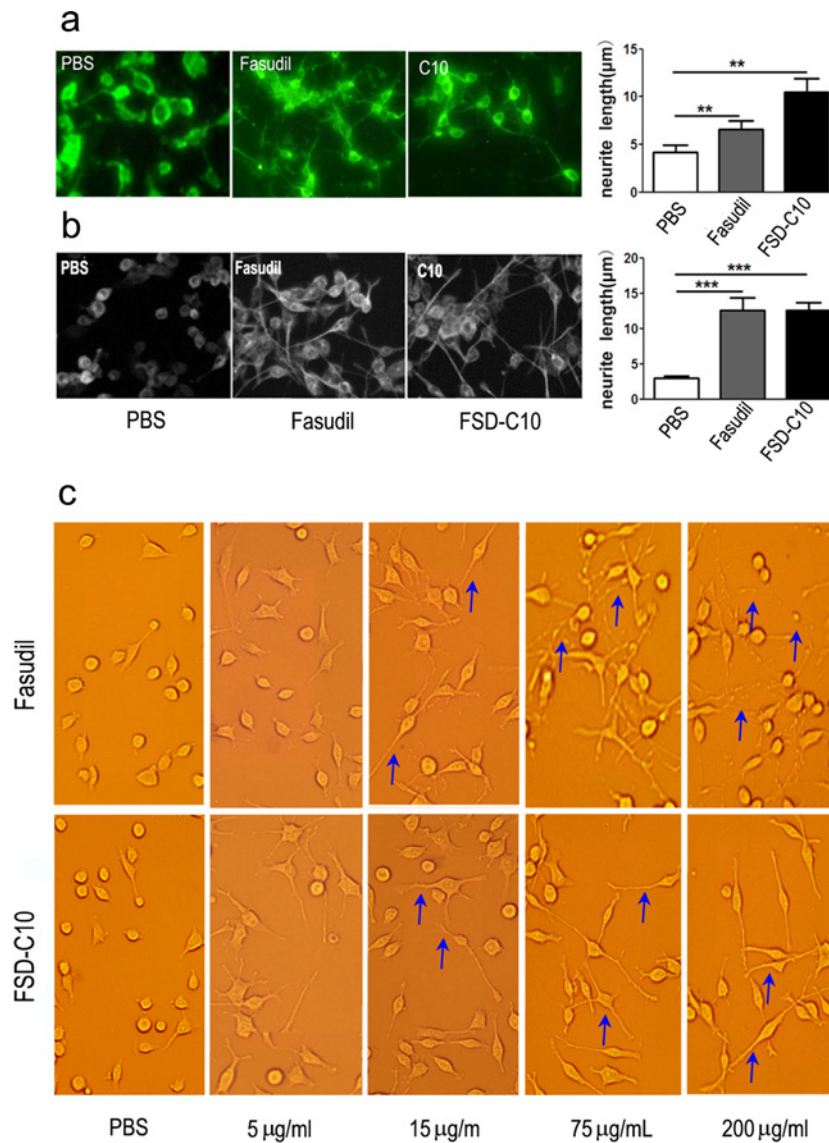
**Figure 4 The viability and death of primary neurons (a) and BV-2 microglia (b) in vitro**

Cell viability was measured using the MTT assay and cell death was detected by LDH release assay. Primary neurons and BV-2 microglia were treated with different concentrations of Fasudil or FSD-C10 (0.6, 3, 15 and 75 µg/ml) for 24 h. The quantitative data are mean ± S.E.M. based on three independent experiments with similar results; \* $P < 0.05$ .

microglia, whereas high concentration of Fasudil (75 µg/ml) caused a significant decrease in BV-2 cell viability ( $P < 0.05$ ) and death ( $P < 0.05$ ). In contrast, FSD-C10 (75 µg/ml) did not lead to significant decrease in BV-2 cell viability and death compared

with control (Figure 4b). As expected, BV-2 microglia treated with higher concentration of Fasudil (75 µg/ml and 200 µg/ml) exhibited obvious cell death after 48 h culture, whereas FSD-C10-exposed cells at similar concentrations showed a good cell





**Figure 5** Fasudil and FSD-C10 promoted neurite outgrowth in primary neurons and dendritic formation in BV-2 microglia. The neurite length of primary neurons (a) and the dendritic length of BV-2 microglia (b) was examined with an inverted Olympus microscope. (c) High concentrations of Fasudil (75 and 200  $\mu\text{g/ml}$ ), but not FSD-C10, caused significant breakage of neurite outgrowth. The length of the longest neurite outgrown or dendritic formation in the cell body was measured by a public-domain image-processing program. The quantitative data are mean  $\pm$  S.E.M. based on three independent experiments with similar results; \*\* $P < 0.01$ , \*\*\* $P < 0.001$ .

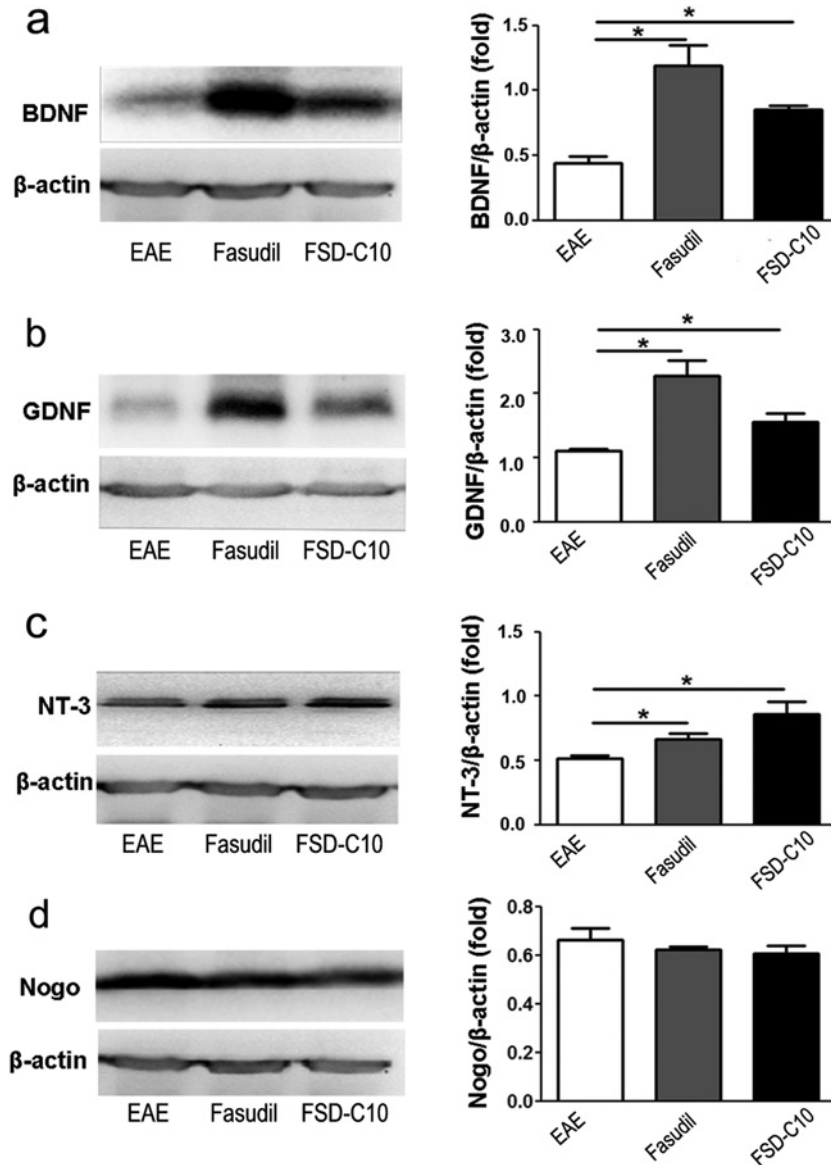
morphology (Figure 4c). However, higher doses of FSD-C10 ( $\geq 200 \mu\text{g/ml}$ ) also caused cell death with the extension of time (result not shown). The results show that FSD-C10 cytotoxicity is lower than Fasudil in *in vitro* experiments.

### The effect of Fasudil and FSD-C10 on neurite outgrowth of neurons and dendritic formation of BV-2 microglia

ROCK has a key role in blocking axon growth and pharmacological ROCK inhibition using small molecules inhibi-

tors has shown to increase axonal regeneration [18,19]. We next explored the effect of Fasudil and FSD-C10 on neurite outgrowth of primary neurons and dendritic formation of BV-2 microglia. As shown in Figure 5(a), the neurite length of primary neurons was significantly prolonged after the treatment of Fasudil (mean = 6.6  $\mu\text{m}$ ) and FSD-C10 (mean = 10.5  $\mu\text{m}$ ) as compared with PBS control (4.1  $\mu\text{m}$ , both  $P < 0.01$ ).

The dendritic length of BV-2 microglia was also significantly prolonged after the treatment of Fasudil (mean = 12.5  $\mu\text{m}$ ) and FSD-C10 (mean = 12.5  $\mu\text{m}$ ) as compared with PBS



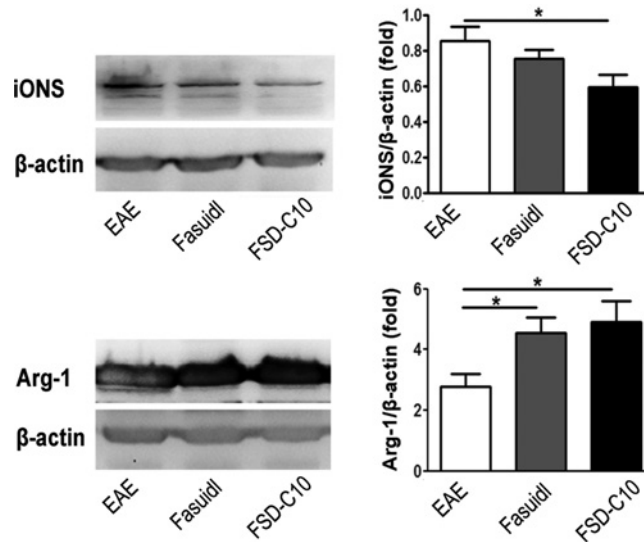
**Figure 6** The effect of Fasudil and FSD-C10 on neuronal growth factors

Chronic EAE was induced in C57BL/6 mice with MOG<sub>35–55</sub> and treated with Fasudil or FSD-C10. On day 28 p.i., brains were harvested to prepare protein for the expression of BDNF (a), GDNF (b), NT-3 (c) and Nogo (d) by Western blot. The results were expressed as the fold change relative to β-actin as the loading control. Quantitative results are mean ± S.E.M. of six mice in each group. \**P* < 0.05.

control (2.94 μm, both *P* < 0.001; Figure 5b). These results indicate that FSD-C10, like Fasudil, exhibited the basic characteristics of ROCK inhibitor, contributing to the neurite outgrowth of neurons and dendritic formation of BV-2 microglia. However, high concentrations of Fasudil (75 and 200 μg/ml) caused significant breakage of dendritic branches (blue arrow), whereas FSD-C10, even at the high concentrations (75 and 200 μg/ml), did not cause significant breakage of dendritic branches on BV2 microglia (Figure 5c).

### The role of Fasudil and FSD-C10 on neurotrophic factors

Previous data suggest that the neurotrophic and neuroprotective molecules directly or/and indirectly contribute to neurite outgrowth of neurons. In the present study, we explored the role of Fasudil and FSD-C10 on production of the neurotrophic factors *in vivo*. Administration of Fasudil or FSD-C10 significantly enhanced expression of BDNF, GDNF and NT-3 proteins in brain compared with control EAE mice (Figures 6a–6c, all *P* < 0.05).



**Figure 7 Fasudil and FSD-C10 shifted M1 to M2 phenotype**

Chronic EAE was induced in C57BL/6 mice with MOG<sub>35–55</sub> and treated with Fasudil or FSD-C10. On day 28 p.i., brains were harvested to prepare protein for the expression of iNOS and Arg-1 by western blot. The results were expressed as the fold change relative to  $\beta$ -actin as the loading control. Quantitative results are mean  $\pm$  S.E.M. of six mice in each group; \* $P < 0.05$ .

In contrast, Fasudil or FSD-C10 administration slightly, although not significantly, declined the expression of Nogo protein level in brain (Figure 6d). These results reveal that the neurite outgrowth of neurons and dendritic formation of BV-2 microglia could be related to increased neurotrophic factors.

### The role of Fasudil and FSD-C10 on the polarization of microglia/macrophages

The classical M1 polarization of macrophages/microglia has been linked with promoting inflammation, whereas the M2 phenotype is anti-inflammatory and promotes tissue repair [1]. Our previous studies clearly demonstrated that Fasudil can convert M1 microglia/macrophages to M2 cells [5,6]. iNOS and Arg-1 are two representatives of M1 and M2 microglia/macrophages respectively and we thus measured their expression in brain of EAE treated with Fasudil or FSD-C10. The results showed that FSD-C10, like Fasudil, inhibited iNOS expression and elevated Arg-1 expression in brain of EAE mice (Figure 7), indicating that FSD-C10, like Fasudil, also converts inflammatory M1 cells toward anti-inflammatory M2 cells *in vivo*, which is consistent with the decrease in inflammatory cytokines described above.

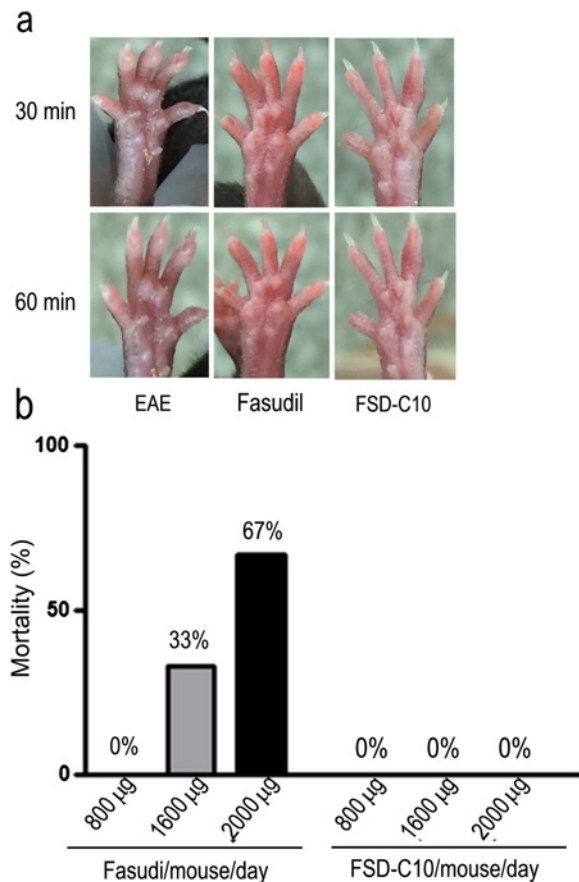
### The influence of Fasudil and FSD-C10 on vasodilation and mortality

The vasodilation and the fluctuation of blood pressure is a side effect and thus limits the long-term use of ROCK inhibitor Fasudil in the nervous system diseases, especially in chronic progressive disorders. We thus compared the role of Fasudil and FSD-C10 on the vasodilation in healthy mice. The results showed

that an obvious vasodilation after 30 and 60 min on the foot of mice was observed in mice that were i.p. injected with Fasudil (800  $\mu$ g/mouse) compared with mice injected with FSD-C10 at the same dose (Figure 8a), revealing that FSD-C10 has a relatively weak vasodilator effect in mice. Simultaneously, the injection of Fasudil (1600 and 2000  $\mu$ g/mouse/i.p) caused nearly 33%–67% mortality within 2 h, whereas none of the mice injected with FSD-C10 died (Figure 8b). Given that FSD-C10 was well tolerant *in vivo*, further pre-clinical studies are valuable for the treatment of MS.

## DISCUSSION

Based on two reasons: (1) ROCK inhibitor should be a promising drug target for preventing neurodegeneration and stimulating neuroregeneration in stroke patients and animal models of stroke, MS, ALS, AD and PD and (2) ROCK inhibitor Fasudil in clinical practice exhibits the following limitations, including a relatively narrow safety window, not suitable for long-term use and poor oral bioavailability, researchers are looking for novel ROCK inhibitors that are more efficient, safer, oral and for long-term use for the treatment of neurological disorders. Our previous studies have demonstrated that a novel ROCK inhibitor FSD-C10 ameliorated the clinical severity of EAE, accompanied by improvement of demyelination and the inhibition of inflammatory cells in the CNS of EAE mice, exhibiting the therapeutic effect in EAE [20]. In the present study, we further compared therapeutic potential, the inhibitory efficiency of ROCK, the cell cytotoxicity,



**Figure 8** The influence of Fasudil and FSD-C10 on vasodilation and mortality

(a) After i.p. injection of FSD-C10 or Fasudil, we observe the vasodilation of mice limbs was observed after 30 and 60 min. A representative of three mice each group is shown. (b) After i.p. injection of FSD-C10 or Fasudil at 800, 1600 and 2000 µg/mouse, animal mortality was recorded with 2 h.

neurite outgrowth and dendritic formation, neurotrophic factors and vasodilation between Fasudil and FSD-C10. Our results first demonstrated that FSD-C10 and Fasudil exerted comparable effect in ameliorating the clinical severity of EAE. Both Fasudil and FSD-C10 induced neurite outgrowth of primary neurons and dendritic formation of BV-2 microglia as well as the production of neurotrophic factor BDNF, GDNF and NT3. Importantly, FSD-C10 treatment in cultured cells and in healthy mice exhibited relatively low cell cytotoxicity and vasodilation respectively, as compared with Fasudil, thus being safer for clinical application.

Consistent with previous findings that the inhibition of ROCK contributes to the neurite outgrowth of neurons [18], our results also showed that both Fasudil and FSD-C10 induced neurite outgrowth of neurons, as well as dendritic formation of BV-2 microglia. The neurite outgrowth is not only a result of a simple expansion and proliferation of neuronal cells, but also entails the neuronal plasticity that is highly dependent on the microenvironment, in which neurotrophic factors promote neurite outgrowth

from cultured adult rat dorsal root ganglion (DRG) neurons [21]. One of them is an involvement of guidance molecules that attract or repulse growing neurite depending on the nerve cell and receptor types. In our study, the expression of three neurotrophic factor BDNF, GDNF and NT-3 was induced by Fasudil or FSD-C10, revealing that neurotrophic factor signalling axis may be a mechanism in neurite outgrowth of primary neurons and dendritic formation of BV-2 microglia.

BDNF is a neurotrophin involved in neuronal survival, differentiation and function, axonal growth and dendritic plasticity in the CNS [22]. BDNF was shown to exert neuroprotective effects in several experimental models of neurological diseases [23–25]. The importance of BDNF for brain function and maintenance is underscored by the fact that BDNF-deficient mice already die during the first weeks of life [26]. Even heterozygous mice with 50% reduced levels of BDNF expression display some behavioural deficits such as aggressiveness, hyperactivity, hyperphagia and obesity [27–29]. GDNF has also been found to be involved in a considerable number of effects in the nervous system, including the survival, migration, differentiation and neurite outgrowth of neurons [30]. NT-3 is a growth factor that modulates glial cell biology and myelination in the CNS and promotes oligodendrocyte precursor proliferation, survival and differentiation [31–33]. Further, NT-3 has significant capacity to provide neuroprotection and reduce astrogliosis, which is an important mechanism underlying the formation of MS plaque [34]. When applied to spinal cord injury, NT-3 overexpression promoted remyelination, axonal regeneration and functional recovery [25]. Following transplantation of NT-3 transduced neural stem cells, NT-3 probably acted not only on donor cells in an autocrine fashion, but also on host cells (paracrine) to enhance neuronal differentiation of both transplanted and endogenous cells and effectively suppressed EAE [35]. In addition, mouse helper T 2 (Th2), but not Th1 cells, expressed a high-affinity receptor for NT-3 and adding NT-3 to cultured Th2 cells to produce a large amount of Interleukin 4 (IL-4) [36], suggesting a direct link between immunomodulation, neuroprotection and therapeutic effects in CNS inflammatory demyelination [37]. Taken together, a set of experiments clearly demonstrate that neurotrophic factor BDNF, GDNF and NT-3 are able to provide excellent environment for neurite outgrowth of neurons that will be ultimately useful for the neural repair and regeneration purposes [38,39].

The role of the Rho/ROCK/LIM domain kinase (LIMK) pathway in neurite outgrowth has been addressed in numerous studies, but the specific contribution of single members of this cascade remained insufficiently understood. Previous studies indicated the inability of adult mammalian CNS axons to regenerate after injury, partly due to the presence of myelin-associated neurite outgrowth inhibitors, including Nogo, myelin-associated glycoprotein (MAG) and MOG. These molecules combine to the same receptor complex and activate downstream signalling molecules, causing the inhibition of neurite growth [40,41]. Rho and ROCK have been identified as downstream signalling molecules of these neurite growth inhibitors and are key elements for neurite growth inhibition and growth cone collapse elicited by these inhibitors [42]. In the present study, the inhibition of ROCK by Fasudil or

FSD-C10 slightly, not significantly though, declined the expression of Nogo. Further studies are necessary to fully evaluate the role of FSD-C10 in the neuroprotection and neuroregeneration.

In addition, ROCK appears to mediate the vasoconstrictor effects and is involved in the regulation of nitric oxide (NO) pathway [43], thus contributing to the maintenance of basal vascular tone [44] and possibly participating in the pathogenesis of human hypertension [45]. Both ROCK I and II are highly homologous and share more than 20 immediate downstream substrates [46]. Two isoforms of ROCK I and II are expressed in vascular smooth muscle cells and inhibit myosin light-chain phosphatase (MLCP) activity [47], whereas ROCK I appears to play a predominant role in vascular inflammation [48]. Additionally, a critical role for ROCK I in mediating cardiac fibrosis was also observed [49]. The activation of Rho/ROCK I signalling disrupts the translocation of occluding/zona occludens-1 (ZO-1), which are the major components of endothelial tight junction. Therefore, we hypothesize that the insensitivity of vascular dilatation in FSD-C10-injected mice may be related to a relatively weak inhibition of ROCK I compared with Fasudil, thus avoiding potential side effect of the latter. Indeed, the role of FSD-C10 in relation to bioavailability, subsequent vascular remodelling and therapeutic potential is still a worthy of further investigation.

#### AUTHOR CONTRIBUTION

Cun-Gen Ma, Bao-Guo Xiao and Guang-Xian Zhang designed the experiments and wrote the manuscript. Yan-Le Xin, Jie-Zhong Yu, Xin-Wang Yang, Chun-Yun Liu, Yan-Hua Li, Ling Feng, Zhi Chai, Wan-Fang Yang, Qing Wang and Wei-Jia Jiang were responsible for performing the cell culture, animal model, immunocytochemistry staining, MTT and LDH assays and western blot assay. Yan-Le Xin performed the statistical analysis.

#### ACKNOWLEDGMENT

We thank Katherine Regan, Department of Neurology and Thomas Jefferson University, Philadelphia, Pennsylvania, U.S.A., for editorial assistance.

#### FUNDING

This work was supported by the National Natural Science Foundation of China [grant numbers 81272163, 81070957, 81070956 and 81371414]; the Department of Science and Technology, Shanxi Province of China [grant number 2013081058]; the Shanxi Scholarship Council of China [grant number 2014-7]; and the Shanxi University of Traditional Chinese Medicine [grant number 2011PY-1].

## REFERENCES

- Guilluy, C., Garcia-Mata, R. and Burrige, K. (2011) Rho protein crosstalk: another social network? *Trends Cell Biol.* **21**, 718–726 [CrossRef PubMed](#)
- Li, Q., Huang, X.J., He, W., Ding, J., Jia, J.T., Fu, G., Wang, H.X. and Guo, L.J. (2009) Injury of neuroprotective potential of fasudil mesylate in brain is chemia-reperfusion rats. *Cell Mol. Neurobiol.* **29**, 169–180 [CrossRef PubMed](#)
- Sun, X., Minohara, M., Kikuchi, H., Ishizu, T., Tanaka, M., Piao, H., Osoegawa, M., Ohyagi, Y., Shimokawa, H. and Kira, J. (2006) The selective Rho kinase inhibitor Fasudil is protective and therapeutic in experimental autoimmune encephalomyelitis. *J. Neuroimmunol.* **180**, 126–134 [CrossRef PubMed](#)
- Huang, X.N., Fu, J. and Wang, W.Z. (2011) The effects of fasudil on the permeability of the rat blood–brain barrier and blood-spinal cord barrier following experimental autoimmune encephalomyelitis. *J. Neuroimmunol.* **239**, 61–67 [CrossRef PubMed](#)
- Hou, S.W., Liu, C.Y., Li, Y.H., Yu, J.Z., Feng, L., Liu, Y.T., Guo, M.F., Xie, Y., Meng, J., Zhang, H.F. et al. (2012) Fasudil ameliorates disease progression in experimental autoimmune encephalomyelitis, acting possibly through antiinflammatory effect. *CNS Neurosci. Ther.* **18**, 909–917 [CrossRef PubMed](#)
- Liu, C.Y., Li, Y.H., Yu, J.Z., Feng, L., Hou, S.W., Liu, Y., Guo, M., Xie, Y., Meng, J., Zhang, H. et al. (2013) Targeting the shift from M1 to M2 macrophages in experimental autoimmune encephalomyelitis mice treated with Fasudil. *PLoS One* **8**, e54844 [CrossRef PubMed](#)
- Takata, M., Tanaka, H., Kimura, M., Nagahara, Y., Tanaka, K., Kawasaki, K., Seto, M., Tsuruma, K., Shimazawa, M. and Hara, H. (2013) Fasudil, a rho kinase inhibitor, limits motor neuron loss in experimental models of amyotrophic lateral sclerosis. *Br. J. Pharmacol.* **170**, 341–351 [CrossRef PubMed](#)
- Song, Y., Chen, X., Wang, L.Y., Gao, W. and Zhu, M.J. (2013) Rho kinase inhibitor fasudil protects against  $\beta$ -amyloid-induced hippocampal neurodegeneration in rats. *CNS Neurosci. Ther.* **19**, 603–610 [CrossRef PubMed](#)
- Tönges, L., Frank, T., Tatenhorst, L., Saal, K.A., Koch, J.C., Szego, É.M., Bähr, M., Weishaupt, J.H. and Lingor, P. (2012) Inhibition of rho kinase enhances survival of dopaminergic neurons and attenuates axonal loss in a mouse model of Parkinson's disease. *Brain* **135**, 3355–3370 [CrossRef PubMed](#)
- Rodriguez-Perez, A.I., Dominguez-Mejide, A., Lanciego, J.L., Guerra, M.J. and Labandeira-Garcia, J.L. (2013) Inhibition of Rho kinase mediates the neuroprotective effects of estrogen in the MPTP model of Parkinson's disease. *Neurobiol. Dis.* **58**, 209–219 [CrossRef PubMed](#)
- Mueller, B.K., Mack, H. and Teusch, N. (2005) Rho kinase, a promising drug target for neurological disorders. *Nat. Rev. Drug. Discov.* **4**, 387–398 [CrossRef PubMed](#)
- Ding, J., Yu, J.Z., Li, Q.Y., Wang, X., Lu, C.Z. and Xiao, B.G. (2009) Rho kinase inhibitor Fasudil Induces neuroprotection and neurogenesis partially through as trocyte-derived G-CSF. *Brain Behav. Immun.* **23**, 1083–1088 [CrossRef PubMed](#)
- Ding, J., Li, Q.Y., Yu, J.Z., Wang, X., Sun, C.H., Lu, C.Z. and Xiao, B.G. (2010) Fasudil, a Rho kinase inhibitor, drives mobilization of adult neural stem cells after hypoxia/reoxygenation injury in mice. *Mol. Cell. Neurosci.* **43**, 201–208 [CrossRef PubMed](#)
- Shimokawa, H., Yamashita, K., Kotani, Y., Nakajima, Y., Shimazawa, M., Yoshimura, S., Nakashima, S., Iwama, T. and Hara, H. (2007) Fasudil, a Rho kinase (ROCK) inhibitor, protects against ischemic neuronal damage *in vitro* and *in vivo* by acting directly on neurons. *Brain Res.* **1154**, 215–224 [CrossRef PubMed](#)
- Chao, M.V. (1992) Neurotrophin receptors: a window into neuronal differentiation. *Neuron* **9**, 583–593 [CrossRef PubMed](#)
- Nilsen, J. and Brinton, R.D. (2002) Impact of progesterone estrogen-induced neuroprotection: synergy by progesterone and 19-norprogesterone and antagonism by medroxyprogesterone acetate. *Endocrinology* **143**, 205–212 [CrossRef PubMed](#)
- Takami, A., Iwakubo, M., Okada, Y., Kawata, T., Odai, H., Takahashi, N., Shindo, K., Kimura, K., Tagami, Y., Miyake, M. et al. (2004) Design and synthesis of Rho kinase inhibitors. *Bioorg. Med. Chem.* **12**, 2115–2137 [CrossRef PubMed](#)



- 18 Lingor, P., Teusch, N., Schwarz, K., Muller, R., Mack, H., Bahr, M. and Mueller, B.K. (2007) Inhibition of Rho kinase (ROCK) increases neurite outgrowth on chondroitin sulphate proteoglycan *in vitro* and axonal regeneration in the adult optic nerve *in vivo*. *J. Neurochem.* **103**, 181–189 [PubMed](#)
- 19 Lingor, P., Tonges, L., Pieper, N., Bermel, C., Barski, E., Planchamp, V. and Bahr, M. (2008) ROCK inhibition and CNTF interact on intrinsic signalling pathways and differentially regulate survival and regeneration in retinal ganglion cells. *Brain* **131**, 250–263 [PubMed](#)
- 20 Li, Y.H., Yu, J.Z., Liu, C.Y., Zhang, H., Zhang, H.F., Yang, W.F., Li, J.L., Feng, Q.J., Feng, L., Zhang, G.X. et al. (2014) Intranasal delivery of FSD-C10, a novel Rho kinase inhibitor, exhibits therapeutic potential in experimental autoimmune encephalomyelitis. *Immunology* **143**, 219–229 [CrossRef PubMed](#)
- 21 Takaku, S., Yanagisawa, H., Watabe, K., Horie, H., Kadoya, T., Sakumi, K., Nakabeppu, Y., Poirier, F. and Sango, K. (2013) GDNF promotes neurite outgrowth and upregulates galectin-1 through the RET/PI3K signaling in cultured adult rat dorsal root ganglion neurons. *Neurochem. Int.* **62**, 330–339 [CrossRef PubMed](#)
- 22 Chao, M.V. (2003) Neurotrophins and their receptors: a convergence point for many signalling pathways. *Nat. Rev. Neurosci.* **4**, 299–309 [CrossRef PubMed](#)
- 23 Hammarberg, H., Lidman, O., Lundberg, C., Eltayeb, S.Y., Gielen, A.W., Muhallab, S., Svenningsson, A., Lindå, H., Meide, P.H., Cullheim, S. et al. (2000) Neuroprotection by encephalomyelitis: rescue of mechanically injured neurons and neurotrophin production by CNS-infiltrating T and natural killer cells. *J. Neurosci.* **20**, 5283–5291 [PubMed](#)
- 24 Hohlfeld, R., Kerschensteiner, M., Stadelmann, C., Lassmann, H. and Wekerle, H. (2000) The neuroprotective effect of inflammation: implications for the therapy of multiple sclerosis. *J. Neuroimmunol.* **107**, 161–166 [CrossRef PubMed](#)
- 25 McTigue, D.M., Horner, P.J., Stokes, B.T. and Gage, F.H. (1998) Neurotrophin-3 and brain-derived neurotrophic factor induce oligodendrocyte proliferation and myelination of regenerating axons in the contused adult rat spinal cord. *J. Neurosci.* **18**, 5354–5365 [PubMed](#)
- 26 Korte, M., Carroll, P., Wolf, E., Brem, G., Thoenen, H. and Bonhoeffer, T. (1995) Hippocampal long-term potentiation is impaired in mice lacking brain-derived neurotrophic factor. *Proc. Natl. Acad. Sci. U.S.A.* **92**, 8856–8860 [CrossRef PubMed](#)
- 27 Kernie, S.G., Liebl, D.J. and Parada, L.F. (2000) BDNF regulates eating behavior and locomotor activity in mice. *EMBO J.* **19**, 1290–1300 [CrossRef PubMed](#)
- 28 Lyons, W.E., Mamounas, L.A., Ricaurte, G.A., Coppola, V., Reid, S.W., Bora, S.H., Wihler, C., Koliatsos, V.E. and Tessarollo, L. (1999) Brain-derived neurotrophic factor-deficient mice develop aggressiveness and hyperphagia in conjunction with brain serotonergic abnormalities. *Proc. Natl. Acad. Sci. U.S.A.* **96**, 15239–15244 [CrossRef PubMed](#)
- 29 Rios, M., Fan, G.P., Fekete, C., Kelly, J., Bates, B., Kuehn, R. and Jaenisch, R. (2001) Conditional deletion of brain-derived neurotrophic factor in the postnatal brain leads to obesity and hyperactivity. *Mol. Endocrinol.* **15**, 1748–1757 [CrossRef PubMed](#)
- 30 Zhou, X., He, X.H., He, B., Zhu, H.W., Zheng, C.B., Xu, J., Jiang, L., Gu, L.Q., Zhu, J.K., Zhu, Q.T. and Liu, X.L. (2013) Etifoxine promotes glial-derived neurotrophic factor-induced neurite outgrowth in PC12 cells. *Mol. Med. Rep.* **8**, 75–80 [PubMed](#)
- 31 Aharoni, R., Eilam, R., Domev, H., Labunskay, G., Sela, M. and Arnon, R. (2005) The immunomodulator glatiramer acetate augments the expression of neurotrophic factors in brains of experimental autoimmune encephalomyelitis mice. *Proc. Natl. Acad. Sci. U.S.A.* **102**, 19045–19050 [CrossRef PubMed](#)
- 32 Kalcheim, C., Carmeli, C. and Rosenthal, A. (1992) Neurotrophin 3 is a mitogen for cultured neural crest cells. *Proc. Natl. Acad. Sci. U.S.A.* **89**, 1661–1665 [CrossRef PubMed](#)
- 33 Barres, B.A., Raff, M.C., Gaese, F., Bartke, I., Dechant, G. and Barde, Y.A. (1994) A crucial role for neurotrophin-3 in oligodendrocyte development. *Nature* **367**, 371–375 [CrossRef PubMed](#)
- 34 Pluchino, S., Muzio, L., Imitola, J., Deleidi, M., Alfaro-Cervello, C., Salani, G., Porcheri, C., Brambilla, E., Cavasinni, F., Bergamaschi, A. et al. (2008) Persistent inflammation alters the function of the endogenous brain stem cell compartment. *Brain* **131**, 2564–2578 [CrossRef PubMed](#)
- 35 Yang, J., Yan, Y., Xia, Y., Kang, T., Li, X., Ciric, B., Xu, H., Rostami, A. and Zhang, G.X. (2014) Neurotrophin 3 transduction augments remyelinating and immunomodulatory capacity of neural stem cells. *Mol. Ther.* **22**, 440–450 [CrossRef PubMed](#)
- 36 Sekimoto, M., Tsuji, T., Matsuzaki, J., Chamoto, K., Koda, T., Nemoto, K., Degawa, M., Nishimura, S. and Nishimura, T. (2003) Functional expression of the TrkC gene, encoding a high affinity receptor for NT-3, in antigen-specific T helper type 2 (Th2) cells. *Immunol. Lett.* **88**, 221–226 [CrossRef PubMed](#)
- 37 Blits, B., Oudega, M., Boer, G.J., Bartlett Bunge, M. and Verhaagen, J. (2003) Adeno-associated viral vector-mediated neurotrophin gene transfer in the injured adult rat spinal cord improves hind-limb function. *Neuroscience* **118**, 271–281 [CrossRef PubMed](#)
- 38 Park, J.W., Kang, Y.D., Kim, J.S., Lee, J.H. and Kim, H.W. (2014) 3D microenvironment of collagen hydrogel enhances the release of neurotrophic factors from human umbilical cord blood cells and stimulates the neurite outgrowth of human neural precursor cells. *Biochem. Biophys. Res. Commun.* **447**, 400–406 [CrossRef](#)
- 39 Montefusco, S., Esposito, R., D'Andrea, L., Monti, M.C., Dunne, C., Dolan, B., Tosco, A., Marzullo, L. and Clyne, M. (2013) Copper promotes TFF1-mediated Helicobacter pylori colonization. *PLoS One* **8**, e79455 [CrossRef PubMed](#)
- 40 Kubo, T., Hata, K., Yamaguchi, A. and Yamashita, T. (2007) Rho-ROCK inhibitors as emerging strategies to promote nerve regeneration. *Curr. Pharm. Des.* **13**, 2493–2499 [CrossRef PubMed](#)
- 41 Wang, X., Chun, S.J., Treloar, H., Vartanian, T., Greer, C.A. and Strittmatter, S.M. (2002) Localization of Nogo-A and Nogo-66 receptor proteins at sites of axon-myelin and synaptic contact. *J. Neurosci.* **22**, 5505–5515 [PubMed](#)
- 42 Schmitz, T. and Chew, L.J. (2008) Cytokines and myelination in the central nervous system. *Scientific World Journal* **8**, 1119–1147 [CrossRef PubMed](#)
- 43 Ming, X.F., Viswambharan, H., Barandier, C., Ruffieux, J., Kaibuchi, K., Rusconi, S. and Yang, Z.H. (2002) Rho GTPase/Rho kinase negatively regulates endothelial nitric oxide synthase phosphorylation through the inhibition of protein kinase B/Akt in human endothelial cells. *Mol. Cell. Biol.* **22**, 8467–8477 [CrossRef PubMed](#)
- 44 Bussemaker, E., Pistrosch, F., Forster, S., Herbrig, K., Gross, P., Passauer, J. and Brandes, R.P. (2007) Rho kinase contributes to basal vascular tone in humans: role of endothelium derived nitric oxide. *Am. J. Physiol. Heart Circ. Physiol.* **293**, H541–H547 [CrossRef PubMed](#)
- 45 Masumoto, A., Hirooka, Y., Shimokawa, H., Hironaga, K., Setoguchi, S. and Takeshita, A. (2001) Possible involvement of Rho-kinase in the pathogenesis of hypertension in humans. *Hypertension* **38**, 1307–1310 [CrossRef PubMed](#)
- 46 Loiran, G. and Pacaud, P. (2010) The of Pho protein in hypertension. *Nat. Rev. Cardiol.* **7**, 637–647 [CrossRef PubMed](#)
- 47 Wang, Y., Zheng, X.R., Riddick, N., Bryden, M., Baur, W., Zhang, X. and Surks, H.K. (2009) ROCK Isoform regulation of myosin phosphatase and contractility in vascular smooth muscle cells. *Circ. Res.* **104**, 531–540 [CrossRef PubMed](#)

48 Noma, K., Rikitake, Y., Oyama, N., Yan, G.J., Alcaide, P., Liu, P.Y., Wang, H.W., Ahl, D., Sawada, N., Okamoto, R. et al. (2008) ROCK1 mediates leukocyte recruitment and neointima formation following vascular injury. *J. Clin. Invest.* **118**, 1632–1644 [CrossRef](#)

49 Zhang, Y.M., Bo, J., Taffet, G.E., Chang, J., Shi, J.J., Reddy, A.K., Michael, L.H., Schneider, M.D., Entman, M.L., Schwartz, R.J. and Wei, L. (2006) Targeted deletion of ROCK1 protects the heart against pressure overload by inhibiting reactive fibrosis. *FASEB J.* **20**, 916–925 [CrossRef](#) [PubMed](#)

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**Received 12 February 2015/7 July 2015; accepted 21 July 2015**

**Accepted Manuscript online 29 July 2015, doi 10.1042/BSR20150032**

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