



REVIEW published: 23 April 2015 doi: 10.3389/fnins.2015.00144

Targeting glutamate uptake to treat alcohol use disorders

P.S.S. Rao^{1†}, Richard L. Bell^{2*}, Eric A. Engleman² and Youssef Sari^{1*}

¹ Department of Pharmacology, College of Pharmacy and Pharmaceutical Sciences, University of Toledo, Toledo, OH, USA, ² Department of Psychiatry, Indiana University School of Medicine, Indianapolis, IN, USA

Alcoholism is a serious public health concern that is characterized by the development of tolerance to alcohol's effects, increased consumption, loss of control over drinking and the development of physical dependence. This cycle is often times punctuated by periods of abstinence, craving and relapse. The development of tolerance and the expression of withdrawal effects, which manifest as dependence, have been to a great extent attributed to neuroadaptations within the mesocorticolimbic and extended amygdala systems. Alcohol affects various neurotransmitter systems in the brain including the adrenergic, cholinergic, dopaminergic, GABAergic, glutamatergic, peptidergic, and serotonergic systems. Due to the myriad of neurotransmitter and neuromodulator systems affected by alcohol, the efficacies of current pharmacotherapies targeting alcohol dependence are limited. Importantly, research findings of changes in glutamatergic neurotransmission induced by alcohol self- or experimenter-administration have resulted in a focus on therapies targeting glutamatergic receptors and normalization of glutamatergic neurotransmission. Glutamatergic receptors implicated in the effects of ethanol include the ionotropic glutamate receptors (AMPA, Kainate, and NMDA) and some metabotropic glutamate receptors. Regarding glutamatergic homeostasis, ceftriaxone, MS-153, and GPI-1046, which upregulate glutamate transporter 1 (GLT1) expression in mesocorticolimbic brain regions, reduce alcohol intake in genetic animal models of alcoholism. Given the hyperglutamatergic/hyperexcitable state of the central nervous system induced by chronic alcohol abuse and withdrawal, the evidence thus far indicates that a restoration of glutamatergic concentrations and activity within the mesocorticolimbic system and extended amygdala as well as multiple memory systems holds great promise for the treatment of alcohol dependence.

Keywords: glutamate, GLT1, EAAT2, dopamine, alcohol, neurotransmitter

Introduction

A direct link has been established between chronic alcohol consumption and at least 50 different diseases and disorders (Reed et al., 1996; Rehm et al., 2003). Moreover, alcohol use disorders (AUDs) are the third leading cause of preventable death, affecting over 18 million adults and resulting in over 100,000 deaths in the U.S. annually (Grant et al., 2004; Mokdad et al., 2004; Johnson, 2010). Economically, AUDs and their associated effects cost society approximately 200 billion dollars or more each year (Harwood et al., 1999). The pattern of drinking and total volume consumed per unit time are important characteristics for diagnosing AUDs and for developing treatment strategies as well as determining epidemiological antecedents and disease

OPEN ACCESS

Edited by:

Thibault Renoir, Florey Institute of Neuroscience and Mental Health, Australia

Reviewed by:

Nicholas W Gilpin, Louisiana State University Health Sciences Center - New Orleans, USA Raman Chandrasekar, Kansas State University, USA Doo-Sup Choi, Mayo Clinic College of Medicine, USA

> *Correspondence: Richard L. Bell,

Department of Psychiatry, Indiana University School of Medicine, Neuroscience Research Building, NB300, 320 W. 15th Street, Indianapolis, IN 46202-2266, USA ribell@iupui.edu; Youssef Sari, Department of Pharmacology, College of Pharmacy and Pharmaceutical Sciences, University of Toledo, Health Science Campus, 3000 Arlington Avenue, HEB282GToledo, OH 43614, USA

youssef.sari@utoledo.edu

[†]Present Address:

P.S.S. Rao, Department of Pharmaceutical Sciences, University of Tennessee Health Science Center, College of Pharmacy, Memphis, TN 38163, USA

Specialty section:

This article was submitted to Neuropharmacology, a section of the journal Frontiers in Neuroscience

Received: 04 June 2014 Accepted: 07 April 2015 Published: 23 April 2015

Citation:

Rao PSS, Bell RL, Engleman EA and Sari Y (2015) Targeting glutamate uptake to treat alcohol use disorders. Front. Neurosci. 9:144. doi: 10.3389/fnins.2015.00144

1

trajectory (Heather et al., 1993; Lancaster, 1994; Zucker, 1995; Shield et al., 2013). These characterizations have led to classifications, typologies and/or drinking profiles for alcoholics (Cloninger, 1987; Babor et al., 1992; Epstein et al., 1995; Lesch and Walter, 1996; Conrod et al., 2000). In addition, it appears that the pharmacological efficacy of some treatments for AUDs is influenced by the phenotypic and/or genotypic characteristics associated with these typologies (Epstein et al., 1995; Johnson et al., 2003; Cherpitel et al., 2004; Dundon et al., 2004; Johnson, 2004, 2010).

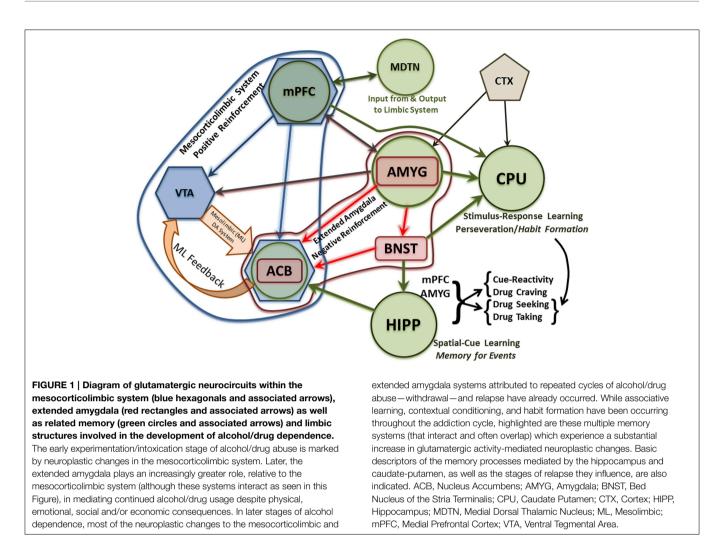
The fact that people from similar environments often differ in their pattern and frequency of alcohol use as well as the welldocumented familial incidence of alcoholism underscores the substantial role of genetics in the development and expression of AUDs (Cotton, 1979; Cloninger, 1987; Enoch and Goldman, 2001).

In general, AUDs define a chronic, progressive, relapsing disorder that advances in stages from experimentation to dependence (Volkow and Li, 2005; Heilig and Egli, 2006; Koob and Le Moal, 2008; Koob, 2009; Spanagel, 2009; Jupp and Lawrence, 2010; Koob and Volkow, 2010). The disease progresses from rewarding, euphoric and positive-reinforcement aspects (e.g., motor and autonomic activation as well as facilitating pro-social behavior) that drive the disease-process in the early stages to the dysphoric and associated negative-reinforcement aspects (e.g., removal of withdrawal-associated effects) of chronic alcohol use that drive the process in later stages. And, while the end-point is the same, the progression for each individual is often different and nonlinear in nature (i.e., individuals will return to earlier stages of the disease cycle with varying frequency and intensity). The change in primacy from positive reinforcement to negative reinforcement across the addiction cycle follows neuroplastic changes in central reward neurocircuitry (e.g., the mesocorticolimbic and extended amygdala systems) induced by chronic alcohol abuse. In the normal functioning brain, a balance (i.e., homeostasis) exists between excitatory and inhibitory neurotransmission. Acute alcohol consumption disturbs this equilibrium by enhancing inhibitory and attenuating excitatory neurotransmission (e.g., Roberto et al., 2003; Basavarajappa et al., 2008; Leriche et al., 2008). However, following long term alcohol consumption, the brain compensates for the depressant effects of alcohol to maintain homeostasis between inhibitory (e.g., GABA) and excitatory (e.g., glutamate) neurotransmission by increasing excitatory activity and reducing inhibitory activity (e.g., Nam et al., 2012; Koob, 2013; Tabakoff and Hoffman, 2013). These changes occur in a number of brain regions within the mesocorticolimbic and extended amygdala reward circuits. And, these brain regions display heavy glutamatergic innervation as shown in Figure 1, which describes some of these glutamatergic projections and is adapted from a number of studies (Aston-Jones et al., 2009; Bonci and Borgland, 2009; Carlezon and Thomas, 2009; Haydon et al., 2009; Kalivas et al., 2009; O'dell, 2009; Renthal and Nestler, 2009; Russo et al., 2009; Taylor et al., 2009; Vezina and Leyton, 2009; Volkow et al., 2009, 2010; Wheeler and Carelli, 2009; Kenny, 2011; Luscher and Malenka, 2011; Potenza et al., 2011; Sulzer, 2011; Kash, 2012; Bass et al., 2013; Chambers, 2013; Cui et al., 2013; Jennings et al., 2013; Volkow and Baler, 2013). In this review, we have focused on alcohol-induced changes in glutamate uptake, the role of glutamate transporters in the development of alcohol dependence, and current and promising glutamate-associated pharmacotherapies for the treatment of alcohol dependence.

The Mesocorticolimbic System, Extended Amygdala and Glutamatergic Neurotransmission

The primary glutamatergic projections, associated with addiction include the mesocorticolimbic dopamine system and the extended amygdala as well brain regions mediating learning and memory (Figure 1). The mesocorticolimbic system is a primary neurocircuit mediating reward and reinforcement salience that is most often, but not always, associated with a positive valence (Tzschentke, 2000; Wise, 2005; Robinson and Berridge, 2008; Brooks and Berns, 2013; Volman et al., 2013). Regarding the dual valence-processing of rewarding/reinforcing stimuli within the mesocorticolimbic system and extended amygdala, it has been shown that activation of the glutamatergic projections from the amygdala through the bed nucleus of the stria terminalis to the ventral tegmental area (VTA) results in aversive and anxiogenic behaviors, whereas activation of the GABAergic parallel projections results in appetitive and anxiolytic behaviors (Jennings et al., 2013). Interestingly, this dissociation between glutamatergic activity and aversion vs. GABAergic activity and reward is also true for similar parallel projections from the VTA to the lateral habenula (Stamatakis et al., 2013; Root et al., 2014). Therefore, from a purely neurochemical perspective within these neurocircuits, it is not surprising that the use of ethanol during the experimentation and early stages of abuse is positively reinforcing and rewarding, given that the acute actions of ethanol are to inhibit glutamatergic activity and potentiate GABAergic activity. During early stages of drug/alcohol abuse positive reinforcement drives continued usage, such that the individual consumes or self-administers the drug or alcohol for the pleasant and euphoric effects they induce. Positive reinforcement is mediated, for the most part, by the mesocorticolimbic dopamine system, which is controlled to a great extent by glutamatergic activity. Similarly, given that the chronic actions of ethanol are to inhibit GABAergic activity and potentiate glutamatergic activity, and a consequent increase in dopaminergic activity within the VTA, accumbens (acb), and prefrontal cortex (PFC), it results in an aversive state especially under conditions of withdrawal. It also recruits increased glutamatergic and corticotrophin releasing factor anxiogenic activity in the amygdala/extended amygdala. Combined, these latter effects elicit continued alcohol and drug usage in an attempt to delay/block/reverse the onset of withdrawal/aversion-associated symptoms.

Early stages of alcohol/drug dependence are also marked by increased glutamatergic activity within the extended amygdala and the presence of neuroplastic changes in the mesocorticolimbic system. Dependence is associated with the development of tolerance and attendant decreases, as a result of neuroplastic changes, in positive reinforcement (e.g., mesocorticolimbic



dopaminergic activity). Dependence is also associated with withdrawal, which is correlated with a number of symptoms linked to increased glutamatergic/excitatory neuronal activity and decreased GABAergic/inhibitory neuronal activity. The extended amygdala mediates, at least in part, withdrawal symptoms related to anxiety and irritability as well as accompanying heightened physiological and autonomic responses. These heightened responses are mediated not only by increased glutamatergic (as opposed to GABAergic) activity but also increased anxiogenic peptidergic activity [i.e., corticotrophin releasing factor (CRF) as opposed to anxiolytic neuropeptide Y (NPY) activity] within the extended amygdala, particularly the AMYG and BNST (Roberto et al., 2012). However, progression through these stages of the addiction cycle is not linear in nature. Rather, there are substantial individual differences in the number and duration of instances where an individual's usage pattern returns to that seen in a previous stage of the addiction cycle.

During late stages of dependence, the neuroplastic changes in the mesocorticolimbic system are retained and the neuroplastic changes of the extended amygdala become more pronounced. As highlighted in **Figure 1**, multiple memory systems (See White, 1996, White, and associated commentaries, for an early review) are also recruited in these neurocircuits with substantial glutamatergic activity. Neuroplastic changes within different memory systems strengthen the behavioral manifestations seen in late stage dependence. Given that alcohol and drug addiction are often characterized as compulsive behaviors, it has been repeatedly postulated, albeit with different terminology, that the caudate-putamen, which mediates habit formation and memory, tends to predominate as a memory circuit in late stage dependence (Koob and Volkow, 2010; Volkow et al., 2010; Everitt and Robbins, 2013; Richard et al., 2013; Zorrilla and Koob, 2013; Wise and Koob, 2014).

Prominent among these behaviors in later stages of dependence is the ubiquitous phenomenon of relapse (Chiauzzi, 1991; Weiss et al., 2001; Barrick and Connors, 2002; Jaffe, 2002). Relapse takes place during withdrawal, which distinguishes it from continued usage in the absence of withdrawal. Relapse is often precipitated by some form of cue-reactivity, which would require contextual processing by the hippocampus as well as autonomic activity modulated by the amygdala and medial PFC. This autonomic/physiological activity can be modulated by all of the brain regions ascribed, in the **Figure 1**, to the mesocorticolimbic system and extended amygdala. The desire to remove associated aversive symptoms/stimuli, or to re-live memories of a previous positive experience, with the former tending to predominate over the latter, leads to drug/alcohol seeking and taking with the recruitment of both habit- and cognitive-memory circuits, again with the former predominating over the latter in late stage dependence.

Glutamatergic Neurotransmission: Interactions with Alcohol

The major excitatory central neurotransmitter, glutamate, acts on two broad categories of receptors, ionotropic and metabotropic. The ionotropic receptors are ligand-gated ion channels and are further classified into N-methyl-D-aspartate (NMDA), alphaamino-3-hydroxy-5-methylisoxazole-4-propionic acid (AMPA) and Kainate receptors based on their sensitivity to respective agonists (Nakanishi, 1992). The metabotropic receptors (mGluR) are G-protein coupled receptors divided into eight mGluR subtypes, having multiple splice variants (Pin and Duvoisin, 1995). As indicated in Section The Mesocorticolimbic System, Extended Amygdala and Glutamatergic Neurotransmission, ethanol exposure alters the central glutamatergic activity and these changes vary depending on the extent of ethanol abuse.

Chronic ethanol exposure (Rossetti and Carboni, 1995; Dahchour and De Witte, 1999, 2000; Melendez et al., 2005; Kapasova and Szumlinski, 2008; Ding et al., 2012), for instance, has been shown to elevate extracellular levels of glutamate in mesocorticolimbic brain areas. Moreover, the effects of ethanol consumption on NMDA- and mGlu-R as well as Homer (a glutamatergic receptor scaffolding protein) subunit expression in the extended amygdala and Acb core are also influenced by the length of ethanolwithdrawal (Obara et al., 2009). Intracellularly, ethanol inhibits, non-competitively, the NMDAR-mediated calcium influx in the cortex, Acb, amygdala, hippocampus, and VTA (Nie et al., 1994; Wirkner et al., 2000; Roberto et al., 2004; Stobbs et al., 2004; Zhu et al., 2007). Additionally, ethanol's effects on NMDA receptor activity are followed by phosphorylation and internalization of NR2 subunits (Suvarna et al., 2005). Ethanol was also found to inhibit NMDA-induced increases in cyclic guanosine monophosphate activity (Hoffman et al., 1989). Furthermore, studies have shown that alcohol withdrawal is associated with increased excitatory amino acid transmission, leading to manifestation of symptoms such as seizures, which can be blocked using NMDA receptor antagonists (Danysz et al., 1992; Snell et al., 1996; Malinowska et al., 1999; Narita et al., 2000; Bienkowski et al., 2001; Nelson et al., 2005).

Glutamate activity at mGluR5, a post-synaptic metabotropic glutamate receptor, has been shown to regulate self-administration of alcohol in rats (Besheer et al., 2008). Similarly, antagonists of mGluR5 are also found to be beneficial in preventing alcohol relapse in a genetic animal model of alcoholism (Schroeder et al., 2005). Thus, pharmacological blockade or very low levels of *mGluR2* mRNA is associated with increased alcohol consumption by alcohol preferring as well as alcohol non-preferring rats (Li et al., 2013; Holmes et al., 2013; Meinhardt et al., 2013; Zhou et al., 2013). Similarly, post-mortem analysis of human frontal cortex from alcohol-dependent patients revealed a

decreased expression levels of mGluR2 mRNA (Meinhardt et al., 2013) further highlighting its involvement in the development and/or expression of alcohol dependence.

Glutamate Transporters: Role in Glutamate Homeostasis

Glutamate transporters are membrane-bound protein pumps found on both neuronal and glial membranes; these transporters regulate glutamate uptake. Increased extracellular glutamate may cause calcium homeostasis dysfunction, increased nitric oxide production, activation of proteases, increased cytotoxic transcription factor levels, and increased free radicals that may subsequently lead to neuronal death (For review, see Wang and Qin, 2010). There are two types of glutamate transporters that regulate extracellular glutamate level: sodium dependent excitatory amino acid transporters (EAATs) and vesicular glutamate transporters (VGLUTs). In addition, there exists a cysteine-glutamate antiporter that regulates the exchange of cysteine and glutamate at the synapse (Bridges et al., 2012).

Excitatory Amino Acid Transporters

EAATs are present in both presynaptic neurons and glial cells and are responsible for modulating glutamate homeostasis (Kanai et al., 1995; Danbolt, 2001). Glutamate is co-transported along with three sodium ions and one proton followed by efflux of one potassium ion (Zerangue and Kavanaugh, 1996). Five subtypes of EAATs have been identified in human and rodent brains. The first three subtypes that were identified in rat/rabbit models are called glutamate/aspartate transporter (GLAST) (Storck et al., 1992), GLT1 (Pines et al., 1992), and excitatory amino acid carrier type 1 (EAAC1) (Kanai and Hediger, 1992); their human counterparts are termed EAAT1, EAAT2, and EAAT3, respectively (Arriza et al., 1994). The other two subtypes were identified in both rodents and humans and named as EAAT4 (Fairman et al., 1995) and EAAT5 (Arriza et al., 1997). The five EAAT subtypes have been found to share 50-60% sequence homology (Seal and Amara, 1999). The localization of these five EAAT subtypes has been studied in detail and described elsewhere (Gegelashvili and Schousboe, 1998; Danbolt, 2001).

Vesicular Glutamate Transporters

VGLUTs are responsible for the uptake and sequestration of glutamate into presynaptic vesicles for storage. The uptake is driven by a proton-dependent electrochemical gradient that exists across the vesicle membrane and depends on the potential gradient created by a vacuolar-type ATPase (Edwards, 2007). Three isoforms of VGLUTs have been identified in the mammalian central nervous system (CNS): VGLUT1, VGLUT2, and VGLUT3 (El Mestikawy et al., 2011). These transporters belong to the type 1 phosphate transporter family. VGLUT1 is expressed in the cortex, hippocampus, and thalamus while VGLUT2 is found in the neocortex, olfactory bulb, dentate gyrus, and subiculum; also, co-expression of VGLUT1 and VGLUT2 has been observed in the hippocampus (Herzog et al., 2006). VGLUT1 is suggested to be expressed at synapses associated with low release rates and long-term potentiation, while VGLUT2 is expressed at synapses with high release rates and long-term potentiation (Fremeau et al., 2001), with others suggesting VGLUT2 is also associated with synapses displaying long-term depression (Kaneko and Fujiyama, 2002). Alternatively, VGLUT3 was identified in neuronal somato-dendrites and glia distributed sparsely throughout the brain. The expression of VGLUTs is believed to be on the extra-presynaptic terminal, suggesting a probable role in modulation of glutamate signaling via endocytosis (Fremeau et al., 2002), although some work has suggested a role for VGLUTs in glutamatergic exocytosis as well (Bellocchio et al., 1998; Fremeau et al., 2004; Seal and Edwards, 2006).

Cysteine–Glutamate Antiporter

Cysteine-glutamate antiporter is a plasma membrane-bound, Na⁺-independent, anionic amino acid transporter that exchanges extracellular cysteine for intracellular glutamate and serves as a source of non-vesicular glutamate release (Danbolt, 2001). It exists as two separate proteins: the light chain cysteine/glutamate exchanger (xCT) that is unique to the cysteine-glutamate antiporter, and the heavy chain 4F2 that is common to many amino acid transporters (Sato et al., 1999). Similar to EAATs, this antiporter is distributed on cells throughout the body and preferentially on glia in the brain (Bridges et al., 2012). Furthermore, it provides tone to mGluRs and cysteine for glutathione synthesis, thereby antagonizing oxidative stress. Given glutamate's role in excitotoxicity, it is not surprising that cysteine-glutamate exchange is thought to play a prominent role in regulating extracellular glutamate levels (Murphy et al., 1989; Bridges et al., 2012).

Studies have shown that a reduction in the cysteine/glutamate exchange increased susceptibility to relapse-seeking behavior (Baker et al., 2003). Also, restoration of the antiporter activity through intracranial perfusion of cysteine or systemic administration of N-acetyl cysteine was shown to decrease cocaine seeking in rat models (Baker et al., 2003). Another study showed that ceftriaxone treatment restored both GLT1 and xCT levels, which in turn inhibited relapse to cocaine-seeking behavior (Sari et al., 2009; Knackstedt et al., 2010). Similarly, we have recently demonstrated chronic alcohol consumption in male alcohol-preferring P rats (a genetic animal model of alcoholism) to be associated with a significant decrease in expression of xCT in Acb and PFC (Alhaddad et al., 2014a). Overall, the cysteine-glutamate antiporter is known to play an important role in maintaining glutamate homeostasis under normal and various pathological conditions (For review, see Bridges et al., 2012).

Role of Glutamate Reuptake in Alcohol Dependence

Glutamate-mediated excitotoxicity is implicated in trauma, ischemia and several other neurodegenerative disorders (Sattler and Tymianski, 2001). Glial sodium dependent transporters, GLAST (EAAT1) and GLT1 (EAAT2), in particular GLT1, are responsible for at least 90% of extracellular glutamate removal (For review, see Anderson and Swanson, 2000). Impaired glutamate uptake due to dysfunction or downregulation of EAAT2 results in several neurological disorders, including Amyotrophic Lateral Sclerosis (ALS), Alzheimer's disease, epilepsy, ischemia and hepatic encephalopathy (Maragakis and Rothstein, 2006). Importantly, we have previously demonstrated that chronic exposure to alcohol results in significant down-regulation of GLT1 expression in the Acb and/or PFC in P rats (Sari and Sreemantula, 2012; Sari et al., 2013).

Furthermore, studies have demonstrated a significant reduction in the levels of EAAT1 and EAAT2 in the basolateral amygdala in postmortem human alcoholic brains as compared to non-alcoholic individuals (Kryger and Wilce, 2010). Acute exposure to alcohol has been found to inhibit glutamatergic neurotransmission. However, chronic exposure to alcohol may elevate glutamate levels, which can lead to withdrawal-associated effects of alcohol deprivation and exposure subject's seeking alcohol-associated negative reinforcement to remove the same (Valenzuela, 1997). Additionally, a reduction in the expression of GLAST level and an increase in GLAST mRNA were found in post-mortem human PFC samples of alcoholics, which might be due to a compensatory mechanism induced by chronic alcohol abuse/dependence (Flatscher-Bader and Wilce, 2008). Furthermore, an increase in VGLUT2 expression in the Acb shell was observed, while VGLUT1 remained unchanged, following chronic alcohol exposure and subsequent deprivation from alcohol (Zhou et al., 2006).

Importantly, GLT1 is currently considered a molecular target for the attenuation of alcohol dependence since it regulates the majority of extracellular glutamate uptake (Rao and Sari, 2012). Previous studies have identified several beta-lactam antibiotics as potent modulators (i.e., elevation) of GLT1 expression (Rothstein et al., 2005). Based on its favorable pharmacokinetic properties, ceftriaxone (a third-generation cephalosporin) was chosen for further study in in vitro models of ischemic injury and motoneuron degeneration and in vivo animal models of ALS (Rothstein et al., 2005). Furthermore, screening of several FDA-approved compounds using a luciferase reporter assay on human astroglial cells identified harmine, a beta-carboline alkaloid, as a potent EAAT2 promoter. Further testing in cell cultures and ALS animal models demonstrated that harmine effectively increased both GLT1 protein and glutamate transporter activity (Li et al., 2011). Thus, harmine may prove efficacious in the treatment of alcohol dependence.

Current Pharmacotherapy Targeting Glutamate Neurotransmission for Alcohol Dependence

Current available therapeutics for alcohol dependence target several different systems due to the non-specific nature of alcohol's action in the brain. In general, the available FDA-approved therapies for alcohol dependence work by blunting the rewarding effects (naltrexone) of ethanol, creating an aversion (disulfiram) toward ethanol or restoring the homeostatic balance between inhibitory and excitatory neurotransmission in the CNS (acamprosate). Ideally, drugs targeting the mesocorticolimbic reward pathway and counteracting chronic ethanol exposure-induced adaptations would present a feasible pharmacological solution.

Acamprosate is a synthetic GABA analog, FDA-approved drug for the prevention of alcohol relapse. It was suggested to

be a functional NMDA antagonist, thereby countering chronic alcohol exposure-induced increases in glutamate concentrations and the subsequent precipitation of a hyperglutamatergic state, which occur during alcohol withdrawal episodes (Dahchour et al., 1998). Acamprosate treatment has also been shown to affect GABAergic neurotransmission by inhibiting presynaptic GABA_B receptors (Berton et al., 1998). Studies have suggested that acamprosate treatment is mildly effective or ineffective in multi-center clinical trials (Anton et al., 2006; Mason et al., 2006). However, further analysis has suggested that, apart from being a well-tolerated drug, acamprosate treatment is associated with significant positive effects on rates of abstinence among both sexes (Mason and Lehert, 2012).

Topiramate, an FDA approved anti-epileptic drug, has shown promising results in attenuating alcohol consumption due to its modulation of glutamatergic neurotransmission. Topiramate has been demonstrated to inhibit the AMPA and kainate glutamate receptors (Skradski and White, 2000; Gryder and Rogawski, 2003). While topiramate treatment was associated with decreased ethanol intake in several animal studies (Hargreaves and Mcgregor, 2007; Nguyen et al., 2007; Breslin et al., 2010), the drug treatment was not associated with changes in ethanol conditioned place preference (Gremel et al., 2006). In addition, topiramate has been shown to be effective in reducing alcohol intake and preventing relapse in human subjects (Baltieri et al., 2008; Florez et al., 2008). Other anticonvulsants [e.g., gabapentin (another synthetic GABA analog altering GABAergic, glutamatergic and adrenergic activity with some efficacy for treating cannabis dependence; Howland, 2013) or valproate] have also been evaluated; with a meta-analysis suggesting anticonvulsants, which generally modulate glutamatergic, GABAergic and possibly other excitatory or inhibitory neuronal activity, have modest but significant efficacy in the treatment of alcohol dependence (Pani et al., 2014).

GLT1 Upregulators: Potential Targets for the Treatment of Alcohol Dependence

Ceftriaxone

Ceftriaxone is a third-generation, semi-synthetic cephalosporin with a broad spectrum of activity against Gram-positive and Gram-negative aerobic, and a few anaerobic, bacteria. Ceftriaxone can be administered either intravenously or intramuscularly and has been successfully used in the treatment of meningitis; urinary tract infections; lower respiratory tract infections; skin, soft tissue, bone, and joint infections; and bacteremia/septicemia (Richards et al., 1984). In addition to its GLT1 modulating effect (Rothstein et al., 2005), ceftriaxone has been studied extensively for its neuroprotective activity. In an animal model of stroke, ceftriaxone treatment was associated with a significant reduction in acute stroke mortality and improved neurological performance (Thone-Reineke et al., 2008). Additionally, a ceftriaxone-mediated increase in GLT1 expression in the spinal cord was found to be beneficial in treating opioid-induced paradoxical pain and neuropathic pain (Ramos et al., 2010). Ceftriaxone has been studied extensively for its potential role in

treating ethanol dependence (for review see Rao and Sari, 2012). We have previously reported that ceftriaxone treatment administered intraperitoneally (i.p.) in P rats attenuated ethanol consumption during five consecutive daily doses of the treatment (Sari et al., 2011). Importantly, attenuation of ethanol consumption following ceftriaxone treatment was associated with upregulation of GLT1 expression levels in the mesocorticolimbic pathway, including the Acb and PFC. Additionally, ceftriaxone administered i.p. was found effective in attenuating ethanol withdrawal and relapse-like drinking behaviors (Qrunfleh et al., 2013; Abulseoud et al., 2014). Furthermore, apart from modulating GLT1 expression, we have demonstrated upregulation of xCT expression as an additional mechanism of ceftriaxone-induced normalization of glutamatergic homeostasis in the Acb and PFC (Alhaddad et al., 2014a; Rao and Sari, 2014b). Ceftriaxone was also effective in attenuating ethanol consumption in P rats subjected to a long-term ethanol drinking paradigm (Rao and Sari, 2014a). Furthermore, our recent findings have demonstrated that ceftriaxone-induced GLT1 upregulation is associated with activation of AKT and NF-kB signaling pathways in the Acb and PFC of P rats (Rao et al., 2015).

GPI-1046

GPI-1046, 3-(3-pyridyl)-1-propyl (2S)-1-(3,3-dimethyl-1,2dioxopentyl)-2-pyrrolidinedinecarboxylate, is a synthetic non-immunosuppressive ligand of FK506 binding protein-12 (Steiner et al., 1997b). FK506 was found to promote neuronal survival. Thus, a series of non-immune suppressive ligands of FKBP-12 were synthesized, including GPI-1046, the prototype for these non-immunosuppressive ligands. GPI-1046 was shown to be effective in neurodegenerative animal models (Steiner et al., 1997a). Along with neuroprotective properties, GPI-1046 was also found to possess anti-retroviral activity and was effective in inhibiting HIV replication (Steiner et al., 2007). Interestingly, GPI-1046 was found to have an upregulatory effect on GLT1 expression both in vitro and in vivo (Ganel et al., 2006). Importantly, we recently reported that GPI-1046 treatment (i.p.) upregulated GLT1 protein expression in the PFC and Acb and concomitantly reduced alcohol intake in male P rats (Sari and Sreemantula, 2012). Further, studies are warranted to reveal the molecular mechanism involved in GPI-1046 mediated changes in GLT1 expression.

MS-153

We have recently reported that a synthetic compound, (R)-(-)-5-methyl-1-nicotinoyl-2-pyrazoline (MS-153) reduced alcohol intake in male P rats (Alhaddad et al., 2014b). This reduction in alcohol intake was associated in part with upregulation of GLT1 expression in the Acb. This compound, in addition to GLT1 upregulation, was shown to activate p-Akt and NF-kB pathways; these signaling pathways were previously suggested to be involved in GLT1 upregulation. In addition, MS-153 mediated changes in GLT1 expression were also examined in the amygdala and hippocampus of P rats (Aal-Aaboda et al., 2015). While ethanol consumption significantly downregulated expression of xCT and GLT1, MS-153 treatment normalized expression of these glutamate transporters in both the amygdala and hippocampus of P rats compared to vehicle control animals.

Conclusion and Closing Thoughts

Alcohol consumption affects a number of neurotransmitters and neuromodulators in the CNS, including dopamine, serotonin and GABA. Pertinent to the present paper, the neurotransmitter glutamate has been shown to be critical in the development and expression of alcohol dependence. In particular, changes in glutamatergic activity are often associated with neuroplastic changes in CNS circuitry (Bliss et al., 2014). These changes occur in brain structures mediating reward, reinforcement, learning and memory (for reviews on some of these changes see White, 1996). In general, based on the existing literature, chronic alcohol and other drugs of

References

- Aal-Aaboda, M., Alhaddad, H., Osowik, F., Nauli, S. M., and Sari, Y. (2015). Effects of (R)-(-)-5-methyl-1-nicotinoyl-2-pyrazoline on glutamate transporter 1 and cysteine/glutamate exchanger as well as ethanol drinking behavior in male, alcohol-preferring rats. *J. Neurosci. Res.* 93, 930–937. doi: 10.1002/jnr. 23554
- Abulseoud, O. A., Camsari, U. M., Ruby, C. L., Kasasbeh, A., Choi, S., and Choi, D. S. (2014). Attenuation of Ethanol withdrawal by ceftriaxone-induced upregulation of glutamate transporter EAAT2. *Neuropsychopharmacology* 39, 1674–1684. doi: 10.1038/npp.2014.14
- Alhaddad, H., Das, S. C., and Sari, Y. (2014a). Effects of ceftriaxone on ethanol intake: a possible role for xCT and GLT-1 isoforms modulation of glutamate levels in P rats. *Psychopharmacology (Berl)*. 231, 4049–4057. doi: 10.1007/s00213-014-3545-y
- Alhaddad, H., Kim, N. T., Aal-Aaboda, M., Althobaiti, Y. S., Leighton, J., Boddu, S. H., et al. (2014b). Effects of MS-153 on chronic ethanol consumption and GLT1 modulation of glutamate levels in male alcohol-preferring rats. *Front. Behav. Neurosci.* 8:366. doi: 10.3389/fnbeh.2014.00366
- Anderson, C. M., and Swanson, R. A. (2000). Astrocyte glutamate transport: review of properties, regulation, and physiological functions. *Glia* 32, 1–14. doi: 10.1002/1098-1136(200010)32:1<1::AID-GLIA10>3.0.CO;2-W
- Anton, R. F., O'malley, S. S., Ciraulo, D. A., Cisler, R. A., Couper, D., Donovan, D. M., et al. (2006). Combined pharmacotherapies and behavioral interventions for alcohol dependence: the COMBINE study: a randomized controlled trial. *JAMA* 295, 2003–2017. doi: 10.1001/jama.295.17.2003
- Arriza, J. L., Eliasof, S., Kavanaugh, M. P., and Amara, S. G. (1997). Excitatory amino acid transporter 5, a retinal glutamate transporter coupled to a chloride conductance. *Proc. Natl. Acad. Sci. U.S.A.* 94, 4155–4160. doi: 10.1073/pnas.94.8.4155
- Arriza, J. L., Fairman, W. A., Wadiche, J. I., Murdoch, G. H., Kavanaugh, M. P., and Amara, S. G. (1994). Functional comparisons of three glutamate transporter subtypes cloned from human motor cortex. *J. Neurosci.* 14, 5559–5569.
- Aston-Jones, G., Smith, R. J., Moorman, D. E., and Richardson, K. A. (2009). Role of lateral hypothalamic orexin neurons in reward processing and addiction. *Neuropharmacology* 56(Suppl. 1), 112–121. doi: 10.1016/j.neuropharm.2008.06.060
- Babor, T. F., Hofmann, M., Delboca, F. K., Hesselbrock, V., Meyer, R. E., Dolinsky, Z. S., et al. (1992). Types of alcoholics, I. Evidence for an empirically derived typology based on indicators of vulnerability and severity. *Arch. Gen. Psychiatry* 49, 599–608. doi: 10.1001/archpsyc.1992.01820080007002
- Baker, D. A., Mcfarland, K., Lake, R. W., Shen, H., Tang, X. C., Toda, S., et al. (2003). Neuroadaptations in cysteine-glutamate exchange underlie cocaine relapse. *Nat. Neurosci.* 6, 743–749. doi: 10.1038/nn1069

abuse result in a pronounced increase in glutamatergic activity within these neuronal circuits. Therefore, any pharmacological treatments that can reverse this hyperglutamatergic state hold great promise in the treatment of alcohol, and possibly illicit drug, dependence. It is noteworthy that compounds that physically and/or functionally upregulate glutamate transporters, in particular GLT1, are ideal candidates for further exploration.

Acknowledgments

This work was supported by Award Number R01AA019458 (YS), UO1AA13522 (RB), and R01AA020396 (EE, RB) from the National Institutes on Alcohol Abuse and Alcoholism. The authors would like to thank Charisse Montgomery for editing this manuscript.

- Baltieri, D. A., Daro, F. R., Ribeiro, P. L., and De Andrade, A. G. (2008). Comparing topiramate with naltrexone in the treatment of alcohol dependence. *Addiction* 103, 2035–2044. doi: 10.1111/j.1360-0443.2008.02355.x
- Barrick, C., and Connors, G. J. (2002). Relapse prevention and maintaining abstinence in older adults with alcohol-use disorders. *Drugs Aging* 19, 583–594. doi: 10.2165/00002512-200219080-00004
- Basavarajappa, B. S., Ninan, I., and Arancio, O. (2008). Acute ethanol suppresses glutamatergic neurotransmission through endocannabinoids in hippocampal neurons. J. Neurochem. 107, 1001–1013. doi: 10.1111/j.1471-4159.2008.05685.x
- Bass, C. E., Grinevich, V. P., Gioia, D., Day-Brown, J. D., Bonin, K. D., Stuber, G. D., et al. (2013). Optogenetic stimulation of VTA dopamine neurons reveals that tonic but not phasic patterns of dopamine transmission reduce ethanol self-administration. *Front. Behav. Neurosci.* 7:173. doi: 10.3389/fnbeh.2013.00173
- Bellocchio, E. E., Hu, H., Pohorille, A., Chan, J., Pickel, V. M., and Edwards, R. H. (1998). The localization of the brain-specific inorganic phosphate transporter suggests a specific presynaptic role in glutamatergic transmission. *J. Neurosci.* 18, 8648–8659.
- Berton, F., Francesconi, W. G., Madamba, S. G., Zieglgansberger, W., and Siggins, G. R. (1998). Acamprosate enhances N-methyl-D-apartate receptor-mediated neurotransmission but inhibits presynaptic GABA(B) receptors in nucleus accumbens neurons. *Alcohol. Clin. Exp. Res.* 22, 183–191. doi: 10.1111/j.1530-0277.1998.tb03636.x
- Besheer, J., Faccidomo, S., Grondin, J. J., and Hodge, C. W. (2008). Regulation of motivation to self-administer ethanol by mGluR5 in alcohol-preferring (P) rats. *Alcohol. Clin. Exp. Res.* 32, 209–221. doi: 10.1111/j.1530-0277.2007.00570.x
- Bienkowski, P., Krzascik, P., Koros, E., Kostowski, W., Scinska, A., and Danysz, W. (2001). Effects of a novel uncompetitive NMDA receptor antagonist, MRZ 2/579 on ethanol self-administration and ethanol withdrawal seizures in the rat. *Eur. J. Pharmacol.* 413, 81–89. doi: 10.1016/S0014-2999(01)00743-9
- Bliss, T. V., Collingridge, G. L., and Morris, R. G. (2014). Synaptic plasticity in health and disease: introduction and overview. *Philos. Trans. R. Soc. Lond. B Biol. Sci.* 369:20130129. doi: 10.1098/rstb.2013.0129
- Bonci, A., and Borgland, S. (2009). Role of orexin/hypocretin and CRF in the formation of drug-dependent synaptic plasticity in the mesolimbic system. *Neuropharmacology* 56 (Suppl. 1), 107–111. doi: 10.1016/j.neuropharm.2008.07.024
- Breslin, F. J., Johnson, B. A., and Lynch, W. J. (2010). Effect of topiramate treatment on ethanol consumption in rats. *Psychopharmacology* 207, 529–534. doi: 10.1007/s00213-009-1683-4
- Bridges, R. J., Natale, N. R., and Patel, S. A. (2012). System xc(-) cysteine/glutamate antiporter: an update on molecular pharmacology and roles within the CNS. *Br. J. Pharmacol.* 165, 20–34. doi: 10.1111/j.1476-5381.2011.01480.x
- Brooks, A. M., and Berns, G. S. (2013). Aversive stimuli and loss in the mesocorticolimbic dopamine system. *Trends Cogn. Sci.* 17, 281–286. doi: 10.1016/j.tics.2013.04.001

- Carlezon, W. A. Jr., and Thomas, M. J. (2009). Biological substrates of reward and aversion: a nucleus accumbens activity hypothesis. *Neuropharmacology* 56 (Suppl. 1), 122–132. doi: 10.1016/j.neuropharm.2008.06.075
- Chambers, R. A. (2013). Adult hippocampal neurogenesis in the pathogenesis of addiction and dual diagnosis disorders. *Drug Alcohol. Depend.* 130, 1–12. doi: 10.1016/j.drugalcdep.2012.12.005
- Cherpitel, C. J., Moskalewicz, J., and Swiatkiewicz, G. (2004). Drinking patterns and problems in emergency services in Poland. *Alcohol Alcohol.* 39, 256–261. doi: 10.1093/alcalc/agh053
- Chiauzzi, E. J. (1991). Preventing Relapse in the Addictions: a Biopsychosocial Approach. New York, NY: Pergamon Press.
- Cloninger, C. R. (1987). Neurogenetic adaptive mechanisms in alcoholism. *Science* 236, 410–416. doi: 10.1126/science.2882604
- Conrod, P. J., Pihl, R. O., Stewart, S. H., and Dongier, M. (2000). Validation of a system of classifying female substance abusers on the basis of personality and motivational risk factors for substance abuse. *Psychol. Addict. Behav.* 14, 243–256. doi: 10.1037/0893-164X.14.3.243
- Cotton, N. S. (1979). The familial incidence of alcoholism: a review. J. Stud. Alcohol. 40, 89-116. doi: 10.15288/jsa.1979.40.89
- Cui, C., Noronha, A., Morikawa, H., Alvarez, V. A., Stuber, G. D., Szumlinski, K. K., et al. (2013). New insights on neurobiological mechanisms underlying alcohol addiction. *Neuropharmacology* 67, 223–232. doi: 10.1016/j.neuropharm.2012.09.022
- Dahchour, A., and De Witte, P. (1999). Acamprosate decreases the hypermotility during repeated ethanol withdrawal. *Alcohol* 18, 77–81. doi: 10.1016/S0741-8329(98)00071-8
- Dahchour, A., and De Witte, P. (2000). Ethanol and amino acids in the central nervous system: assessment of the pharmacological actions of acamprosate. *Prog. Neurobiol.* 60, 343–362. doi: 10.1016/S0301-0082(99)00031-3
- Dahchour, A., De Witte, P., Bolo, N., Nedelec, J. F., Muzet, M., Durbin, P., et al. (1998). Central effects of acamprosate: part 1. Acamprosate blocks the glutamate increase in the nucleus accumbens microdialysate in ethanol withdrawn rats. *Psychiatry Res.* 82, 107–114. doi: 10.1016/S0925-4927(98)00016-X
- Danbolt, N. C. (2001). Glutamate uptake. Prog. Neurobiol. 65, 1–105. doi: 10.1016/S0301-0082(00)00067-8
- Danysz, W., Dyr, W., Jankowska, E., Glazewski, S., and Kostowski, W. (1992). The involvement of NMDA receptors in acute and chronic effects of ethanol. *Alcohol. Clin. Exp. Res.* 16, 499–504. doi: 10.1111/j.1530-0277.1992.tb01407.x
- Ding, Z. M., Engleman, E. A., Rodd, Z. A., and Mcbride, W. J. (2012). Ethanol increases glutamate neurotransmission in the posterior ventral tegmental area of female wistar rats. *Alcohol. Clin. Exp. Res.* 36, 633–640. doi: 10.1111/j.1530-0277.2011.01665.x
- Dundon, W., Lynch, K. G., Pettinati, H. M., and Lipkin, C. (2004). Treatment outcomes in type A and B alcohol dependence 6 months after serotonergic pharmacotherapy. *Alcohol. Clin. Exp. Res.* 28, 1065–1073. doi: 10.1097/01.ALC.0000130974.50563.04
- Edwards, R. H. (2007). The neurotransmitter cycle and quantal size. *Neuron* 55, 835–858. doi: 10.1016/j.neuron.2007.09.001
- El Mestikawy, S., Wallen-Mackenzie, A., Fortin, G. M., Descarries, L., and Trudeau, L. E. (2011). From glutamate co-release to vesicular synergy: vesicular glutamate transporters. *Nat. Rev. Neurosci.* 12, 204–216. doi: 10.1038/ nrn2969
- Enoch, M. A., and Goldman, D. (2001). The genetics of alcoholism and alcohol abuse. *Curr. Psychiatry Rep.* 3, 144–151. doi: 10.1007/s11920-001-0012-3
- Epstein, E. E., Kahler, C. W., Mccrady, B. S., Lewis, K. D., and Lewis, S. (1995). An empirical classification of drinking patterns among alcoholics: binge, episodic, sporadic, and steady. *Addict. Behav.* 20, 23–41. doi: 10.1016/0306-4603(94)00043-X
- Everitt, B. J., and Robbins, T. W. (2013). From the ventral to the dorsal striatum: devolving views of their roles in drug addiction. *Neurosci. Biobehav. Rev.* 37, 1946–1954. doi: 10.1016/j.neubiorev.2013.02.010
- Fairman, W. A., Vandenberg, R. J., Arriza, J. L., Kavanaugh, M. P., and Amara, S. G. (1995). An excitatory amino-acid transporter with properties of a ligand-gated chloride channel. *Nature* 375, 599–603. doi: 10.1038/375599a0
- Flatscher-Bader, T., and Wilce, P. A. (2008). Impact of alcohol abuse on protein expression of midkine and excitatory amino acid transporter 1 in the human prefrontal cortex. *Alcohol. Clin. Exp. Res.* 32, 1849–1858. doi: 10.1111/j.1530-0277.2008.00754.x

- Florez, G., Garcia-Portilla, P., Alvarez, S., Saiz, P. A., Nogueiras, L., and Bobes, J. (2008). Using topiramate or naltrexone for the treatment of alcoholdependent patients. *Alcohol.Clin. Exp. Res.* 32, 1251–1259. doi: 10.1111/j.1530-0277.2008.00680.x
- Fremeau, R. T. Jr., Burman, J., Qureshi, T., Tran, C. H., Proctor, J., Johnson, J., et al. (2002). The identification of vesicular glutamate transporter 3 suggests novel modes of signaling by glutamate. *Proc. Natl. Acad. Sci. U.S.A.* 99, 14488–14493. doi: 10.1073/pnas.222546799
- Fremeau, R. T. Jr., Troyer, M. D., Pahner, I., Nygaard, G. O., Tran, C. H., Reimer, R. J., et al. (2001). The expression of vesicular glutamate transporters defines two classes of excitatory synapse. *Neuron* 31, 247–260. doi: 10.1016/S0896-6273(01)00344-0
- Fremeau, R. T. Jr., Voglmaier, S., Seal, R. P., and Edwards, R. H. (2004). VGLUTs define subsets of excitatory neurons and suggest novel roles for glutamate. *Trends Neurosci.* 27, 98–103. doi: 10.1016/j.tins.2003.11.005
- Ganel, R., Ho, T., Maragakis, N. J., Jackson, M., Steiner, J. P., and Rothstein, J. D. (2006). Selective up-regulation of the glial Na+-dependent glutamate transporter GLT1 by a neuroimmunophilin ligand results in neuroprotection. *Neurobiol. Dis.* 21, 556–567. doi: 10.1016/j.nbd.2005.08.014
- Gegelashvili, G., and Schousboe, A. (1998). Cellular distribution and kinetic properties of high-affinity glutamate transporters. *Brain Res. Bull.* 45, 233–238. doi: 10.1016/S0361-9230(97)00417-6
- Grant, J., Potenza, M., Hollander, E., Kim, S. W., and Cunningham-Williams, R. (2004). A multicenter investigation of fixed-dose nalmefene in the treatment of pathological gambling. *Neuropsychopharmacology* 29, S122–S122.
- Gremel, C. M., Gabriel, K. I., and Cunningham, C. L. (2006). Topiramate does not affect the acquisition or expression of ethanol conditioned place preference in DBA/2J or C57BL/6J mice. *Alcohol.Clin. Exp. Res.* 30, 783–790. doi: 10.1111/j.1530-0277.2006.00091.x
- Gryder, D. S., and Rogawski, M. A. (2003). Selective antagonism of GluR5 kainatereceptor-mediated synaptic currents by topiramate in rat basolateral amygdala neurons. J. Neurosci. 23, 7069–7074.
- Hargreaves, G. A., and Mcgregor, I. S. (2007). Topiramate moderately reduces the motivation to consume alcohol and has a marked antidepressant effect in rats. *Alcohol. Clin. Exp. Res.* 31, 1900–1907. doi: 10.1111/j.1530-0277.2007. 00485.x
- Harwood, H. J., Fountain, D., and Fountain, G. (1999). Economic cost of alcohol and drug abuse in the United States, 1992: a report. Addiction 94, 631-635.
- Haydon, P. G., Blendy, J., Moss, S. J., and Rob Jackson, F. (2009). Astrocytic control of synaptic transmission and plasticity: a target for drugs of abuse? *Neuropharmacology* 56 (Suppl. 1), 83–90. doi: 10.1016/j.neuropharm.2008.06.050
- Heather, N., Tebbutt, J. S., Mattick, R. P., and Zamir, R. (1993). Development of a scale for measuring impaired control over alcohol consumption: a preliminary report. J. Stud. Alcohol. 54, 700–709. doi: 10.15288/jsa.1993.54.700
- Heilig, M., and Egli, M. (2006). Pharmacological treatment of alcohol dependence: target symptoms and target mechanisms. *Pharmacol. Ther.* 111, 855–876. doi: 10.1016/j.pharmthera.2006.02.001
- Herzog, E., Takamori, S., Jahn, R., Brose, N., and Wojcik, S. M. (2006). Synaptic and vesicular co-localization of the glutamate transporters VGLUT1 and VGLUT2 in the mouse hippocampus. *J. Neurochem.* 99, 1011–1018. doi: 10.1111/j.1471-4159.2006.04144.x
- Hoffman, P. L., Rabe, C. S., Moses, F., and Tabakoff, B. (1989). N-methyl-Daspartate receptors and ethanol: inhibition of calcium flux and cyclic GMP production. J. Neurochem. 52, 1937–1940. doi: 10.1111/j.1471-4159.1989.tb07280.x
- Holmes, A., Spanagel, R., and Krystal, J. H. (2013). Glutamatergic targets for new alcohol medications. *Psychopharmacology (Berl)* 229, 539–554. doi: 10.1007/s00213-013-3226-2
- Howland, R. H. (2013). Gabapentin for the treatment of substance use disorders. J. Psychosoc. Nurs. Ment. Health Serv. 51, 11–14. doi: 10.3928/02793695-20131120-01
- Jaffe, S. L. (2002). Treatment and relapse prevention for adolescent substance abuse. *Pediatr. Clin. North Am.* 49, 345–352, vi. doi: 10.1016/S0031-3955(01)00008-6
- Jennings, J. H., Sparta, D. R., Stamatakis, A. M., Ung, R. L., Pleil, K. E., Kash, T. L., et al. (2013). Distinct extended amygdala circuits for divergent motivational states. *Nature* 496, 224–228. doi: 10.1038/nature12041

- Johnson, B. A. (2004). Role of the serotonergic system in the neurobiology of alcoholism: implications for treatment. CNS Drugs 18, 1105–1118. doi: 10.2165/00023210-200418150-00005
- Johnson, B. A. (2010). Medication treatment of different types of alcoholism. Am. J. Psychiatry 167, 630–639. doi: 10.1176/appi.ajp.2010.08101500
- Johnson, B. A., Ait-Daoud, N., Ma, J. Z., and Wang, Y. (2003). Ondansetron reduces mood disturbance among biologically predisposed, alcohol-dependent individuals. *Alcohol. Clin. Exp. Res.* 27, 1773–1779. doi: 10.1097/01.ALC.0000095635.46911.5D
- Jupp, B., and Lawrence, A. J. (2010). New horizons for therapeutics in drug and alcohol abuse. *Pharmacol. Ther.* 125, 138–168. doi: 10.1016/j.pharmthera.2009.11.002
- Kalivas, P. W., Lalumiere, R. T., Knackstedt, L., and Shen, H. (2009). Glutamate transmission in addiction. *Neuropharmacology* 56 (Suppl. 1), 169–173. doi: 10.1016/j.neuropharm.2008.07.011
- Kanai, Y., Bhide, P. G., Difiglia, M., and Hediger, M. A. (1995). Neuronal highaffinity glutamate transport in the rat central nervous system. *Neuroreport* 6, 2357–2362. doi: 10.1097/00001756-199511270-00020
- Kanai, Y., and Hediger, M. A. (1992). Primary structure and functional characterization of a high-affinity glutamate transporter. *Nature* 360, 467–471. doi: 10.1038/360467a0
- Kaneko, T., and Fujiyama, F. (2002). Complementary distribution of vesicular glutamate transporters in the central nervous system. *Neurosci. Res.* 42, 243–250. doi: 10.1016/S0168-0102(02)00009-3
- Kapasova, Z., and Szumlinski, K. K. (2008). Strain differences in alcohol-induced neurochemical plasticity: a role for accumbens glutamate in alcohol intake. *Alcohol. Clin. Exp. Res.* 32, 617–631. doi: 10.1111/j.1530-0277.2008.00620.x
- Kash, T. L. (2012). The role of biogenic amine signaling in the bed nucleus of the stria terminals in alcohol abuse. *Alcohol* 46, 303–308. doi: 10.1016/j.alcohol.2011.12.004
- Kenny, P. J. (2011). Reward mechanisms in obesity: new insights and future directions. *Neuron* 69, 664–679. doi: 10.1016/j.neuron.2011.02.016
- Knackstedt, L. A., Melendez, R. I., and Kalivas, P. W. (2010). Ceftriaxone restores glutamate homeostasis and prevents relapse to cocaine seeking. *Biol. Psychiatry* 67, 81–84. doi: 10.1016/j.biopsych.2009.07.018
- Koob, G. F. (2009). Dynamics of neuronal circuits in addiction: reward, antireward, and emotional memory. *Pharmacopsychiatry* 42 (Suppl. 1), S32–S41. doi: 10.1055/s-0029-1216356
- Koob, G. F. (2013). Negative reinforcement in drug addiction: the darkness within. *Curr. Opin. Neurobiol.* 23, 559–563. doi: 10.1016/j.conb.2013.03.011
- Koob, G. F., and Le Moal, M. (2008). Neurobiological mechanisms for opponent motivational processes in addiction. *Philos. Trans. R. Soc. Lond. B Biol. Sci.* 363, 3113–3123. doi: 10.1098/rstb.2008.0094
- Koob, G. F., and Volkow, N. D. (2010). Neurocircuitry of addiction. Neuropsychopharmacology 35, 217–238. doi: 10.1038/npp.2009.110
- Kryger, R., and Wilce, P. A. (2010). The effects of alcoholism on the human basolateral amygdala. *Neuroscience* 167, 361–371. doi: 10.1016/j.neuroscience.2010.01.061
- Lancaster, F. E. (1994). Gender differences in the brain: implications for the study of human alcoholism. Alcohol. Clin. Exp. Res. 18, 740–746. doi: 10.1111/j.1530-0277.1994.tb00940.x
- Leriche, M., Mendez, M., Zimmer, L., and Berod, A. (2008). Acute ethanol induces Fos in GABAergic and non-GABAergic forebrain neurons: a double-labeling study in the medial prefrontal cortex and extended amygdala. *Neuroscience* 153, 259–267. doi: 10.1016/j.neuroscience.2008.01.069
- Lesch, O. M., and Walter, H. (1996). Subtypes of alcoholism and their role in therapy. *Alcohol Alcohol.* Suppl. 31, 63–67. doi: 10.1093/oxfordjournals.alcalc.a008221
- Li, J., Cheng, Y., Bian, W., Liu, X., Zhang, C., and Ye, J. H. (2010). Region-specific induction of FosB/DeltaFosB by voluntary alcohol intake: effects of naltrexone. *Alcohol. Clin. Exp. Res.* 34, 1742–1750. doi: 10.1111/j.1530-0277.2010.01261.x
- Li, Y., Sattler, R., Yang, E. J., Nunes, A., Ayukawa, Y., Akhtar, S., et al. (2011). Harmine, a natural beta-carboline alkaloid, upregulates astroglial glutamate transporter expression. *Neuropharmacology* 60, 1168–1175. doi: 10.1016/j.neuropharm.2010.10.016
- Luscher, C., and Malenka, R. C. (2011). Drug-evoked synaptic plasticity in addiction: from molecular changes to circuit remodeling. *Neuron* 69, 650–663. doi: 10.1016/j.neuron.2011.01.017

- Malinowska, B., Napiorkowska-Pawlak, D., Pawlak, R., Buczko, W., and Gothert, M. (1999). Ifenprodil influences changes in mouse behaviour related to acute and chronic ethanol administration. *Eur. J. Pharmacol.* 377, 13–19. doi: 10.1016/S0014-2999(99)00393-3
- Maragakis, N. J., and Rothstein, J. D. (2006). Mechanisms of Disease: astrocytes in neurodegenerative disease. *Nat. Clin. Pract. Neurol.* 2, 679–689. doi: 10.1038/ncpneuro0355
- Mason, B. J., Goodman, A. M., Chabac, S., and Lehert, P. (2006). Effect of oral acamprosate on abstinence in patients with alcohol dependence in a doubleblind, placebo-controlled trial: the role of patient motivation. *J. Psychiatr. Res.* 40, 383–393. doi: 10.1016/j.jpsychires.2006.02.002
- Mason, B. J., and Lehert, P. (2012). Acamprosate for alcohol dependence: a sexspecific meta-analysis based on individual patient data. *Alcohol. Clin. Exp. Res.* 36, 497–508. doi: 10.1111/j.1530-0277.2011.01616.x
- Meinhardt, M. W., Hansson, A. C., Perreau-Lenz, S., Bauder-Wenz, C., Stahlin, O., Heilig, M., et al. (2013). Rescue of infralimbic mGluR2 deficit restores control over drug-seeking behavior in alcohol dependence. J. Neurosci. 33, 2794–2806. doi: 10.1523/JNEUROSCI.4062-12.2013
- Melendez, R. I., Hicks, M. P., Cagle, S. S., and Kalivas, P. W. (2005). Ethanol exposure decreases glutamate uptake in the nucleus accumbens. *Alcohol. Clin. Exp. Res.* 29, 326–333. doi: 10.1097/01.ALC.0000156086.65665.4D
- Mokdad, A. H., Marks, J. S., Stroup, D. F., and Gerberding, J. L. (2004). Actual causes of death in the United States, 2000. JAMA 291, 1238–1245. doi: 10.1001/jama.291.10.1238
- Murphy, T. H., Miyamoto, M., Sastre, A., Schnaar, R. L., and Coyle, J. T. (1989). Glutamate toxicity in a neuronal cell line involves inhibition of cysteine transport leading to oxidative stress. *Neuron* 2, 1547–1558. doi: 10.1016/0896-6273(89)90043-3
- Nakanishi, S. (1992). Molecular diversity of glutamate receptors and implications for brain function. *Science* 258, 597–603. doi: 10.1126/science.1329206
- Nam, H. W., Mciver, S. R., Hinton, D. J., Thakkar, M. M., Sari, Y., Parkinson, F. E., et al. (2012). Adenosine and glutamate signaling in neuron-glial interactions: implications in alcoholism and sleep disorders. *Alcohol. Clin. Exp. Res.* 36, 1117–1125. doi: 10.1111/j.1530-0277.2011.01722.x
- Narita, M., Soma, M., Mizoguchi, H., Tseng, L. F., and Suzuki, T. (2000). Implications of the NR2B subunit-containing NMDA receptor localized in mouse limbic forebrain in ethanol dependence. *Eur. J. Pharmacol.* 401, 191–195. doi: 10.1016/S0014-2999(00)00428-3
- Nelson, T. E., Ur, C. L., and Gruol, D. L. (2005). Chronic intermittent ethanol exposure enhances NMDA-receptor-mediated synaptic responses and NMDA receptor expression in hippocampal CA1 region. *Brain Res.* 1048, 69–79. doi: 10.1016/j.brainres.2005.04.041
- Nguyen, S. A., Malcolm, R., and Middaugh, L. D. (2007). Topiramate reduces ethanol consumption by C57BL/6 mice. Synapse 61, 150–156. doi: 10.1002/syn.20350
- Nie, Z., Madamba, S. G., and Siggins, G. R. (1994). Ethanol inhibits glutamatergic neurotransmission in nucleus accumbens neurons by multiple mechanisms. *J. Pharmacol. Exp. Ther.* 271, 1566–1573.
- Obara, I., Bell, R. L., Goulding, S. P., Reyes, C. M., Larson, L. A., Ary, A. W., et al. (2009). Differential effects of chronic ethanol consumption and withdrawal on homer/glutamate receptor expression in subregions of the accumbens and amygdala of P rats. *Alcohol. Clin. Exp. Res.* 33, 1924–1934. doi: 10.1111/j.1530-0277.2009.01030.x
- O'dell, L. E. (2009). A psychobiological framework of the substrates that mediate nicotine use during adolescence. *Neuropharmacology* 56 (Suppl. 1), 263–278. doi: 10.1016/j.neuropharm.2008.07.039
- Pani, P. P., Trogu, E., Pacini, M., and Maremmani, I. (2014). Anticonvulsants for alcohol dependence. *Cochrane Database Syst. Rev.* 2:CD008544. doi: 10.1002/14651858.CD008544.pub2
- Pin, J. P., and Duvoisin, R. (1995). The metabotropic glutamate receptors: structure and functions. *Neuropharmacology* 34, 1–26. doi: 10.1016/0028-3908(94)00129-G
- Pines, G., Danbolt, N. C., Bjoras, M., Zhang, Y., Bendahan, A., Eide, L., et al. (1992). Cloning and expression of a rat brain L-glutamate transporter. *Nature* 360, 464–467. doi: 10.1038/360464a0
- Potenza, M. N., Sofuoglu, M., Carroll, K. M., and Rounsaville, B. J. (2011). Neuroscience of behavioral and pharmacological treatments for addictions. *Neuron* 69, 695–712. doi: 10.1016/j.neuron.2011.02.009

- Qrunfleh, A. M., Alazizi, A., and Sari, Y. (2013). Ceftriaxone, a beta-lactam antibiotic, attenuates relapse-like ethanol-drinking behavior in alcohol-preferring rats. J. Psychopharmacol. 27, 541–549. doi: 10.1177/0269881113482529
- Ramos, K. M., Lewis, M. T., Morgan, K. N., Crysdale, N. Y., Kroll, J. L., Taylor, F. R., et al. (2010). Spinal upregulation of glutamate transporter GLT-1 by ceftriaxone: therapeutic efficacy in a range of experimental nervous system disorders. *Neuroscience* 169, 1888–1900. doi: 10.1016/j.neuroscience.2010.06.014
- Rao, P., and Sari, Y. (2014a). Effectiveness of ceftriaxone treatment in preventing relapse-like drinking behavior following long-term Ethanol dependence in P Rats. J. Addict. Res. Ther. 5:1000183. doi: 10.4172/2155-6105.1000183
- Rao, P. S., and Sari, Y. (2012). Glutamate transporter 1: target for the treatment of alcohol dependence. *Curr. Med. Chem.* 19, 5148–5156. doi: 10.2174/092986712803530511
- Rao, P. S., and Sari, Y. (2014b). Effects of ceftriaxone on chronic Ethanol consumption: a potential role for xCT and GLT1 modulation of glutamate levels in Male P Rats. J. Mol. Neurosci. 54, 71–77. doi: 10.1007/s12031-014-0251-5
- Rao, P. S., Saternos, H., Goodwani, S., and Sari, Y. (2015). Effects of ceftriaxone on GLT1 isoforms, xCT and associated signaling pathways in P rats exposed to ethanol. *Psychopharmacology (Berl)*. doi: 10.1007/s00213-015-3868-3. [Epub ahead of print].
- Reed, T., Page, W. F., Viken, R. J., and Christian, J. C. (1996). Genetic predisposition to organ-specific endpoints of alcoholism. *Alcohol. Clin. Exp. Res.* 20, 1528–1533. doi: 10.1111/j.1530-0277.1996.tb01695.x
- Rehm, J., Room, R., Graham, K., Monteiro, M., Gmel, G., and Sempos, C. T. (2003). The relationship of average volume of alcohol consumption and patterns of drinking to burden of disease: an overview. *Addiction* 98, 1209–1228. doi: 10.1046/j.1360-0443.2003.00467.x
- Renthal, W., and Nestler, E. J. (2009). Histone acetylation in drug addiction. *Semin. Cell. Dev. Biol.* 20, 387–394. doi: 10.1016/j.semcdb.2009.01.005
- Richard, J. M., Castro, D. C., Difeliceantonio, A. G., Robinson, M. J., and Berridge, K. C. (2013). Mapping brain circuits of reward and motivation: in the footsteps of Ann Kelley. *Neurosci. Biobehav. Rev.* 37, 1919–1931. doi: 10.1016/j.neubiorev.2012.12.008
- Richards, D. M., Heel, R. C., Brogden, R. N., Speight, T. M., and Avery, G. S. (1984). Ceftriaxone. A review of its antibacterial activity, pharmacological properties and therapeutic use. *Drugs* 27, 469–527. doi: 10.2165/00003495-198427060-00001
- Roberto, M., Gilpin, N. W., and Siggins, G. R. (2012). The central amygdala and alcohol: role of gamma-aminobutyric acid, glutamate, and neuropeptides. *Cold Spring Harb. Perspect. Med.* 2:a012195. doi: 10.1101/cshperspect.a012195
- Roberto, M., Madamba, S. G., Moore, S. D., Tallent, M. K., and Siggins, G. R. (2003). Ethanol increases GABAergic transmission at both pre- and postsynaptic sites in rat central amygdala neurons. *Proc. Natl. Acad. Sci. U.S.A.* 100, 2053–2058. doi: 10.1073/pnas.0437926100
- Roberto, M., Schweitzer, P., Madamba, S. G., Stouffer, D. G., Parsons, L. H., and Siggins, G. R. (2004). Acute and chronic ethanol alter glutamatergic transmission in rat central amygdala: an *in vitro* and *in vivo* analysis. *J. Neurosci.* 24, 1594–1603. doi: 10.1523/JNEUROSCI.5077-03.2004
- Robinson, T. E., and Berridge, K. C. (2008). Review. The incentive sensitization theory of addiction: some current issues. *Philos. Trans. R. Soc. Lond. B. Biol. Sci.* 363, 3137–3146. doi: 10.1098/rstb.2008.0093
- Root, D. H., Mejias-Aponte, C. A., Qi, J., and Morales, M. (2014). Role of glutamatergic projections from ventral tegmental area to lateral habenula in aversive conditioning. *J. Neurosci.* 34, 13906–13910. doi: 10.1523/JNEUROSCI.2029-14.2014
- Rossetti, Z. L., and Carboni, S. (1995). Ethanol withdrawal is associated with increased extracellular glutamate in the rat striatum. *Eur. J. Pharmacol.* 283, 177–183. doi: 10.1016/0014-2999(95)00344-K
- Rothstein, J. D., Patel, S., Regan, M. R., Haenggeli, C., Huang, Y. H., Bergles, D. E., et al. (2005). Beta-lactam antibiotics offer neuroprotection by increasing glutamate transporter expression. *Nature* 433, 73–77. doi: 10.1038/nature 03180
- Russo, S. J., Mazei-Robison, M. S., Ables, J. L., and Nestler, E. J. (2009). Neurotrophic factors and structural plasticity in addiction. *Neuropharmacology* 56 (Suppl. 1), 73–82. doi: 10.1016/j.neuropharm.2008.06.059
- Sari, Y., Sakai, M., Weedman, J. M., Rebec, G. V., and Bell, R. L. (2011). Ceftriaxone, a beta-lactam antibiotic, reduces ethanol consumption in alcoholpreferring rats. *Alcohol Alcohol.* 46, 239–246. doi: 10.1093/alcalc/agr023

- Sari, Y., Smith, K. D., Ali, P. K., and Rebec, G. V. (2009). Upregulation of GLT1 attenuates cue-induced reinstatement of cocaine-seeking behavior in rats. J. Neurosci. 29, 9239–9243. doi: 10.1523/JNEUROSCI.1746-09.2009
- Sari, Y., and Sreemantula, S. N. (2012). Neuroimmunophilin GPI-1046 reduces ethanol consumption in part through activation of GLT1 in alcohol-preferring rats. *Neuroscience* 227, 327–335. doi: 10.1016/j.neuroscience.2012.10.007
- Sari, Y., Sreemantula, S. N., Lee, M. R., and Choi, D. S. (2013). Ceftriaxone treatment affects the levels of GLT1 and ENT1 as well as Ethanol Intake in Alcohol-Preferring Rats. J. Mol. Neurosci. 51, 779–787. doi: 10.1007/s12031-013-0064-y
- Sato, H., Tamba, M., Ishii, T., and Bannai, S. (1999). Cloning and expression of a plasma membrane cysteine/glutamate exchange transporter composed of two distinct proteins. *J. Biol. Chem.* 274, 11455–11458. doi: 10.1074/jbc.274.17.11455
- Sattler, R., and Tymianski, M. (2001). Molecular mechanisms of glutamate receptor-mediated excitotoxic neuronal cell death. *Mol. Neurobiol.* 24, 107–129. doi: 10.1385/MN:24:1-3:107
- Schroeder, J. P., Overstreet, D. H., and Hodge, C. W. (2005). The mGluR5 antagonist MPEP decreases operant ethanol self-administration during maintenance and after repeated alcohol deprivations in alcohol-preferring (P) rats. *Psychopharmacology (Berl)* 179, 262–270. doi: 10.1007/s00213-005-2175-9
- Seal, R. P., and Amara, S. G. (1999). Excitatory amino acid transporters: a family in flux. Annu. Rev. Pharmacol. Toxicol. 39, 431–456. doi: 10.1146/annurev.pharmtox.39.1.431
- Seal, R. P., and Edwards, R. H. (2006). The diverse roles of vesicular glutamate transporter 3. Handb. Exp. Pharmacol. 137–150. doi: 10.1007/3-540-29784-7_7
- Shield, K. D., Rylett, M., Gmel, G., Kehoe-Chan, T. A., and Rehm, J. (2013). Global alcohol exposure estimates by country, territory and region for 2005–a contribution to the Comparative Risk Assessment for the 2010 Global Burden of Disease Study. Addiction 108, 912–922. doi: 10.1111/add.12112
- Skradski, S., and White, H. S. (2000). Topiramate blocks kainate-evoked cobalt influx into cultured neurons. *Epilepsia* 41, S45–S47. doi: 10.1111/j.1528-1157.2000.tb02171.x
- Snell, L. D., Nunley, K. R., Lickteig, R. L., Browning, M. D., Tabakoff, B., and Hoffman, P. L. (1996). Regional and subunit specific changes in NMDA receptor mRNA and immunoreactivity in mouse brain following chronic ethanol ingestion. *Brain Res. Mol. Brain Res.* 40, 71–78. doi: 10.1016/0169-328X(96)00038-1
- Spanagel, R. (2009). Alcoholism: a systems approach from molecular physiology to addictive behavior. *Physiol. Rev.* 89, 649–705. doi: 10.1152/physrev. 00013.2008
- Stamatakis, A. M., Jennings, J. H., Ung, R. L., Blair, G. A., Weinberg, R. J., Neve, R. L., et al. (2013). A unique population of ventral tegmental area neurons inhibits the lateral habenula to promote reward. *Neuron* 80, 1039–1053. doi: 10.1016/j.neuron.2013.08.023
- Steiner, J. P., Connolly, M. A., Valentine, H. L., Hamilton, G. S., Dawson, T. M., Hester, L., et al. (1997a). Neurotrophic actions of nonimmunosuppressive analogues of immunosuppressive drugs FK506, rapamycin and cyclosporin A. *Nat. Med.* 3, 421–428. doi: 10.1038/nm0497-421
- Steiner, J. P., Galey, D., Haughey, N. J., Asch, D., and Nath, A. (2007). Neuroprotective and antiretroviral effects of the immunophilin ligand GPI 1046. J. Neuroimmune. Pharmacol. 2, 49–57. doi: 10.1007/s11481-006-9060-0
- Steiner, J. P., Hamilton, G. S., Ross, D. T., Valentine, H. L., Guo, H., Connolly, M. A., et al. (1997b). Neurotrophic immunophilin ligands stimulate structural and functional recovery in neurodegenerative animal models. *Proc. Natl. Acad. Sci. U.S.A.* 94, 2019–2024. doi: 10.1073/pnas.94.5.2019
- Stobbs, S. H., Ohran, A. J., Lassen, M. B., Allison, D. W., Brown, J. E., and Steffensen, S. C. (2004). Ethanol suppression of ventral tegmental area GABA neuron electrical transmission involves N-methyl-D-aspartate receptors. J. Pharmacol. Exp. Ther. 311, 282–289. doi: 10.1124/jpet.104.071860
- Storck, T., Schulte, S., Hofmann, K., and Stoffel, W. (1992). Structure, expression, and functional analysis of a Na(+)-dependent glutamate/aspartate transporter from rat brain. *Proc. Natl. Acad. Sci. U.S.A.* 89, 10955–10959. doi: 10.1073/pnas.89.22.10955
- Sulzer, D. (2011). How addictive drugs disrupt presynaptic dopamine neurotransmission. *Neuron* 69, 628–649. doi: 10.1016/j.neuron.2011.02.010
- Suvarna, N., Borgland, S. L., Wang, J., Phamluong, K., Auberson, Y. P., Bonci, A., et al. (2005). Ethanol alters trafficking and functional N-methyl-D-aspartate receptor NR2 subunit ratio via H-Ras. J. Biol. Chem. 280, 31450–31459. doi: 10.1074/jbc.M504120200

- Tabakoff, B., and Hoffman, P. L. (2013). The neurobiology of alcohol consumption and alcoholism: an integrative history. *Pharmacol. Biochem. Behav.* 113, 20–37. doi: 10.1016/j.pbb.2013.10.009
- Taylor, J. R., Olausson, P., Quinn, J. J., and Torregrossa, M. M. (2009). Targeting extinction and reconsolidation mechanisms to combat the impact of drug cues on addiction. *Neuropharmacology* 56 (Suppl. 1), 186–195. doi: 10.1016/j.neuropharm.2008.07.027
- Thone-Reineke, C., Neumann, C., Namsolleck, P., Schmerbach, K., Krikov, M., Schefe, J. H., et al. (2008). The beta-lactam antibiotic, ceftriaxone, dramatically improves survival, increases glutamate uptake and induces neurotrophins in stroke. J. Hypertens. 26, 2426–2435. doi: 10.1097/HJH.0b013e32831 3e403
- Tzschentke, T. M. (2000). The medial prefrontal cortex as a part of the brain reward system. Amino Acids 19, 211–219. doi: 10.1007/s007260 070051
- Valenzuela, C. F. (1997). Alcohol and neurotransmitter interactions. Alcohol Health Res. World 21, 144–148.
- Vezina, P., and Leyton, M. (2009). Conditioned cues and the expression of stimulant sensitization in animals and humans. *Neuropharmacology* 56 (Suppl. 1), 160–168. doi: 10.1016/j.neuropharm.2008.06.070
- Volkow, N. D., and Baler, R. D. (2013). Brain imaging biomarkers to predict relapse in alcohol addiction. *JAMA Psychiatry* 70, 661–663. doi: 10.1001/jamapsychiatry.2013.1141
- Volkow, N. D., Fowler, J. S., Wang, G. J., Baler, R., and Telang, F. (2009). Imaging dopamine's role in drug abuse and addiction. *Neuropharmacology* 56 (Suppl. 1), 3–8. doi: 10.1016/j.neuropharm.2008.05.022
- Volkow, N. D., and Li, T. K. (2005). Drugs and alcohol: treating and preventing abuse, addiction and their medical consequences. *Pharmacol. Ther.* 108, 3–17. doi: 10.1016/j.pharmthera.2005.06.021
- Volkow, N. D., Wang, G. J., Fowler, J. S., Tomasi, D., Telang, F., and Baler, R. (2010). Addiction: decreased reward sensitivity and increased expectation sensitivity conspire to overwhelm the brain's control circuit. *Bioessays* 32, 748–755. doi: 10.1002/bies.201000042
- Volman, S. F., Lammel, S., Margolis, E. B., Kim, Y., Richard, J. M., Roitman, M. F., et al. (2013). New insights into the specificity and plasticity of reward and aversion encoding in the mesolimbic system. *J. Neurosci.* 33, 17569–17576. doi: 10.1523/JNEUROSCI.3250-13.2013
- Wang, Y., and Qin, Z. H. (2010). Molecular and cellular mechanisms of excitotoxic neuronal death. *Apoptosis* 15, 1382–1402. doi: 10.1007/s10495-010-0481-0
- Weiss, F., Ciccocioppo, R., Parsons, L. H., Katner, S., Liu, X., Zorrilla, E. P., et al. (2001). Compulsive drug-seeking behavior and relapse. Neuroadaptation, stress, and conditioning factors. *Ann. N.Y. Acad. Sci.* 937, 1–26. doi: 10.1111/j.1749-6632.2001.tb03556.x
- Wheeler, R. A., and Carelli, R. M. (2009). Dissecting motivational circuitry to understand substance abuse. *Neuropharmacology* 56 (Suppl. 1), 149–159. doi: 10.1016/j.neuropharm.2008.06.028

- White, N. M. (1996). Addictive drugs as reinforcers: multiple partial actions on memory systems. *Addiction* 91, 921–949; discussion 951–965. doi: 10.1046/j.1360-0443.1996.9179212.x
- Wirkner, K., Eberts, C., Poelchen, W., Allgaier, C., and Illes, P. (2000). Mechanism of inhibition by ethanol of NMDA and AMPA receptor channel functions in cultured rat cortical neurons. *Naunyn Schmiedebergs Arch. Pharmacol.* 362, 568–576. doi: 10.1007/s002100000262
- Wise, R. A. (2005). Forebrain substrates of reward and motivation. J. Comp. Neurol. 493, 115–121. doi: 10.1002/cne.20689
- Wise, R. A., and Koob, G. F. (2014). The development and maintenance of drug addiction. *Neuropsychopharmacology* 39, 254–262. doi: 10.1038/npp.2013.261
- Zerangue, N., and Kavanaugh, M. P. (1996). Flux coupling in a neuronal glutamate transporter. *Nature* 383, 634–637. doi: 10.1038/383634a0
- Zhou, F. C., Sahr, R. N., Sari, Y., and Behbahani, K. (2006). Glutamate and dopamine synaptic terminals in extended amygdala after 14-week chronic alcohol drinking in inbred alcohol-preferring rats. *Alcohol* 39, 39–49. doi: 10.1016/j.alcohol.2006.06.013
- Zhou, Z., Karlsson, C., Liang, T., Xiong, W., Kimura, M., Tapocik, J. D., et al. (2013). Loss of metabotropic glutamate receptor 2 escalates alcohol consumption. *Proc. Natl. Acad. Sci. U.S.A.* 110, 16963–16968. doi: 10.1073/pnas.1309839110
- Zhu, W., Bie, B., and Pan, Z. Z. (2007). Involvement of non-NMDA glutamate receptors in central amygdala in synaptic actions of ethanol and ethanol-induced reward behavior. J. Neurosci. 27, 289–298. doi: 10.1523/JNEUROSCI.3912-06.2007
- Zorrilla, E. P., and Koob, G. F. (2013). Amygdalostriatal projections in the neurocircuitry for motivation: a neuroanatomical thread through the career of Ann Kelley. *Neurosci. Biobehav. Rev.* 37, 1932–1945. doi: 10.1016/j.neubiorev.2012.11.019
- Zucker, R. A. (1995). "Pathways to alcohol problems and alcoholism: a developmental account of the evidence for multiple alcoholisms and for contextual contributions to risk," in *The Development of Alcohol Problems: Exploring the Biopsychosocial Matrix of Risk*, eds R. A. Zucker, G. M. Boyd, J. Howard (Rockville, MD: NIAAA Research Monograph 26;U.S. Department of Health and Human Services;National Institute on Alcohol Abuse and Alcoholism), 255–289.

Conflict of Interest Statement: The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Copyright © 2015 Rao, Bell, Engleman and Sari. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) or licensor are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.