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2 3	Three years of alendronate treatment results in similar levels of				
4	vertebral microdamage as after one year of treatment				
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Three years of daily alendronate treatment increases microdamage in vertebral bone but does not significantly increase it beyond levels of microdamage found after 1 year of treatment. This suggests microdamage accumulation peaks during the early period of bisphosphonate treatment, and does not continue to accumulate with longer periods of treatment.

Introduction: Clinically-relevant doses of alendronate increase vertebral microdamage by 4- to 43 5-fold in skeletally mature beagles after 1 year of treatment. The goal of this study was to 44 45 determine if microdamage would continue to accumulate with three years of alendronate treatment in an intact beagle dog model. 46 Methods: One-year-old female beagles were treated with daily oral doses of vehicle (VEH, 1 47 ml/kg/day) or alendronate (ALN, 0.2 mg/kg/day or 1.0 mg/kg/day) for three years. These ALN 48 doses were chosen to approximate, on a mg/kg basis, those used to treat osteoporosis (ALN0.2) 49 50 and Paget's disease (ALN1.0). Microdamage accumulation, static and dynamic 51 histomorphometry, densitometry, and mechanical properties of lumbar vertebrae were assessed. Comparisons were made among the three groups treated for three years, and also within each 52 treatment group, to animals treated under the same conditions for one year (Allen et al. Bone, 53 2006). 54 55 Results: Overall microdamage accumulation (crack surface density) was not significantly higher in animals treated for three years with either dose of ALN, while crack density increased 56 significantly (100%; p < 0.05) with the higher dose of ALN when compared to VEH. Both ALN 57 doses significantly suppressed the rate of bone turnover (-60% versus VEH). There was no 58 difference among groups for any of the structural biomechanical properties - ultimate load, 59 stiffness, or energy absorption. However, when adjusted for areal bone mineral density ALN-60 treated animals had significantly lower energy absorption (-20%) compared to VEH. Toughness, 61 the energy absorption capacity of the bone tissue, was significantly lower than VEH for both 62 ALN0.2 (-27%) and ALN1.0 (-33%). Compared to animals treated for one year, there was no 63 significant difference in microdamage accumulation for either ALN dose. VEH-treated animals 64 had significantly lower bone turnover (-58%) and significantly higher levels of microdamage (+ 65 300%) compared to values in 1 year animals. Toughness was significantly lower in animals 66 treated for 3 years with ALN1.0 (-18%) compared to animals treated for 1 year while there was no 67 difference in toughness between the two treatment durations for either VEH or ALN0.2. 68 Conclusions: Although three years of alendronate-treatment resulted in higher microcrack 69 density in vertebral trabecular bone compared to control dogs, the amount of microdamage was 70 not significantly higher than animals treated for 1 year with similar doses. This suggests that 71 bisphosphonate-associated increases in microdamage occur early in treatment. Because 72 toughness continued to decline significantly over three years of treatment at the higher ALN dose, 73 decreases in toughness are probably not dependent on damage accumulation. 74 75

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76 INTRODUCTION

Microdamage accumulates with age [1,2] and may play an important role in age-associated bone fragility [3,4]. Microdamage formation occurs in response to mechanical loads [5-7], preferentially at sites of increased tissue mineralization [8-10], and is removed by remodeling [6,7]. The level of skeletal microdamage is determined by the balance between microdamage formation and its removal. Therefore conditions that either increase microdamage formation, or decrease its removal, can have a significant impact on the accumulation of microdamage and bone fragility.

84 Bisphosphonates are efficacious for reducing fractures due to their suppression of bone 85 remodeling [11-13]. However, as reductions in remodeling are permissive for the accumulation of microdamage, bisphosphonate treatment also increases skeletal microdamage. Numerous 86 87 animal studies have noted significant increases in microdamage following bisphosphonate 88 treatment [14-18]. This accumulation of microdamage occurs with alendronate and risedronate doses comparable to those used for the treatment of post-menopausal osteoporosis, although the 89 accumulation is greater when higher doses are given (e.g. those approximating doses used for 90 treatment of Paget's disease) [18]. 91

92 Whether microdamage accumulation continues or plateaus with extended bisphosphonate treatment is not known, yet has significant implications as some patients now enter their second 93 decade of treatment. Studies to date have assessed microdamage at a single time point, most 94 often one year. Recently, Komatsubara et al. [16,17] reported that 3 years of daily incadronate, 95 at 2.5 or 5x the clinical dose, significantly increased the accumulation of microdamage in both the 96 vertebrae and rib of dogs. As no data are available concerning microdamage levels with shorter 97 term incadronate treatment (< 3 years), this study was not able to address whether microdamage 98 99 accumulation continues or plateaus with prolonged bisphosphonate treatment.

Animal studies documenting increased microdamage with bisphosphonate-treatment have consistently shown increases in vertebral bone strength and stiffness, leading to questions regarding the implications of increased microdamage with bisphosphonates. However, in all but one of these studies [16] bisphosphonate treatment reduced bone toughness, the energy

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absorption capacity of the bone tissue [14,15,17,18]. Furthermore, when normalized for
 increases in BMD, energy absorption capacity at the whole bone (structural) level was
 significantly compromised following 1 year of alendronate treatment [19]. If microdamage
 continues to accumulate with prolonged bisphosphonate treatment it is possible that this could
 lead to further reductions in work to failure and toughness.

The goal of this study was to test the hypothesis that microdamage continues to accumulate 109 throughout the duration of bisphosphonate treatment and that this continued accumulation is 110 accompanied by a progressive decline in energy absorption and toughness. We have recently 111 112 documented that clinically-relevant doses of alendronate reduce vertebral bone turnover by more 113 than 70%, increase microdamage by 4- to 5-fold and non-significantly reduce vertebral toughness by 14-17% in skeletally mature beagles after 1 year of treatment [18]. The current study reports 114 results from animals treated for three years with the same doses of alendronate used in the one 115 year study. This allows both an across-treatment analysis at the three-year time point (vehicle 116 versus alendronate) as well as a within-treatment analysis across time points (1 versus 3 years). 117

118

119 MATERIALS AND METHODS

120 Animals

All procedures were approved prior to the study by the Indiana University School of Medicine 121 Animal Care and Use Committee. Thirty-six female beagles (1-2 years old upon arrival) were 122 123 purchased from LBL (Reelsville, IN). Upon arrival, lateral X-rays of all dogs were obtained to confirm skeletal maturity (closed proximal tibia and lumbar vertebra growth plates). Animals were 124 housed two per cage in environmentally controlled rooms at Indiana University School of 125 Medicine's AALAC accredited facility and provided standard dog chow and water. Two dogs 126 127 (both in the ALN 0.2 group) developed hernias, both in year 2, that required surgery. One of these animals developed a second hernia which progressed to the point of needing to be 128 terminated early (month 34 of treatment); this animal was still included in all analyses. All other 129 animals completed the 36 month treatment without serious complication. 130

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132 Experimental Design

133 Following two weeks of acclimatization, animals were assigned to treatment groups (n=12/group) 134 by matching body weights. All dogs were treated daily for 3 years with oral doses of vehicle (saline, 1 ml/kg/day) or alendronate sodium (0.20 or 1.00 mg/kg/day; Merck and Co., Inc). 135 Alendronate doses were chosen to approximate, on a mg/kg basis, the doses used for treatment 136 of post-menopausal osteoporosis and Paget's disease, respectively. Alendronate was dissolved 137 in saline and administered to the dogs orally with a syringe. Vehicle-treated animals received 1 138 ml/kg/day of saline. Dosing was performed each morning after an overnight fast and at least 2 139 140 hours prior to feeding. 141 Prior to necropsy, animals were injected with calcein (0.20 mL/kg, IV) using a 2-12-2-5 labeling schedule. Animals were euthanized by intravenous administration of sodium 142 pentobarbital (0.22mg/kg Beuthanasia-D Special). After death, lumbar vertebrae were dissected 143 144 and saved for analyses. The second and third lumbar vertebrae were fixed in 10% neutral buffered formalin while the fourth lumbar vertebra was wrapped in saline-soaked gauze and 145 frozen (-20°C). All tissue preparation, processing, and analyses were similar to those used for 146 dogs treated for one year [18]. 147

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149 Histology (Static, dynamic, and microdamage)

Static and dynamic histomorphometric measures of trabecular bone were obtained on second
 lumbar vertebrae (L2). Bones were embedded undecalcified in methyl methacrylate (MMA;
 Aldrich). Mid-sagittal (4 µm) sections were cut using a Reichert-Jung 2050 microtome (Magee
 Scientific, Inc) and stained with McNeal's tetrachrome for static histomorphometry. Mid-sagittal (8
 µm) sections were cut and left unstained for dynamic histomorphometry and wall thickness
 measures.

Third lumbar vertebrae (L3) were processed for microdamage assessment by bulk staining in basic fuchsin as previously described [18,20]. Using 1% basic fuchsin dissolved in increasing concentrations of ethanol, specimens were stained according to the following schedule: 8 hours 80% (with one change to fresh 80% after 4 hours), overnight in 95% (with one change to fresh

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95%), 8 hours in 100% (with one change to fresh 100% after 4 hours). Bones were placed under 160 vacuum (20 in Hg) for all stages during the day and left on the bench top overnight. Following 161 staining, bones were washed in 100% ethanol and embedded undecalcified in MMA. Mid-sagittal 162 (80-100 µm) sections were cut using a diamond wire saw (Histosaw; Delaware Diamond Knives). 163 Histological measurements were made using a semiautomatic analysis system (Bioquant 164 OSTEO 7.20.10, Bioguant Image Analysis Co.) attached to a microscope equipped with an 165 ultraviolet light source (Nikon Optiphot 2 microscope, Nikon). A 5 x 5 mm region of interest, 166 located 1 mm below the cranial plateau, was used for sampling. Static and dynamic variables 167 168 were measured and calculated in accordance with ASBMR recommended standards [21]. 169 Microdamage was assessed using UV fluorescence as previously described [22]. Measurements included crack length (Cr.Le, µm) and crack number (Cr.N), with calculations of crack density 170 (Cr.Dn, #/mm²; Cr.N / bone area) and crack surface density (Cr.S.Dn, µm/mm²; Cr.N * Cr.Le / 171 172 bone area). 173 Densitometry 174 Areal bone mineral density (aBMD, g/cm²) of the fourth lumbar vertebra (L4), without the posterior 175 elements or cranial/caudal endplates, was quantified using a PIXImus II densitometer (Lunar 176

Corp.). Volumetric bone density and geometry of the L4 vertebra was quantified using a Norland
Stratec XCT Research SA+ pQCT (Stratec Electronics). One slice (0.07 X 0.07 x 0.50 mm voxel
size) was taken at three locations (25, 50 and 75% of total vertebra height). Total, trabecular,
and cortical volumetric bone mineral density (vBMD, mg/cm³) and cross-sectional area (CSA,

mm²) were obtained for each slice and then averaged together to obtain a single representative
 value for each specimen.

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184 Biomechanical Testing

The biomechanical properties of L4 vertebrae were quantified using a servohydraulic testing system (MTS Bionix, MTS Corporation). Compression to failure was carried out on saline soaked specimens using displacement control mode (20 mm/min). Load vs displacement data were

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digitally recorded at a sampling rate of 10Hz. Plots were analyzed for determination of ultimate force (F), stiffness (k) and work to ultimate force (w). Apparent material-level properties ultimate stress (σ_{ult}), modulus (E), and toughness (U) were estimated using the following equations: $\sigma_{ult} =$ (F / CSA) / BV/TV; E = (k * (height / CSA)) / BV/TV; U = (w / (height * CSA)) / BV/TV, where cross sectional area (CSA) is from pQCT, height measured using digital calipers, and BV/TV from L2 histomorphometry.

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195 Statistics

196 All statistical tests were performed using SAS software (SAS Institute, Inc.). To determine 197 whether variables were different among treatment groups after 3 years, data were evaluated 198 using a one-way analysis of variance (ANOVA) with Fisher's protected least-significant difference (PLSD) post-hoc tests. Strength-density and energy absorption-density relationships from three-199 year treated animals were compared between VEH and ALN treatments using analyses of 200 covariance with least square means (LSM) used to determine differences in parameters after 201 accounting for aBMD. To determine whether changes occurred within treatment groups across 202 203 time, t-tests were used to compare data from animals treated for three years with results from an earlier study in our lab which treated animals under the same conditions for one year [18]. For all 204 205 tests, $p \leq 0.05$ was considered statistically significant. All data are presented as mean \pm standard error. 206

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208 RESULTS

At the conclusion of the study there was no significant difference in body mass among the three groups (VEH: 12.6 ± 0.6 kg; ALN0.2: 12.4 ± 0.5 kg; ALN1.0: 11.6 ± 0.7 kg; p = 0.492).

Crack density, the number of microcracks per mm bone tissue, was significantly higher than VEH for ALN1.0 (+100%, p = 0.01), but not ALN0.2 (+50%; p = 0.12)(Figure 1A). Mean crack length was significantly smaller in both ALN-treated groups compared to VEH (-20% for both) (Figure 1B). Crack surface density, the product of crack density and crack length, was not significantly different among groups (Figure 1C).

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Activation frequency (Ac.f) was significantly lower than VEH in both ALN0.2 (-59%) and ALN1.0 (-60%) treated animals. The reduction in Ac.f resulted from significant suppression of both mineral apposition rate (MAR) and mineralizing surface (MS/BS), with no change in wall thickness. MAR was 17% lower than VEH for both doses of ALN while MS/BS was -51% and -62% for ALN0.2 and ALN1.0 groups, respectively (Table 1).

Structural biomechanical properties – ultimate load, stiffness, and energy to ultimate load 221 - were not significantly different among the treatment groups (Table 2). When normalized for 222 aBMD, there was no difference in the strength-density relationship between VEH- and ALN-223 224 treated animals (Figure 2A). The slope of the energy absorption-density relationship was similar 225 between treatments yet at a given aBMD the energy absorption capacity was significantly lower in vertebrae from ALN-treated animals (-20%, p = 0.01) compared to VEH (Figure 2B). For both the 226 strength-density and energy absorption-density relationships the two doses of ALN were pooled 227 228 as the results were similar when doses were assessed separately.

Toughness, the energy absorption capacity of the bone tissue, was significantly lower in 229 both ALN0.2 (-26%) and ALN1.0 (-33%) groups compared to VEH (Table 2). There was no 230 difference among groups for the other two material-level properties, ultimate stress and modulus. 231 232 Vertebral aBMD was not significantly different among groups while vBMD tended to be higher (p=0.056) in both ALN0.2 and ALN1.0 groups (both +7%) versus VEH (Table 3). 233 Trabecular vBMD, cortical vBMD, and cross-sectional area were not different among the three 234 235 treatment groups. Trabecular bone volume, assessed by histology, was significantly greater in both ALN0.2 (+23%) and ALN1.0 (+31%) treatment groups compared to VEH (Table 3). 236 After three years of treatment Ac.f. was significantly lower in ALN0.2 (-40%, p = 0.01), but 237 not ALN1.0 (-30%, p = 0.30), compared to similar treatment groups at 1 year (Figure 3a). The 238 239 level of microdamage (both Cr.Dn and Cr.S.Dn) was not significantly different at 3 years compared to 1 year for either ALN group (Figure 3b, 3c). Compared to 1-year of treatment, 240 ALN0.2 had higher ultimate load (+21%), stiffness (+55%) and modulus (+65%) at 3 years while 241 ALN1.0 had significantly higher stiffness (+42%) and modulus (+30%) and lower toughness (-242 18%) (Table 4 and Figure 3). VEH-treated animals had significantly lower Ac.f. (-58%), higher 243

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- microdamage accumulation (+301%), and higher structural- and material-level strength and
 stiffness at 3 years compared to VEH-treated animals after 1 year (Table 4 and Figure 3).
- 246

247 DISCUSSION

Animal studies have consistently documented higher levels of microdamage in bisphosphonate-248 treated animals [14-18] yet it has remained unclear whether microdamage accumulation 249 continues or plateaus with extended bisphosphonate treatment. Recently, we have documented 250 that clinically-relevant doses of alendronate increase microdamage by 4- to 5-fold in skeletally 251 252 mature beagles after 1 year of treatment [18]. We now present data to show that the level of 253 microdamage in vertebral trabecular bone does not significantly increase with an additional two years of alendronate treatment (3 years total treatment duration) at doses approximating those 254 used to treat post menopausal osteoporosis or Paget's disease. 255

As remodeling is necessary to remove microdamage [6,7], bisphosphonate-treatment would 256 be expected to allow accumulation of damage due to turnover suppression. While the degree of 257 turnover suppression is correlated to the degree of microdamage accumulation [15,17,18], even 258 mild suppression of turnover (~40%) with bisphosphonate-treatment is sufficient to allow 259 260 significant increases in microdamage [18]. The current study shows that the initial suppression of turnover with bisphosphonate treatment has the greatest influence on microdamage 261 accumulation. Following one year of ALN-treatment, vertebral bone turnover is suppressed by 262 263 ~70%, associated with a 4 to 5-fold increase in microdamage [18]. With an additional 2 years of treatment, and a continued decline in turnover (-30 to -40% compared to values in 1 year 264 animals), microdamage was not significantly increased (1.3- and 1.6-fold higher than VEH). The 265 most plausible explanations for this finding are 1) microdamage can be controlled at a new 266 267 equilibrium level even with only 30% of normal bone turnover and/or 2) there is a reduced formation of microdamage. The latter could result from the lowering of trabecular strains due to 268 the 20-30% increase in bone volume (Table 3). 269 Consistent with the relationship between turnover suppression and microdamage 270

accumulation, animals treated for 3 years with vehicle had significantly lower turnover (-58%) and

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significantly higher levels of microdamage (+300%) compared to those treated for 1 year. These
data highlight that microdamage accumulation is not due to bisphosphonates, per se, but rather
the reduction in turnover brought about by bisphosphonate treatment.

275 Toughness, the energy absorption capacity of the material, is consistently reduced in bisphosphonate-treated animals [14,15,17,18]. This change has often been attributed to 276 microdamage accumulation although a cause and effect has yet to be established. The current 277 results provide two pieces of evidence to suggest microdamage accumulation is not 'causing' 278 reduced toughness in bisphosphonate-treated bone. First, despite higher levels of microdamage 279 280 in VEH-treated animals after three years (compared to 1 year), there was no change in bone 281 toughness. Second, despite no significant difference in microdamage accumulation between animals treated for one and three years with either dose of ALN, animals treated with ALN1.0 had 282 significantly lower bone toughness at three years compared to one year. While these data do not 283 284 disprove a cause/effect relationship they strongly suggest bisphosphonate-associated reductions in bone toughness extend beyond simply the accumulation of microdamage. 285

Structural biomechanical properties – ultimate load, stiffness, and energy absorption – were 286 not significantly different than VEH after 3 years of ALN treatment. These results differ from 287 288 those at one year, where both doses of ALN significantly increased vertebral stiffness [18] and the higher dose of ALN increased strength [15]. The absence of difference among these groups 289 treated for 3 years is likely the result of significant increases in VEH-treated animals, which had 290 291 significantly higher ultimate load (+24%) and stiffness (+68%) compared to values in 1 year treated animals. These higher structural-level mechanical properties in 3-year VEH-treated 292 animals compared to 1-year VEH-treated animals likely result from age-associated periosteal 293 expansion. Vertebral cross-sectional area, which plays a significant role in determining structural 294 295 parameters and results from continued periosteal expansion, was significantly higher (+16%) in the 3-year VEH group compared to the 1 year group. Material-level properties – ultimate stress 296 and modulus – were also higher in VEH-treated animals at 3 years compared to one year. We 297 have recently documented increases in collagen cross-linking and collagen maturity of vertebrae 298 that are attributable to turnover suppression [23]. As the organic matrix is known to affect 299

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material properties, we hypothesize that the reduction in turnover between 1 year and 3 years in
 vehicle-treated animals (-58%) results in an increase in collagen cross-linking and maturity which,
 in conjunction with other parameters such as mineralization and microdamage, determine
 material-level biomechanical properties [24].

An alternative approach to investigate the effects of bisphosphonate treatment on 304 biomechanical properties is to compare the relationships between bone density and 305 biomechanical properties. Proposed by Hernandez and Keaveny [25], these relationships allow 306 the determination of changes in bone strength or energy to fracture that are not accounted for by 307 a change in bone mass (aBMD). ALN-treated animals had 20% lower energy absorption capacity 308 309 at a given aBMD, indicating that an increase in BMD is necessary with alendronate treatment to maintain energy absorption capacity at a level comparable to non-treated bone. This result is 310 consistent with the 22% lower energy absorption at a given aBMD following one year of treatment 311 312 with doses of ALN approximating those used to treat osteoporosis [19].

Given the invasive nature of both microdamage and biomechanical property measures, it 313 proves difficult to determine if the changes noted in the current study extend to humans treated 314 with bisphosphonates. Higher levels of microdamage exist in bisphosphonate-treated women 315 316 [26], although there is no data to support whether there exists a similar treatment-duration accumulation pattern as noted in the current study. Bisphosphonates have clear anti-fracture 317 efficacy suggestive of improved biomechanical properties [11-13]. However, given the multi-318 319 factorial nature of fractures it remains possible that reduced toughness or lower energy absorption at a given aBMD could exist even in light of an overall population reduction in fracture 320 risk with bisphosphonates. Indeed both toughness and energy absorption are compensated for 321 by the increased bone density that routinely occurs with bisphosphonate treatment, but the 322 323 material properties of the tissue are nevertheless compromised.

In conclusion, three years of alendronate-treatment resulted in higher microcrack density in vertebral trabecular bone of intact beagle dogs, yet the amount of microdamage was not significantly higher than in animals treated with equivalent doses for 1 year. This suggests that increased skeletal microdamage associated with turnover suppression occurs early in treatment,

- 328 and does not progress with longer treatment duration. Because toughness continued to decline
- significantly over three years of treatment at the higher ALN dose, decreases in toughness are
- 330 probably not dependent on damage accumulation.

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342 FIGURE LEGENDS

343 Figure 1. Microdamage parameters in vertebral trabecular bone following three years of daily vehicle (VEH) or alendronate (ALN) treatment (0.20 or 1.00 mg/kg/day). (A) Crack density, the 344 345 number of microcracks normalized to bone area, was significantly higher (p = 0.032) in animals treated with the higher dose of alendronate (ALN1.0). (B) Mean crack length was significantly 346 lower (p = 0.013) with both doses of ALN. (C) Crack surface density, the product of crack 347 density and crack length, was not significantly different among groups (p = 0.149). There was no 348 significant difference between doses of ALN for any microdamage parameter. * p < 0.05 versus 349 VEH. 350

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Figure 2. Strength-density (A) and energy absorption-density (B) relationships of vertebral bone 352 from beagles treated for 3 years with vehicle (VEH) or alendronate (ALN). Areal bone mineral 353 density (aBMD) was assessed by densitometry while strength and energy absorption were 354 assessed by monotonic compression biomechanical tests. The strength-density relationship was 355 similar for vehicle (\circ , y = 20397x - 2065) and alendronate-treated animals (\bullet , y = 23385x - 3033). 356 The slope of the energy absorption-density relationship was similar yet the intercepts differed 357 358 significantly between vehicle (\circ , y = 9912x – 1306) and alendronate-treated animals (pooled (\bullet), y = 11489x - 2228). After adjusting for aBMD, the energy absorption capacity was significantly 359 lower (-20%) in ALN-treated specimens compared to VEH. ALN-treated groups were combined 360 as there was no difference between the two doses for either relationship. 361

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Figure 3. Differences in activation frequency (A) crack density (B) and toughness (C) between animals treated for 1 and 3 years with vehicle (VEH) or alendronate (ALN0.2 and ALN1.0). (A) Activation frequency was significantly lower in both VEH and ALN0.2 after 3 years of treatment compared to animals at 1 year. (B) Crack density, the number of microcracks normalized to bone area, was not significantly different for either dose of ALN but was significantly higher in VEH-treated animals after 3 years compared to animals treated for 1 year. (C) Toughness, the material-level energy absorption capacity, was significantly lower in animals treated with the

- higher dose of alendronate (ALN1.0) after 3 years compared to values at 1 year. * p < 0.05
- versus 1 year animals within treatment.

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	Vehicle	Alendronate	Alendronate	
	1 ml/kg/day	0.20 mg/kg/day	1.00 mg/kg/day	p value
MAR, um/day	1.36 ± 0.08	1.13 ± 0.06 *	1.11 ± 0.07 *	0.028
MS/BS, %	7.63 ± 0.99	4.15 ± 0.51 *	3.30 ± 0.92 *	0.002
BFR/BS, µm ³ /µm ² /year	36.75 ± 0.10	16.70 ± 0.04 *	14.31 ± 0.09 *	0.0004
Ac.f, #/year	0.793 ± 0.097	0.328 ± 0.037 *	0.319 ± 0.094 *	0.0002

N= 12 animals per treatment group. MAR, mineral apposition rate; MS/BS, mineralizing surface per unit bone surface; BFR/BS, bone formation rate normalized to bone surface; Ac.f, activation frequency. * p < 0.05 vs vehicle.

Table 2. Compressive biomechanical properties of the fourth lumbar vertebrae

	Vehicle	Alendronate	Alendronate	
	1 ml/kg/day	0.20 mg/kg/day	1.00 mg/kg/day	p value
Ultimate Load, N	4656 ± 234	4966 ± 254	4847 ± 304	0.710
Stiffness, N/mm	11889 ± 1153	14241 ± 1146	13622 ± 956	0.299
Energy to Ultimate Load, Nmm	1961 ± 198	1764 ± 140	1581 ± 136	0.260
Ultimate Stress/(BVTV)	1.78 ± 0.11	1.64 ± 0.14	1.51 ± 0.07	0.229
Modulus/(BVTV)	67.3 ± 6.9	74.1 ± 9.1	62.9 ± 3.2	0.518
Toughness/(BVTV)	0.049 ± 0.004	0.036 ± 0.003 *	0.033 ± 0.002 *	0.004

N= 12 animals per treatment group. BV/TV, bone volume normalized to tissue volume. * p < 0.05 vs vehicle.

Table 3. Lumber vertebrae bone mineral density, geometry, and bone volume

	Vehicle 1 ml/kg/day	Alendronate 0.20 mg/kg/day	Alendronate 1.00 mg/kg/day	p value
Whole aBMD, g/cm ²	0.330 ± 0.010	0.343 ± 0.011	0.337 ± 0.009	0.644
Total vBMD, mg/cm ³	554 ± 14	591 ± 13	597 ± 12	0.056
Trabecular vBMD, mg/cm ³	329 ± 7	349 ± 7	341 ± 6	0.111
Cortical vBMD, mg/cm ³	1020 ± 10	1019 ± 7	1027 ± 6	0.746
CSA, mm ²	136 ± 4.5	131 ± 5.3	127 ± 5.1	0.461
Trabecular BV/TV, %	19.9 ± 1.3	24.5 ± 1.7 *	25.8 ± 1.7 *	0.029

N= 12 animals per treatment group. aBMD, areal bone mineral density; vBMD, volumetric bone mineral density; CSA, cross sectional area; BV/TV, bone volume normalized to tissue volume. * p < 0.05 vs vehicle. Table 4. Percent difference between animals treated for 1 and 3 years within treatment

	Vehicle 1 ml/kg/day	Alendronate 0.20 mg/kg/day	Alendronate 1.00 mg/kg/day
aBMD, g/cm ²	-1	-5	-4
CSA, mm ²	+17	+15	+15
Ultimate Load, N	+24	+21	+14
Ultimate Stress/(BVTV)	+20	+18	-1
Stiffness, N/mm	+68	+55	+42
Modulus/(BV/TV)	+62	+65	+30
Energy to Ultimate Load, Nmm	+13	+7	-3

Comparisons between parameters of lumbar vertebrae after 1 and 3 years of treatment (1 year data from Allen et al, Bone 2006). Values represent percent difference between 1 and 3 year animals. **Bold** denotes significance (p < 0.05). N= 12 animals per treatment group per time point. aBMD, areal bone mineral density; CSA, cross sectional area; BV/TV, bone volume normalized to tissue volume.

Figure 1.



Figure 2.



Figure 3.

