



NIH PUBLIC ACCESS

Author Manuscript

Arterioscler Thromb Vasc Biol. Author manuscript; available in PMC 2015 August 01.

Published in final edited form as:

Arterioscler Thromb Vasc Biol. 2014 August ; 34(8): 1643–1649. doi:10.1161/ATVBAHA.114.303033.

Perivascular adipose tissue and coronary vascular disease

Meredith Kohr Owen¹, Jillian N. Noblet², Daniel J. Sassoon², Abass M. Conteh², Adam G. Goodwill², and Johnathan D. Tune²

¹Department of Cell Biology and Physiology, University of North Carolina School of Medicine, Chapel Hill, NC 27599

²Department of Cellular and Integrative Physiology, Indiana University School of Medicine, Indianapolis, IN 46202

Abstract

Coronary perivascular adipose tissue (PVAT) is a naturally occurring adipose tissue depot that normally surrounds the major coronary arteries on the surface of the heart. While originally thought to promote vascular health and integrity, there is a growing body of evidence to support that coronary PVAT displays a distinct phenotype relative to other adipose depots and is capable of producing local factors with the potential to augment coronary vascular tone, inflammation, and the initiation and progression of coronary artery disease. The purpose of the present review is outline previous findings regarding the cardiovascular effects of coronary PVAT and the potential mechanisms by which adipose-derived factors may influence coronary vascular function and the progression of atherogenesis.

Introduction

Coronary perivascular adipose tissue (PVAT) is a visceral adipose tissue of mesothelial origin that normally surrounds the major coronary arteries on the surface of the heart^{1, 2}. Coronary PVAT is functionally distinct from the adipose tissue found on the surface of the myocardium, which is defined as myocardial (epicardial) adipose tissue (mEAT)^{3, 4}. In addition to adipocytes and pre-adipocytes, coronary PVAT contains fibroblasts, macrophages, leukocytes, as well as blood vessels and autonomic nerves. With no fascia separating PVAT from the coronary circulation and myocardium, these essential components of the heart share the same microcirculation¹. Originally perceived as a relatively ubiquitous and benign tissue that largely provides structural support and insulation^{5, 6}, it is becoming clear that factors derived from PVAT (adipokines) are capable of influencing a variety of key (patho)physiologic parameters. In particular, recent data support that cardiac adiposity expands with obesity⁷, that atherosclerotic plaques occur predominately in coronary arteries that are encased in PVAT⁷⁻¹⁰, and that coronary PVAT volume is positively associated with underlying plaque burden¹¹. Patients with high mEAT volume have also been shown to have a higher incidence of atrial fibrillation, independent of

Author for correspondence: Johnathan D. Tune, Ph.D. Department of Cellular and Integrative Physiology Indiana University School of Medicine 635 Barnhill Drive Indianapolis, IN 46202 Phone: 317-274-3433 Fax: 317-274-3318 jtune@iu.edu.

Disclosure None.

left atrium enlargement¹²⁻¹⁴. As such, cardiac adiposity has been identified as an independent risk factor for coronary artery disease^{8, 15, 16} and a predictor of future coronary events¹⁷. While specific adipokines can serve to promote vascular health and integrity^{5, 18, 19}, evidence is mounting in support of marked up-regulation of pro-atherogenic mRNA and protein expression profiles in coronary PVAT and mEAT in the setting of obesity²⁰⁻²⁵. This aberrant regulation of coronary PVAT also correlates with underlying vascular dysfunction and disease in obesity^{23, 26-30}. Thus, there is growing evidence to support the hypothesis that local alterations in PVAT-derived factors contribute to the initiation, progression and expansion of coronary disease²⁴, independent of changes in visceral adipose tissue and/or systemic adipokine levels that may occur in the setting of obesity³¹. The purpose of the present review is to outline current data regarding the cardiovascular effects of coronary PVAT and the potential mechanisms by which adipose-derived factors may influence coronary endothelial and smooth muscle function and the progression of atherogenesis.

Vascular Effects of Peripheral vs. Coronary PVAT

Initial studies in to the vascular effects of peripheral (non-cardiac) PVAT demonstrated significant reductions in contractile responses to a variety of agonists in aorta³²⁻³⁵, mesenteric³⁶⁻³⁸, and human internal thoracic arteries^{39, 40}. This “anti-contractile” (or ADRF) vasodilator effect has been attributed to PVAT-derived adiponectin⁴¹, hydrogen sulfide (H₂S)³⁷, hydrogen peroxide (H₂O₂)³³, and Ang1-7⁴² mediated vasodilation via the opening of voltage-dependent K_v7 channels³⁷, BK_{Ca} channels^{40, 43} and/or Kir channels³³. In contrast, the presence of peripheral PVAT has also been shown to potentiate contraction of mesenteric arteries to electrical field stimulation via increased production of angiotensin II and superoxide^{44, 45}. Recent data from Watts *et al.* implicate chemerin as a PVAT-derived constricting factor in aortic and mesenteric vascular beds⁴⁶. Thus, non-cardiac PVAT is capable of producing factors that illicit both vasodilation and vasoconstriction.

Experiments to elucidate the vascular effects of coronary PVAT are rather limited and somewhat conflicting. Studies in isolated coronary arteries from lean or hypercholesterolemic swine show little to no effect of coronary PVAT on endothelial-dependent vasodilation or coronary contractile responses to endothelin-1, angiotensin II, or the thromboxane A2 mimetic U46619⁴⁷⁻⁴⁹. Alternatively, coronary PVAT has been found to diminish endothelial-dependent dilation in dogs^{29, 50} and to significantly exacerbate underlying coronary endothelial dysfunction in obese swine⁴⁸. Further studies in “clean” (PVAT free) conduit coronary arteries revealed that the addition of coronary PVAT from lean swine augments contractile responses to KCl-induced depolarization and to prostaglandin F_{2α} in proportion to the amount of PVAT added to the bath²³. Interestingly, this effect was also observed in response to mesenteric PVAT, but not subcutaneous PVAT. Furthermore, the constricting effect of coronary PVAT was markedly exaggerated in endothelium intact and denuded coronary arteries from obese swine. Additional findings support that these enhanced effects are associated with substantial alterations in the protein expression of obese coronary PVAT^{23, 24} and with inherent differences in the phenotype of obese smooth muscle cells^{51, 52}. Taken together, these findings indicate that factors derived from coronary PVAT can act to impair endothelial-dependent dilation and potentiate

contractions of coronary vascular smooth muscle, especially in the setting of obesity. Potential mediators and mechanisms of these influences are discussed below.

In summary, the findings to date indicate that the vascular effects of PVAT are highly dependent upon anatomical location of the artery/adipose tissue depot, the species being studied, the pharmacologic agonist(s) used, and the underlying phenotype of the endothelium and smooth muscle in relation to the overall health status of the studied model^{23, 53}. Generally, PVAT from peripheral beds exerts vasodilator “anti-contractile” influences whereas coronary PVAT tends to induce vasoconstrictor effects, which includes attenuation of endothelial-dependent dilation. It is important to recognize that the experimental evidence thus far derives from *in vitro* examination of isolated arteries. Thus, the functional (physiologic) relevance of these vascular influences on the regulation of blood pressure, organ blood flow, and/or progression of disease remains a critical and experimentally difficult question to address moving forward. In addition, more careful examination of the precise cell types and mediators responsible for these effects is also warranted.

Expression Profiles in Coronary PVAT

Recent evidence supports that there are substantial differences in gene and protein expression in different adipose tissue depots (e.g. subcutaneous vs. coronary) and that these profiles are significantly altered in the setting of disease. Examination of PVAT surrounding the major coronary arteries suggests that this adipose depot is phenotypically consistent with both white and brown adipose tissue^{54, 55}. Data from the Weintraub laboratory indicate that adipocytes from human coronary PVAT exhibit a reduced state of adipogenic differentiation compared to adipocytes from other depots from the same subjects (e.g. subcutaneous or perirenal-visceral)⁵⁶ and that expression of pro-inflammatory genes and secretion of cytokines such as IL-6, IL-8, and monocyte chemoattractant protein (MCP-1) is markedly elevated in coronary PVAT vs. other adipose tissue depots and/or in the presence of coronary artery disease^{20, 56} (see Table). Furthermore, recent findings from our laboratory as well as others support that this heightened pro-inflammatory environment of coronary PVAT is markedly exacerbated by obesity and/or with the progression of coronary artery disease^{21-23, 26, 31, 48, 57-59}. In particular, increased expression of “pro-atherogenic” factors including leptin, resistin, tumor necrosis factor- α , IL-6, chemerin and calpastatin have been documented to date^{9, 23, 26, 46, 48, 57, 60-64}. Diminished expression of potentially “vasculoprotective” proteins such as adiponectin, which has been associated with improvements in endothelial function⁶⁵, has also been demonstrated in human coronary PVAT in the setting of obesity and coronary artery disease^{26, 31, 58, 66, 67, 68} (see Figure). Interestingly, augmented expression of the osteogenic factors osteoprotegerin²⁰ and osteoglycin²³ were also recently identified in coronary PVAT. These factors have been previously linked with atherosclerosis and the severity of coronary artery disease^{69, 70}. Accordingly, strong and growing evidence supports that coronary PVAT displays a distinct phenotype relative to other adipose tissue depots and is capable of locally producing factors with the potential to influence the initiation and progression of coronary vascular dysfunction and disease.

Within the context of coronary PVAT expression profiles it is important to consider how factors produced in the coronary adventitia are able to traverse the arterial wall to influence the endothelium and/or vascular smooth muscle. The current hypothesis is that the vasa vasorum, a network of small vessels that supply blood to the walls of large blood vessels, is interspersed within the PVAT and thus is capable of delivering adventitial-derived factors to conduit coronary arteries⁷¹⁻⁷³. This hypothesis is supported by prior studies which have demonstrated that neovascularization of the coronary vasa vasorum precedes the development of overt endothelial dysfunction in swine fed a high cholesterol diet⁷² and by experiments which found increases in blood flow through the vasa vasorum to the intima of atherosclerotic coronary arteries of monkeys⁷⁴. Neovascularization originating from the adventitia has also been associated with the extent of inflammation and coronary disease in humans⁷⁵. Although the temporal association between expansion of the coronary vasa vasorum and the development endothelial dysfunction and atherosclerosis is intriguing, further studies to directly examine this hypothesis for the transit of PVAT-derived factors across the coronary wall are needed.

Pathways Influenced by Coronary PVAT

As outlined above, initial studies regarding the vascular effects of coronary PVAT have shown that factors produced by this depot can impair endothelial-dependent vasodilation and augment coronary smooth muscle constriction, especially in the setting of obesity^{23, 24}. At present we are far from understanding the precise factors and signaling pathways responsible for the vascular effects of coronary PVAT. However, there are recent investigations which provide insight regarding potential mechanisms of PVAT-induced coronary vascular dysfunction.

Data from our laboratory support that coronary PVAT significantly attenuates endothelial dependent dilation of isolated coronary arteries in the setting of obesity⁴⁸. This endothelial dysfunction was associated with elevated expression of the adipokine leptin, which we have demonstrated induces significant reductions in coronary endothelial nitric oxide production via a PKC- β dependent phosphorylation of eNOS at the Thr⁴⁹⁵ inhibitory site^{48, 50, 62}. This hypothesis is supported by additional studies that found that the endothelial effects of obese coronary PVAT are abrogated by the inhibition of leptin receptors with a recombinant, pegylated leptin antagonist or by the inhibition of PKC- β with ruboxistaurin⁴⁸. These findings are corroborated by data from other laboratories which have documented increased activation of PKC- β in obesity⁷⁶⁻⁷⁹. Prior studies have also implicated leptin in other key aspects of atherogenesis, including: 1) monocyte chemattraction⁸⁰; 2) promotion of cholesterol ester accumulation in foam cells⁸¹; 3) reduction of plasma high density lipoprotein cholesterol and apolipoprotein A-I concentrations^{82, 83}; 4) activation of acute phase reactants^{84, 85}; 5) elevation of oxidative stress and modification of plasma lipoproteins⁸⁶; 6) augmented DNA-binding activity of proinflammatory transcription factors⁸⁷.

Alternatively, reductions in adiponectin expression in obese coronary PVAT could facilitate inflammation, endothelial dysfunction, and atherogenesis as recent data from Karastergiou *et al.* indicate that administration of recombinant adiponectin successfully reversed PVAT-

mediated increases in endothelial adhesion molecule expression (ICAM-1) and adhesion of monocytic cells to human coronary artery endothelial cells⁶¹. PVAT-derived adiponectin has also been shown to improve the bioavailability of nitric oxide in gluteal arteries obtained from healthy, but not obese humans⁴¹. Prior studies also demonstrate that adiponectin administration diminishes oxidative stress, inflammation, and improves endothelial function via adenosine monophosphate-activated protein kinase (AMPK)-induced phosphorylation of eNOS at Thr¹⁷⁶^{65, 88, 89}. Taken together, these findings suggest that an imbalance between pro-atherogenic vs. anti-atherogenic PVAT-derived adipokines could serve to activate a number of key regulatory pathways to promote obesity-induced coronary artery disease at a local level. Alterations in these pathways, along with other adipokines such as resistin and tumor necrosis factor- α that are known to negatively impact endothelial function and vascular remodeling⁹⁰⁻⁹⁵ should be further explored.

Recently, Owen *et al.* documented that coronary PVAT is capable of releasing factors that initiate and/or potentiate coronary contraction via activation of voltage-dependent ion channels (i.e. Ca_v1.2 channels)²³. This effect of PVAT was substantially augmented in tissues obtained from obese relative to lean swine, thus suggesting that obesity increases production of “adipose-derived constricting factors” from coronary PVAT. A global proteomic assessment of coronary PVAT supernatant from lean and obese swine revealed substantial alterations in key regulatory pathways, including cellular growth and proliferation (51 molecules) and cellular movement (39 molecules). Of particular interest were increases in RhoA (2.9-fold) and calpastatin (1.6-fold) which are directly linked to smooth muscle contraction, Ca²⁺ sensitization, and the progression of hypertension^{96, 97}. Further studies to examine the effects of calpastatin, a known endogenous calpain inhibitor^{97, 98} revealed that this protein dose-dependently augments contractions of isolated coronary arteries similarly to that of coronary PVAT. Interestingly, interrogation of the Rho-kinase pathway revealed that coronary contractions to lean PVAT are largely mediated via a Rho-dependent pathway, whereas enhanced coronary contractions to obese coronary PVAT occurred independent of Rhokinase signaling (was unaffected by the inhibition of Rho-kinase). These data, along with concurrent evidence that PVAT-derived factors significantly impair coronary vasodilation of H₂O₂-sensitive K⁺ channels²³, indicate that the effects of coronary PVAT are related not only to inherent alterations in coronary PVAT expression profiles but also to underlying mechanistic differences in obese coronary artery smooth muscle cells. This hypothesis is supported by earlier studies from our laboratory and others which have demonstrated that obesity decreases the functional expression of coronary K⁺ channels⁹⁹⁻¹⁰³ and increases coronary Ca_v1.2 channel current, expression, and contraction^{51, 52, 104}.

Implications and Conclusions

Taken together, there is a growing body of evidence to support that changes in the phenotypic expression patterns in coronary PVAT occur concomitantly with mechanistic alterations in endothelium and vascular smooth muscle. These changes appear to be dependent on the unique characteristics of the cell types involved and the underlying environment/milieu in which they reside. However, the extent to which PVAT-derived factors “causally” contribute to changes in vascular expression of K⁺ channels, Ca²⁺

channels, Rho-signaling, macrophage/foam cell formation, and/or regional heterogeneity of smooth muscle differentiation/proliferation and atheroma progression has not been determined. Future research to delineate the involvement of specific adipose tissue cell types, how adipose tissue-derived factors are delivered to the vascular wall and possibly systemic circulation (i.e. vasa vasorum), identity of precise mediators, as well as signaling pathways and end-effector mechanisms influenced by coronary perivascular and epicardial adipose tissue beds remain central questions moving forward.

Acknowledgments

None.

Sources of Funding This publication was made possible by funding from NIH HL092245, HL117620 and by the Indiana University Health-Indiana University School of Medicine Strategic Research Initiative (JDT). Additional support was also provided by the American Heart Association 13POST1681001813 (AGG), HL117620-S1 (AMC), and TL1 TR000162 (DJS).

Abbreviations

ADRF	adipose derived relaxing factor
AMPK	adenosine monophosphate-activated protein kinase
H₂O₂	hydrogen peroxide
H₂S	hydrogen sulfide
mEAT	myocardial epicardial adipose tissue
MCP-1	monocyte chemoattractant protein-1
PKC	protein kinase C
PVAT	perivascular adipose tissue

REFERENCES

1. Iacobellis G, Willens HJ. Echocardiographic epicardial fat: A review of research and clinical applications. *Journal of the American Society of Echocardiography : official publication of the American Society of Echocardiography*. 2009; 22:1311–1319. quiz 1417-1318. [PubMed: 19944955]
2. Iozzo P. Myocardial, perivascular, and epicardial fat. *Diabetes care*. 2011; 34(Suppl 2):S371–379. [PubMed: 21525485]
3. Company JM, Booth FW, Laughlin MH, Arce-Esquivel AA, Sacks HS, Bahouth SW, Fain JN. Epicardial fat gene expression after aerobic exercise training in pigs with coronary atherosclerosis: Relationship to visceral and subcutaneous fat. *Journal of applied physiology*. 2010; 109:1904–1912. [PubMed: 20947714]
4. Verhagen SN, Vink A, van der Graaf Y, Visseren FL. Coronary perivascular adipose tissue characteristics are related to atherosclerotic plaque size and composition. A post-mortem study. *Atherosclerosis*. 2012; 225:99–104. [PubMed: 23022141]
5. Ouchi N, Shibata R, Walsh K. Targeting adiponectin for cardioprotection. *Expert opinion on therapeutic targets*. 2006; 10:573–581. [PubMed: 16848693]
6. Sacks HS. Weight loss in obesity reduces epicardial fat thickness; so what? *Journal of applied physiology*. 2009; 106:1–2. [PubMed: 18948441]

7. Sarin S, Wenger C, Marwaha A, Qureshi A, Go BD, Woomert CA, Clark K, Nassef LA, Shirani J. Clinical significance of epicardial fat measured using cardiac multislice computed tomography. *The American journal of cardiology*. 2008; 102:767–771. [PubMed: 18774004]
8. Ding J, Hsu FC, Harris TB, Liu Y, Kritchevsky SB, Szklo M, Ouyang P, Espeland MA, Lohman KK, Criqui MH, Allison M, Bluemke DA, Carr JJ. The association of pericardial fat with incident coronary heart disease: The multi-ethnic study of atherosclerosis (mesa). *The American journal of clinical nutrition*. 2009; 90:499–504. [PubMed: 19571212]
9. Greif M, Becker A, von Ziegler F, Leberherz C, Lehrke M, Broedl UC, Tittus J, Parhofer K, Becker C, Reiser M, Knez A, Leber AW. Pericardial adipose tissue determined by dual source ct is a risk factor for coronary atherosclerosis. *Arteriosclerosis, thrombosis, and vascular biology*. 2009; 29:781–786.
10. Sacks HS, Fain JN. Human epicardial adipose tissue: A review. *American heart journal*. 2007; 153:907–917. [PubMed: 17540190]
11. Mahabadi AA, Reinsch N, Lehmann N, Altenbernd J, Kalsch H, Seibel RM, Erbel R, Mohlenkamp S. Association of pericoronary fat volume with atherosclerotic plaque burden in the underlying coronary artery: A segment analysis. *Atherosclerosis*. 2010; 211:195–199. [PubMed: 20223460]
12. Al Chekatie MO, Welles CC, Metoyer R, Ibrahim A, Shapira AR, Cytron J, Santucci P, Wilber DJ, Akar JG. Pericardial fat is independently associated with human atrial fibrillation. *Journal of the American College of Cardiology*. 2010; 56:784–788. [PubMed: 20797492]
13. Hatem SN, Sanders P. Epicardial adipose tissue and atrial fibrillation. *Cardiovascular research*. 2014
14. Shin SY, Yong HS, Lim HE, Na JO, Choi CU, Choi JI, Kim SH, Kim JW, Kim EJ, Park SW, Rha SW, Park CG, Seo HS, Oh DJ, Kim YH. Total and interatrial epicardial adipose tissues are independently associated with left atrial remodeling in patients with atrial fibrillation. *Journal of cardiovascular electrophysiology*. 2011; 22:647–655. [PubMed: 21235672]
15. Schlett CL, Massaro JM, Lehman SJ, Bamberg F, O'Donnell CJ, Fox CS, Hoffmann U. Novel measurements of periaortic adipose tissue in comparison to anthropometric measures of obesity, and abdominal adipose tissue. *International journal of obesity*. 2009; 33:226–232. [PubMed: 19139753]
16. Lehman SJ, Massaro JM, Schlett CL, O'Donnell CJ, Hoffmann U, Fox CS. Peri aortic fat, cardiovascular disease risk factors, and aortic calcification: The framingham heart study. *Atherosclerosis*. 2010; 210:656–661. [PubMed: 20152980]
17. Kunita E, Yamamoto H, Kitagawa T, Ohashi N, Oka T, Utsunomiya H, Urabe Y, Tsushima H, Awai K, Budoff MJ, Kihara Y. Prognostic value of coronary artery calcium and epicardial adipose tissue assessed by non-contrast cardiac computed tomography. *Atherosclerosis*. 2014; 233:447–453. [PubMed: 24530777]
18. Essick EE, Ouchi N, Wilson RM, Ohashi K, Ghobrial J, Shibata R, Pimentel DR, Sam F. Adiponectin mediates cardioprotection in oxidative stress-induced cardiac myocyte remodeling. *American journal of physiology. Heart and circulatory physiology*. 2011; 301:H984–993. [PubMed: 21666115]
19. Ouchi N, Shibata R, Walsh K. Cardioprotection by adiponectin. *Trends in cardiovascular medicine*. 2006; 16:141–146. [PubMed: 16781946]
20. Chatterjee TK, Aronow BJ, Tong WS, Manka D, Tang Y, Bogdanov VY, Unruh D, Blomkalns AL, Piegore MG Jr, Weintraub DS, Rudich SM, Kuhel DG, Hui DY, Weintraub NL. Human coronary artery perivascular adipocytes overexpress genes responsible for regulating vascular morphology, inflammation, and hemostasis. *Physiological genomics*. 2013; 45:697–709. [PubMed: 23737535]
21. Gnacinska M, Malgorzewicz S, Lysiak-Szydłowska W, Sworczak K. The serum profile of adipokines in overweight patients with metabolic syndrome. *Endokrynologia Polska*. 2010; 61:36–41. [PubMed: 20205102]
22. Monzillo LU, Hamdy O, Horton ES, Ledbury S, Mullooly C, Jarema C, Porter S, Ovalle K, Moussa A, Mantzoros CS. Effect of lifestyle modification on adipokine levels in obese subjects with insulin resistance. *Obesity research*. 2003; 11:1048–1054. [PubMed: 12972674]

23. Owen MK, Witzmann FA, McKenney ML, Lai X, Berwick ZC, Moberly SP, Alloosh M, Sturek M, Tune JD. Perivascular adipose tissue potentiates contraction of coronary vascular smooth muscle: Influence of obesity. *Circulation*. 2013; 128:9–18. [PubMed: 23685742]
24. Payne GA, Kohr MC, Tune JD. Epicardial perivascular adipose tissue as a therapeutic target in obesity-related coronary artery disease. *British journal of pharmacology*. 2012; 165:659–669. [PubMed: 21545577]
25. Silha JV, Krsek M, Skrha JV, Sucharda P, Nyomba BL, Murphy LJ. Plasma resistin, adiponectin and leptin levels in lean and obese subjects: Correlations with insulin resistance. *European journal of endocrinology / European Federation of Endocrine Societies*. 2003; 149:331–335. [PubMed: 14514348]
26. Cheng KH, Chu CS, Lee KT, Lin TH, Hsieh CC, Chiu CC, Voon WC, Sheu SH, Lai WT. Adipocytokines and proinflammatory mediators from abdominal and epicardial adipose tissue in patients with coronary artery disease. *International journal of obesity*. 2008; 32:268–274. [PubMed: 17878891]
27. Fontana L, Eagon JC, Trujillo ME, Scherer PE, Klein S. Visceral fat adipokine secretion is associated with systemic inflammation in obese humans. *Diabetes*. 2007; 56:1010–1013. [PubMed: 17287468]
28. McKenney ML, Schultz KA, Boyd JH, Byrd JP, Alloosh M, Teague SD, Arce-Esquivel AA, Fain JN, Laughlin MH, Sacks HS, Sturek M. Epicardial adipose excision slows the progression of porcine coronary atherosclerosis. *Journal of cardiothoracic surgery*. 2014; 9:2. [PubMed: 24387639]
29. Payne GA, Borbouse L, Bratz IN, Roell WC, Bohlen HG, Dick GM, Tune JD. Endogenous adipose-derived factors diminish coronary endothelial function via inhibition of nitric oxide synthase. *Microcirculation*. 2008; 15:417–426. [PubMed: 18574744]
30. Tesauro M, Cardillo C. Obesity, blood vessels and metabolic syndrome. *Acta physiologica*. 2011; 203:279–286. [PubMed: 21439028]
31. Karastergiou K, Fried SK. Multiple adipose depots increase cardiovascular risk via local and systemic effects. *Current atherosclerosis reports*. 2013; 15:361. [PubMed: 23982264]
32. Dubrovskaja G, Verlohren S, Luft FC, Gollasch M. Mechanisms of adrf release from rat aortic adventitial adipose tissue. *American journal of physiology. Heart and circulatory physiology*. 2004; 286:H1107–1113. [PubMed: 14644761]
33. Gao YJ, Lu C, Su LY, Sharma AM, Lee RM. Modulation of vascular function by perivascular adipose tissue: The role of endothelium and hydrogen peroxide. *British journal of pharmacology*. 2007; 151:323–331. [PubMed: 17384669]
34. Lohn M, Dubrovskaja G, Lauterbach B, Luft FC, Gollasch M, Sharma AM. Periadventitial fat releases a vascular relaxing factor. *FASEB journal : official publication of the Federation of American Societies for Experimental Biology*. 2002; 16:1057–1063. [PubMed: 12087067]
35. Soltis EE, Cassis LA. Influence of perivascular adipose tissue on rat aortic smooth muscle responsiveness. *Clinical and experimental hypertension. Part A, Theory and practice*. 1991; 13:277–296.
36. Galvez B, de Castro J, Herold D, Dubrovskaja G, Arribas S, Gonzalez MC, Aranguiz I, Luft FC, Ramos MP, Gollasch M, Fernandez Alfonso MS. Perivascular adipose tissue and mesenteric vascular function in spontaneously hypertensive rats. *Arteriosclerosis, thrombosis, and vascular biology*. 2006; 26:1297–1302.
37. Schleifenbaum J, Kohn C, Voblova N, Dubrovskaja G, Zavarirskaya O, Gloe T, Crean CS, Luft FC, Huang Y, Schubert R, Gollasch M. Systemic peripheral artery relaxation by kcnq channel openers and hydrogen sulfide. *Journal of hypertension*. 2010; 28:1875–1882. [PubMed: 20577128]
38. Verlohren S, Dubrovskaja G, Tsang SY, Essin K, Luft FC, Huang Y, Gollasch M. Visceral periaortic adipose tissue regulates arterial tone of mesenteric arteries. *Hypertension*. 2004; 44:271–276. [PubMed: 15302842]
39. Gao YJ, Zeng ZH, Teoh K, Sharma AM, Abouzahr L, Cybulsky I, Lamy A, Semelhago L, Lee RM. Perivascular adipose tissue modulates vascular function in the human internal thoracic artery. *The Journal of thoracic and cardiovascular surgery*. 2005; 130:1130–1136. [PubMed: 16214530]

40. Malinowski M, Deja MA, Janusiewicz P, Golba KS, Roleder T, Wos S. Mechanisms of vasodilatory effect of perivascular tissue of human internal thoracic artery. *Journal of physiology and pharmacology : an official journal of the Polish Physiological Society*. 2013; 64:309–316. [PubMed: 23959727]
41. Greenstein AS, Khavandi K, Withers SB, Sonoyama K, Clancy O, Jeziorska M, Laing I, Yates AP, Pemberton PW, Malik RA, Heagerty AM. Local inflammation and hypoxia abolish the protective anticontractile properties of perivascular fat in obese patients. *Circulation*. 2009; 119:1661–1670. [PubMed: 19289637]
42. Lee RM, Lu C, Su LY, Gao YJ. Endothelium-dependent relaxation factor released by perivascular adipose tissue. *Journal of hypertension*. 2009; 27:782–790. [PubMed: 19516177]
43. Lynch FM, Withers SB, Yao Z, Werner ME, Edwards G, Weston AH, Heagerty AM. Perivascular adipose tissue-derived adiponectin activates bk(ca) channels to induce anticontractile responses. *American journal of physiology. Heart and circulatory physiology*. 2013; 304:H786–795. [PubMed: 23292715]
44. Lu C, Su LY, Lee RM, Gao YJ. Superoxide anion mediates angiotensin ii-induced potentiation of contractile response to sympathetic stimulation. *European journal of pharmacology*. 2008; 589:188–193. [PubMed: 18538762]
45. Gao YJ, Takemori K, Su LY, An WS, Lu C, Sharma AM, Lee RM. Perivascular adipose tissue promotes vasoconstriction: The role of superoxide anion. *Cardiovascular research*. 2006; 71:363–373. [PubMed: 16756966]
46. Watts SW, Dorrance AM, Penfold ME, Rourke JL, Sinal CJ, Seitz B, Sullivan TJ, Charvat TT, Thompson JM, Burnett R, Fink GD. Chemerin connects fat to arterial contraction. *Arteriosclerosis, thrombosis, and vascular biology*. 2013; 33:1320–1328.
47. Bunker AK, Laughlin MH. Influence of exercise and perivascular adipose tissue on coronary artery vasomotor function in a familial hypercholesterolemic porcine atherosclerosis model. *Journal of applied physiology*. 2010; 108:490–497. [PubMed: 19959766]
48. Payne GA, Borbouse L, Kumar S, Neeb Z, Alloosh M, Sturek M, Tune JD. Epicardial perivascular adipose-derived leptin exacerbates coronary endothelial dysfunction in metabolic syndrome via a protein kinase c-beta pathway. *Arteriosclerosis, thrombosis, and vascular biology*. 2010; 30:1711–1717.
49. Reifemberger MS, Turk JR, Newcomer SC, Booth FW, Laughlin MH. Perivascular fat alters reactivity of coronary artery: Effects of diet and exercise. *Medicine and science in sports and exercise*. 2007; 39:2125–2134. [PubMed: 18046183]
50. Payne GA, Bohlen HG, Dincer UD, Borbouse L, Tune JD. Periadventitial adipose tissue impairs coronary endothelial function via pkc-beta-dependent phosphorylation of nitric oxide synthase. *American journal of physiology. Heart and circulatory physiology*. 2009; 297:H460–465. [PubMed: 19482966]
51. Berwick ZC, Dick GM, O’Leary HA, Bender SB, Goodwill AG, Moberly SP, Owen MK, Miller SJ, Obukhov AG, Tune JD. Contribution of electromechanical coupling between kv and ca v1.2 channels to coronary dysfunction in obesity. *Basic research in cardiology*. 2013; 108:370. [PubMed: 23856709]
52. Berwick ZC, Moberly SP, Kohr MC, Morrical EB, Kurian MM, Dick GM, Tune JD. Contribution of voltage-dependent k+ and ca2+ channels to coronary pressure-flow autoregulation. *Basic research in cardiology*. 2012; 107:264. [PubMed: 22466959]
53. Lim S, Meigs JB. Ectopic fat and cardiometabolic and vascular risk. *International journal of cardiology*. 2013; 169:166–176. [PubMed: 24063931]
54. Police SB, Thatcher SE, Charnigo R, Daugherty A, Cassis LA. Obesity promotes inflammation in periaortic adipose tissue and angiotensin ii-induced abdominal aortic aneurysm formation. *Arteriosclerosis, thrombosis, and vascular biology*. 2009; 29:1458–1464.
55. Sacks HS, Fain JN, Holman B, Cheema P, Chary A, Parks F, Karas J, Optican R, Bahouth SW, Garrett E, Wolf RY, Carter RA, Robbins T, Wolford D, Samaha J. Uncoupling protein-1 and related messenger ribonucleic acids in human epicardial and other adipose tissues: Epicardial fat functioning as brown fat. *The Journal of clinical endocrinology and metabolism*. 2009; 94:3611–3615. [PubMed: 19567523]

56. Chatterjee TK, Idelman G, Blanco V, Blomkalns AL, Piegore MG Jr, Weintraub DS, Kumar S, Rajshaker S, Manka D, Rudich SM, Tang Y, Hui DY, Bassel-Duby R, Olson EN, Lingrel JB, Ho SM, Weintraub NL. Histone deacetylase 9 is a negative regulator of adipogenic differentiation. *The Journal of biological chemistry*. 2011; 286:27836–27847. [PubMed: 21680747]
57. Baker AR, Silva NF, Quinn DW, Harte AL, Pagano D, Bonser RS, Kumar S, McTernan PG. Human epicardial adipose tissue expresses a pathogenic profile of adipocytokines in patients with cardiovascular disease. *Cardiovascular diabetology*. 2006; 5:1. [PubMed: 16412224]
58. Langheim S, Dreas L, Veschini L, Maisano F, Foglieni C, Ferrarello S, Sinagra G, Zingone B, Alfieri O, Ferrero E, Maseri A, Ruotolo G. Increased expression and secretion of resistin in epicardial adipose tissue of patients with acute coronary syndrome. *American journal of physiology. Heart and circulatory physiology*. 2010; 298:H746–753. [PubMed: 20061546]
59. Lau DC, Dhillon B, Yan H, Szmilko PE, Verma S. Adipokines: Molecular links between obesity and atherosclerosis. *American journal of physiology. Heart and circulatory physiology*. 2005; 288:H2031–2041. [PubMed: 15653761]
60. Chatterjee TK, Stoll LL, Denning GM, Harrelson A, Blomkalns AL, Idelman G, Rothenberg FG, Neltner B, Romig-Martin SA, Dickson EW, Rudich S, Weintraub NL. Proinflammatory phenotype of perivascular adipocytes: Influence of high-fat feeding. *Circulation research*. 2009; 104:541–549. [PubMed: 19122178]
61. Karastergiou K, Evans I, Ogston N, Miheisi N, Nair D, Kaski JC, Jahangiri M, Mohamed-Ali V. Epicardial adipokines in obesity and coronary artery disease induce atherogenic changes in monocytes and endothelial cells. *Arteriosclerosis, thrombosis, and vascular biology*. 2010; 30:1340–1346.
62. Knudson JD, Dincer UD, Dick GM, Shibata H, Akahane R, Saito M, Tune JD. Leptin resistance extends to the coronary vasculature in prediabetic dogs and provides a protective adaptation against endothelial dysfunction. *American journal of physiology. Heart and circulatory physiology*. 2005; 289:H1038–1046. [PubMed: 15894577]
63. Mazurek T, Zhang L, Zalewski A, Mannion JD, Diehl JT, Arafat H, Sarov-Blat L, O'Brien S, Keiper EA, Johnson AG, Martin J, Goldstein BJ, Shi Y. Human epicardial adipose tissue is a source of inflammatory mediators. *Circulation*. 2003; 108:2460–2466. [PubMed: 14581396]
64. Shibasaki I, Nishikimi T, Mochizuki Y, Yamada Y, Yoshitatsu M, Inoue Y, Kuwata T, Ogawa H, Tsuchiya G, Ishimitsu T, Fukuda H. Greater expression of inflammatory cytokines, adrenomedullin, and natriuretic peptide receptor-c in epicardial adipose tissue in coronary artery disease. *Regulatory peptides*. 2010; 165:210–217. [PubMed: 20691218]
65. Beltowski J, Jamroz-Wisniewska A, Widomska S. Adiponectin and its role in cardiovascular diseases. *Cardiovascular & hematological disorders drug targets*. 2008; 8:7–46. [PubMed: 18336252]
66. Iacobellis G, Corradi D, Sharma AM. Epicardial adipose tissue: Anatomic, biomolecular and clinical relationships with the heart. *Nature clinical practice. Cardiovascular medicine*. 2005; 2:536–543.
67. Eiras S, Teijeira-Fernandez E, Shamagian LG, Fernandez AL, Vazquez-Boquete A, Gonzalez-Juanatey JR. Extension of coronary artery disease is associated with increased il-6 and decreased adiponectin gene expression in epicardial adipose tissue. *Cytokine*. 2008; 43:174–180. [PubMed: 18562207]
68. Spiroglou SG, Kostopoulos CG, Varakis JN, Papadaki HH. Adipokines in periaortic and epicardial adipose tissue: Differential expression and relation to atherosclerosis. *Journal of atherosclerosis and thrombosis*. 2010; 17:115–130. [PubMed: 20145358]
69. Ikeda T, Shirasawa T, Esaki Y, Yoshiki S, Hirokawa K. Osteopontin mRNA is expressed by smooth muscle-derived foam cells in human atherosclerotic lesions of the aorta. *The Journal of clinical investigation*. 1993; 92:2814–2820. [PubMed: 8254036]
70. Tousoulis D, Siasos G, Maniatis K, Oikonomou E, Kioufis S, Zaromitidou M, Paraskevopoulos T, Michalea S, Kollia C, Miliou A, Kokkou E, Papavassiliou AG, Stefanadis C. Serum osteoprotegerin and osteopontin levels are associated with arterial stiffness and the presence and severity of coronary artery disease. *International journal of cardiology*. 2013; 167:1924–1928. [PubMed: 22640692]

71. Gossel M, Versari D, Mannheim D, Ritman EL, Lerman LO, Lerman A. Increased spatial vasa vasorum density in the proximal lad in hypercholesterolemia--implications for vulnerable plaque-development. *Atherosclerosis*. 2007; 192:246–252. [PubMed: 16919638]
72. Herrmann J, Lerman LO, Rodriguez-Porcel M, Holmes DR Jr, Richardson DM, Ritman EL, Lerman A. Coronary vasa vasorum neovascularization precedes epicardial endothelial dysfunction in experimental hypercholesterolemia. *Cardiovascular research*. 2001; 51:762–766. [PubMed: 11530109]
73. Moreno PR, Purushothaman KR, Sirol M, Levy AP, Fuster V. Neovascularization in human atherosclerosis. *Circulation*. 2006; 113:2245–2252. [PubMed: 16684874]
74. Heistad DD, Armstrong ML. Blood flow through vasa vasorum of coronary arteries in atherosclerotic monkeys. *Arteriosclerosis*. 1986; 6:326–331. [PubMed: 3707431]
75. Kumamoto M, Nakashima Y, Sueishi K. Intimal neovascularization in human coronary atherosclerosis: Its origin and pathophysiological significance. *Human pathology*. 1995; 26:450–456. [PubMed: 7535741]
76. Bohlen HG. Mechanisms for early microvascular injury in obesity and type ii diabetes. *Current hypertension reports*. 2004; 6:60–65. [PubMed: 14972096]
77. Casellini CM, Barlow PM, Rice AL, Casey M, Simmons K, Pittenger G, Bastyr EJ 3rd, Wolka AM, Vinik AI. A 6-month, randomized, double-masked, placebo-controlled study evaluating the effects of the protein kinase c-beta inhibitor ruboxistaurin on skin microvascular blood flow and other measures of diabetic peripheral neuropathy. *Diabetes care*. 2007; 30:896–902. [PubMed: 17392551]
78. Mehta NN, Sheetz M, Price K, Comiskey L, Amrutia S, Iqbal N, Mohler ER, Reilly MP. Selective pkc beta inhibition with ruboxistaurin and endothelial function in type-2 diabetes mellitus. *Cardiovascular drugs and therapy / sponsored by the International Society of Cardiovascular Pharmacotherapy*. 2009; 23:17–24. [PubMed: 18949545]
79. Tinsley JH, Hunter FA, Childs EW. Pkc and mlck-dependent, cytokine-induced rat coronary endothelial dysfunction. *The Journal of surgical research*. 2009; 152:76–83. [PubMed: 18621396]
80. Gruen ML, Hao M, Piston DW, Hasty AH. Leptin requires canonical migratory signaling pathways for induction of monocyte and macrophage chemotaxis. *American journal of physiology. Cell physiology*. 2007; 293:C1481–1488. [PubMed: 17728393]
81. O'Rourke L, Yeaman SJ, Shepherd PR. Insulin and leptin acutely regulate cholesterol ester metabolism in macrophages by novel signaling pathways. *Diabetes*. 2001; 50:955–961. [PubMed: 11334438]
82. Rainwater DL, Comuzzie AG, VandeBerg JL, Mahaney MC, Blangero J. Serum leptin levels are independently correlated with two measures of hdl. *Atherosclerosis*. 1997; 132:237–243. [PubMed: 9242970]
83. Hergenc G, Schulte H, Assmann G, von Eckardstein A. Associations of obesity markers, insulin, and sex hormones with hdl-cholesterol levels in turkish and german individuals. *Atherosclerosis*. 1999; 145:147–156. [PubMed: 10428305]
84. Kazumi T, Kawaguchi A, Hirano T, Yoshino G. C-reactive protein in young, apparently healthy men: Associations with serum leptin, qtc interval, and high-density lipoprotein-cholesterol. *Metabolism: clinical and experimental*. 2003; 52:1113–1116. [PubMed: 14506615]
85. Shamsuzzaman AS, Winnicki M, Wolk R, Svatikova A, Phillips BG, Davison DE, Berger PB, Somers VK. Independent association between plasma leptin and c-reactive protein in healthy humans. *Circulation*. 2004; 109:2181–2185. [PubMed: 15117839]
86. Porreca E, Di Febbo C, Moretta V, Angelini A, Guglielmi MD, Di Nisio M, Cuccurullo F. Circulating leptin is associated with oxidized ldl in postmenopausal women. *Atherosclerosis*. 2004; 175:139–143. [PubMed: 15186958]
87. Bouloumie A, Marumo T, Lafontan M, Busse R. Leptin induces oxidative stress in human endothelial cells. *FASEB journal : official publication of the Federation of American Societies for Experimental Biology*. 1999; 13:1231–1238. [PubMed: 10385613]
88. Deng G, Long Y, Yu YR, Li MR. Adiponectin directly improves endothelial dysfunction in obese rats through the ampk-enos pathway. *International journal of obesity*. 2010; 34:165–171. [PubMed: 19823181]

89. Lee S, Zhang H, Chen J, Dellsperger KC, Hill MA, Zhang C. Adiponectin abates diabetes-induced endothelial dysfunction by suppressing oxidative stress, adhesion molecules, and inflammation in type 2 diabetic mice. *American journal of physiology. Heart and circulatory physiology.* 2012; 303:H106–115. [PubMed: 22561304]
90. Dick GM, Katz PS, Farias M 3rd, Morris M, James J, Knudson JD, Tune JD. Resistin impairs endothelium-dependent dilation to bradykinin, but not acetylcholine, in the coronary circulation. *American journal of physiology. Heart and circulatory physiology.* 2006; 291:H2997–3002. [PubMed: 16905596]
91. Gao X, Picchi A, Zhang C. Upregulation of tnf-alpha and receptors contribute to endothelial dysfunction in zucker diabetic rats. *American journal of biomedical sciences.* 2010; 2:1–12. [PubMed: 20559450]
92. Kougas P, Chai H, Lin PH, Lumsden AB, Yao Q, Chen C. Adipocyte-derived cytokine resistin causes endothelial dysfunction of porcine coronary arteries. *Journal of vascular surgery.* 2005; 41:691–698. [PubMed: 15874935]
93. Picchi A, Gao X, Belmadani S, Potter BJ, Focardi M, Chilian WM, Zhang C. Tumor necrosis factor-alpha induces endothelial dysfunction in the prediabetic metabolic syndrome. *Circulation research.* 2006; 99:69–77. [PubMed: 16741160]
94. Verma S, Li SH, Wang CH, Fedak PW, Li RK, Weisel RD, Mickle DA. Resistin promotes endothelial cell activation: Further evidence of adipokine-endothelial interaction. *Circulation.* 2003; 108:736–740. [PubMed: 12874180]
95. Zhang C, Wu J, Xu X, Potter BJ, Gao X. Direct relationship between levels of tnf-alpha expression and endothelial dysfunction in reperfusion injury. *Basic research in cardiology.* 2010; 105:453–464. [PubMed: 20091314]
96. Uehata M, Ishizaki T, Satoh H, Ono T, Kawahara T, Morishita T, Tamakawa H, Yamagami K, Inui J, Maekawa M, Narumiya S. Calcium sensitization of smooth muscle mediated by a rho-associated protein kinase in hypertension. *Nature.* 1997; 389:990–994. [PubMed: 9353125]
97. Letavernier E, Perez J, Bellocq A, Mesnard L, de Castro Keller A, Haymann JP, Baud L. Targeting the calpain/calpastatin system as a new strategy to prevent cardiovascular remodeling in angiotensin ii-induced hypertension. *Circulation research.* 2008; 102:720–728. [PubMed: 18258859]
98. Minobe E, Asmara H, Saud ZA, Kameyama M. Calpastatin domain I is a partial agonist of the calmodulin-binding site for channel activation in cav1.2 ca2+ channels. *The Journal of biological chemistry.* 2011; 286:39013–39022. [PubMed: 21937422]
99. Berwick ZC, Dick GM, Moberly SP, Kohr MC, Sturek M, Tune JD. Contribution of voltage-dependent k(+) channels to metabolic control of coronary blood flow. *Journal of molecular and cellular cardiology.* 2012; 52:912–919. [PubMed: 21771599]
100. Borbouse L, Dick GM, Asano S, Bender SB, Dincer UD, Payne GA, Neeb ZP, Bratz IN, Sturek M, Tune JD. Impaired function of coronary bk(ca) channels in metabolic syndrome. *American journal of physiology. Heart and circulatory physiology.* 2009; 297:H1629–1637. [PubMed: 19749164]
101. Lu T, Ye D, He T, Wang XL, Wang HL, Lee HC. Impaired ca2+-dependent activation of large-conductance ca2+-activated k+ channels in the coronary artery smooth muscle cells of zucker diabetic fatty rats. *Biophysical journal.* 2008; 95:5165–5177. [PubMed: 18790848]
102. Mokolke EA, Dietz NJ, Eckman DM, Nelson MT, Sturek M. Diabetic dyslipidemia and exercise affect coronary tone and differential regulation of conduit and microvessel k+ current. *American journal of physiology. Heart and circulatory physiology.* 2005; 288:H1233–1241. [PubMed: 15528227]
103. Yang Y, Jones AW, Thomas TR, Rubin LJ. Influence of sex, high-fat diet, and exercise training on potassium currents of swine coronary smooth muscle. *American journal of physiology. Heart and circulatory physiology.* 2007; 293:H1553–1563. [PubMed: 17526655]
104. Lin YC, Huang J, Kan H, Castranova V, Frisbee JC, Yu HG. Defective calcium inactivation causes long qt in obese insulin-resistant rat. *American journal of physiology. Heart and circulatory physiology.* 2012; 302:H1013–1022. [PubMed: 22198168]

Significance

There is a growing body of evidence to support that changes in the phenotypic expression patterns in coronary perivascular adipose tissue (PVAT) occur concomitantly with mechanistic alterations in endothelium and vascular smooth muscle in the setting of cardiovascular disease. These changes appear to be dependent on the unique characteristics of the cell types involved and the underlying environment/milieu in which they reside. This review summarizes current findings regarding the cardiovascular effects of coronary PVAT and outlines potential mechanisms by which adipose-derived factors may influence coronary disease.

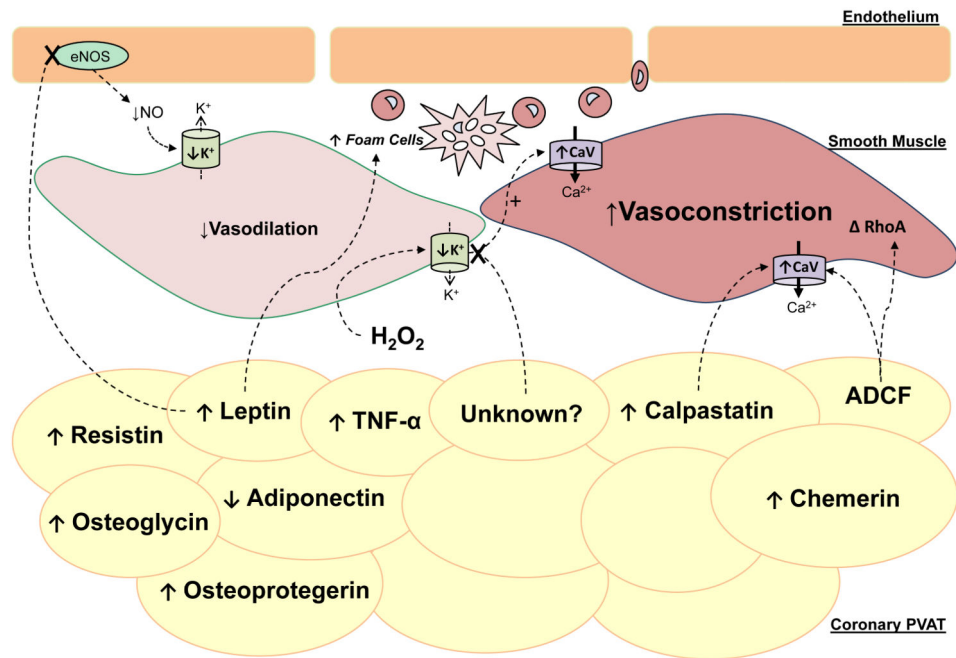


Figure 1.

Schematic diagram outlining known alterations in coronary PVAT-derived adipokines and potential downstream effector mechanisms in endothelium and vascular smooth muscle. Leptin released from coronary PVAT diminishes eNOS activity, preventing nitric oxide mediated dilation of vascular smooth muscle via activation of K⁺ channels and contributes to the recruitment of macrophages and retention of foam cells in the extravascular space. Calpastatin and an unknown adipose-derived constricting factor(s) (ADCF) increase vasoconstriction via Ca_v1.2 channels and may function to increase RhoA activity in healthy coronary smooth muscle. Other adipokines implicated in other vascular beds may also play a role in promoting coronary vascular endothelial and smooth muscle dysfunction, including, but not limited to: increases in resistin, chemerin, osteoglycin, osteoprotegerin, and decreases in adiponectin production.

Table

Comparison of coronary perivascular and subcutaneous adipose tissue adipokine expression.

Adipokine	Condition	Coronary PVAT Expression Relative to Subcutaneous	References
Leptin	NCAD	↓ mRNA	60
	CAD	↓ mRNA	57
Adiponectin	NCAD	↓ mRNA, ↓ protein secretion	60
	CAD	↑ protein secretion	26
TNF- α	NCAD+CAD	↑ mRNA	64
	CAD	↑ mRNA, ↑ protein secretion	63
		↓ protein secretion	26
IL-6	NCAD	↑ mRNA	60, 20
	NCAD+CAD	↑ mRNA	64
	CAD	↓ mRNA	57
		↑ protein secretion	63
IL-1 β	NCAD+CAD	↑ mRNA	64
	CAD	↑ mRNA, ↑ protein secretion	63
MCP-1	NCAD	↑ protein secretion	60
	NCAD+CAD	↑ mRNA	64
	CAD	↑ mRNA, ↑ protein secretion	63
PAI-1	CAD	↓ mRNA	57

NCAD, no coronary artery disease; CAD, coronary artery disease; NCAD+CAD, grouped population of NCAD and CAD; TNF- α , tumor necrosis factor-alpha; IL-6, interleukin-6; IL-1 β , interleukin-1 beta; MCP-1, monocyte chemoattractant protein-1; PAI-1, plasminogen activator inhibitor-1.