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A dosimetry and radiobiological model for intravascular brachytherapy treatment planning with radioisotope emitting stents

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FLORIDA INTERNATIONAL UNIVERSITY

Miami, Florida

A DOSIMETRY AND RADIOBIOLOGICAL MODEL FOR INTRAVASCULAR BRACHYTHERAPY TREATMENT PLANNING WITH

RADIOISOTOPE EMITTING STENTS

A thesis submitted **in partial** fulfillment of the

requirements for the degree of

MASTER OF SCIENCE

in

BIOMEDICAL ENGINEERING

by

Maria Alejandra **Caceres**

To: Dean Vish Prasad College of Engineering

This thesis, written by Maria Alejandra Caceres, and entitled A Dosimetry and Radiobiological Model for Intravascular Brachytherapy Treatment Planning with Radioisotope Emitting Stents, having been approved in respect to style and intellectual content, is referred to you for judgment.

We have read this thesis and recommend that it be approved.

Richard Schoephoerster

Anthony McGoron

Juan Franquiz, *Major Professor*

Date of Defense: July 24, 2003

The thesis of Maria Alejandra Caceres is approved.

Dean Vish Prasad College of Engineerip

Dean Doyglas Wartzok University Graduate School

Florida International University, 2003

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ABSTRACT OF THE THESIS

A DOSIMETRY AND RADIOBIOLOGICAL MODEL FOR INTRAVASCULAR BRACHYTHERAPY TREATMENT PLANNING WITH RADIOISOTOPE EMITTING STENTS

by

Maria **Alejandra Caceres**

Florida International University, 2003

Miami, Florida

Professor Juan Franquiz, Major Professor

The aim of this study was to develop a practical, versatile and fast dosimetry and radiobiological model for calculation of the 3D dose distribution and radiobiological effectiveness of radioactive stents. The algorithm was written in Matlab 6.5 programming language and is based on the dose point kernel convolution. The dosimetry and radiobiological model was applied for evaluation of the 3D dose distribution of ³²P, ⁹⁰Y, Re and ¹⁷⁷Lu stents. Of the four, ³²P delivers the highest dose, while ⁹⁰Y, ¹⁸⁸Re and ¹⁷⁷Lu require high levels of activity to deliver a significant therapeutic dose in the range of 15-30 Gy. **Results of the radiobiological model demonstrated that the same physical dose delivered by different radioisotopes produces significantly different radiobiological effects. This type of theoretical dose calculation can be useful in the development of new stent designs, the planning of animal studies and clinical** trials, **and clinical decisions involving individualized treatment plans.**

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CHAPTER 1 INTRODUCTION

Coronary artery **disease** (CAD) is **the major cause of death in most western countries.** Approximately 12,800,000 Americans suffer from CAD and nearly 500,000 die from heart attacks caused by CAD (2003 Heart and Stroke Statistical Update, **American Heart Association). Therefore its treatment and prevention is of great interest** to the cardiology community. CAD has been successfully treated with percutaneous **transluminal coronary angioplasty** (PTCA). **However, restenosis, or re-narrowing of the** artery, remains the major limitation of this technique with rates of approximately *30-50%* (Fischell et al., 2000; Fox, 2002; King, 2002). The use of coronary stents in conjunction **with balloon angioplasty has reduced restenosis rates** by **20-30% as a result of their** positive effect on elastic recoil and constrictive remodeling (Fox, 2002; King, 2002). **Despite their success, in-stent restenosis is still a significant problem with stents, and the stent presence may in fact aggravate neointima formation. High in-stent restenosis rates** represent a major impediment to the success of PTCA (Hehrlein and Kubler, 1997; Fischell, 1998; Nath et al., 1999; Fox, 2002).

Vascular irradiation, otherwise known as intravascular brachytherapy (IVBT) has **emerged as a prophylaxis for the prevention of restenosis after angioplasty, and is considered a very promising anti-restenosis therapy (Kuntz and Baim, 2000; Serruys and** Kay, 2000; **Sims** et al., 2002; Fox, 2002). A radiation delivery platform **such as a radioactive stent provides a mechanism in which mechanical and irradiation benefits can be combined. The three main components of restenosis identified as early recoil,**

remodeling and neointimal hyperplasia, can all be successfully treated with a radioactive stent. The stent itself serves as a scaffold, thus maintaining the lumen of the artery open, even after early recoil soon after the angioplasty procedure. In addition, the ionizing radiation emitted from the stent has been shown to have positive effects on the reduction of neointimal growth and negative vascular remodeling (Mintz et al., 1996; Schwartz and Holmes, 2002).

The safety and efficacy of radioactive stents has been well demonstrated in several animal studies (Hehrlein et al., *1995;* **Carter and Laird, 1996; Fischell, 1998; Carter et al., 1999; John et al., 2000), yet experimental results are inconsistent and restenosis at stent edges, otherwise known as the "candy wrapper effect", remains a major problem with this treatment modality (Albeiro et al., 2000b; Fischell et al., 2000; Serruys and Kay, 2000; Fox, 2002). This can be attributed to geographic miss, excessive barotrauma during the stent implantation procedure, or inadequate irradiation at the stent edges due to the rapid dose falloff of the radiation source (Fischell et al., 2000; Serruys and Kay, 2000; Fox, 2002). It is clear that for radioactive stents to be successful, the problem of edge restenosis must be overcome. This requires an in-depth evaluation of different candidate radioisotopes and possibly a combined dose delivery strategy to maximize radiation at stent edges.**

The planning of any radiotherapy treatment is a crucial step in the overall success of the treatment. Dosimetric analysis is a first and necessary step as it is important to characterize and optimize the dose that will be delivered to the arterial wall tissue. While the overall benefits of intravascular brachytherapy are well documented, uncertainties still remain with uniform radiation delivery, optimal radioisotope, radiation dose, dose

rate, initial activity and target tissue (Fischell et al., 2000; Fox, 2002). Ultimately, the goal of a radioactive stent is to deliver an appropriate dose and dose rate to prevent restenosis with an acceptable level of toxicity.

In addition to an in-depth dosimetric analysis, an intravascular brachytherapy treatment planning system should include an a priori evaluation of the radiobiological effect of the treatment. In IVBT, the dose values reported are those of physically absorbed dose. However, the real therapeutic effect depends on the radiobiology of the tissue being irradiated, and therefore other parameters should be considered, such as the initial dose rate, half-life of the radioisotope being used, repair capability of the tissue, and the type of tissue being irradiated (Dale, 1985, 1993; Brenner et al., 1996; Hall et al., 1988, 1999; Brenner and Hall, 2001; Mitchell, 2002). The concept of BED, or biological equivalent dose, takes these parameters into account for dose calculation. This comparison of physical absorbed dose and biological equivalent dose is very important, particularly when comparing the effectiveness of different radioisotopes and could be a significant factor in dose prescription. Thus far, there **is no** treatment planning system in intravascular brachytherapy specifically designed for radioactive stents that **takes into** account both the physical dose and the biologically equivalent dose.

1.1 Objectives and Specific Aims

The main objectives and specific aims of this work are as follows:

1. To develop a practical and fast algorithm based on the dose point kernel (DPK) convolution for calculation of the 3D dose distribution of radioactive stents. This

dosimetry model will be equipped to accommodate variations in stent geometry and dimension, radioisotope(s) and initial activity. Results will be shown as depth-dose profiles in the longitudinal and axial directions.

- 2. To develop an algorithm that calculates the 3D radiobiological effectiveness of the dose distribution of radioactive stents in the form of BED (Biological equivalent dose).
- 3. To apply the dosimetry and radiobiological models to stents with the following radioisotopes: ${}^{32}P$, ${}^{90}Y$, 188 Re and ${}^{177}Lu$, and combinations of these, to characterize the 3D dosimetry of each and demonstrate the potential for use in intravascular brachytherapy.
- 4. To validate the dosimetry model by comparing the results of the dose point kernel convolution with those of Monte Carlo simulations in a ^{32}P cylinder.

1.2 Significance of this Study

This is a development work in which we have implemented an algorithm based on the DPK convolution that calculates both the physical absorbed dose and BED of radioactive stents. This model offers the advantage of fast computation times and versatility in radioisotope selection, stent geometry and dimensions, and initial activity. It allows the evaluation of different radioisotopes, or combinations of such, for addressing the problem of edge restenosis and as well as to improve the overall dose distribution of a radioactive stent. The intention of this model is twofold. It serves as a dosimetry tool to calculate the three-dimensional dose distribution of the radiation emitted from a

radioactive stent, and functions as a radiotherapy treatment planning system by evaluating the radiobiological damage to the tissue. The model's final results can be used to aid in the development of industrial designs of a radioactive stents, the planning of animal and clinical trials, and possibly in the planning of an individualized intravascular brachytherapy treatment with a radioactive stent.

CHAPTER 2 BACKGROUND AND RATIONALE

2.1 Angioplasty and Stenting

Atherosclerotic artery disease is primarily caused by accumulation of fatty debris from the blood in the arteries resulting in a narrowing or stenosis of the arterial lumen, which consequently compromises blood flow (Nath et al., 1999). Many arteries are subject to this condition, particularly the coronary arteries, which supply blood to the heart muscle itself. Stenosis in the coronary arteries can cause much damage to heart as a result of poor oxygenation of tissues as well as by impairing the heart's electrical conduction system, which can potentially lead to a life-threatening arrhythmia, ischemia and myocardial infarction (Fox, 2002).

Figure 2-1. The heart and its arteries

The late 1950's witnessed the advent of bypass surgery for treatment of diseased coronary arteries. In 1977, the method of coronary artery bypass graft was introduced and consisted of taking a segment of a vein from the patient's leg and inserting the graft into the diseased artery (Fox, 2002). However, these two methods are very invasive since they

require a major operation that involves opening the patient's chest. In 1977, Andreas Gruentzig introduced PTCA, or percutaneous transluminal coronary angioplasty, which consists of the inflation of a balloon catheter in the narrowed area of the coronary artery, thereby restoring a stable lumen with a diameter similar to that of a normal artery (Figure 2-2) (Fox, 2002; King, 2002).

Figure 2-2. Overview of **PTCA procedure** (from: http://www.incirculationimagebank.net/03/).

PTCA, a less invasive and more affordable procedure, proved to be an attractive alternative to bypass surgery. When compared with coronary bypass graft surgery, patients treated with PTCA have lower initial costs and fewer complications. However, restenosis is angioplasty's major setback, limiting the long-term success of the procedure often requiring repeat revascularization in approximately 30% of patients within six months (Wurdeman et al., 1998). Of the more than 400,000 angioplasties performed

every year in the United States, 30 to 40% of them result in a post-angioplasty restenotic lesion (King, 2002).

The use of intracoronary stenting has revolutionized catheter-based revascularization of stenosed vessels. Stents are hollow mesh-wire tubes made of **stainless** steel or a nickel-titanium alloy that are delivered via the angioplasty balloon or can be self-expandable and serve as scaffolding to hold the artery open, thereby restoring blood flow (Fox, 2002). They account for approximately 70-90% of all interventional cardiology procedures and in conjunction with balloon angioplasty have been found to significantly reduce restenosis rates as shown by two major trials reported by Serruys et al. (1994) and Fischman et al. (1994). Stents are purely a mechanical means of preventing early recoil of the artery, thus creating a larger lumen than PTCA alone. They have proven effective in treating the first two components of restenosis: elastic recoil and remodeling. Nevertheless, stents have not been entirely successful in eliminating restenosis due to tissue proliferation, and have actually been found to stimulate neointimal hyperplasia in 20-30% of the cases within six to twelve months following angioplasty (Fox, 2002). In-stent restenosis remains problematic, particularly in smaller vessels and longer lesions (Janicki et al., 1999) with rates ranging anywhere from 31- *50%* as reported by several animal and clinical studies (Fischell et al., 2000).

As a result of these findings, the interventional cardiology community has come up with a variety of techniques in an attempt to treat or prevent restenosis. Such techniques include balloon angioplasty, additional stenting or supplementing repeat balloon angioplasty with excimer laser, high-speed rotational atherectomy and directional atherectomy (King, 2002). Numerous pharmacological agents with antiproliferative

properties have been tested for their potential to inhibit restenosis with mostly disappointing results (Serruys et al., 2001). Strategies consisting of local delivery of cytotoxic drugs and suicide gene therapy have been unsuccessful because of the risk of undesired toxicity to surrounding healthy tissues (King, 2002). These genetic and pharmacological interventions have great potential for success but mechanisms of drug delivery need much improvement before they can be considered the standard treatment modality. The use of ionizing radiation in intravascular brachytherapy has significantly reduced restenosis rates in several randomized trials (Sims, 2002; Kuntz, 2002), and is therefore considered to be one of the most promising methods for treatment and prevention of coronary artery restenosis.

2.2 Restenosis

Restenosis is clinically defined as a narrowing of the arterial lumen that is equal to or exceeds *50%* of the lumen diameter in adjacent normal segments of that artery (King, 2002). The expanding angioplasty balloon can apply pressures of up to 20 atmospheres for time periods ranging from seconds to minutes. Not only does this expand the lesion site narrowed by plaque, but it can also disrupt and injure the tunicae intima and media and even the tunica adventitia of the arterial wall (Figure 2-3). Restenosis is the result of the overstretch injury and exaggerated wound healing response of the damaged artery wall stimulated by the application of these pressures during the angioplasty procedure. It is a complex problem, although three main mechanisms are identified as leading causes (King, 2002).

Figure 2-3. Anatomy **of the artery wall**

The first and most immediate mechanism is elastic recoil, which occurs promptly after the initial overstretch of the artery and does not seem to progress much beyond the first few minutes after the deflation of the angioplasty balloon (King, 2002). It represents the slow return of the overstretched tissue from the expansion of the balloon to **its initial** diameter. On average, this has been quantified at 50% of the artery cross-sectional area or one-third of the lumen diameter (King, 2002).

A second mechanism of restenosis has been identified as that of wound contracture or remodeling, which occurs in subsequent weeks following PTCA. The entire artery can become contracted so that the external elastic lamina occupies a much smaller circumference compared to what it did immediately following the procedure (Fox, 2002; King, 2002; Wilcox, 2002). Remodeling **is in** essence the change in artery diameter following coronary angioplasty, which can either be favorable or unfavorable. A favorable remodeling occurs when the artery is partially enlargened to compensate for neointimal growth and plaque. On the contrary, an unfavorable or negative remodeling occurs when the artery contracts, that is, the diameter decreases in addition to lumen loss caused by neointimal thickening (Pasterkamp et al., 2000; Schwartz and Holmes, 2002).

It has been concluded through animal studies that the tunica adventitia plays an extremely important role in vascular remodeling, because it is this outermost layer that forms a ring of connective tissue around the artery (Rubin, 2002). The adventitia contributes to remodeling through a constriction of the external elastic lamina and an accumulation of myofibroblasts (Wilcox et al., 1996). Furthermore, several animal studies, as well as some clinical studies supported by intravascular ultrasound indicate that remodeling may be even more important than neointimal growth in the onset of restenosis (Mintz et al., 1996).

The third and last mechanism associated with restenosis is that of neointimal hyperplasia, which normally occurs within the first six months after stent implantation. Neointimal hyperplasia is characterized by an active and aggressive migration of smooth muscle cells from the outer layers of the artery and into the lumen, which can cause a very severe obstruction of the artery (Figure 2-4). It is mainly a proliferative response to overstretch balloon injury, which causes cracks and tears in the vessel wall (Fox, 2002; King, 2002). This injury leads to a series of immunological responses in which monocytes, macrophages and lymphocytes infiltrate the site of injury.

Figure 2-4. Restenosis as a result of neointimal hyperplasia

Platelet aggregation follows, stimulating the release of certain cytokines such as PDGF, or platelet derived growth factor, which ultimately results in the proliferation of smooth muscle cells and myofibroblasts (King, 2002; Sims, 2002). This cascade of events eventually ends in the migration of smooth muscle cells from their usual location in the media and/or adventitia, to the intima. In a matter of weeks to months, this healing site begins to resemble a fibrous plaque as neointimal growth extends in to the arterial lumen resulting in restenosis.

2.3 Radiation to Treat Restenosis

Intravascular brachytherapy has emerged as an attractive candidate therapy for the treatment of restenosis and is considered a promising anti-restenosis technology (Kuntz and Baim, 2000; Serruys and Kay, 2000; Fox, 2002). It involves treating restenosis with a radioactive source within the arterial lumen. "Brachy" which means "short" in Greek, implies that a highly localized radiation is applied within short distances in the artery. Preclinical and clinical studies have generally shown that applying ionizing radiation in the dose range of 15-30 Gy can significantly reduce the incidence of restenosis in patients who have undergone angioplasty (Nath et al., 1999). The effects of ionizing radiation have been thoroughly studied for the past half-century, and its effects on cell death can be attributed to DNA damage in the form of single-stranded or more importantly, doublestranded breaks that ultimately result in chromosomal aberrations that prove to be lethal when the cell attempts division (Mitchell, 2002). This concept has been successfully, adopted by radiation oncologists for the treatment of cancer, and has also been applied for the treatment of restenosis. The rationale for use of intravascular irradiation to inhibit

restenosis is based on the increased radiosensitivity of actively proliferating cells to the lethal effects of ionizing radiation. In intravascular brachytherapy, the most important mechanistic endpoint is the inability of a cell to clonogenically divide, thereby preventing restenosis due to neointimal hyperplasia and remodeling (Mitchell, 2002).

2.4 Radiation Delivery Systems

One method of irradiation is that of external beam treatments. Major advantages of this type of treatment include an extremely uniform dose delivery to the arterial wall as well as greater accuracy of dose calculations (Fox, 2002). However, because of the nature of this treatment, it is almost impossible to avoid radioactively treating significant portions of the heart and other tissues beyond it (Fox, 2002). This presents an undesired risk since it is well documented that irradiation can potentially induce coronary artery disease (Stewart et al., *1995).* Furthermore, it has been documented that a late effect of external beam irradiation, when applied at a high rate can result in a fibrosis, leading to severe carotid or coronary artery restenosis (Silverberg at al, 1978).

2.4.1 Catheter-based systems

Brachytherapy of an intravascular nature for treatment of arterial restenosis can be delivered via two main platforms: temporary catheter-based systems such as radioactive ribbons and balloons and permanent implants in the form of radioactive stents. In catheter-based systems, high-intensity radioactive sources in the form of thin wires, seeds, or ribbons attached to the end of a specially designed catheter are introduced to the site of angioplasty (Fox, 2002; Sims, 2002). For example, in a catheter-based y-

irradiation system, a ribbon containing sealed sources of 192 Iridium is inserted into a catheter. An angiogram confirms that the region is in place, covering in entirety the region of interest, and after 10-20 minutes, the ribbon is removed. The Cordis Checkmate IRT \Box is one such system involving 192 Ir that consists of an array of cylindrical-shaped seeds contained in a nylon ribbon (Jani et al., 2002). Alternatively, a catheter-based β irradiation system delivers β -emitting radioisotopes such as ³²P or ⁹⁰Y to the treatment area for a period of 2-5 minutes. The Guidant Galileo[™] Intravascular Radiotherapy System is an example of a catheter-based system that uses a ^{32}P wire as the radiation source (Raizner et al., 2002). Radioactive sources are also encased to form a "seed train" that can be delivered to the lesion site using a manual or hydraulic system. The Novoste Beta-CathTM is one another device that uses a hydraulic system to deliver the betaemitting $90\text{Sr}^{90}\text{Y}$ radioactive seeds to the lesion site using a closed catheter (Jani et al., 2002). The Cordis Checkmate™ system and Novoste's Beta-Cath™ system were FDA approved in 2000, and the Guidant Galileo™ system was approved in 2001. All of these systems are currently implemented in a clinical setting. These methods have given satisfactory results, although centering and geographic miss pose some problems.

Since the angioplasty procedure **entails** the insertion of saline-filled balloon into the artery, this suggests that a balloon filled with a radioactive liquid, such as liquid ¹⁸⁶Re can be a feasible method of delivering radiation to the artery wall. The main advantage of radioactive liquid-filled balloons, perhaps over all of the other types of brachytherapy, is that it can be applied to all arteries, regardless of size, degree of calcification and tortuosity (Fox, 2002; Weinberger and Knapp, 2002). In addition, a radioactive liquidfilled balloon guarantees adequate centering in the arterial lumen, feat that is not as easily

achieved in catheter-based systems. However, there are some concerns about balloon rupture, which could deliver high doses of radiation directly into the patient's blood stream. Because there is such risk in spillage of radioactive liquid, and the potential damage to the patient and lab personnel, as well as the logistics involved in filling, handling and disposing of radioactive solutions, radioactive liquid-filled balloons proves to be a cumbersome technique and not the method of choice of many clinicians (Fox, 2002).

The use of gas-filled balloons in intravascular brachytherapy has also been explored. This approach **is similar** to the liquid-filled balloon approach but with the radioactive material in a gaseous state. 133 Xenon has been investigated as a candidate for this type of radiation delivery system (Apple and Waksman, 2002). The balloon dilation catheter is filled with the radioactive gas and dilated at the lesion site so as to deliver the radiation dose to the arterial wall. The inert chemical property of 133 Xe offers enormous safety and limits radiation exposure to the medical staff. Other novel approaches being tested for intravascular brachytherapy include the catheter-based soft x-ray generator and the 99mTc injecting system (Fox, 2002; Apple and Waksman, 2002).

2.4.2 Radioactive Stents

Permanent implants in the form of radioactive stents have been a very attractive proposal since their foundation (Hehrlein and Kubler, 1997; Fischell, 1888;2000; Fox, 2002). A radioactive stent eliminates the need of a repeat revascularization, as the angioplasty/stenting procedure is done in concurrence with the irradiation procedure. Furthermore, procedures in the catheterization lab are identical to those already

performed, as radioactive stents are delivered via the femoral artery to the target site in the same manner as non-radioactive stents. Radioactive beta-emitting stents offer the advantage of uniform dosimetry, safety for laboratory personnel, as well as easy storage (Fox, 2002). Because it is a permanent implant, problems with adequate disposal of radioactive materials are eliminated. Adequate centering of the radioactive source, which is a problem with radioactive wires and seeds, is not an issue with radioactive stents because of the direct contact with the vessel circumference (Fischell, 1998; Fox, 2002). Artery wall irradiation after radioactive stent implantation is maintained in the days and weeks following the procedure, thereby covering the whole period of cell proliferation (Fischell, 1998; John et al., 2001).

The activity of a radioactive stent is up to 10,000 times lower than the activity levels of other sources used in catheter-based vascular brachytherapy (Hehrlein and Kubler, 1997; Fischell, 1998). Although stent activities are much lower, the total dose delivered by a permanently implanted stent is similar to that of a high activity radioactive seed. The main distinction between the two is the time sequence in which the radiation is delivered. For example, a 1μ Ci ³²P stent, which has a half-life of 14 days, delivers the same total dose as a 5-minute exposure of a 6 mCi ^{32}P wire (Amols, 1999). Furthermore, this low level of activities allows manual handling and eliminates the need for radioisotope licensing in the catheterization lab. Radioactive stents have an inherent dosimetry advantage in that the radioactive source is in intimate contact with the artery wall. However, the dose distribution is extremely nonuniform due to the gridded nature of stent geometry.

Radioactive stents combine both mechanical and therapeutical benefits for restenosis prevention. The stent itself, due to its scaffolding action, supports the artery wall, while radiation prevents growth of smooth muscle cells that can occlude the lumen of the artery. In addition, it has been observed that ionizing radiation, or vascular brachytherapy can have some favorable effects on treating the remodeling aspect of restenosis by inhibiting the proliferation of the adventitial myofibroblast (Rubin, 2002). When looking at the post-angioplasty artery as a wound undergoing a process analogous to wound contraction, the adventitial myofibroblast is believed to occupy a pivotal role in constricting the blood vessel, thus leading to a lumen narrowing in the case of a negative remodeling (Scott et al., 1996; Wilcox et al., 1996; Rubin, 2002). In consequence, it can be postulated that a radioactive stent, with proper usage, radioisotope selection and initial activity, can be extremely promising in treating the three mechanisms associated with restenosis: early recoil, due to the scaffolding action, and constrictive remodeling and neointimal hyperplasia as a consequence of ionizing radiation.

2.4.2.1 Preclinical Studies (Animal Studies)

Stent-based coronary brachytherapy to prevent restenosis has been studied in preclinical and clinical trials for almost a decade. Several animal models have given testimony to the fact that radioactive stents are capable of reducing or even eliminating neointimal hyperplasia. In 1994, Fischell et al. demonstrated that a ³²P impregnated stent wire with an activity of 0.006 µCi inhibited smooth muscle cell proliferation in cell culture. Hehrlein et al. (1995) were the first to describe the use of radioactive stents, which they implanted in non-diseased rabbit iliac arteries. The results of their study, using radioactive stents with low doses (max. 35 μ Ci) potently inhibited smooth muscle cell proliferation and neointimal hyperplasia in the rabbit model (Hehrlein et al., 1995). They found that neointima formation was markedly reduced by the implantation of the radioactive stent in a dose-dependent fashion. In the control model, in which a nonradioactive stent was implanted, there was a $30 \pm 2\%$ neointimal growth compared to the $0.5 \pm 0.1\%$ seen in the model with the radioactive stent implanted (Hehrlein et al., 1995).

Carter and Laird (1996) reported studies in swine models using low-dose irradiation from β -particle emitting stents. The initial stent activity was 0.14 µCi and the dose delivered to the arterial wall was approximately 3 Gy over the 4-week study period. A 37% reduction in neointimal area and a 32% reduction in stenosis for β -particle emitting stents were observed when compared to non-radioactive control stents at the end of the 4-week period. In another set of experiments by Carter et al. (1999), the effects of beta particle-emitting stents with activities ranging from $3-23$ μ Ci were evaluated in normal pig coronary arteries. At 28 days from implantation a reduction in the medial and neointimal cell density was observed which inversely correlated with the radiation dose. Neointimal proliferation was also lower for the radioactive stent as compared to the nonradioactive stent. Several other studies have been performed in different animal models such as the porcine and canine model with overall inconsistent results (Carter and Fischell, 1998; Virmani et al., 2002).

2.4.2.2 Clinical Studies

Animal models can only partially mimic the restenosis process after angioplasty and stent implantation in humans. In 1996, IRIS (Isostent for Restenosis Intervention Study) was the first study done on patients using the $32P$ Palmaz-Schatz stent. In this study, 30 patients underwent successful stent placement with an activity between *0.5* and 1.5 pCi (Hehrlein and Fischell, 2002). Thirty days post-treatment, there were no adverse effects, and in fact a considerable expansion of the arterial lumen was observed. However, a 6-month follow-up revealed a 31% restenosis rate.

The Milan dose-response study (Albeiro et al., 2000a), which ran from October 1997 to October 1998, explored the short and intermediate term results of the ^{32}P stent in patients with coronary artery disease. Eighty-one patients were used for this study, and they were divided into three groups depending on the activity level of the stent: 0.75-3.0 μ Ci (Group 1), 3.0-6.0 μ Ci (Group 2) and 6.0-12.0 μ Ci (Group 3). This trial demonstrated the safety of using these ranges of activity levels. The low incidence of late thrombosis was also observed. After a 6-month follow-up, no deaths occurred and intra-stent neointimal hyperplasia was reduced in a dose-related manner, but intra-lesion restenosis rates were 52% in Group 1, 41% in Group 2 and *50%* in Group 3 (Albeiro et al., 2000a). This trial generated disappointing results in that they identified the "candy wrapper effect", or edge restenosis. It is hypothesized that decreased radiation doses at the edges of the stent might actually stimulate restenosis, while higher doses in the middle inhibit it (van der Giessen et al., 2001). Barotrauma at stent edges during the implantation procedure might be a factor as well. A second phase of this trial was initiated using higher activity levels, but yielded similar results despite less aggressive implantation techniques using pressures of only 8 to 10 atm (Albeiro et al., 2000b).

Other European clinical studies involving the implantation of ³²P radioactive stents were conducted in Vienna and Rotterdam. The Rotterdam group did not observe cases of edge restenosis in ^{32}P radioactive stents of 0.75 to 1.5 µCi (Hehrlein and Fischell, 2002). However, they reported a restenosis rate of 43% occurring only at stent edges in an additional group of lesions treated by $32P$ radioactive stents of higher activities in the range of 6 to 12 μ Ci. The Vienna group treated 36 patients with ³²P radioactive stents with activities between 5.36 and 20.77 μ Ci. At a 6-month follow-up, 15 patients showed signs of edge restenosis (Hehrlein and Fischell, 2002).

2.4.2.3 **Edge Restenosis or "Candy Wrapper Effect"**

Despite much hope and hype and an overall initial **success in** reduction of neointimal **area, some** clinical and animal trials have exposed several pitfalls pertaining to the radioactive stent. Most notably, the biggest hindrance of radioactive stents, is that of edge restenosis, nicknamed the "candy-wrapper effect" because of its appearance in an angiogram. Figure *2-5* shows the tight stenosis at the stent edges, which are marked by arrows. This phenomenon has generally been observed 2 to 3 mm distal and proximal to the stent (Albeiro et al., 2000b; Serruys and Kay, 2000; van der Giessen et al., 2001). This is believed to be the cause of inadequate dose delivery to target tissues after they have been injured by the angioplasty procedure. The angioplasty balloon that delivers the radioactive stent is usually longer than the stent itself resulting in barotrauma at the edges to which there is no radiation and therefore no inhibition of cellular growth. Experimental results from animal models have also suggested that low doses of radiation might in fact

Figure 2-5. Edge restenosis as seen in an angiogram *(Circulation,* Albeiro et **al., 2000b).**

stimulate neointimal growth (Amols, 1999a). Another cause is geographic **miss,** or failure of completely irradiating the target lesion. Finally, edge restenosis can also be a result of the basic physics of a radioactive stent. At the ends of the stent, the dose will be very close to 50% of its value at the center due to steep dose gradients, especially in the case of beta emitters (Amols, 1999a; Serruys and Kay, 2000; van der Giessen et al., 2001).

2.5 Beta and Gamma Emitters: Radioisotopes in Intravascular Brachytherapy

The radioisotopes used in intravascular brachytherapy include both beta and gamma emitters. Radiation energy from beta emitters in characterized by high energy with low tissue penetration applied in short time periods, which diminishes rapidly with distance from the source. Nevertheless, beta particle emitters in contact with the arterial wall are for the most part sufficient to induce cell death of superficial smooth muscle cell layers in the intima and media, but not the adventitia (Amols, 1999a; Fox, 2002).
However, the limited depth of penetration of beta emitters poses the advantage of **minimal exposure to adjacent tissues.**

Yet, despite the fact that beta emitters are practical for use in the catheterization **lab because of minimal exposure to remote tissues and staff, they can also present some** challenges for effective delivery of radiation to the arterial wall in the case of large, highly calcified vessels, and the rapid fall-off dose may also be problematic. Gamma emitters, on the other hand, offer the advantage of higher tissue penetration, making it ideal for larger vessels with highly calcified plaques. Gamma radiation generally has lower energy, but it must be applied for larger periods of time for maximum effectiveness (Amols, 1999a; Fox, 2002). As a result, much more precaution must be taken to shield the surrounding tissues as well as the catheterization lab personnel. Beta emitters also yield higher doses per emission than y-sources and hence the specific activities required for β -emitters are at least an order of magnitude lower than for γ -sources. It is also important to note that the dose rate for a given activity of a beta-emitter is much greater than for a similar activity of a gamma-emitter (Fox, 2002). Therefore, a beta-emitting source has a much lower activity than a gamma-emitting source. Table 2-1 shows the properties of some the radioisotopes used in IVBT.

In IVBT, particularly in the case of a permanent implant such as a radioactive stent, the selection of the radioisotope in critical. A compromise between an acceptable half-life, energy, range and ease of production is necessary for a cost-efficient and feasible production of a radioactive stent. The ideal radioactive source would be one that displays a low energy gamma emission, which would provide a uniform dose distribution

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Isotope	Emission Type	Half-life	Average Energy (MeV)	Maximum Energy (MeV)
$32\overline{p}$	Pure β -	14.28 d	0.695	1.709
$\overline{^{90}Y}$	β-	2.67d	0.761	2.282
192 Ir	β - γ	73.8 d	0.224 NA	0.672 0.317
133 Xe	β -	5.243 d	0.115 NA	0.346 0.081
188 _{Re}	β-	16.94 _{hr}	0.706 NA	2.118 0.155
$\overline{^{103}}Pd$	x-rays	16.99 d	NA	0.040
125 I	x-rays	59.4 d	NA	0.046
48 _V	$\beta +$ ν	16d	0.230 NA	0.690 0.511
^{131}Cs	\boldsymbol{v}	9.7 _d	NA	0.030

Table 2-1. Properties of Radioisotopes used in IVBT (animal trials, clinical trials, theoretical calculations).

and negligible doses to surrounding healthy tissues. Similar dose characteristics could also be obtained with a very high-energy beta emitter (Amols, 1999a). Unfortunately, these ideal sources do not exist as most commonly available gamma sources are of too high an energy, or too low an activity and dose rate. Similarly, most beta sources have too low an energy, too short a half-life, or excessive amounts of gamma contamination (Amols, 1999a). In addition, half-life considerations are extremely important. It should be long enough to cover the period of cell proliferation after the onset of restenosis, yet short enough to not cause any detrimental effects as a result of permanently remaining in the body.

2.6 Treatment Planning for Radioactive Stents

Unlike the situation with a cancerous tumor, where one is interested in killing all cells, intravascular brachytherapy does not set out to completely liquidate all the cells responsible for causing artery obstruction. Doing so would mean application of higher than wanted doses which would inevitable lead to injury to unwanted **tissues,** leading to a number of complications and adverse effects. Instead, the inherent focus of this treatment is to limit the proliferative potential of a group of clonogenic cells, whether they are smooth muscle cells from the media or adventitia (Brenner et al., 1996; Hall et al., 1999). Because smooth muscle cells are somatic cells, they have limited cycles of cell division before they naturally senesce, and therefore do not require the high doses of radiation that cancerous cells for effective treatment of the problem. It is only necessary to inhibit the proliferation of a considerable portion of the cell population so that the surviving fraction of cells runs out of reproductive ability before sufficient progeny are produced to cause restenosis (Brenner et al., 1996; Hall et al., 1999). This presents the notion of a therapeutic window, a rather narrow margin between the radiation dose required to effectively treat the problem, and that which can result in undesirable damage to the artery wall. A very low dose, such as 10-12 Gy might not be enough to effectively treat the problem, or might delay rather than prevent restenosis, while a dose near 30 Gy might be excessive, causing unwanted injury to healthy tissues. It is believed that a dose somewhere between *15-30* Gy might be enough to inhibit restenosis (Hall et al., 1999; Nath et al., 1999).

The use of a treatment planning system in vascular brachytherapy can provide the interventional cardiologist with a tool for prescribing, evaluating and reporting the dose

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given to a patient, or an animal in the case of animal studies. It can provide a means of documenting the treatment given, as well as further customizing the radiation dose for an individualized treatment plan (Fox, 1999). It may also allow an opportunity to retrospectively evaluate the influence of dose on the success or side effects of the treatment. Most importantly, this type of treatment planning process can be used to evaluate different treatments and give an a priori evaluation of the overall effectiveness of the treatment.

2.7 Radiobiological Considerations

Designing an optimal treatment for brachytherapy, or any other radiotherapy treatment for that matter, requires a detailed knowledge of radiobiological principles. Although animal studies and clinical trials provide the physical evidence of the efficacy of a treatment, a radiobiological perspective is necessary in understanding the fundamentals of any in vivo study. Radiation can inhibit restenosis because it kills cells by direct ionization of DNA, or by the creation of free radicals, which can interact with nearby water molecules (John et al., 2001; Mitchell et al., 2002). The free radicals and reactive ions produced by indirect effects, or radiation deposited directly can alter the purines and pyrimidines (bases) in DNA resulting in point mutations and cross-links, as well as disruption in the integrity of the phosphodiester bonds which join the nucleotides in each of the DNA strands. This can lead to double or single-stranded breaks in the DNA, translocations, and chromosome aberrations in the form of dicentrics and rings, all of which can potentially limit the proliferative potential of the cell eventually resulting in cell death (Hall, 1988, 1999; Mitchell, 2002). These **ideas are** illustrated in figures 2-6 and 2-7.

Figure 2-6. Mechanism of Damage of Ionizing Radiation

Figure 2-7. Direct and Indirect Action of Ionizing Radiation

Several studies in the 1960's demonstrated that certain portions of the cell cycle are more susceptible to radiation effects, with cells in G2 and mitosis stages of the cell cycle being the most radiosensitive (Hall et al., 1988). Damage occurs more readily when the chromosomes are greatly condensed in mitosis. This is important because tumor and cancer cells, or rapidly proliferating cells, undergo division more often than normal cells, and are therefore more susceptible to damage. This property is the foundation of why radiation is so successful in treating cancer. Therefore, rapidly dividing cells in the neointima are more susceptible to radiation and the reason why this treatment is successful in treating restenosis (Mitchell, 2002).

Biological tissues have the ability to accumulate and repair radiation damage. In terms of modeling the lesions to DNA, there are two main kinds of lesions: repairable lesions and irreversible lesions (Mitchell, 2002). A repairable lesion has a good probability of being repaired or misrepaired by at least one biochemical repair pathway. An irreversible lesion, on the other hand, has no chance of being repaired, and can be further sub-divided into fatal lesions and non-lethal mutations, which eventually result in cell death (Mitchell, 2002). Low levels of radiation can lead to a delay in cell division from which the cell can gradually recover and continue to divide as normal. Higher doses of radiation may inhibit cell division permanently, thus not allowing the cells to divide, ultimately resulting in death.

Central to the subject of radiobiology is the linear quadratic formalism or LQ model. Dose-response or dose-survival curves for all dose rates are fit to the linear quadratic formalism, which expresses the surviving fraction (S) of cells as a function of dose (D) (Dale, 1985; Brenner, 1997).

$$
S=e^{-aD-\beta D^2}
$$

The constants α and β are characteristic of a given cell type and have units of Gy⁻¹ and $Gv²$ respectively. The ratio α/B is expressed in units of Gy and is of great clinical significance because it is a measure of the fractionated sensitivity of a tissue (Dale, 1993). They are also measures of two separate cell kill processes, type A and B (Giap et al., 1999). Type A damage is characterized by two critical targets (DNA strands) being hit simultaneously by a single radiation event resulting in cell death without repair (Dale, 1985; Giap et al., 1999). Type A damage is directly related to the linear component of the linear-quadratic formalism, αd . In Type B, which is related to βd^2 damage, each of the two critical sites is hit in sequence by different radiation events. When one of the two critical **sites is** damaged, there is evidence of sublethal damage. Over a period of time, the cell can either be killed by accumulation of sublethal damage or it can be repaired. At lower doses, the linear component dominates, while at higher doses, the quadratic term starts to dominate (Mitchell, 2002).

The LQ model is used for the radiobiological assessment of a particular radiation therapy. The biological effects of radiation are dependent on the parameters BED and RE which are derived from the LQ model, as they take into account α and B constants of a given tissue, or more importantly the ratio of the two. Biological equivalent dose (or (BED) (also referred to as ERD or extrapolated response dose), is a single-figure measure of the biological effectiveness of a dosing strategy on a particular tissue (Dale 1985, 1993). The concept of BED is key in comparing different treatments, but the calculation of BED is made through another fundamental LQ parameter RE, or relative effectiveness per unit dose (Dale, 1985, 1993; Giap et al., 1999). RE is dimensionless and measures the effectiveness of a given unit of dose to cause a specific biological endpoint taking into account the dose rate and repair capability of the tissue (Giap et al., 1999). BED (units are Gy) is equal to the product of RE and the absorbed dose at a specific point (Dale, *1985,* 1993; Giap et al., 1999). Thus far, there **is no** radiobiological assessment of radioactive stents employing the concepts of BED and RE for comparing dosing strategies with different radioisotopes.

2.8 Target Tissue

There is yet to be a consensus as to what is the target tissue to be irradiated. In addition, a tolerance dose for the artery wall has not been established. It had long been speculated that neointimal growth was the result of proliferation of smooth muscle cells in the tunica media (Hehrlein et al., 1995). This fact has been challenged by the tunica adventitia as the source of proliferating cells, in particular the adventitial myofibroblast, which migrates through the media in to the arterial lumen (Scott et al., 1996; Wilcox et al., 1996, 2002). The different layers of the artery wall and their respective thicknesses are shown in Figure 2-8.

Figure 2-8. Thickness of artery wall layers

If this in fact is the case, then an effective radiation dose needs to be delivered at least 1 to 2 mm from the artery lumen (Fox, 2002). Further studies on the cellular cascade of events associated with restenosis present two main hypotheses. One points to the myofibroblast as being the target cell of interest, while the other focuses on the monocyte-derived macrophage (Rubin et al., 2002). The monocyte-derived macrophage has received much attention because it is a key manufacturer of a number of cytokines

and growth factors responsible for the activation and promotion of myofibroblast or smooth muscle cell proliferation (Rubin et al., 1998, 2002). At present, most investigators believe the target lies in the adventitia with either macrophages or myofibroblasts being the cells responsible for triggering the complex biological processes that ultimately lead to medial smooth muscle cell proliferation and restenosis (Wilcox et al., 1996, 2002; Fox, 2002; Rubin et al., 2002). This further illustrates the overall complexity of the restenosis problem, and the interplay of factors that must come together in order to render any antirestenosis treatment successful. In general, for restenosis modeling purposes, it is correct to deliver radiation up to 3 mm from the arterial lumen (Fox, 2002).

2.9 Dosimetry of Radioactive Stents

Dosimetric considerations in all forms of brachytherapy are extremely important. In the case of the radioactive stent, the matter is further complicated by the fact that the **stent is a** permanently implanted device. This implies a careful consideration of a particular radioisotope's half-life, as well as the initial activity of the stent. The dosimetric characterization of the radiation field in both longitudinal and radial directions, of any brachytherapy source is generally the first step prior to animal and clinical studies that might pave the way for future commercialization of a particular treatment.

In conventional brachytherapy dose delivered to a target point is specified at a distance of 1 cm from the source (Nath et al., 1999). Consequently a large volume of experimental and theoretical dosimetry data is available for source to target distances of 1 cm or more and the dosimetry of both gamma and beta emitters is well established at

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these distances. In vascular brachytherapy, the distribution of radiation dose around the source is difficult to measure because the sources have very small diameters and the dose distribution must be determined very close to the source, usually less than 5mm (Amols, 1999a; Fox, 1999; Nath et al., 1999). At such small distances the dosimetry is complicated due to the steep dose gradients, low energy secondary radiations and selfshielding effects of the radioactive source. The uncertainties in dosimetry are larger for low-energy beta-emitting sources than for gamma sources due to the limited range of the beta particles and the effect of heterogeneities in the medium (Amols, 1999a; Nath et al., 1999).

The near-filed dose distributions of a radioactive stent can be determined by both theoretical and experimental methods. Theoretical calculations are obtained from dose point kernel calculations and from Monte Carlo simulations. The dose point kernel (DPK) describes the distribution of absorbed dose around point-isotropic sources of electrons and beta particles in water (Berger, 1971; Prestwich, 1989; Cross, 1992). In 1995, Prestwich et al. presented the results of a dose distribution study around a uniform cylinder of $32P$ using the DPK method. In this model, the source of radioactivity was uniformly distributed on the two-dimensional surface of a cylinder. The stent can be modeled as a cylinder, although it in fact possesses a quite intricate and detailed geometry. The gridded architecture of a stent is an important consideration of these dosimetric determinations as the radiation is expected to be higher in the proximity of the struts than in the spaces in between struts. To overcome this problem and to elaborate on the model presented by Prestwich et al. (1995), another group of researchers calculated the near-field dose of a ^{32}P stent using the dose point kernel and a cylindrical coordinate system to simulate the wire mesh geometry of the stent (Janicki et al., 1997). The dose distribution generated by the beta particles emitted from the radioisotope was computed at distances ranging from 0.1 to 3 mm from the stent surface. A later study by Janicki et al. (1999) introduced a new model taking into account the scaling property of the dose point kernel function which can be extended to a heterogeneous medium with a density different to that of water. Because in a realistic artery the presence of plaque can allow attenuation of the beta particles, incorporating plaque density in a theoretical model has some advantageous effects.

Monte Carlo simulations have also been used by several groups of investigators to study the dose distributions surrounding radioactive stents (Li et al., 1998; Reynaert et al., 1999, 2001; Fox and Henson, 2000). Monte Carlo codes, in particular the MCNP (Monte Carlo N-Particle Transport Code System), is a multimaterial, 3-D, arbitrary geometry, electron/photon/neutron transport code developed at the Los Alamos National Laboratory (Li et al., 1998). This type of theoretical simulation can be achieved by specifying the specific energy assumed for the radioactive source, the architecture assumed for the stent, the specific depth assumed for the radioactive species in the stent material, and the inclusion or exclusion of nonhomogenous material or the absorption or scatter of stent struts (Coffey and Duggan, 2002).

2.10 AAPM Task Group 60 Report

The AAPM (American Association of Physicists in Medicine) Task Group 60 has developed **a series** of recommendations based on several early dosimetric studies for dose specifications in intravascular brachytherapy (Nath et al., 1999). It is recommended to

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use 0.5 mm from the surface of the stent as the reference depth and a total treatment time of 28 days as a common framework for the dose field characterization (Nath et al., 1999). A time period of 28 days is reasonable because it falls within the time frame of restenosis onset, namely the first few weeks to two months after stent placement. The desired objective of the dosimetric characterization of a radioactive stent is to determine the 3D dose rate per unit of activity around the stent deployed in an angioplasty balloon catheter, as well as to demonstrate the circumferential and longitudinal uniformity of the dose delivered by the stent (Nath et al., 1999). Furthermore, the AAPM recommends that the three-dimensional dose distributions around the stent be carefully determined before animal and clinical trials. The model described in this study will take into account the recommendations of the AAPM, although it will be equipped to calculate the dose at a depth less than 0.5 mm and times greater or less than 28 days. The results presented in this work are for lifetime doses delivered by the stent under the assumption that the stent is a permanent implant unless otherwise stated.

CHAPTER 3 MATERIALS AND METHODS

This chapter provides an outline of the development of the radiation dosimetry and radiobiological model that was used to calculate the 3D dose distribution and radiobiological effectiveness of a radioactive stent. It includes the set of equations that govern the dose point kernel convolution method, the calculation of beta and gamma dose point kernels, and the equations of the radiobiological model. In addition, information on the stent geometry that was used, as well as details on the algorithm using the dose point kernel convolution and the different dose calculations that were done is also provided in this section. This chapter closes with the validation of results obtained from the dose point kernel convolution method to those obtained through Monte Carlo simulations.

3.1 Radiation Dosimetry Model

The radiation dosimetry model is based on the dose point kernel (DPK) convolution. Several investigators have used the dose point kernel for calculation of 3D dose distribution of radioactive stents (Prestwich, et al.., 1995; Duggan et al., 1998; Janicki et al., 1997, 1999). The dosimetry model is equipped for the calculation of absorbed dose at any specified point defined radially and longitudinally from the surface of the stent. In addition to its versatility regarding stent geometries, it will also allow dose calculation of stents with different radioisotopes or combinations of radioisotopes and initial activities. In this project, the dosimetry of a 177 Lu, 32 P, 188 Re, and 90 Y stent was determined as well as that of combinations of these radioisotopes.

3.1.1 Dose Point Kernel Convolution

Absorbed dose due to any source can be computed by splitting the extended source into a series of point sources and summing up the individual contributions (Berger, 1971; Janicki et al., 1997). In other words, the dose at each target point in tissue will receive contributions from each point source. The initial activity is distributed uniformly over the surface or the volume of the stent. The stent is the source of radiation, and is represented as a series of point sources defined by a set of coordinates (x,y,z) . This operation can be expressed in the form of a convolution integral as follows:

$$
D(r,t) = \frac{A_0}{S\lambda} \times [1 - \exp(-\lambda t)] \times \int DPK\big(r - r'\big| \big| \, ds\,,\qquad(1)
$$

where $D(r, t)$ is the absorbed dose at a particular point in space and time, A_0 is initial activity of the stent uniformly distributed over the stent surface, λ (h⁻¹) is the decay constant of the radionuclide, S is the total active stent surface area $\text{(mm}^2)$, t (h) is the time interval after stenting, *DPK* is the dose point kernel function for a specific radioisotope, and $r - r'$ is the distance (mm) between a source point on the stent and the source point where the dose is calculated. The dose rate at any time **t** after stent implantation is dependent on the initial activity and is given by:

$$
\dot{D}(t) = \dot{D}_0 e^{-\lambda t}, \qquad (2)
$$

where \dot{D}_0 is the initial dose rate given by:

$$
\dot{D}_0(r) = \frac{A_0}{S} \times \int DPK\left(r - r\right)ds\tag{3}
$$

The total absorbed dose at time t is obtained by integrating equation (2).

$$
D(t) = \int_0^t \dot{D}_0 e^{-\lambda t} dt
$$
\n
$$
= \dot{D}_0 \times \left(\frac{1 - e^{-\lambda t}}{\lambda}\right)
$$
\n(4)

In the case of a radioactive stent that is inserted permanently in the patient, time **t** is equal to infinity. The dose values obtained are the lifetime doses of the stent until complete decay of the radioisotope. Therefore, Equation (4) is converted to:

$$
D(r) = \frac{D_0}{\lambda} \tag{5}
$$

The source points, which are the coordinates defining the stent itself, are designated **as r' (mm)=** (x', y', z'). The target points, or the points defining the artery wall depth at which the dose is calculated, are defined by the coordinates r (mm)= (x, y, z) z). The z coordinates will cover the length of the stent and extend *5* mm beyond the edges of the stent at intervals of 0.125 mm. The x and y coordinates extend radially from the surface of the stent up to a distance of *5* mm from the surface of the stent. Absorbed dose was calculated at intervals of 0.125 mm. Dose values were calculated starting from 0.1 mm from the stent surface. The distance between source and target points is given by the following equation:

$$
r = \sqrt{(x - x')^{2} + (y - y')^{2} + (z - z')^{2}}
$$
 (6)

3.1.2 Dose Point Kernel

The dose point kernel (DPK) describes the spherically symmetrical dose-rate distribution of absorbed dose around an isotropic source obtained from Monte Carlo simulations of beta or gamma radiations in water (Berger 1971; Giap et al., 1995). The DPK function $K(r, r')$ defines the dose rate at a point r delivered by a point source of unit cumulative activity located at r'. For radioisotopes having beta and gamma emissions, the dose point kernel consists of two components. Therefore the absorbed dose in the target tissue has contributions from both beta and gamma radiations. This additive expression for DPK is as follows:

$$
DPK(r,r') = DPKr(r,r') + DPK\beta(r,r'),
$$
\n(7)

where $DPK_r(r,r')$ and $DPK_\beta(r,r')$ are the DPK for the gamma and beta radiations respectively. The following table illustrates the main properties of the radioisotopes used in the radiation dosimetry model. Of note is the fact that ^{32}P and ^{90}Y emit pure beta energy, therefore the dose delivered is given only by the convolution over the target points of the beta DPK. On the other hand, even though ¹⁷⁷Lu and ¹⁸⁸Re decay by mainly by beta emissions, they also emit a low energy gamma radiation and also have a gamma DPK component contributing to the total dose.

Radio- isotope	Half-life	Energy	Maximum Energy (MeV)	Average Energy (MeV)
177 Lu	6.734d	beta gamma	0.498 $0.113(6.4\%)$ $0.208(11.0\%)$	0.134 NA
188 Re	16.94 h	beta gamma	2.118 0.155(14.9%) 0.633(1.25%) $0.478(1.01\%)$	0.706 NA
32 _P	14.28 d	beta	1.709	0.695
$\boldsymbol{^{90} \mathbf{V}}$	2.67d	beta	2.282	0.761

Table **3-1. Properties of** Radioisotopes **used in Radiation Dosimetry Model**

3.1.3 Beta Dose Point Kernel

Beta DPK of ^{32}P , 188 Re and ^{90}Y were obtained from the tabulated data of Cross et al. (1992). These data were derived from Monte Carlo radiation transport simulations of beta particles in water considering the beta spectrum of different radioisotopes. The DPK represents the dose expressed in units of $\frac{nGy \cdot cm}{Bq \cdot hr}$ as a function of a distance r (mm) emitted by an isotropic point source. Figures 3-1 through 3-3 show the DPK curves of $32P$, $90Y$ and 188 Re. For these radioisotopes, DPK data was fit to an analytical expression shown in Table 3-2.

Figure 3-1. Beta DPK of ³²P

Figure 3-2. Beta DPK of ⁹⁰Y

Figure 3-3. Beta DPK of ¹⁸⁸Re

Radio- isotope	Equation	Value
32 _P	$DPK(r)B = -0.016r5 + 0.0866r4 + 1.8508r3 - 15.253r2 + 9.889r + 104.6$	0.9999
\mathbf{P}^{\bullet}	$DPK(r)B = -0.0058r5 + 0.1258r4 - 0.655r3 - 0.7175r2 - 5.0275r + 101$	0.9997
188 Re	$DPK(r)_{\beta} = 0.013 \text{ l}r^6 - 0.373 \text{ l}r^5 + 4.046 \text{ l}r^4 - 20.505r^3 + 48.83 \text{ l}r^2 - 66.652r + 126.89$	0.9964

Table 3-2. Functions and r 2 values for DPK **expressions of selected radioisotopes**

3.1.4 Dose Point Kernel of Beta Emissions of '"Lu

Since 177 Lu is just recently being used in radiotherapy applications, there is not readily available DPK data in published literature for this radioisotope. The DPK of the beta emissions of 177 Lu was calculated by using the following equation:

$$
DPK(r,r')_{\beta} = \sum_{i=1}^{N} \frac{n_i(E_i)}{N} \times DPK(r,r',E_i),
$$
 (8)

where $DPK(r,r)_{\beta}$ is the DPK of the beta radiations, $DPK(r,r'E_i)$ is the DPK of the beta particles of energy *E;, N* is the total number of electrons emitted by disintegration of the radioisotope, and $n_i(E_i)$ is the number of particles of a particular energy E_i . This method makes use of DPK values for beta particles of specific energies over the energy spectrum of ¹⁷⁷Lu. The DPK of beta particles $K(r, rE)$ at different energies E has been calculated by Cross et al. (1992) by means of Monte Carlo radiation transport simulations. The calculation of the spectrum of beta emissions of 177 Lu $(n_1(E_i)/N)$ is shown in Appendix A. The calculations for determining the DPK of beta **emissions** of 1"Lu are provided in

Appendix B. Figure 3-4 shows the beta DPK for 177 Lu. Because the fitting of the DPK data of ¹⁷⁷Lu to an analytical expression yielded a high error percentage, beta DPKs were stored as a lookup table in the dosimetry model. The beta DPK for each distance *r* was calculated by logarithmic linear interpolation according to the following equation:

$$
DPK(r) = \frac{\ln(DPK_1) - [\ln(DPK_1) - \ln(DPK_2)] \times (dis_1 - r)}{dis_1 - dis_2} \quad , \tag{9}
$$

where r is the distance of interest between the source and target points, DPK_1 and DPK_2 are the DPK values for distances *dis₁* and *dis₂* respectively, and *DPK (r)* is the dose point kernel for the distance of interest.

Figure 3-4. Beta DPK of '"Lu

3.1.5 Dose Point Kernel of Gamma emissions of '"Lu and 188Re

Dose point kernel data of gamma emissions are not readily available in a tabulated fashion like the beta DPK provided by Cross et al. (1992). Both ¹⁷⁷Lu and ¹⁸⁸Re emit non-negligible gamma radiation that must be accounted for in the total dose calculation.

Isotope	Half-life	Energy	Maximum Energy (MeV)	Average Energy (MeV)
177 Lu	6.734d	beta gamma	0.498 $0.208(11.0\%)$ $0.113(6.4\%)$	0.134 NA
188 Re	16.94h	beta gamma	2.118 0.155(14.9%) 0.633(1.25%) 0.478(1.01%)	0.706 NA

Table 3-3. Beta and Gamma energies of 1"Lu and 1 88Re

Furhang et al. (1996) have derived a mathematical expression of photon or gamma dose kernels for several radioisotopes, including 188 Re. The gamma DPK was generated in water for the full gamma **emission** spectrum of the radioisotope through Monte Carlo simulations (Furhang et al., 1996). For the calculation of gamma DPK of ¹⁸⁸Re, the following expression was used:

$$
DPK(r)_{\gamma} = \sum_{i}^{i_{MAX}} \left(\frac{a_{-2}}{r^2} + \frac{a_{-1}}{r} + a_0 + a_1 \cdot r + a_2 \cdot r^2 \right)_i e^{-m_i \cdot r}, \quad (10),
$$

where r is the distance in cm, and DPK is expressed in $cGy/Bq \cdot s$. The coefficients for this expression are listed in Table 3-4.

a_{2}	2.162×10^{-12} 1.444×10^{-12}
a ₋₁	3.455×10^{-13}
ao	1.298×10^{-14}
m	0.10739 5.197
Error _{max}	1.2%

Table 3-4. Coefficients of the gamma DPK expression of '"8 Re

Table 3-5. Functions and r 2 values for gamma DPK expressions of ' 88Re and 17 Lu Radio- **Equation** \mathbf{r}^2 **isotope 188**Re $K(r)_r = \left[\frac{2.162 \times 10^{-12}}{r^2} + \frac{3.455 \times 10^{-13}}{r} + 1.298 \times 10^{-14} \right] e^{-0.10739r} + \left[\frac{1.444 \times 10^{-12}}{r^2} \right] e^{-5.197}$ N/A $K(r)$ _r = 0.0071r⁶ -0.0447r⁵ +0.1107r⁴ -0.1377r³ +0.0896r² -0.0253r +0.0428 0.9669 177 **Lu**

¹⁷⁷Lu is a relatively new in radiotherapy treatment and as a result was not among the radioisotopes whose gamma DPK was calculated by Furhang et al., (1996). In this case the gamma DPK of 177 Lu was calculated by using the gamma energy tables of Luxton and Jozsef (1999), which were calculated through EGS4 Monte Carlo simulations for gamma rays with energies between 10 keV and 2 MeV. These tables present the dose of a gamma ray of particular energy emitted in water as a function of distance radial distance from a point source (Luxton and Jozsef, 1999). The dose for gamma emissions of 0.208 MeV(11%) and 0.113 MeV (6.4%) was calculated using these tables and a logarithmic linear interpolation according to equation 9. Dose is expressed in nGy $cm²/Bq-hr$. The total dose from the two gamma energies is obtained by multiplying each dose value at a specific energy by its fraction and summing up the two components according to the following equation:

$$
DPK(r)_{r} = (f_1 \times DPK_1(r)) + (f_2 \times DPK_2(r))
$$
\n(11)

Refer to Appendix C for results of the interpolation calculation and the final gamma DPK data. This data was plotted and fit to the analytical expression shown in table 3-5. Figure 3-5 shows the curve of the gamma DPK of ¹⁷⁷Lu.

Figure 3-5. Gamma DPK of **¹⁷ Lu**

3.2 Radiobiological Model

The main purpose of the radiobiological model is to compare the physical absorbed dose at a specific target point with the biological equivalent dose at that same point taking into account the initial dose rate, repair probability of the tissue, duration of treatment and biological endpoint $(\alpha/\beta \text{ ratio})$ (Dale, 1985, 1993; Giap et al., 1999). The radiobiological impact of the different radioisotope distribution **is assessed** by means of two main parameters, BED and RE. Biological equivalence dose (BED) and relative effectiveness per unit dose (RE) are the focus of the radiobiological model. BED is a measure of the biological consequences to the tissue receiving a particular dose. RE measures the effectiveness of a given dose. These parameters are derived from the linear-quadratic (LQ) formalism, which expresses the surviving fraction (S) of cells as a function of dose (D).

$$
S = e^{-\alpha D - \beta D^2} \tag{12}
$$

The constants α and β are characteristic of a given cell type and have units of Gy⁻¹ and Gy⁻² respectively. BED and RE are related to the physical absorbed dose by the following expression:

$$
BED (Gy) = RE x (Absorbed dose)
$$
 (13)

3.2.1 Relative Effectiveness per unit Dose for a Permanent Implant

For a permanently implanted exponentially decaying source such as treatment of coronary artery disease with radioactive stents, the LQ parameter **RE is represented** by:

$$
(RE)_{\infty} = 1 + \frac{2R_0\lambda}{\mu - \lambda} \left(\frac{\beta}{\alpha}\right) \left[\frac{1}{2\lambda} - \frac{1}{\mu + \lambda}\right]
$$

$$
= 1 + \frac{2R_0\lambda}{\mu - \lambda} \left(\frac{\beta}{\alpha}\right) \frac{\mu - \lambda}{2\lambda(\mu + \lambda)} \tag{14}
$$

which reduces to:

$$
RE = 1 + \frac{\dot{D}_0}{\mu + \lambda} \left(\frac{\beta}{\alpha}\right) \tag{15}
$$

where \dot{D}_0 is the initial dose rate delivered to the tissue and λ is the radioactive decay constant of the implanted radionuclide. μ is the sublethal repair constant, which describes

the probability for cells to repair the radiation damage per hour and is related to the repair half-life $T_{1/2}$ by $\mu = \ln 2/T^{1/2}$. The value of the repair half-life to be used in this model is taken from published literature (Brenner and Hall, 2001) and has been experimentally determined to be 0.5 hours for smooth muscle cells. The α/β ratio for early-responding vascular tissue has been estimated as 8 Gy (Brenner and Hall, 2001). This is assuming that the cells in the arterial wall responsible for the neointimal hyperplasia component of restenosis are rapidly proliferating cells and can be estimated more accurately as tumor or cancer cells than normal tissue. A summary of the values used in the radiobiological model is shown in Table 3-6. Included is also the values of λ for each of the radioisotopes, which was calculated according to $\lambda = \ln 2/t^{1/2}$, where $t^{1/2}$ is the half-life of the radioisotope.

α/β ratio	8 Gy
Repair half-life	0.5 hours
μ (sublethal repair constant)	$1.386 h^{-1}$
λ (decay constant of the radionuclide)	$32P:0.00202 h-1$ $^{90}Y: 0.0108 h^{-1}$ 188 Re: 0.0409 h ⁻¹ 177 Lu: 0.00428 h ⁻¹
Dose rate	varies for each point
Absorbed dose	varies for each point

Table 3-6. Summary of Values for Variables **used in Radiobiological Model**

3.2.2 RE for Implant Containing Two Different Radioisotopes

The dosimetry model will also be used to calculate the dose distribution of a stent containing two radioisotopes, one in the center or body of the stent, and another in the edges of the stent or a uniform distribution of a combination of radioisotopes. In this **case,** equation (14) will not be adequate because it only takes into account the biological effects of the dose rate and half-life of one radioisotope only, where in fact the two radioisotopes might have important effects, particularly if the half-lives differ greatly. For a permanent implant in which the dose is a result of a combination of two radioisotopes as in the case of the hybrid stent, the following equation was used (Chen, 2003):

$$
RE = 1 + \frac{\beta}{\alpha} \times \frac{\frac{\dot{D}_{01}^2}{\lambda_1(\lambda_1 + \mu)} + \frac{\dot{D}_{02}^2}{\lambda_2(\lambda_2 + \mu)} + 2\frac{\dot{D}_{01}\dot{D}_{02}(\lambda_1 + \lambda_2 + 2\mu)}{(\lambda_1 + \lambda_2)(\lambda_1 + \mu)(\lambda_2 + \mu)}{\frac{\dot{D}_{01}}{\lambda_1} + \frac{\dot{D}_{02}}{\lambda_2}},
$$
 (16)

where \dot{D}_{01} and \dot{D}_{02} is the dose rate for isotopes 1 and 2 respectively, and λ_1 and λ_2 are the decay constants. The biological equivalent dose or BED is calculated in the same manner as that for a stent containing one radioisotope and is the product of RE and the absorbed dose.

3.3 Stent Model

The stent design used in this research **is an** example of a stent geometry that can be used for calculation of the dose distribution surrounding the stent. The stent (patent No. 6,187,037) was designed by Bionucleonics Inc, of Miami, FL. Cartesian coordinates (x,y,z) are used in our model and arranged in a Microsoft Excel file and loaded into the program. The file with these stent source coordinates can be found in Appendix D. The dose calculation model described in this work can accommodate any set of coordinates that define the geometry of the stent. The stent has deployed dimensions of 15.25 mm in length and 4mm in diameter with a strut thickness of 0.1527mm (Satz, 2001). The stent is divided into 1840 pieces, or strut points, which are considered the point sources in the DPK convolution calculation. The detailed gridded architecture of the stent used in this research project is shown in Figure 3-6. It represents a projection of the stent into the (θ, θ) z) plane.

Figure 3-6. Stent geometry and dimensions

The stent developed by Bionucleonics Inc., has 177 Lu incorporated into a nickeltitanium alloy which when subsequently activated, yields an entirely uniform dispersion of the radioisotope, much more uniform than coating the alloy surface by ion implantation, vacuum deposition or other deposition technique (Satz, 2001). 177 Lu shows an optimal combination of properties that make it very attractive in the preparation in this type of stents, including a wide cross-section in barns, which facilitates activation in a reactor and requires a shorter reaction time to achieve the preferred levels of radioactivity. 177 Lu has a half-life of 6.734 days with a maximum energy of 498 keV, an average energy of 134 keV and gamma emissions of 113 keV (6.4%) and 208 keV (11.0%) (Satz, 2001). The 177 Lu stent is 0.1% by weight lutetium. This stent design will be used for all other dose calculations with different radioisotopes. For radioisotopes other than '7Lu, the radiation was assumed as being on the surface of the stent, not embedded within the nickel-titanium alloy.

3.4 Algorithm for Radiation Dosimetry Model

The algorithm is written in Matlab 6.5 programming language (The MathWorks, Inc., Natick, Massachusetts) and is based on the dose point kernel (DPK) convolution. The DPK convolution method divides the extended source of radiation, in this case the stent, into a series of point sources. The dose at a target point is made up of the contributions of each of the individual point sources. For calculation purposes, the stent will be represented by a set of contiguous transaxial **slices.** Figure 3-7 represents a slice along the transaxial plane of the stent and the corresponding cross-section of stent struts. Each slice corresponds to one value of z along the **z-axis** (length of stent), and contains the strut point (x,y) for that value of z. The dose calculation algorithm calculates the dose

delivered from each of the source point defining the stent architecture on a slice-by-slice basis.

Figure 3-7. Stent cross section at midplane $(z = 0)$

3.4.1 Dose Calculation Algorithm Flowchart

Each slice along the z axis of the stent depends on the coordinates x and y. Therefore, each stent coordinates can be represented by the following notation: $z(x,y)$, where z is -7.75 mm to 7.75 mm at intervals of 0.125 mm. For each slice z there are 16 sets of (x, y) coordinates. Computation time was decreased by excluding the source points at which distances between source and target points extended beyond the range of the DPK for a particular radioisotope. Figure 3-8 shows a flow diagram of the algorithm for the radiation dosimetry model developed in this project. A sample of the algorithm code is provided in appendices E and F.

Figure 3-8. Diagram of Dose Calculation Algorithm

3.4.2 Model Assumptions

The algorithm for calculation of dose distributions around radioactive stents is based on

the following assumptions:

- 1) Absorbed dose is calculated for stent with deployed dimensions of 4 mm in diameter and 15.25 mm in length.
- 2) Medium (soft arterial tissue) is isotropic and homogeneous with attenuation properties close to that of water. The presence of plaque is not accounted for.
- 3) Dose is symmetrical around the stent, so for calculation efficiency purposes, only the dose in the first quadrant of the stent is calculated.
- 4) Absorbed dose is expressed per unit of activity in $cGy/\mu Ci$. cGy describes the dose delivered per gram of tissue. μ Ci is the unit of activity, where activity is a measurement of the number of radioactive disintegrations or transformations an amount of radioactive material undergoes in a given period of time.
- *5)* Stent is a permanently implanted therefore the convolution equation is integrated to time $t = \infty$.
- 6) For ' 77Lu, source of radiation is embedded within the NiTi alloy. In this case, the dose delivered to tissue is corrected by a self-attenuation factor described in Section 3.4.3. For all other radioisotopes the source of radiation is simulated on the stent surface as a coating.

3.4.3 Conversion of Units

The DPK data in the tables of Cross et al. (1992) are expressed in units of nGy-cm²/Bq-h. In our calculations, it was expressed in cGy-mm²/ μ Ci-h. The conversion of units was performed as follows:

$$
k = \frac{nGy \cdot cm^2}{Bq \cdot h} \times \frac{10^{-7} \, \text{cGy}}{nGy} \times \frac{37,000Bq}{\mu Ci} \times \frac{100mm^2}{cm^2} = \frac{0.37 \, \text{cGy} \cdot mm^2}{\mu Ci \cdot h} \tag{17}
$$

Therefore, to express the dose in units of $cGy \cdot mm^2/\mu Ci \cdot h$, the dose values obtained were multiplied by the conversion factor $k = 0.37$.

3.4.4 Attenuation Correction Factor for Dose Point Kernel Calculation of 1"Lu

The 17Lu stent is made radioactive by a different **manner** altogether, one that has the potential for a more uniform distribution of the radioisotope, leading to a more uniform dose delivery to the artery wall. The lutetium **is an** integral part of the nickeltitanium alloy prior to its activation. For this reason, a correction factor needs to be introduced to account for the self-attenuation of the stent material. The self-attenuation by uniform dispersion of the radioactive source in the stent material is given by the following expression:

$$
Actual dose = D \times \left[1 - \left(\frac{1}{T}\right) \rho \frac{dT}{dx} \Delta x\right],\tag{18}
$$

where the percentage loss of energy due to attenuation of beta particles is given by $\left(\frac{1}{T}\right)\rho\frac{dT}{dx}\Delta x$, in which $\frac{dT}{dx}$ is the total stopping power(the sum of the collision and radiative stopping powers) for NiTi alloy equal to 2.391 MeV-cm²/g (Berger et la, 2000), T is the average beta **emission** energy equal to 0.134 MeV, and p is the density of the NiTi alloy equal to 6.45 α /cm³.

The CSDA range (continuous slow down approximation) is a very close approximation to the average path length traveled by a charged particle **as it** slows down to rest (Attix, 1986, p.180; Berger et al., 2000). The R_{CSDA} of NiTi for beta particles with an average energy of 0.134 MeV is 3.443 x 10^{-2} g/cm² (Berger et al., 2000). This value, divided by the density of NiTi results in the range expressed only in units of distance 0.0534 mm. This value was used to exclude those source points on the stent that have no contribution to the total dose at a particular target point. Assuming that the source of radioactivity **is a point** source located in the center of the stent strut, then the distance traveled by beta particles to the target point, which is equal to 0.07635, or half the stent strut thickness, is greater than the CSDA range. For this **reason** the location of the point source was assumed to be in the center of half of the strut width, as shown in Figure 3-8. As a result, a quantity of 0.038175 mm, or one fourth the strut width was added to each x and y coordinate defining the stent geometry in order to consider the point source in the center of the stent. In equation 18, Δx is equal to 0.038175 mm. This is the only segment that will be attenuated because the others are so out of range that none of the radiations will make it to the target point. In addition, the dose is multiplied by a factor of 0.5 taking into account the fact that the activity of half of the strut will have no contribution to the dose whatsoever.

Figure 3-9. Portion of stent strut with attenuation correction

The attenuation due to gamma absorption was also calculated according to the exponential attenuation equation for gamma rays and x-rays (Attix, 1986, p.39). In this case Δx is $\frac{0.1527}{2}$ = 0.076350 or half of the stent strut thickness, under the assumption that the gamma radiation is emitted from the center of the stent strut.

$$
attention = e^{-\mu\Delta x} \qquad (19)
$$

$$
\frac{\mu}{\rho}_{NiTi} = \left(\frac{\mu}{\rho}\right)_{Ni} + \left(\frac{\mu}{\rho}\right)_{Ti} f_{Ti}, \quad (20)
$$

where $\left(\frac{\mu}{\rho}\right)_{N_i}$ and $\left(\frac{\mu}{\rho}\right)_{T_i}$ are the linear mass attenuation coefficients for Ni and Ti

respectively, which were obtained from the tables provided by Chantler et al. (2003). f_{Ni} and f_{Ti} are the fractions of each material that contribute to the total composition of the NiTi, which is *50%* Ni and *50%* Ti. The attenuation factors for both beta and gamma energies will be multiplied by the calculated dose values to obtain the corrected dose values due to the attenuation of the stent material. This was applied only in the case of 17 Lu.

3.5 Model **Simulations**

One of the aims of this study is to theoretically evaluate different radioisotopes as potential candidates in a radioactive stent for the treatment and prevention of restenosis, as well as for the prevention of edge restenosis. In addition to characterizing the 3D dose distribution of ^{32}P , ^{90}Y , ^{188}Re and ^{177}Lu stents, combinations of these radioisotopes will also be explored such as shown in table 3-7. The geometric distribution of radioisotopes can be varied with ease. Two radioisotopes can be combined throughout the entire length of the stent. The 3 mm at both ends of the stent were simulated to include a radioisotope that is different from that in the center of the stent as shown in Figure 3-10. This also includes the "dumbbell effect" or the incorporation of higher activities of the radioactive source at the stent ends.

Figure 3-10. Radioisotope distribution at stent edges

Stent Simulations	Stent Center	Stent Edges
	$177Lu + 32P$	$177Lu + 32P$
2	$177Lu + {}^{90}Y$	$177Lu + {}^{90}Y$
3	$177Lu + 188Re$	$177Lu + 188Re$
	32 _P	$3 \times \frac{32}{9}P$
5	90 _Y	$3x^{90}Y$
6	188 Re	$3x^{188}$ Re
	177 Lu	$3x^{177}$ Lu
8	32 _P	188 _{Re}
9	32 _P	90 _V

Table **3-7. Radioisotope Distribution of Stent** Geometry
3.6 Model Validation

The validation of the DPK convolution for dosimetry calculation of radioactive stent was compared against the results obtained from Monte Carlo simulations using the MCNP code. Monte Carlo simulations were done by Michael G. Stabin of Vanderbilt University. The stent was simulated as a hollow cylinder with 32P embedded within the volume of the cylinder. The dimensions of this cylinder are shown in Figure 3-11.

Figure **3-11. Dimensions of 32P cylinder used** in MCNP **and DPK calculations**

The dose was calculated at distances of 0.01, 0.2, *0.5,* 0.7, 1.0, 2.0, and 4.0 mm from the surface of the stent at $z = 0$ or the midplane of the stent. The Monte Carlo simulation was done under the following assumptions:

- 1) Cylinder **is made** of NiTi *(50%* Ni, *50%* Ti).
- 2) The full beta energy spectrum of $32P$ was used.
- 3) Dose was calculated as dose per electron emitted from the whole stent, not per unit, and was calculated for 3.7×10^{-4} dps at each emission point.
- 4) Dose is expressed in units of cGy/mCi-h.
- *5)* Dose calculation included the production of secondary electrons and bremssthralung.
- 6) The number of simulations yields a mean error of 6.1% in the calculations of the dose at the specified points.

The radiation dose from a $3^{2}P$ cylinder was calculated at the same distances described above using the DPK dosimetry model developed in this project. The cylinder was defined by cylindrical coordinates with a radius of 0.48-1.08 mm. The radius was divided to discrete intervals of 0.2 mm, the length in intervals of 0.115 mm from -9 to 9 mm, and the angle in intervals of 5 degrees, from 0 to *355* degrees. This is equivalent to 31,913 units of volume. The activity was 1 mCi per unit. To make this calculation comparable to the Monte Carlo results, the issue of attenuation had to be addressed. Unlike Monte Carlo simulations, the DPK convolution method relies on using average values of attenuation. Therefore, in this type of calculation, the attenuation correction for the dose delivered by $32P$ embedded within the volume of the NiTi cylinder relied on two main points: a) the elimination of all those source points of the cylinder whose distance traveled within the stent to the target point is greater than the CSDA range of the mean energy of ³²P in the NiTi material, and b) calculation and implementation of an attenuation factor for beta particles as described in equation 18, section 3.4.3. First and foremost, the CSDA range **in mm** was calculated for the mean beta energy of 32P

traveling across a NiTi material. This calculation was done using the values shown in

Table 3-8.

Table **3-8. Stopping Power** (dT/dx) and CSDA **range for the mean beta energy of 32P emissions in a NiTi alloy.**

		Energy (MeV) dT/dx (Mev-cm ³ /g) CSDA Range (g/cm ²)
0.695	1.382	0.3942

The range, calculated as:

$$
\frac{0.3942 g/cm^2}{6.45 g/cm^3} = 6.11 cm = 0.611 mm,
$$

meaning that the width of the cylinder is greater than the range. The following assumptions were made based on this result, and were included in the selection of source coordinates defining the source components of the cylinder:

1) The angle was reduced to a range between -45° and 45°. This approximation is justified because the mean distance within the stent source points to the target points is greater than the CSDA range of 0.611 mm.

Figure 3-12. Angle range defining contributing points to dose at target

mean distance
within the stent
$$
0.78mm - 0.01\sqrt{2}mm = 0.77mm
$$

- 2) Only those points in the z range from -0.23 mm to 0.23 mm contributed to the radiation dose. This assumption is justified by the fact that most of the source points outside of this range have distances within the stent greater than the CSDA range.
- 3) Approximately only half of the source points in the range $z \in [-0.23mm,0.23mm]$ contributed to the radiation dose. The final dose values were thus multiplied by a factor of 0.5.

Figure 3-13. Contributing points to dose at target for a segment along the stent z axis

4) The dose was divided by 31,913 units of volume under the assumption that each unit has 1 mCi of activity.

Attenuation correction for $32P$ particles embedded within the volume of the stent was calculated according to:

$$
AF=1-\frac{1}{T}\rho\frac{dT}{dx}\Delta x,
$$

where Δx is the mean distance from the source points included in the calculation to the **stent surface. It was approximated by:**

$$
\Delta x = \overline{d} - r \,, \tag{21}
$$

where \overline{d} is the mean distance from source to target points, which was calculated by the **code in Appendix F, and r is the distance from the surface of the stent to the target point.**

CHAPTER 4 RESULTS

Results presented are for the lifetime dose delivered by the stent under the assumption that the stent is permanently implanted. Dose is expressed per unit of activity as cGy/pCi. Radioactivity is assumed on the stent surface unless stated otherwise as in the case of 177 Lu. A target distance of 2.0 mm is taken as a reference point with adventitial layer of the artery as the target tissue for prevention of cell proliferation due to neointimal hyperplasia and negative vascular remodeling.

4.1 Dosimetry of a 3P stent

Figure 4-1 represents a 2D dosimetry plot for a stent coated with ^{32}P as a function of radial distances along the entire axis of the stent. The different curves are for doses at different depths into the arterial wall. In the near-field or 0.1 mm from the surface of the stent, the dose varies from 3000 to 4700 cGy/ μ Ci at the stent strut wires to 1000 cGy/ μ Ci in the interstices between struts. At a distance of 1.0 mm, dose delivery is more homogenous with a cumulative dose of \sim 500 cGy/ μ Ci.

Figures 4-2 to 4-5 represent 3D dose maps extending from -10 mm $\le z \le 10$ mm at distances $d = 0.1, 0.5, 1.0$ and 2.0 mm from the stent surface. These plots are projections over the stent in the (θ, z) plane as shown in Figure 3-6 (Chapter 3). Shown is the dose for a segment over the stent between 0 and 45 degrees. These plots provide a clear view of the nonuniformity of the dose at very close distances to the stent. Evident is also the fall-off of the dose at the edges of the stent at \sim -8.0 and 8.0 mm on the z axis.

Figure 4-1. 2D plot of dose distribution of a ³²P stent

Figure 4-2. 3D map of dose to tissue after total decay of a ^{32}P stent at 0.1 mm exterior to the stent surface.

Figure 4-3. 3D map of dose to tissue after total decay of a 32P stent at 0.5 mm exterior to the stent surface.

Figure 4-4. 3D map of dose to tissue after total decay of a 32P stent at 1.0 mm exterior to the stent surface.

Figure 4-5. 3D map of dose to tissue after total decay of a 32P stent at 2.0 mm exterior to the stent surface.

Figure 4-6 represents the dose as a function of radial distance up to 5 mm into the arterial wall calculated at the stent midplane or $z = 0$. For ³²P, the dose delivered at 0.1 mm is 4797 $cGy/\mu Ci$, while at 1.0 mm it falls off by 88% to 531.84 $cGy/\mu Ci$. At a target distance of 2mm, the dose is 172.26 cGy/ μ Ci, or 3.7% of the dose at 0.1 mm. The dose delivered to the arterial wall at a fixed radial distance of 2.0mm along the axis of the stent is shown in Figure 4-7 for half a stent from 0 to 12 mm. At the stent ends, the dose is 88.78 cGy/ μ Ci or 51.54% of the dose at $z = 0$.

Figure 4-6. The variation **of dose with radial distance of a 32P stent**

Figure 4-7. The variation **of dose with axial distance of a 32P stent**

4.2 Dosimetry of a '"Y stent

Figure 4-8 represents a 2D dosimetry plot for a stent coated with ^{90}Y as a function of increasing radial distance along the entire axis of the stent. The different curves are for doses at different depths into the arterial wall. At 0.1 mm from the surface of the stent, the dose varies from 900 to 600 $cGy/\mu Ci$ at close proximity to the stent strut wires to \sim 200 cGy/ μ Ci in the areas between struts. At a distance of 1.0 mm, dose delivery is more homogenous with a cumulative dose of \sim 100 cGy/ μ Ci. At 2.0 and 3.0 mm, the dose is completely uniform, although it has fallen to less than 5% of the dose at 0.1 mm.

Figure 4-8. 2D plot of dose distribution of a "Y stent

Figures 4-9 to 4-5 represent 3D dose maps extending from -10 mm $\le z \le 10$ mm at distances $d = 0.1, 0.5, 1.0$ and 2.0 mm from the stent surface. These plots are projections over the stent in the (θ, z) plane shown in Figure 3-6 (Chapter 3). Shown is the dose for a segment over the stent between 0 and 45 degrees of the total stent circumference. At a distance of 0.1 mm, the effect of the stent wires is pronounced. This effect is reduced significantly at distances of 1.0 and 2.0 mm as seen in Figures 4-11 and 4-12.

Figure 4-9. 3D map of dose to tissue after total decay of a ⁹⁰Y stent at 0.1 mm exterior to the **stent surface.**

Figure 4-10. 3D map of dose to tissue after total decay of a "Y stent at 0.5 mm exterior to the stent surface.

Figure 4-11. 3D map of dose to tissue after total decay of a ⁹⁰Y stent at 1.0 mm exterior to **the stent surface.**

Figure 4-12. 3D map of dose to tissue after total decay of a 9Y stent **at 2.0 mm exterior to the stent surface.**

Figure 4-13 represents the dose as a function of radial distance up to 5 mm into the arterial wall calculated at the stent midplane or $z = 0$. For ⁹⁰Y, the dose delivered at 0.1 mm is 888.81 cGy/ μ Ci, while at 1.0 mm it falls by 85.42% to 129.60 cGy/ μ Ci. At a target distance of 2mm, the dose is 58.19 $cGy/\mu Ci$, or 6.5% of the dose at 0.1 mm. The dose delivered to the arterial wall at a fixed radial distance of 2.0 mm along the axis of the stent is shown in Figure 4-14 for half a stent from 0 to 12 mm. At the stent ends, the dose is 29.89 cGy/ μ Ci or 51.38% of the dose at $z = 0$.

Figure 4-13. The variation of dose with radial distance of a ^{90}Y stent

Figure 4-14. The variation of dose with axial distance of a ⁹⁰Y stent

4.3 Dosimetry of a '"Re stent

Figure 4-15 represents the 2D dosimetry plot for a stent coated with ¹⁸⁸Re as a function of increasing radial distance along the entire axis of the stent. The different curves are for doses at different depths into the arterial wall. At 0.1 mm from the surface of the stent, the dose varies from 290 to 195 cGy/µCi at close proximity to the stent strut wires down to \sim 60 cGy/ μ Ci in the areas between struts. At a distance of 1.0 mm, dose delivery is more homogenous with a cumulative dose of \sim 30 cGy/ μ Ci. At 2.0 and 3.0 mm, the dose is completely uniform, although it has fallen to about 6% of the dose at 0.1 mm.

Figure 4-15. 2D plot of dose distribution of a ¹⁸⁸Re stent

Figures 4-16 to 4-19 provide a view of 3D dose maps extending from -10 mm $\le z \le 10$ mm at distances $d = 0.1, 0.5, 1.0$ and 2.0 mm from the stent surface. These plots are projections over the stent in the (θ, z) plane shown in Figure 3-6 (Chapter 3). Shown is the dose for a segment over the stent between 0 and 45 degrees of the total stent circumference. At a distance of 0.1 mm, the effect of the stent wires is very pronounced, indicating the nonuniformity of dose. This effect is significantly reduced at distances of 1.0 and **2.0 mm as seen** in Figures 4-18 and 4-19.

Figure 4-16. 3D map of dose to tissue after total decay of a "8Re stent at 0.1 mm exterior to the stent surface.

Figure 4-17. 3D map of dose to tissue after total decay of a ' ⁸ Re stent at 0.5 mm exterior to the stent surface.

Figure 4-18. 3D map of dose to tissue after total decay of a ¹⁸⁸Re stent at 1.0 mm exterior to the stent surface.

Figure 4-19. 3D map of dose to tissue after total decay of a ¹⁸⁸Re stent at 2.0 mm exterior to the stent surface.

Figure 4-20. The variation of dose with radial distance of a ¹⁸⁸Re stent

The dose delivered to the arterial wall at a fixed radial distance of 2.0mm along the axis of the stent is shown in Figure 4-21 for half a stent from 0 to 12 mm. At the stent edge, the dose is 7.6 cGy/ μ Ci or 53 % of the dose at $z = 0$. Figure 4-20 represents the dose delivered by a ¹⁸⁸Re stent as a function of radial distance up to 5 mm into the arterial wall calculated at the stent midplane or $z = 0$. At 0.1 mm, the dose delivered is 289.75 $cGy/\mu Ci$, while at 1.0 mm it falls by 88.35% to 33.76 $cGy/\mu Ci$. At a target distance of 2mm, the dose is 14.30 cGy/µCi , or 4.9% of the dose at 0.1 mm.

Figure 4-21. The variation of dose with **axial distance for a " ⁸ Re stent**

 188 Re decays by both beta and gamma emissions. Although its highest energy is provided by beta decay, it has gamma decay of 0.155 MeV at 14.9%, 0.633 MeV at 1.25% and 0.478 at 1.01%. The relative percentage of this gamma decay to the total dose delivered by a ¹⁸⁸Re stent as a function of radial distance is shown in Figure 4-22. The beta DPK of

¹⁸⁸Re has a maximum range of 8.7 mm, at which point the dose is composed of gamma **emissions** entirely. At the target distance of 2 **mm, gamma emissions** contribute about a 20% to the total dose.

Figure 4-22. Contribution of gamma energy to total dose of a 188Re stent

Figure 4-23 provides a view of the total dose as a function of radial distance and the contributions of both beta and gamma **emissions.** From a distance of 4.0 mm the greatest contribution is provided by the **gamma emissions.** Note that the dose from beta **emissions** falls off more quickly than that of gamma emissions, which is of a much smaller magnitude, but with a less steep dose gradient. Numerical values are presented in Table 4-1.

Figure 4-23. Beta and gamma components of dose delivered by a 188Re stent

Radial distance (mm)	BETA $(cGy/\mu Ci)$	GAMMA $(cGy/\mu Ci)$	TOTAL DOSE $(cGy/\mu Ci)$
0.1	259.68	30.07	289.75
0.5	54.81	8.19	63.00
1.0	28.61	5.14	33.76
1.5	17.65	3.82	21.47
2.0	11.28	3.02	14.30
2.5	7.22	2.48	9.70
3.0	4.56	2.08	6.64
3.5	2.83	1.77	4.61
4.0	1.72	1.54	3.26
4.5	1.04	1.34	2.38
5.0	0.62	1.19	1.80

Table 4-1. Beta and gamma components of total dose of a ¹⁸⁸Re stent

4.4 Self-Attenuation Correction Factor for DPK Calculation of a 177Lu stent

The ¹⁷⁷Lu stent, unlike all others simulated in this work has the radioactivity embedded within the volume of the stent. This required the calculation of an attenuation correction factor of the beta and gamma emissions as they travel from the inside of the material to the target distances. The self-attenuation factor for both beta and gamma emissions was calculated according to equations 18 and 19 in (Chapter 3). Selfattenuation factors were 0.28 for beta emissions and 0.991 for gamma emissions. The final dose results obtained from the dosimetry model was then multiplied by these factors for beta and gamma emissions respectively. Figure 4-24 shows a comparison of the dose delivered from the midstent, or $z = 0$, to target points ranging from 0.1 to 5 mm from the stent. The dose from the beta emissions is significantly attenuated with only 28% of the dose reaching the target tissue. As the distance increases from the stent surface, the dose from beta emissions becomes negligible and the dose is composed entirely of gamma emissions. At these points the dose with and without attenuation correction is almost identical, because for gamma energy the self-attenuation is negligible. Table 4-2 shows numerical values of the components of the dose delivered by ¹⁷⁷Lu at increasing radial distances from the surface of the stent.

Figure 4-24. Comparison of dose with and without attenuation correction for a ¹⁷⁷Lu stent.

Radial distance (mm)	Beta	Beta (with attenuation correction)	Gamma	Gamma (with attenuation correction)	Total dose	Total dose (with attenuation correction)
0.1	2,774.4	776.8	142.42	141.14	2,916.80	917.96
0.5	55.7	15.6	33.91	33.61	89.61	49.20
1.0	0.4914	0.1376	20.99	20.81	21.49	20.94
1.5	0.0123	0.0035	17.10	16.95	17.11	16.95
2.0	0.0000	0.0000	13.34	13.22	13.34	13.22
2.5	0.0000	0.0000	11.56	11.46	11.56	11.46
3.0	0.0000	0.0000	9.55	9.46	9.55	9.46
3.5	0.0000	0.0000	8.50	8.42	8.50	8.42
4.0	0.0000	0.0000	7.23	7.17	7.23	7.17
4.5	0.0000	0.0000	6.54	6.48	6.54	6.48
5.0	0.0000	0.0000	5.68	5.63	5.68	5.63

Table 4-2. Comparison of dose with and without self-attenuation correction factors for dose delivered by a 177 Lu stent.

4.5 Dosimetry of a 1"Lu **stent**

Figure 4-25 shows the 2D dose distribution of '"Lu emitting stent along the entire axis of the stent with distances ranging from 0.1 to 2.0 mm from the stent surface. Due to the steep dose gradient of 177 Lu, the dose delivered at a distance of 0.1 mm ranges between \sim 900 cGy/ μ Ci at points in close proximity to the stent struts and \sim 50 cGy/ μ Ci in areas between the struts. As the distance from the stent increases, the dose uniformity increases, although the dose delivered decreases dramatically up to insignificant values with 1.4% of the dose at 0.1 mm, at a target distance of 2.0 mm into the artery wall.

Figure 4-25. 2D plot of dose distribution of a 177Lu stent

Figures 4-26 to 4-29 provide a view of 3D dose maps extending from -10 mm $\le z \le 10$ mm at distances d = 0.1, *0.5,* 1.0 and 2.0 mm from the stent surface. These plots are projections over the stent in the (θ, z) plane shown in Figure 3-6 (Chapter 3). Shown is the dose for a segment over the stent between 0 and 45 degrees of the total stent circumference. At a distance of 0.1 mm, the effect of the stent wires is very pronounced, indicating the nonuniformity of dose. This effect is significantly reduced at distances of 1.0 and 2.0 mm as seen in Figures 4-28 and 4-29.

Figure 4-26. 3D map of dose to tissue after total decay of a ¹⁷⁷Lu stent at 0.1 mm exterior to **the stent surface.**

Figure 4-27. 3D map of dose to tissue after total decay of a 1 77Lu stent at 0.5 mm exterior to the stent surface.

Figure 4-28. 3D map of dose to tissue after total decay of a 1 77Lu stent at 1.0 mm exterior to the stent surface.

Figure 4-29. 3D map of dose to tissue after total decay of a 177Lu stent at 2.0 mm exterior to the stent surface.

Figure 4-30 shows the variation of dose as a function of radial distance delivered by a ¹⁷⁷Lu calculated at stent midplane $z = 0$. A maximum dose of 917.96 cGy/ μ Ci is delivered at a distance of 0.1 mm from the stent, however it falls off steeply by 95% to 49.2 cGy/µCi at a distance of 0.5 mm. The high energy delivered at such short distances is the result of the beta energy. At a target distance of 2 mm from the stent, the dose is only 13.73 $cGy/\mu Ci$, or 1.5% of the initial dose, and is composed entirely of gamma energy.

Figure 4-30. The variation **of dose with radial distance** of a ¹⁷ Lu stent

Figure 4-31 shows the variation of dose with axial displacement for a fixed radial distance of 2.0 mm. The graph shows only half the stent as the dose distribution is symmetrical on the negative z axis of the stent. The dose tends to drop gradually at the

edges of the stent and beyond. At the stent edge, the dose is 8.1562 cGy/4Ci or 60% of the dose at $z = 0$. The contribution to this dose is made solely by gamma emissions of 177 Lu. $\,$

Figure 4-31. The variation of dose with axial distance of a '"Lu stent

Like 188 Re, 177 Lu decays by both beta and gamma emissions. Figure 4-32 shows the percentage of contribution of gamma energy to total dose delivered by the ¹⁷⁷Lu as a function of radial distance from the stent surface. At a radial distance greater than 1 mm the contribution of gamma to total dose is 100%. Figure 4-33 shows the breakdown of the dose into beta and gamma components. It is clear that the dose by beta energy falls off significantly and makes very little contribution to the total dose at a radial distance greater than 1.0 mm. Specific values are shown in Table 4-3.

Figure 4-32. Contribution of gamma energy to total dose delivered by a ¹⁷⁷Lu stent.

Figure 4-33. Beta and **gamma** components of dose delivered by a '"Lu stent

Radial distance (mm)	BETA $(cGy/\mu Ci)$	GAMMA $(cGy/\mu Ci)$	TOTAL DOSE $(cGy/\mu Ci)$
0.1	776.83	141.14	917.96
0.5	15.59	33.61	49.20
1.0	0.31	22.08	22.40
1.5	0.0035	16.95	16.95
2.0	0.00	13.73	13.73
2.5	0.00	11.46	11.46
3.0	0.00	9.75	9.75
3.5	0.00	8.42	8.42
4.0	0.00	7.36	7.36
4.5	0.00	6.48	6.48
5.0	0.00	5.76	5.76

Table 4-3. Beta and gamma components of total dose of a ¹⁷⁷Lu stent

4.6 Dosimetry Comparison of ³² P, 9Y, 1 8Re, and 7Lu stents

Figures 4-34 shows a comparison of the dose distribution of the four radioisotope stents as a function of radial distance. Figure 4-35 represents the ratio of the dose at the point of interest to the dose at the middle of the stent along the stents axis for a fixed radial distance of 2.0 mm for each of the radioisotope stents.

Figure 4-34. Comparison of radial dose distribution of four radioisotopes.

Figure 4-35. Comparison of axial dose distribution of four radioisotopes

4.7 Radiobiological Modeling for 32P,Y, "Re, and 17Lu **Stents**

The purpose of the radiobiological model was to establish a comparison between the physical absorbed dose and the biological equivalent dose (BED) at the same target point. This was done by employing equations 13 and 15 (Chapter 3). Figures 4-36 to 4-39 represent this comparison taken as a function of radial distance from the stent midplane z = 0 for ${}^{32}P$, ${}^{90}Y$, 188 Re, and 177 Lu stents respectively. In all cases, it is clear that the BED is always higher than the physical dose. Tables 4-4 to 4-7 provide the values for dose rate, physical absorbed dose, relative effectiveness (RE) and BED for a set of radial distances ranging from 0.1 to 5.0 mm in intervals of 0.5 mm.

4.7.1 Radiobiological Modeling of a ³²P stent

Figure 4-36. Comparison of BED and physical absorbed dose of a 32P stent

4.7.2 Radiobiological Modeling of a ⁹⁰Y stent

Figure 4-37. Comparison of BED and physical absorbed dose of a ⁹⁰Y stent

Radial distance (mm)	Dose rate (cGy/h)	Absorbed $dose$ (cGy)	Relative Effectiveness	Biological equivalent dose
0.1	9.62	888.81	87.06	77,381.13
0.5	2.47	228.36	23.11	5,277.61
1.0	1.40	129.60	13.55	1,755.93
1.5	0.9209	85.11	9.24	786.44
2.0	0.6296	58.19	6.63	386.08
2.5	0.4359	40.29	4.90	197.43
3.0	0.3017	27.89	3.70	103.19
3.5	0.2074	19.17	2.86	54.74
4.0	0.1409	13.02	2.26	29.45
4.5	0.0945	8.73	1.85	16.11
5.0	0.0624	5.77	1.56	8.98

Table 4-5. Calculation of BED for a ⁹⁰Y stent at increasing radial distance

4.7.3 Radiobiological Modeling of a 1 8Re stent

Figure 4-38. Comparison of BED and physical absorbed dose of a ¹⁸⁸Re stent

Radial distance (mm)	Dose rate (cGy/h)	Absorbed $dose$ (cGy)	Relative Effectiveness	Biological equivalent dose
0.1	11.85	289.75	104.81	30369.83
0.5	2.57	63.00	23.57	1485.20
1.0	1.38	33.76	13.10	442.05
1.5	0.8782	21.47	8.69	186.65
2.0	0.5850	14.30	6.12	87.60
2.5	0.3966	9.70	4.47	43.38
3.0	0.2717	6.64	3.38	22.45
3.5	0.1884	4.61	2.65	12.21
4.0	0.1334	3.26	2.17	7.07
4.5	0.0974	2.38	1.85	4.41
5.0	0.0738	1.80	1.65	2.97

Table 4-6. Calculation of BED for a ¹⁸⁸Re stent at increasing radial distance

4.7.4 Radiobiological Modeling of a 1'Lu stent

Figure 4-39. Comparison of BED and physical absorbed dose of a 1"Lu stent

Radial distance (mm)	Dose rate (cGy/h)	Absorbed $dose$ (cGy)	Relative Effectiveness	Biological equivalent dose
0.1	3.95	917.96	36.49	33,495.70
0.5	0.21	49.20	2.90	142.79
1.0	0.10	22.40	1.87	41.79
1.5	0.07	16.95	1.66	28.06
2.0	0.06	13.73	1.53	21.02
2.5	0.05	11.46	1.44	16.53
3.0	0.04	9.75	1.38	13.43
3.5	0.04	8.42	1.33	11.17
4.0	0.03	7.36	1.28	9.45
4.5	0.03	6.48	1.25	8.11
5.0	0.02	5.76	1.22	7.04

Table 4-7. Calculation of BED for a '"Lu stent at increasing radial distance
The rate at which the dose is delivered is a very important factor in IVBT. Figure 4-40 shows a comparison of the dose rates of the four radioisotope stents. The curves represent the initial dose rate as a function of radial distance from the stent midplane $z = 0$. This is important in the resulting BED for each of the radioisotopes.

Figure 4-40. Variation of dose rate with radial distance for ³²P, ⁹⁰Y, ¹⁸⁸Re, and ¹⁷⁷Lu stents

BED serves an important function in comparing different treatments. Table 4-8 shows the calculation of BED for ${}^{32}P$, ${}^{90}Y$, 188 Re and 177 Lu stents delivered to the same target point of 2.0 mm into the arterial wall. The third column represents the value of the dose at 2 mm, while the fourth column is the activity required for each to reach a therapeutically significant dose of 2000 cGy or 20 Gy. This calculation of BED shows how for a same physical dose, the BED can vary greatly, with that of ¹⁸⁸Re being 20 times the value of BED of $32P$.

Radio- isotope	Half-life	Physical Absorbed Dose at 2 mm $(cGy/\mu Ci)$	Activity (μCi)	Dose Rate (cGy/h)	RE	BED
32 _p	14.28 d	172.26	11.6	4.036	37.35	74,631
$\overline{^{90} \mathrm{V}}$	2.67d	58.19	34.4	21.66	194.83	390,010
188 _{Re}	16.94 h	14.30	139.8	81.781	717.42	1,434,500
177 Lu	6.734d	13.73	145.7	8.601	78.33	156,685

Table 4-8. Calculation of BED of each radioisotope for the same physical dose of 20 Gy

4.8 Combinations of Isotopes within the Same Stent

The following results describe the dosimetry of a radioactive stent containing two radioisotopes. Figures 4-41, 4-43, and 4-45 show the 2D dose distribution of a 177 Lu stent combined with ${}^{32}P$, ${}^{90}Y$ and 188 Re respectively along the entire axis of the stent with distances ranging from 0.1 to 2.0 mm from the stent surface. Figures 4-42, 4-44 and 4-46 represent a comparison of the physical absorbed dose and the biological equivalent dose (BED) for each of the combinations with 177 Lu at increasing radial distance from the stents midplane. Tables 4-9 to 4-11 present the results of the radiobiological modeling according to equation 16 (Chapter 3). The values of RE and BED are for the effects of the combined radioisotopes.

4.8.1 Uniform distribution of 177 Lu and 32 P

Figure 4-41. 2D plot of dose from a combined ¹⁷⁷Lu and ³²P stent

Figure 4-42. Comparison of BED and physical absorbed dose of a combined ¹⁷⁷Lu and ³²P stent.

Radial distance (mm)	Dose rate (cGy/h) 177 Lu	Dose rate (cGy/h) ^{32}P	Total dose rate (cGy/h)	Absorbed dose (cGy)	Relative Effectiveness	Biological equivalent dose
0.1	3.947	9.691	13.638	5,715.60	118.07	675,000.00
0.5	0.212	2.186	2.397	1,131.30	22.24	25,158.00
1.0	0.096	1.074	1.171	554.23	11.38	6,308.20
1.5	0.073	0.605	0.678	316.53	6.99	2,211.50
2.0	0.059	0.348	0.407	185.99	4.57	850.04
2.5	0.049	0.197	0.246	109.03	3.14	342.61
3.0	0.042	0.108	0.150	63.03	2.29	144.05
3.5	0.036	0.056	0.092	36.00	1.78	64.07
4.0	0.032	0.027	0.059	20.67	1.49	30.83
4.5	0.028	0.012	0.040	12.32	1.33	16.44
5.0	0.025	0.005	0.029	8.04	1.25	10.07

Table 4-9. Calculation of BED for a combined ¹⁷⁷Lu and ³²P stent at increasing radial **distance**

4.8.2 Uniform Distribution of 177Lu and Y

Figure 4-43. 2D plot of dose distribution of a combined ¹⁷⁷Lu and ⁹⁰Y stent

Figure 4-44. Comparison of BED and physical absorbed dose of a combined ""Lu and 9Y stent.

4.8.3 Uniform distribution of ¹⁷⁷Lu and ¹⁸⁸Re

Figure 4-45. 2D plot of dose distribution of a combined ¹⁷⁷Lu and ¹⁸⁸Re stent

Figure 4-46. Comparison of BED and physical absorbed dose of a combined ¹⁷⁷Lu and ¹⁸⁸Re stent.

Radial distance (mm)	Dose rate (cGy/h) 177 Lu	Dose rate (cGy/h) 188 _{Re}	Total dose rate (cGy/h)	Absorbed dose (cGy)	Relative Effectiveness	Biological equivalent dose
0.1	3.95	11.85	15.80	1,207.70	83.30	101,000
0.5	0.21	2.58	2.79	112.20	18.33	2,056.20
1.0	0.096	1.381	1.477	56.15	10.48	588.27
1.5	0.073	0.878	0.951	38.42	6.90	264.99
2.0	0.059	0.585	0.644	28.03	4.84	135.74
2.5	0.049	0.397	0.446	21.15	3.56	75.26
3.0	0.042	0.272	0.314	16.40	2.73	44.83
3.5	0.036	0.188	0.225	13.03	2.21	28.74
4.0	0.032	0.133	0.165	10.62	1.87	19.83
4.5	0.028	0.097	0.125	8.86	1.65	14.65
5.0	0.025	0.074	0.099	7.56	1.51	11.45

Table **4-11. Calculation of BED for a combined** ¹⁷ Lu and 188Re **stent at increasing radial distance**

4.9 Modification of Radioisotope Distribution at Stent Edges

The problem of edge restenosis is the results of underdosing at the stent edges as a result of excessive barotraumas during the angioplasty procedure as well as the radiation dose fall-off at the edges. Figures 4-7, 4-14, 4-21 and 4-31 show that the dose at the stent edges is roughly 50% of that in the center of the stent for ³²P, ⁹⁰Y, ¹⁸⁸Re and ¹⁷⁷Lu respectively. The dosimetry model developed in this work was used to modify the radioactivity at the stent edges in order to increase the dose to greater than 50%. This modification was done at the 3 mm proximal and distal segments along the z axis of the stent.

4.9.1 Different **radioisotopes at stent edges**

One alternative is that of combining different radioisotopes at the stent edges. The simulated stents consisted of uniform ^{32}P with added 188 Re and ^{90}Y at 3 mm in the edges.

These isotopes were chosen because of their higher energy compared to ³²P alone. Figure 4-47 shows this comparison as a function of axial distance from the stent midplane at a fixed radial distance of 2 mm. Figure 4-48 shows the same information but with the values normalized at the stent edge of 7.625 **mm in** order to observe the dose gradient extending beyond the **stent** edge. The effect of incorporating different radioisotopes at the stent edges can be further amplified by increasing the activity of the radioisotopes at the stent edges. Figure 4-49 shows the results of increasing the activity of 188 Re and 90 Y to 10 μ Ci at the stent edges. For a ³²P stent, the dose at the edge is 52% of that in the center. For the stent with uniform ^{32}P with 10 μ Ci of 188 Re at the edges, the dose at the edges is 83% higher than the dose at the stent center. In the case of $90Y$, the dose at the edges is 168% higher than the dose at the center.

Figure 4-47. Comparison of a ³²P stent with ¹⁸⁸Re and ⁹⁰Y at stent edges

Figure 4-48. Comparison of a ^{32}P stent with 188 Re and ^{90}Y at stent edges normalized to the stent edge

Figure 4-49. Comparison of a ³²P stent with 10x the activity of ¹⁸⁸Re and ⁹⁰Y at stent edges

4.9.2 Increase of activity at stent edges ("dumbbell effect")

Another way of increasing the dose at the stent edges was by increasing the activity of at the edges of the same radioisotope in order to achieve the "dumbbell effect". For ³²P, ^{90}Y , 188 Re and 177 Lu stents the activity at the stent edges was increased by an order of three. Therefore the stent edges had 3μ Ci of activity.

4.9.2.1 3P **stent with 3x the** activity **at the stent edges**

Figure 4-50 shows the 2D dose distribution of a ^{32}P stent with weighted activity at the edges (3 mm) along the entire axis of the stent with distances ranging from 0.1 to 2.0 mm from the stent surface. The dose is higher at the stent ends compared to the stent center.

Figure 4-50. 2D plot of dose distribution of a 32P stent with 3x the activity on **the stent edges.**

Figure 4-51 shows a comparison of a uniform ^{32}P stent with a ^{32}P stent with 3 times the activity at the edges. The graph shows the curves as the ratio of the dose at a particular point to the dose at the stent center for a fixed radial distance of 2 mm. At the stent edge, the dose delivered by the stent with the weighted activity is 150% of the dose at the stent center, compared to 50% with a uniform ³²P stent.

Figure 4-51. Comparison of dose as a function of axial distance from a uniform 32P stent and one with 3x the activity at the stent edges.

Radial		Position Along Stent Axis						
Distance from Stent Surface	Stent	Stent Center	Stent Edge	1.0 mm Distal	2.0 mm Distal	3.0 mm distal		
	^{32}P	1,082.00	573.24	177.33	58.30	17.50		
0.5 mm	$3 \times \frac{32}{P}$	1,086.80	1,689.50	525.11	173.87	52.45		
	^{32}P	531.79	276.52	121.52	13.47	13.47		
1.0 mm	$3 \times \frac{32}{P}$	535.42	806.28	359.27	40.36	40.36		
	^{32}P	154.04	79.38	43.19	18.11	5.86		
2.0 mm	$3 \times \frac{32}{P}$	155.53	227.93	127.31	54.01	17.58		

Table 4-12. Dose calculation of a ³²P stent with variation in activity at stent edges

3 x "P indicates **nonuniform** activity (three times the activity at the stent edges)

Table 4-12 shows the dose delivered from a uniform ³²P stent and one with 3 times the activity at the stent edges for radial distances of 0.5, 1 and 2 mm from the surface of the stent. The values of dose are taken for points at the stent center, or midplane, at the stent edge, and 1, 2 and 3 mm distal to the edge of the stent.

4.9.2.2. 9Y stent with 3x the activity at the stent edges

Figure 4-52 shows the 2D dose distribution of a ^{90}Y stent with weighted activity at the edges along the entire axis of the stent with distances ranging from 0.1 to 2.0 mm from the stent surface.

Figure 4-52. 2D plot of dose distribution of a "Y stent with 3x the activity **at the stents edges**

Figure 4-53 shows a comparison of a uniform $90Y$ stent with a $90Y$ stent with 3 times the activity at the edges. The graph shows the curves as the ratio of the dose at a particular point to the dose at the stent center for a fixed radial distance of 2 mm. At the stent edge, the dose delivered by the stent with the weighted activity is 130% of the dose at the stent center, compared to 50% with a uniform ⁹⁰Y stent.

Figure 4-53. Comparison of dose as a function of axial distance from a uniform ⁹⁰Y stent **and a "Y stent with 3 times the activity at the stent edges.**

Table 4-13 shows the dose delivered from a uniform $90Y$ stent and one with 3 times the activity at the stent edges for radial distances of 0.5, 1 and 2 mm from the surface of the stent. The values of dose are taken for points at the stent center, or midplane, at the stent edge, and 1, 2 and 3 mm distal to the stent edge.

Radial		Position Along Stent Axis					
Distance from Stent Surface	Stent	Stent Center	Stent Edge	1.0 _{mm} Distal	2.0 mm Distal	3.0 mm distal	
0.5 mm	$90\mathrm{V}$	228.34	120.19	47.21	21.78	10.34	
	$3 \times \frac{90}{Y}$	237.90	341.30	132.88	61.64	29.57	
1.0 mm	90 Y	129.59	67.06	35.77	17.85	8.70	
	$3x^{90}Y$	137.75	184.92	99.83	50.36	24.83	
2.0 mm	$^{90}\mathrm{V}$	58.19	29.90	19.07	10.77	5.59	
	90 V	63.66	79.19	52.23	30.12	15.84	

Table **4-13** . **Dose calculation of a "Y stent with variation in activity at stent edges**

3 x Y indicates **nonuniform** activity (three times the activity at the stent edges)

4.9.2.31 8Re stent with 3x the activity at the stent edges

Figure 4-54 shows the 2D dose distribution of a 188 Re stent with weighted activity at the edges along the entire axis of the stent with distances ranging from 0.1 to 2.0 mm from the stent surface.

Figure 4-54. 2D plot of dose distribution of a 188Re stent with 3x the activity **at the stent edges.**

Figure 4-55 shows a comparison of a uniform 188 Re stent with a 188 Re stent with 3 times the activity at the edges. The graph shows the curves as the ratio of the dose at a particular point to the dose at the stent center for a fixed radial distance of 2 mm. At the stent edge, the dose delivered by the stent with the weighted activity is 125% of the dose at the stent center, compared to 50% with a uniform 188 Re stent.

Figure 4-55. Comparison of dose as a function of axial distance from a uniform ¹⁸⁸Re stent **and a ¹ "Re stent with 3 times the activity at the stent edges.**

Table 4-14 shows the dose delivered from a uniform ¹⁸⁸Re stent and one with 3 times the activity at the stent edges for radial distances of 0.5, 1 and 2 mm from the surface of the stent. The values of dose are taken for points at the stent center, or midplane, at the stent edge, and 1, 2 and 3 mm distal to the stent edge.

Radial		Position Along Stent Axis					
Distance from Stent Surface	Stent	Stent Center	Stent Edge	1.0 _{mm} Distal	2.0 mm Distal	3.0 mm Distal	
0.5 mm	188 Re	63.00	33.52	12.30	5.50	2.66	
	$3x^{188}$ Re	65.45	95.84	34.45	15.04	7.00	
1.0 mm	188 Re	33.76	17.74	9.21	2.30	1.28	
	3×188 Re	35.95	49.13	25.43	5.98	3.12	
2.0 mm	188 Re	14.30	7.59	4.86	2.83	1.63	
	3×188 Re	15.98	19.91	12.86	7.33	4.06	

Table 4-14. Dose calculation of a ¹⁸⁸Re stent with variation in activity at stent edges

 $3 \times$ ¹⁸⁸Re indicates nonuniform activity (three times the activity at the stent edges)

4.9.2.4. " ⁷ Lu with **3x the** activity **at the stent edges**

Figure 4-56 shows the 2D dose distribution of a ¹⁷⁷Lu stent with weighted activity at the edges along the entire axis of the stent with distances ranging from 0.1 to 2.0 mm from the stent surface.

Figure 4-56. 2D plot of dose distribution of a 177Lu emitting stent with 3x the activity at the **stent ends.**

Figure 4-57 shows a comparison of a uniform 177 Lu stent with a 177 Lu stent with 3 times the activity at the edges. The graph shows the curves as the ratio of the dose at a particular point to the dose at the stent center for a fixed radial distance of 2 mm. At the stent edge, the dose delivered by the stent with the weighted activity is 67% of the dose at the stent center, compared to 50% with a uniform ¹⁷⁷Lu stent.

Figure 4-57. Comparison of dose as a function of axial distance from a uniform¹⁷⁷Lu stent **and a** 17"Lu **stent with 3 times the** activity **at the stent edges.**

Table 4-15 shows the dose delivered from a uniform ¹⁷⁷Lu stent and one with 3 times the activity at the stent edges for radial distances of 0.5, 1 and 2 mm from the surface of the stent. The values of dose are taken for points at the stent center, or midplane, at the stent edge, and 1, 2 and 3 mm distal to the stent edge.

Radial		Position Along Stent Axis					
Distance from Stent Surface	Stent	Stent Center	Stent Edge	1.0 mm Distal	2.0 mm Distal	3.0 mm Distal	
0.5 mm	177 Lu	49.10	27.66	10.05	6.74	4.97	
	177 Lu 3x	55.29	74.43	23.62	15.05	10.73	
1.0 mm	177 Lu	21.71	12.47	8.57	6.16	4.67	
	177 Lu 3x	27.55	29.42	19.51	13.52	9.96	
2.0 mm	177 Lu	13.50	8.14	6.50	5.13	4.10	
	177 Lu $+$ 3x	18.66	17.48	13.94	10.85	8.51	

Table 4-15. Dose calculation of a 17 Lu stent with variation in activity at stent edges

3 x ¹⁷⁷ Lu indicates nonuniform activity (three times the activity at the stent edges)

4.10. Validation of DPK convolution method with Monte Carlo simulation of a 3P Cylinder

The validation of the DPK convolution used in the radiation dosimetry model was done for the dose emitted by a cylinder of $32P$ as a function of radial distance from the surface of the cylinder. Table 4-16 summarizes the parameters that were needed for the calculation of the attenuation correction factor AF for the radioactivity of the ^{32}P within the volume of the cylinder.

Table **4-16. Tabulated parameters for calculation of attenuation correction factor for 32P cylinder used in validation** of DPK **convolution.**

r (mm)	\boldsymbol{d} (mm)	Δx (mm)	$-\Delta x$ $\overline{\mathbf{T}}^{\mathbf{p}}$ dx	AF
0.01	0.489	0.479	0.609	0.391
0.2	0.629	0.429	0.55	0.45
0.5	0.888	0.388	0.494	0.506
0.7	1.07	0.37	0.471	0.529
1.0	1.36	0.36	0.458	0.542
2.0	2.33	0.33	0.4201	0.5799
4.0	4.32	0.32	0.407	0.593

The Monte Carlo simulation of the ³²P cylinder was done by Dr. Michael Stabin of Vanderbilt University. Table 4-17 shows the dose values at specific target points obtained from Monte Carlo and DPK methods as well as the % error. This data is plotted in

Figure 4-58.

r (mm)	Monte Carlo	DPK	% Error
0.01	10,400	10,699.7	2.88
0.2	6,000	6,297.9	4.96
0.5	3,680	3,276.4	10.97
0.7	2,580	2,276.6	11.76
1.0	1,900	1,907.3	0.38
2.0	514	371.4	27.74
4.0	35.2	27.8	21.02

Table 4-17. Comparison between Monte Carlo and DPK dose values for increasing radial distance.

Figure 4-58. Monte Carlo versus DPK convolution method for dose calculation as a function of radial distance from surface of cylinder.

The dose emitted from a ³²P stent was also compared to the dose calculated by Janicki et al. (1997), which was also based on a DPK convolution model. The dose was calculated for increasing radial distance r up to 2 mm from the surface of the stent. The stent used by Janicki et al. (1997) was a Palmaz-Schatz stent with expanded dimensions of 5 mm in diameter and 13 mm in length. The sample stent used in the radiation dosimetry model developed in this work had deployed dimensions of 4 mm in diameter and 15 mm in length. Dose was calculated for stents with an initial activity of $1 \mu Ci$ cumulated over a one half-life time interval of 14.28 days for 32 P. Figure *4-59* shows the comparison of the radial dose distribution curves obtained from the DPK radiation dosimetry model developed in this work and the results presented by Janicki et al. (1997). Table 4-18 provides tabulated values of these results.

Figure 4-59. Comparison of DPK radiation dosimetry model with published results (Jancki et al. (1997)).

Table 4-18. Dose at increasing radial distance for DPK radiation dosimetry model and published results from Janicki et al. (1997)

CHAPTER 5 **DISCUSSION**

The results from the DPK radiation dosimetry model of the ³²P, ⁹⁰Y, ¹⁸⁸Re and ¹⁷⁷Lu stents reveal several important findings. These radioisotopes are commonly used in IVBT, with the exception of ¹⁷⁷Lu, and are examples of the different radioisotopes that can be used in the radiation dosimetry model. Relevant to all is the nonuniformity of the dose at close proximity to the stent struts. There is a great variation of the dose at areas immediately adjacent to the struts and that between the struts, resulting in the peaks and valleys of the curves at distances of 0.1 mm shown in Figures 4-1, 4-8, 4-15 and *4-25.* This effect is also evident in the 3D dose maps for each of the stents. At distances of 1-2 mm from the stent surface, the dose distribution becomes more homogenous, although falling to very low, almost negligible doses. It is believed that a dose in the range of *15-* 30 Gy is significant in the treatment and prevention of neointimal hyperplasia. Dose is expressed per unit of activity so to obtain a desired dose it is necessary to multiply by a factor of activity. However, to deliver dose in this range to the adventitial layer at a depth of 2 mm into the arterial wall, the dose at closer proximities to the stent surface will be massive.

This pronounced nonuniformity of dose at such close distances is more evident with ¹⁷⁷Lu than with other isotopes investigated, and quite unfavorable. Because the beta energy of 177 Lu is so low and falls off so steeply to negligible doses at \sim 2.0 mm, the dose at distances greater than 2.0 mm results mostly from gamma emissions. The dose from gamma energy is very low, and not enough to produce any significant effect on its own.

The lutetium is embedded within the volume of the stent and a self-attenuation correction factor of 0.28 was calculated for the beta emissions of this radioisotope. The self-attenuation correction factor for gamma emissions was 0.991. This means that only 28% of the beta emissions will reach the target point, while most of the gamma energy will be accounted for. Furthermore, in the presence of plaque, the dose will be attenuated even further. This is not the most favorable radioisotope for IVBT, even less so for incorporation into the NiTi alloy of the stent.

The radiation dosimetry model also allows the modification of dose distribution at the stent edges. The problem of edge restenosis has prompted the investigation of other alternatives within the radioactive stent. One such alternative is the increase of activity at the stent ends. As can be seen from the results obtained for the uniform stents, the dose at the stent ends is roughly *50%* of that at the center of the stent. This is because the center of the stent receives dose from the center as well as from the stent segment on either side. At the stent ends, the dose is delivered by only half of a stent, as there is no source of radiation in the areas proximal and distal from the stent ends. Figures *4-50, 4-52, 4-54,* and 4-6 illustrate 2D axial dose plots for ${}^{32}P$, ${}^{90}Y$, ${}^{188}Re$ and 11 'Lu stents respectively with 3 times the activity at the stent ends. This did increase the percentage of the dose at the stent ends compared to the dose in the center of the stent as well as for 3 mm distal to the stent edges. However, the dose at the stent ends and beyond will never be the same as that in the center because of the dose fall-off particularly in the case of beta emitters. Even though clinical trials using these modifications have given unsatisfactory results, self-expanding radioactive stents, as well as radioactive angioplasty balloons could help improve these efforts by reducing barotraumas at the stent edges (Serruys and Kay, 2000; Hehrlein and Fischell, 2002).

Along with increasing the activity at the stent ends, different radioisotopes were also simulated. A uniform ^{32}P stent was reinforced with ^{90}Y and ^{188}Re . The selection of these radioisotopes at the stent edges was based on their higher energy. Figures 4-46 and 4-47 show the results of this simulation and reveal that the dose is slightly increased at the stent ends. This effect is amplified by increasing the activity of $90Y$ and 188 Re yielding 162% and 83% higher dose at the ends respectively. The dosimetry of the 177 Lu stent established in this work has demonstrated that ¹⁷⁷Lu is a poor candidate for IVBT due to its rapid fall-off dose and extremely low beta energy that is almost entirely attenuated when incorporated into the metal alloy of the stent. As previously stated, the gamma energy of ¹⁷⁷Lu is attenuated by only 0.01%, and at distances greater than 2 mm from the stent, the dose is composed in its entirety of gamma emissions. Figure 4-35 shows a comparison of the axial dose distribution of the ^{32}P , ^{90}Y , ^{188}Re and ^{177}Lu calculated at a fixed radial distance of 2 mm from the surface of the stent and measured as the ratio of the dose to the dose at the center of the stent. Of the four radioisotopes, 177 Lu has the smoothest dose fall-off at the stent edges, with over 60% of the dose at the center at the edge of the stent, compared to values of *50%* and below for the other radioisotopes. At this distance, the smoothness of the curve of 177 Lu is a result of the gamma emissions. The low gamma energy of 177 Lu produces a smooth reduction in the dose fall-off at the edges of the stent. This effect, which could in theory reduce the candy wrapper effect, makes ¹⁷⁷Lu a potentially useful candidate for use at the stent edges.

The concept of BED is very important in radiotherapy treatment planning and is widely used in the treatment of cancerous tumors (Dale, 1985,1993; Giap et al., 1999). **BED is more complete than physical absorbed dose in that it takes into account the repair** capability of the tissue, half-life and dose rate, all of which are not included in the **physical dose. Even though establishing a treatment based on BED is difficult due to** limited experimental data, it is physiologically more relevant. The results of the **radiobiological model for the four radioactive stents modeled in this project reveal that** the BED is much higher than the physical absorbed dose, ranging from 100 times higher **in the case of 1 88Re to 36 times higher in the case of '"Lu. This can be explained by the** much higher energy and shorter half-life of ¹⁸⁸Re. Furthermore, it is interesting to note **that the relative effectiveness (RE) and hence the BED vary greatly as a function of radial distance. At a smaller radial distance, the dose rate is much higher, resulting in a higher value of RE. At a distance of 5.0 mm, the dose rate has decreased significantly and RE is** close to unity, therefore there is not much difference between the physical absorbed dose **and the BED. Table 4-8 offers an interesting analysis of BED for the same physical dose** delivered by the four radioisotope **stents.** The underlying **premise** is that for **the same physical dose delivered by different radioisotopes, the biological equivalent dose BED is** much different, because of the difference in dose rates, and half-lives of the **radioisotopes. These results illustrate the importance of this type of calculation and provide an insight on the actual effect of this dose on the tissue. The goal is to irradiate target tissue without causing significant damage to healthy tissue.**

In the case of IVBT, the concept of BED may be even more important. Unlike the **treatment of cancer, where the interest is in irradiating and killing the tumor, in restenosis** it is only necessary to irradiate a given population of cells so as to limit their proliferative potential. Also, the range of target tissue is very small, only in the order of a few millimeters thick. In the case of cancer, a higher BED is more favorable. In IVBT applications, this is not as clear. Radiotherapy treatments are not an exact science. Even today, dose prescription is done rather empirically. Because there is a lack of information in the subject of BED related to IVBT and restenosis, the theoretical calculation of BED is not widely used. It is not clear up to what point is a high BED desirable, and what are the dose thresholds for BED that would prove effective in the prevention of restenosis. The practical consequences of BED are unknown and worthy of further experimental investigation.

An attempt was made in this work to investigate the radiobiological effects of the uniform combination of radioisotopes **in one** stent. Figures 4-41, 4-43 and 4-45 show the dose distribution of a 177 Lu stent combined with ^{32}P , ^{90}Y and 188 Re respectively. This results in an increase in the total dose than what would be delivered by 177 Lu alone. A radiobiological modeling was done for each case as well, and is illustrated in figures 4- 42, 4-44 and 4-46. Similar to the simulation of a one-radioisotope stent, the BED is greater than the physical dose. The dose rate **is also** increased. Unfortunately, the advantage of incorporating more than one radioisotope in a stent is unknown and the radiobiological data obtained from this analysis does not provide much enlightenment. The use of multiple radioisotopes in a permanent implant has been used in radioactive seeds for use in head and neck cancer and prostate carcinomas. For tumors containing both fast growing and slow growing cells, the use of different radioisotopes with different half-lives is advantageous (Chen et al., 2003). The details of the radiobiology and cellular kinetics in IVBT is not as clear, and therefore, conclusions that are made in the use of radiation for treatment of cancer cannot be extended to IVBT with the same conviction. This **area is** worthy of further experimental study. Nevertheless, the DPK radiation dosimetry model has demonstrated to have the versatility of allowing the calculation of dose and radiobiological equivalent dose of a stent containing more than one radioisotope with **similar ease** and dexterity.

The validation of the DPK convolution method was done for the dose emitted from a cylinder of $32P$ as a function of radial distance from the midplane of the cylinder and compared to the results obtained from a Monte Carlo simulation done by Dr. Michael Stabin of Vanderbilt University. Figure 4-57 shows that the curves for DPK and Monte Carlo are in close agreement. The percentage error between the two ranges between 2.88% at the points closest to the surface of the cylinder to 21.02% at the furthest distance of 4.0 mm. The error is greater at larger distances because the number of radiations reaching the target point is considerably less. In addition there is a statistical difference between the dose calculated from DPK and Monte Carlo methods, as the DPK relies on the CSDA range and Monte Carlo follows the trajectory of each individual electron and its respective attenuation along path length. Nevertheless, the correlation between the two methods is most important at smaller distances, which are of greatest significance in IVBT dosimetry.

In addition, the results of the DPK convolution method used in the radiation dosimetry model in this work was also compared with the results of a paper published by Janicki et al. (1997) which calculated dosimetry of a ^{32}P Palmaz-Schatz stent using the DPK. Figure 4-58 shows the comparison of these two methods for increasing radial distance calculated from the center of the stent. Dose was calculated from stents with an initial activity of 1 μ Ci cumulated at one half life interval of 14.28 days for ³²P. Table 4-18 provides the tabulated data of these results. This table, as well as figure 4-58, shows that the dose values obtained in the radiation dosimetry model developed in this work are comparable to those obtained by Janicki et al. (1997). The error percentage between the two can be explained by the different stent geometries and stent dimensions that were used in each respective dose calculation. Additionally, the surface area or surface area units in the convolution equation represented by the number of stent struts or coordinates could be quite different for each stent.

The use of the dose point kernel is attractive because it is relatively simple to accommodate different stent geometries, radioisotopes and activities, a task that is more difficult and time consuming with Monte Carlo simulations. The calculation of dose distribution of a radioactive stent in this project revealed fast computation times of less than 15 minutes. Determination of the dose distribution around a radioactive source simply requires inputting radiation source coordinates, initial activity, and the analytical expression for a specific DPK, or a lookup table. Even the combination of isotopes is done quickly and easily. Even though the Monte Carlo method can be very accurate, it is generally not used for radiotherapy treatment planning because of the amount of time required. Instead, most dose calculations rely on the use of tabulated data or dose point kernel tables generated from Monte Carlo radiation transport simulations in water to perform a very fast and generally accurate assessment of the dose distribution.

Monte Carlo simulations do hold some advantages over DPK methods. Monte Carlo simulations can take into account different densities and materials, such as the

presence of plaque to simulate more closely a realistic diseased artery. With DPK, this becomes much more difficult, as the DPK describes the dose emitted by an isotropic point source in water. However, calculations have been done using a scaling factor of DPK that takes into account different densities such as that of plaque surrounding an artery (Janicki et al., 1999).

The most important disadvantage of the DPK convolution method relevant in this study is the problem of self-attenuation when radioactive material is embedded within the NiTi material of the stent, such as the case of the 177 Lu stent. In this case, the lutetium is an integral part of the nickel-titanium alloy prior to its activation; therefore problems of leaching, flaking or peeling of the radioactive coatings of the stent are essentially eliminated (Satz, 2001). As a result, a more uniform distribution of the dose is expected. However, attenuation is a big problem with beta particle irradiation, particularly in the case of 177 Lu, which has a very low energy and range. It is recommended that if a stent is to be made in this manner that the radioactive material is of much higher beta energy, or a low gamma energy, so that it is still capable of delivering significant dose to the target tissue without being completely attenuated within the stent. In the case of ¹⁷⁷Lu, only 28% of the dose from the beta energy is actually delivered to the target tissue. Even by increasing the activity of 177 Lu, the dose still would not be optimal, as there would have to be an enormous dose at close proximity to the stent struts in order to deliver a therapeutically significant dose to tissue at a target distance of 2 mm. Furthermore, increasing the initial activity would also prove problematic since the cost of irradiation would increase significantly, and there is a limit as to how much the stent can be activated. The attenuation factor calculated in this work was based on approximations **and average values. This issue of attenuation correction could be more exact and tractable** through Monte Carlo simulations.

Calculation of the dose within a specified volume is a standard means employed by radiation oncologists/radiation physicists for planning brachytherapy treatments (Fox, 1999). These dose calculation methods provide clinicians with a tool to make pretreatment evaluations of the dose distribution and customize the treatment for the individual patient. Treatment planning in IVBT poses some difficulty because the distances of interest are so small. In conventional brachytherapy applications, distances are usually in a much greater range. Furthermore, there are many uncertainties related to the optimal dose, radiation source, activity, and target tissue to be irradiated. In vascular brachytherapy, the use of dose calculation methods for pre-treatment evaluation is not widely carried out.

Even though radioactive stents have had their share of problems, particularly edge **effects, IVBT is considered a promising and effective anti-restenosis therapy (Kuntz and** Baim, 2000; Sims et al., 2002; Fox, 2002). In 2002, IVBT was the standard of care for **the treatment of in-stent restenosis, and is currently in practice in over 500 catheterization** labs in the United States (Waksman, 2002b). Catheter-based systems such as the Guidant Galileo[™], Cordis Checkmate[™] and Novoste Beta-Cath™ systems are all implemented clinically with much success. The radioactive stent as a delivery platform does need **much improvement, but a device that is so efficient in inhibiting in-stent neointimal** hyperplasia, with such a low incidence of thrombosis and simple in terms of delivery technique should not be abandoned (Colombo, 2000).

Improvement of stent design is a viable option, as well as a radioactive stent specifically tailored for radiation delivery. Stents with fewer, thicker struts make poor radiation delivery systems because of the accentuated difference of dose delivery **between the areas adjacent to the struts and the areas between the struts (Amols, 1999b).** This **can be improved by decreasing stent strut thickness, increasing the number of struts, and placing them closer together in order to deliver a more uniform dose at closer proximities to the stent. It is possible to incorporate radiation considerations into stent design but perhaps not without compromising mechanical properties. A stent with more struts placed closer together could inhibit re-endothelialization that is** an **important healing process necessary to minimize stent thrombosis. Nevertheless, it is questionable if such a stent design would comply with the mechanical properties of the stent.**

Radioisotope selection could also further improve the dose delivery of radioactive stents. Some uncertainty exists as to whether 32P, the most commonly used radioisotope in radioactive stents, which has a maximum energy of 1.71 MeV, provides sufficient penetration to deliver adequate amounts of radiation to the media and adventitia (Amols, 1999a). Doses from all radioactive sources decrease by at least 30-50% per millimeter, with the dose being very high near the source, but rapidly decreasing with increasing distance. The dose fall-off rate exceeds 70% per millimeter for 32P (Amols, 1999a) If an **adequate dose is delivered to the media, the adventitia will receive a negligible dose. On the other hand, for** an **adequate dose to be delivered to the adventitia, the dose to the intima may be excessive (Amols, 1999a; Fischell et al., 2000). Other radioisotopes such as positron emitter** ⁴⁸ V, and **gamma emitters 103Pd and ¹³¹ Cs have received some attention as candidates for use in radioactive stents (Hehrlein and Fischell, 2002). However, these**

gamma emitters are of relatively low energies and would require higher activities to deliver therapeutically effective doses. There also exists the concern of irradiating healthy **tissue as** a result of the more penetrating gamma rays. Still, there is uncertainty as to the optimal activity of the radioisotope, dose and dose rate to be delivered to the tissue, as well as the half-life that best matches the growth rates of the **tissues** involved in neointimal hyperplasia and restenosis.

Drug-eluting stents have been acclaimed by many to be one of the greatest breakthroughs in interventional cardiology (Waksman, 2002a; Indolfi et al., 2003). With the advent of drug-eluting stent technology and the reported lower restenosis rates, it would appear that the future of intravascular brachytherapy (IVBT) would be in jeopardy, particularly in the case of radioactive stents. Despite the fact that radioactive stents need much improvement, IVBT of a catheter-based nature is used clinically with much success. After redilatation of in-stent restenosis, brachytherapy reduces the risk of subsequent recurrence by over *50%* (Teirstein and King, 2003). While IVBT has proven to be an effective treatment in cases of in-stent restenosis, its fate in the drug-eluting stent world appears to be uncertain.

At present, there is sufficient data from numerous trials including SCORES, ELUTES, FIM, RAVEL, SIRIUS, TAXUS I, and TAXUS II to convincingly indicate the promise and effectiveness of drug-eluting stents in reducing restenosis rates to single digit percentages. A target lesion restenosis rate of only *5.5%* was observed in the Boston Scientific TAXUS II trial with a slow-release paclitaxel stent. In the Cordis SIRIUS trial where sirolimus-eluting stents were compared to bare stents, restenosis of the target lesion was observed in only 8.9% of treated patients compared with 36.6% of placebo

patients (Teirstein and King, 2003). Moreover, the Cordis CYPHER drug-eluting stent received FDA approval in April 2003, and the Boston Scientific TAXUS drug-eluting stent has been given an expedited review status by the FDA (Endovascular Today, May/June 2003).

Despite the enthusiasm surrounding drug-eluting stents, the technology is still not perfect. The overall success of drug-eluting stents relies on inconsistent results between animal and clinical trials. The animal models have generally yielded positive results, although it is important to note that the atherosclerotic and healing processes in animals and humans are quite different, with the healing process in humans being much longer and complex. In addition, cases of edge restenosis in drug eluting stents, as well as thrombosis, polymer-induced inflammation, drug-leaching and toxicity, nonabsorbable polymers, and late stent malapposition have been reported (Waksman, 2002a, Indolfi et al., 2003). Late malapposition, or separation of the stent from the vessel wall, has been observed in both the RAVEL and the multi-center SIRIUS BX Velocity sirolimus coated stent trials in a small cohort of patients that were subject to intravascular ultrasound analysis (Indolfi et al., 2003). Other problems could emerge in the long-term results of clinical trials.

A major shortcoming of drug-eluting stents is the cost. At present, drug-eluting stents are costly, which might have a negative impact on their widespread applicability (Waksman, 2002a). Each drug-eluting stent costs more than twice as much as a bare metal stent. In Europe, where drug-eluting stents have been approved since 2002, the use of drug-eluting stents ranged between 5 and 10% of cases, mainly for economic reasons. Adequate reimbursement could also be a problem (Waksman, 2002a). For hospitals

receiving a budget on drug-eluting stents, not all patients and all lesions can be treated with this technology.

Clinical trials have confirmed the benefit and efficacy of IVBT in treatment of instent restenosis lesions of varying lengths and vessel diameters, native artery and aortocoronary saphenous vein graft targets, bifurcation lesions, and/or the presence of diabetes (Waksman and Weinberger, 2003). Recent trials have provided excellent-quality data supporting the effectiveness of y-radiation for the treatment of saphenous vein graft in-stent restenosis, superficial femoral artery obstructions, and diseased dialysis grafts. Results from the SVG BRITE (Saphenous Vein Graft Beta Radiation to Prevent In-Stent Restenosis) study with the use of a ^{32}P source train are encouraging, with a 0% restenosis rate (Waskman, 2002b). In the majority of clinical trials, drug-eluting stents have been used mostly for smaller and simpler lesions. In the SIRIUS study, a *35%* restenosis rate was reported in insulin-dependent diabetic patients, despite the sirolimus-eluting stent. Similarly, rates between 16 and 18% were observed in patients with long lesions and small vessels (Waksman, 2002a). The risk of restenosis is much higher in longer and complex lesions, and the initial data on drug-eluting stents is not very encouraging with higher rates of restenosis in this group of patients (Massat, 2002; Waksman, 2002a; Yajnik et al., 2003). Additionally, there exists no data regarding the effectiveness of drug-eluting stents for saphenous vein graft stenoses (Yajnik et al., 2003).

It is the opinion of some experts that drug-eluting stents will more than double the current number of patients undergoing stent implantation to approximately 2 million a year by 2005 (Yajnik et al., 2003). If this **is the case, even a** very small rate of failure such as *5%* will translate into a considerable number of candidates for IVBT, up to

100,000 per year, even in a drug-eluting stent era (Yajnik et al., 2003). Even though drugeluting stents might become the treatment of choice for in-stent restenosis, restenotic lesions in a more difficult subset of patients with diabetes, long lesions and smaller vessels, and restenosis in peripheral vessels will likely benefit from IVBT therapy. Intravascular brachytherapy should not be dismissed completely as it has proven to be an effective means of treating restenosis and may be needed for the treatment of in-stent restenosis due to drug-eluting stent failures (Waksman, 2002a). A randomized study of IVBT vs. drug-eluting stents in peripheral and more complex lesions is warranted and might provide more conclusive information on this subject.

Experimental work using radiochromic film as well as complete Monte Carlo simulations is recommended for further characterization of the ¹⁷⁷Lu stent. In addition, the calculation of self-attenuation correction factor by experimental means might also be useful and could be a more accurate than theoretical approximations. This is specifically significant for the cases in which the radioisotope is embedded within the volume of the stent. The design of the experiment would consist of measuring the stent activity in a radioisotope calibrator. This activity would be for the radiations emitted from the whole stent and designated as Asa, or the activity of the self-attenuated radiations. The stent would then be pulverized and its activity measured in a beta liquid scintillator with a photomultiplier tube, and corrected for the time decay. This activity, Ao, is the actual activity of the stent. The self-attenuation correction factor would be given by the quotient of the two measured activities or AsA/Ao. Once determined, this self-attenuation correction factor would be multiplied by the dose calculated from the stent. This is the **experimental method used** by NIST **(National Institute of Standards and Technology) for determining the activity** of any **material.**

CHAPTER 6 CONCLUSIONS

The DPK method developed in this work proved to be a fast, easy and versatile way to estimate the 3D distribution of the dose emitted from radioactive stents. The radiation dosimetry model developed in this work has the versatility to accommodate different radioisotopes, stent geometries and activities. This model has fast computation times of 15 minutes or less per simulation. It characterized the dose at different distances into the target tissue in both radial and axial directions. The dosimetry of ${}^{32}P$, ${}^{90}Y$, ${}^{188}Re$ and ¹⁷⁷Lu was investigated. Of the four, ³²P delivers the highest dose, while ⁹⁰Y, ¹⁸⁸Re and ¹⁷⁷Lu require high levels of activity to deliver a significant therapeutic dose in the range of 15-30 Gy. At very short distances from the stent, the dose is highly nonuniform. This nonuniformity decreases with increasing distance, although the dose falls off significantly.

Two different kinds of stents were simulated in this study. The radioactivity was assumed on the stent surface for ${}^{32}P$, ${}^{90}Y$ and ${}^{188}Re$. In the case of ${}^{177}Lu$, the radioactivity was embedded within the volume of the stent. This required the calculation of selfattenuation correction factors for both beta and gamma emissions of ¹⁷⁷Lu. The total dose delivered by ¹⁷⁷Lu was attenuated by 72% for beta emissions and 0.9 % for gamma emissions. Even though the gamma emissions were hardly attenuated, the energy is too low to be significant. In addition, the self-attenuation of beta particles by the stent when the '7Lu is incorporated into the alloy, make this radioisotope a poor candidate for intravascular brachytherapy. For a radioactive stent in which the radioactive material is
an integral part of the metal alloy of the stent, a radioisotope with a much higher energy should be used for a significant dose to be delivered to the target tissue. The main limitation of the DPK model is the calculation of the self-attenuation of the beta emissions when the radioisotope is incorporated into the volume of the stent. This particular case could require the experimental determination of a self-attenuation factor.

The rapid fall-off dose of beta particles of radioisotopes used in this work results in the dose at the stent edges being about fifty percent less than the dose at the center of the stent. The low energy of the beta emissions does not allow adequate delivery of radiation dose to the critical targets. Nevertheless, the low gamma energy of 177 Lu produces a smooth reduction in the dose at the edges of the stent compared to the other radioisotopes studied. This effect, which in theory could reduce the candy wrapper effect, makes ¹⁷⁷Lu a potentially useful radioisotope to be incorporated at the edges of the stent. This also implies that the use of a gamma emitter at the stent edges could also be favorable.

A radiobiological assessment provides interesting insights on the dose delivered at a particular target point by different radioisotopes. It was shown that for the same physical dose delivered by ^{32}P , ^{90}Y , ^{188}Re and ^{177}Lu stents, the BED is quite different. Physical absorbed dose does not say much about the quality and characteristics of the dose. However, BED takes into account the rate at which the dose is delivered, the halflife of the radioisotope and the repair capability of the tissue. BED can be up to 100 times higher than physical absorbed dose. Results of the radiobiological model demonstrated that the same physical dose delivered by different radioisotopes could produce significantly different biological effects. The practical consequence of this effect is unknown and is worthy of further experimental investigation.

The dosimetry and radiobiological models developed in this work serve as a tool to characterize the dosimetry of a radioactive stent, and give an idea of the radiobiological effect. This type of theoretical dose calculation can be useful for the development of new stent designs in efforts to improve the radioactive stent technology, and in the planning of animal studies and clinical trials.

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APPENDIX A **Determination of the Spectrum of Beta Emissions of 17 Lu**

The beta spectrum of 177 Lu was determined according to the formalism of Dillman et al. (1969) and Prestwich et al. (1989). The disintegration of ¹⁷⁷Lu includes three beta emissions (Table A.1).

	Table A.1. Nuclear Properties of " Lu
Atomic Mass	177
Atomic Number	71
Beta Decay	0.498 MeV (79%)
	0.385 MeV (9.1%)
	0.177 MeV (12%)
Gamma Decay	0.208 MeV (11%)
	1.113 MeV (6.4%)

Table A.1. Nuclear Properties of **177Lu**

For each emission an individual spectrum was calculated by using the following equation (Prestwich et al., 1989):

$$
n(E_i) = CF(Z, W_i) P_i W_i (E_M - E_i)^2,
$$

where E_i is the energy bin, $n(E_i)$ is the relative number of beta emissions corresponding to the energy E_i . E_M is the maximum energy of the beta emission.

$$
W_i = E_i + m_o c^2 - V_o,
$$

in which $m_{o}c^{2}$ is the electron rest mass expressed in energy units (0.511 MeV) and V_{o} is the screened potential, given by:

$$
V_o = \frac{1,13 \frac{1}{Z} \Big|^{413}}{\alpha^2} = 0.0092 \text{ MeV},
$$

where Z is the atomic number of daughter nucleus $(Z = 72)$ and α is the fine structure constant (α = 1/137) (Evans, p. 601, 1982).

Pi is the screened momentum given by :

$$
P_i = \sqrt{W_i^2 - m_o c^4}.
$$

F(Z,Wi) **is the Fermi function (Evans, p.548, 1982) expressed by:**

$$
F(Z, W_i) = F_N(Z, W_i) \times F_2
$$

in which F_2 is given by:

$$
\left(\frac{\alpha^2 Z^2 w_i^2 + (w_i^2 - 1)}{4}\right)^s,
$$

$$
F_N(Z, W_i) = \frac{2\pi v_i}{\left(1 - e^{-2\pi v_i}\right)},
$$

$$
v_i = \frac{\alpha Z w_i}{\eta_i},
$$

$$
w_i = \frac{W_i}{m_o c^2},
$$

$$
\eta_i = \frac{P_i}{m_o c^2},
$$

and
$$
S = \sqrt{1 - \alpha^2 Z^2} - 1 = -0.1492
$$
.

C is a correction factor (Prestwich et al., 1989) given by:

$$
C = 1 + \left[a(Z - 50) + b[Z - 50]^2 \right] \times (1 - e^{-k v_i}),
$$

where $a = 4.05 \times 10^{-4}$

$$
b = 2.23 \times 10^{-5}
$$

$$
k = 1.26
$$

For calculating the beta spectrum, the maximum energy was divided into 100 energy bins. The relative number of emissions was determined for the average energy of each energy bin. Tables A.2, A.3 and A.4 show the calculations for each beta disintegration of 177 Lu. The graphs of the beta spectra are shown in Figures A.1, A.2 and A.3.

Energy										
bin number	E (MeV)	W	$\mathbf P$	W	η	ν	$\mathbf{F}_{\mathbf{n}}$	F ₂	$\mathbf C$	n(E)
$\bf{0}$	0.00249	511.0	3.20	1.0000	0.0063	84.0067	527.8307	1.2119	1.0197	0.02998
1	0.00747	511.1	9.04	1.0002	0.0177	29.7039	186.6355	1.2118	1.0197	0.02938
$\mathbf 2$	0.01245	514.3	57.77	1.0064	0.1131	4.6779	29.3923	1.2076	1.0169	0.02897
3	0.01743	519.2	92.11	1.0161	0.1803	2.9625	18.6140	1.2011	0.9953	0.02816
$\overline{\mathbf{4}}$	0.02241	524.2	117.0	1.0259	0.2289	2.3554	14.7993	1.1948	0.9673	0.02718
5	0.02739	529.2	137.6	1.0356	0.2692	2.0216	12.7019	1.1887	0.9399	0.02622
6	0.03237	534.2	155.6	1.0454	0.3046	1.8038	11.3337	1.1828	0.9146	0.02532
$\overline{7}$	0.03735	539.2	172.0	1.0551	0.3365	1.6478	10.3541	1.1771	0.8918	0.02450
8	0.04233	544.1	187.0	1.0648	0.3659	1.5294	9.6101	1.1715	0.8713	0.02374
$\boldsymbol{9}$	0.04731	549.1	201.0	1.0746	0.3934	1.4356	9.0215	1.1661	0.8526	0.02304
10	0.05229	554.1	214.2	1.0843	0.4193	1.3592	8.5418	1.1608	0.8358	0.02239
11	0.05727	559.1	226.8	1.0941	0.4439	1.2954	8.1418	1.1556	0.8204	0.02178
12	0.06225	564.1	238.8	1.1038	0.4674	1.2412	7.8021	1.1506	0.8063	0.02121
13	0.06723	569.0	250.4	1.1136	0.4899	1.1945	7.5094	1.1456	0.7933	0.02067
14	0.07221	574.0	261.5	1.1233	0.5117	1.1537	7.2541	1.1408	0.7814	0.02016
15	0.07719	579.0	272.2	1.1331	0.5328	1.1177	7.0291	1.1361	0.7703	0.01968
16	0.08217	584.0	282.7	1.1428	0.5532	1.0857	6.8292	1.1316	0.7601	0.01921
17	0.08715	589.0	292.8	1.1526	0.5730	1.0570	6.6501	1.1271	0.7505	0.01876
18	0.09213	593.9	302.7	1.1623	0.5924	1.0311	6.4887	1.1227	0.7416	0.01833
19	0.09711	598.9	312.4	1.1720	0.6113	1.0076	6.3424	1.1184	0.7332	0.01792
20	0.10209	603.9	321.8	1.1818	0.6298	0.9862	6.2091	1.1142	0.7254	0.01752
21	0.10707	608.9	331.1	1.1915	0.6479	0.9665	6.0870	1.1101	0.7180	0.01713
22	0.11205	613.9	340.1	1.2013	0.6656	0.9485	5.9747	1.1060	0.7111	0.01675
23	0.11703	618.8	349.1	1.2110	0.6831	0.9317	5.8712	1.1021	0.7045	0.01637
24	0.12201	623.8	357.8	1.2208	0.7002	0.9163	5.7753	1.0982	0.6983	0.01601
25	0.12699	628.8	366.4	1.2305	0.7171	0.9019	5.6863	1.0944	0.6924	0.01566
26	0.13197	633.8	374.9	1.2403	0.7337	0.8884	5.6034	1.0907	0.6868	0.01531
27	0.13695	638.8	383.3	1.2500	0.7500	0.8759	5.5260	1.0870	0.6815	0.01497
28	0.14193	643.7	391.5	1.2598	0.7662	0.8641	5.4535	1.0834	0.6765	0.01463
29	0.14691	648.7	399.6	1.2695	0.7821	0.8531	5.3855	1.0798	0.6716	0.01430
30	0.15189	653.7	407.7	1.2792	0.7978	0.8427	5.3216	1.0763	0.6671	0.01398
31	0.15687	658.7	415.6	1.2890	0.8133	0.8329	5.2614	1.0729	0.6627	0.01365

Table A.2. Spectrum calculation and distribution of the beta emission with maximum energy of 0.498 MeV (79%)

81	0.40587	907.7	750.2	1.7763	1.4680	0.6359	4.0703	0.9499	0.5621	0.00144
82	0.41085	912.7	756.2	1.7860	1.4798	0.6343	4.0609	0.9481	0.5612	0.00130
83	0.41583	917.6	762.2	1.7958	1.4916	0.6327	4.0516	0.9463	0.5603	0.00116
84	0.42081	922.6	768.2	1.8055	1.5033	0.6312	4.0426	0.9445	0.5594	0.00103
85	0.42579	927.6	774.2	1.8153	1.5150	0.6297	4.0338	0.9427	0.5585	0.00091
86	0.43077	932.6	780.1	1.8250	1.5266	0.6283	4.0252	0.9410	0.5577	0.00080
87	0.43575	937.6	786.1	1.8347	1.5383	0.6268	4.0168	0.9392	0.5568	0.00069
88	0.44073	942.5	792.0	1.8445	1.5499	0.6254	4.0085	0.9375	0.5560	0.00059
89	0.44571	947.5	797.9	1.8542	1.5615	0.6241	4.0005	0.9358	0.5552	0.00049
90	0.45069	952.5	803.8	1.8640	1.5730	0.6228	3.9927	0.9341	0.5544	0.00041
91	0.45567	957.5	809.7	1.8737	1.5846	0.6215	3.9850	0.9324	0.5537	0.00033
92	0.46065	962.5	815.6	1.8835	1.5961	0.6202	3.9775	0.9307	0.5529	0.00026
93	0.46563	967.4	821.5	1.8932	1.6076	0.6189	3.9701	0.9291	0.5522	0.00019
94	0.47061	972.4	827.3	1.9030	1.6190	0.6177	3.9629	0.9275	0.5515	0.00014
95	0.47559	977.4	833.2	1.9127	1.6305	0.6165	3.9559	0.9258	0.5508	0.000095
96	0.48057	982.4	839.0	1.9225	1.6419	0.6153	3.9490	0.9242	0.5501	0.000058
97	0.48555	987.4	844.8	1.9322	1.6533	0.6142	3.9423	0.9226	0.5494	0.000030
98	0.49053	992.3	850.7	1.9419	1.6647	0.6131	3.9357	0.9210	0.5487	0.000011
99	0.49551	997.3	856.5	1.9517	1.6760					
						0.6120	3.9292	0.9195	0.5481	0.000001

Figure A.1 Beta Spectrum of '"Lu with Maximum Energy of 0.498 MeV (79%)

Table A.3. Spectrum calculation and distribution of the beta emission with maximum energy of 0.385 MeV (9.1%)

Energy bin										
number	E (MeV)	W	$\mathbf P$	W	η	$\mathbf v$	$\mathbf{F}_{\mathbf{n}}$	$\mathbf{F_{2}}$	$\mathbf C$	n(E)
$\pmb{0}$	0.00249	511.0	3.20	1.0000	0.0063	84.0067	527.8307	1.2119	1.0197	0.03951
$\mathbf{1}$	0.00747	511.1	9.04	1.0002	0.0177	29.7039	186.6355	1.2118	1.0197	0.03849
$\mathbf 2$	0.01245	514.3	57.77	1.0064	0.1131	4.6779	29.3923	1.2076	1.0169	0.03772
3	0.01743	519.2	92.11	1.0161	0.1803	2.9625	18.6140	1.2011	0.9953	0.03644
4	0.02241	524.2	117.0	1.0259	0.2289	2.3554	14.7993	1.1948	0.9673	0.03494
5	0.02739	529.2	137.6	1.0356	0.2692	2.0216	12.7019	1.1887	0.9399	0.03349
$\boldsymbol{6}$	0.03237	534.2	155.6	1.0454	0.3046	1.8038	11.3337	1.1828	0.9146	0.03213
$\overline{7}$	0.03735	539.2	172.0	1.0551	0.3365	1.6478	10.3541	1.1771	0.8918	0.03087
8	0.04233	544.1	187.0	1.0648	0.3659	1.5294	9.6101	1.1715	0.8713	0.02970
$\boldsymbol{9}$	0.04731	549.1	201.0	1.0746	0.3934	1.4356	9.0215	1.1661	0.8526	0.02862
10	0.05229	554.1	214.2	1.0843	0.4193	1.3592	8.5418	1.1608	0.8358	0.02760
11	0.05727	559.1	226.8	1.0941	0.4439	1.2954	8.1418	1.1556	0.8204	0.02665
12	0.06225	564.1	238.8	1.1038	0.4674	1.2412	7.8021	1.1506	0.8063	0.02575
13	0.06723	569.0	250.4	1.1136	0.4899	1.1945	7.5094	1.1456	0.7933	0.02489
14	0.07221	574.0	261.5	1.1233	0.5117	1.1537	7.2541	1.1408	0.7814	0.02407
15	0.07719	579.0	272.2	1.1331	0.5328	1.1177	7.0291	1.1361	0.7703	0.02329
16	0.08217	584.0	282.7	1.1428	0.5532	1.0857	6.8292	1.1316	0.7601	0.02254
17	0.08715	589.0	292.8	1.1526	0.5730	1.0570	6.6501	1.1271	0.7505	0.02182
18	0.09213	593.9	302.7	1.1623	0.5924	1.0311	6.4887	1.1227	0.7416	0.02112
19	0.09711	598.9	312.4	1.1720	0.6113	1.0076	6.3424	1.1184	0.7332	0.02045
20	0.10209	603.9	321.8	1.1818	0.6298	0.9862	6.2091	1.1142	0.7254	0.01979
21	0.10707	608.9	331.1	1.1915	0.6479	0.9665	6.0870	1.1101	0.7180	0.01916
22	0.11205	613.9	340.1	1.2013	0.6656	0.9485	5.9747	1.1060	0.7111 0.7045	0.01854
23	0.11703	618.8	349.1	1.2110	0.6831	0.9317	5.8712	1.1021 1.0982	0.6983	0.01793 0.01734
24	0.12201	623.8	357.8	1.2208	0.7002	0.9163 0.9019	5.7753 5.6863	1.0944	0.6924	0.01676
25	0.12699	628.8	366.4	1.2305	0.7171	0.8884	5.6034	1.0907	0.6868	0.01620
26	0.13197	633.8	374.9	1.2403	0.7337	0.8759	5.5260	1.0870	0.6815	0.01564
27	0.13695	638.8	383.3	1.2500	0.7500	0.8641	5.4535	1.0834	0.6765	0.01510
28	0.14193	643.7	391.5	1.2598	0.7662 0.7821	0.8531	5.3855	1.0798	0.6716	0.01456
29	0.14691	648.7	399.6	1.2695	0.7978	0.8427	5.3216	1.0763	0.6671	0.01404
30	0.15189	653.7	407.7	1.2792 1.2890	0.8133	0.8329	5.2614	1.0729	0.6627	0.01352
31	0.15687	658.7	415.6 423.5	1.2987	0.8287	0.8237	5.2046	1.0696	0.6585	0.01302
32	0.16185	663.7	431.2	1.3085	0.8439	0.8149	5.1509	1.0662	0.6545	0.01252
33	0.16683	668.6	438.9	1.3182	0.8589	0.8066	5.1001	1.0630	0.6506	0.01203
34	0.17181	673.6	446.5	1.3280	0.8738	0.7987	5.0519	1.0598	0.6470	0.01155
35	0.17679	678.6	454.0	1.3377	0.8885	0.7912	5.0062	1.0566	0.6434	0.01108
36	0.18177	683.6 688.6	461.5	1.3475	0.9031	0.7841	4.9627	1.0535	0.6400	0.01061
37	0.18675	693.5	468.9	1.3572	0.9176	0.7773	4.9213	1.0505	0.6368	0.01015
38	0.19173 0.19671	698.5	476.2	1.3670	0.9320	0.7708	4.8818	1.0474	0.6336	0.00970
39	0.20169	703.5	483.5	1.3767	0.9462	0.7647	4.8442	1.0445	0.6306	0.00926
40	0.20667	708.5	490.7	1.3864	0.9603	0.7587	4.8082	1.0415	0.6277	0.00883
41			497.9	1.3962	0.9744	0.7531	4.7738	1.0387	0.6249	0.00840
42	0.21165	713.5								

Figure A.2 Beta Spectrum of 177Lu **with Maximum Energy of 0.385** MeV (9.1%)

Table **A.4. Spectrum calculation and distribution of the beta emission with maximum energy of** 0.177 **MeV (12.0%)**

Energy bin number	E(MeV)	W	P	W	η	v	F,	F ₂	$\mathbf C$	n(E)
$\bf{0}$	0.00249	511.0	3.20	1.0000	0.0063	84.0067	527.8307	1.2119	1.0197	0.08591
1	0.00747	511.1	9.04	1.0002	0.0177	29.7039	186.6355	1.2118	1.0197	0.08109
2	0.01245	514.3	57.77	1.0064	0.1131	4.6779	29.3923	1.2076	1.0169	0.07687
3	0.01743	519.2	92.11	1.0161	0.1803	2.9625	18.6140	1.2011	0.9953	0.07174
4	0.02241	524.2	117.0	1.0259	0.2289	2.3554	14.7993	1.1948	0.9673	0.06635
5	0.02739	529.2	137.6	1.0356	0.2692	2.0216	12.7019	1.1887	0.9399	0.06122
6	0.03237	534.2	155.6	1.0454	0.3046	1.8038	11.3337	1.1828	0.9146	0.05645
7	0.03735	539.2	172.0	1.0551	0.3365	1.6478	10.3541	1.1771	0.8918	0.05203
8	0.04233	544.1	187.0	1.0648	0.3659	1.5294	9.6101	1.1715	0.8713	0.04791
9	0.04731	549.1	201.0	1.0746	0.3934	1.4356	9.0215	1.1661	0.8526	0.04408
10	0.05229	554.1	214.2	1.0843	0.4193	1.3592	8.5418	1.1608	0.8358	0.04050
11	0.05727	559.1	226.8	1.0941	0.4439	1.2954	8.1418	1.1556	0.8204	0.03714
12	0.06225	564.1	238.8	1.1038	0.4674	1.2412	7.8021	1.1506	0.8063	0.03399
13	0.06723	569.0	250.4	1.1136	0.4899	1.1945	7.5094	1.1456	0.7933	0.03102
14	0.07221	574.0	261.5	1.1233	0.5117	1.1537	7.2541	1.1408	0.7814	0.02822
15	0.07719	579.0	272.2	1.1331	0.5328	1.1177	7.0291	1.1361	0.7703	0.02557
16	0.08217	584.0	282.7	1.1428	0.5532	1.0857	6.8292	1.1316	0.7601	0.02308
17	0.08715	589.0	292.8	1.1526	0.5730	1.0570	6.6501	1.1271	0.7505	0.02073
18	0.09213	593.9	302.7	1.1623	0.5924	1.0311	6.4887	1.1227	0.7416	0.01852
19	0.09711	598.9	312.4	1.1720	0.6113	1.0076	6.3424	1.1184	0.7332	0.01644
20	0.10209	603.9	321.8	1.1818	0.6298	0.9862	6.2091	1.1142	0.7254	0.01449
21	0.10707	608.9	331.1	1.1915	0.6479	0.9665	6.0870	1.1101	0.7180	0.01266
22	0.11205	613.9	340.1	1.2013	0.6656	0.9485	5.9747	1.1060	0.7111	0.01096
23	0.11703	618.8	349.1	1.2110	0.6831	0.9317	5.8712	1.1021	0.7045	0.00937

24	0.12201	623.8	357.8	1.2208	0.7002	0.9163	5.7753	1.0982	0.6983	0.00791
25	0.12699	628.8	366.4	1.2305	0.7171	0.9019	5.6863	1.0944	0.6924	0.00657
26	0.13197	633.8	374.9	1.2403	0.7337	0.8884	5.6034	1.0907	0.6868	0.00535
27	0.13695	638.8	383.3	1.2500	0.7500	0.8759	5.5260	1.0870	0.6815	0.00426
28	0.14193	643.7	391.5	1.2598	0.7662	0.8641	5.4535	1.0834	0.6765	0.00328
29	0.14691	648.7	399.6	1.2695	0.7821	0.8531	5.3855	1.0798	0.6716	0.00243
30	0.15189	653.7	407.7	1.2792	0.7978	0.8427	5.3216	1.0763	0.6671	0.00170
31	0.15687	658.7	415.6	1.2890	0.8133	0.8329	5.2614	1.0729	0.6627	0.00110
32	0.16185	663.7	423.5	1.2987	0.8287	0.8237	5.2046	1.0696	0.6585	0.00063
33	0.16683	668.6	431.2	1.3085	0.8439	0.8149	5.1509	1.0662	0.6545	0.00028
34	0.17181	673.6	438.9	1.3182	0.8589	0.8066	5.1001	1.0630	0.6506	$7.4E - 0.5$
35	0.17679	678.6	446.5	1.3280	0.8738	0.7987	5.0519	1.0598	0.6470	$1.2E-07$

Figure A.3 Beta Spectrum of '"Lu with Maximum Energy of 0.177 MeV **(12.0%)**

The complete **spectrum** including the three **beta emissions** was determined by adding the individual contributions of each beta **emission according** to the following **equation:**

$$
n(E_i) = 0.79n_1(E_i) + 0.091n_2(E_i) + 0.12n_3(E_i),
$$

where $n_1(E_v)$, $n_2(E_v)$ and $n_3(E_v)$ correspond to the spectrum emissions of the maximum **energies** of 0.498 MeV, 0.385 MeV and 0.177 MeV respectively. Table A.5 shows the

numerical data of the complete beta energy spectrum of ' 77 Lu. Figure A.4 shows the graph of the complete spectrum.

E (MeV)	n(E)
0.00249	0.03759
0.00747	0.03645
0.01245	0.03554
0.01743	0.03417
0.02241	0.03261
0.02739	0.03111
0.03237	0.02970
0.03735	0.02841
0.04233	0.02721
0.04731	0.02610
0.05229	0.02506
0.05727	0.02409
0.06225	0.02318
0.06723	0.02232
0.07221	0.02150
0.07719	0.02073
0.08217	0.02000
0.08715	0.01930
0.09213	0.01863
0.09711	0.01799
0.10209	0.01738
0.10707	0.01679
0.11205	0.01623
0.11703	0.01569
0.12201	0.01518
0.12699	0.01468
0.13197	0.01421
0.13695	0.01376
0.14193	0.01333
0.14691	0.01291
0.15189	0.01252
0.15687	0.01215
0.16185	0.01180
0.16683	0.01146
0.17181	0.01115
0.17679	0.01085
0.18177	0.01057
0.18675	0.01029
0.19173	0.01001

Table A.5. Complete beta energy spectrum of 17 Lu

E (MeV)	n(E)
0.43575	0.00054
0.44073	0.00046
0.44571	0.00039
0.45069	0.00032
0.45567	0.00026
0.46065	0.00020
0.46563	0.00015
0.47061	0.00011
0.47559	0.00007
0.48057	0.00005
0.48555	0.00002
0.49053	0.00001
0.49551	0.00000

Figure A.4 Beta Energy Spectrum of 17 Lu

APPENDIX B Calculation of Beta Dose Point Kernel of ¹⁷⁷Lu

The DPK for ¹⁷⁷Lu was calculated according to equation 9 (Chapter 3, p.). The **DPK of beta particles was derived from the dimensionless dose distribution** $j(r/R, E)$ tabulated by Cross et al. (1992), where:

$$
j(r/R, E) = 4\pi \rho r^2 \times DPK(r, E) \times R/E,
$$

in which ρ is the density of water equal to 1 g/cm^2 , r^2 is the distance from the point source **to the target point and R is the range of beta particles in water for a specific energy E. The DPK was calculated from the following equation:**

$$
DPK(r,E) = \frac{j(r/R,E)}{4\pi \rho r^2 R} \times E
$$

The range R was determined from the beta range tables in water (Attix, **Appendix E, p.575, 1986). DPK was determined at intervals of 25 keV. Those values of j(r,E) or range R, which were not available in the tables, were calculated by logarithmic linear interpolation. Table B-1 shows the contribution to the DPK of each energy bin at intervals** of 25 keV. **Table B-2 shows the value of the DPK of** 1 77Lu **at distances ranging up to 2 mm.**

Table B-1. Contribution of each energy bin to DPK of'77Lu for distances **ranging from** 0.01 to 0.2 cm

Energy <mark>I</mark> r (cm)																							
(MeV)	ā	Q.015	Q.02	0.025	S.	0.035	Mio	0.045	8.	0.055	ä	0.065	0.07	ADD	a.	0.085	00	COO1	្ធ	0.125	615	0.175	2
0.025					\bullet																		
0.050					0																		
Q100					0																		
OST	០០០ឬឆ្នំមិននៃ និដ្ឋ និង		o o o o 5 8 2 9 5 8 2 7 1	-----														------------		-------------	----------------	-----------	-------
0.125					\bullet																		
0.150					$2 = R$																		
0.175																							
0.200								្ឆ	1006														
0.25		倒 作 及 9 4 公 女 女 な		ちゅおああはのだ	₽		8 + = 2 8 8 8			0.018													
0.250					\overline{r} 5			$28.5 - 5.5 = 7$	$\frac{17}{5}$ = $\frac{19}{5}$ $\frac{29}{5}$ $\frac{29}{5}$ $\frac{25}{5}$	197	$\mathbf{3}$	ខ្ល	흥										
0.275										35	<u>হর</u>												0
0.300										34.5	23.74	85 a	$\overline{5}$ = $\overline{5}$									\circ \circ \circ	$\overline{}$
0.325					22 23	6.45 4.5 2.6						$\ddot{ }$				<u>ង្គ</u> ង នី	BE 25 85	SS	0.002				\bullet \bullet
		12	\blacktriangleright	္က	29					$\frac{53}{2}$		Ξ	85 08 0					0.046	0.002	Ξă			
	$\frac{8}{10}$	3.	$rac{5}{4}$	\mathbb{Z}	\mathfrak{F}_1		23 \approx $-$			ខ្ម	$-$ g	\mathbf{S}		ិ និ					B	អូ		$\overline{}$ $\overline{}$	\bullet
0.350 0.375 0.400	12	\bullet	332		\mathbf{r}	프 드		ន្ល	$\overline{0.74}$	ਲੂ		ន្ទ	$\frac{8}{10}$					<u>ទី ទី ទី</u>	\tilde{c}	COD			0
0.45	553	25	$\ddot{11}$	\sim $\frac{12}{15}$	္ထိ	\mathbf{S}	3.0	$\mathbf{3}$	53	\mathbf{g}	3	50	នី	\tilde{c}	3	ី	ះ		5 ₁	ទី	ੱ ਮੌ	\blacksquare	0
0.450		\mathbf{r}	$\mathbf{5}$	50	\overline{a}	027	0.24	$\frac{9}{2}$	51	$\frac{4}{10}$	ី	등	5	g	g	go	go	go	go	0.001	7E-04	Š	\bullet
0.475	80	ន	\mathbf{c}	50 ₀	g	0.065	0.055	SE	ECO	0.03	ශ	g	\mathbf{g}	0.023	g	ខ្ល	0.019	0.017	0.015	0.07	0.002	1E-04	\$
Newg	1465.5	802.7	Ŕ,	198.5	11.8 ≂	69.08	4.94	33	20.963 [15.15]		11.11	<u>757</u>	5.18	230	2356	1.63	1.064	A.SOS	ij	MS	QUO	3E.04	45.05

			$nGy-cm^2/$
r (cm)	MeV/g	nGy/Bq-h	Bq-h
0			212.51
0.01	1465.6	845240.8	84.52408
0.015	802.7	462933.1	104.16
0.02	356.63	205675.7	82.27026
0.025	198.5	114478.9	71.54933
0.03	111.83	64494.6	58.04514
0.035	69.1	39851.35	48.81791
0.04	41.9	24164.57	38.66331
0.045	29.3	16897.9	34.21824
0.05	20.9	12053.45	30.13362
0.055	15.2	8766.144	26.51759
0.06	11.11	6407.359	23.06649
0.065	7.52	4336.934	18.32355
0.07	5.18	2987.41	14.63831
0.075	3.39	1955.081	10.99733
0.08	2.36	1361.059	8.710779
0.085	1.63	940.0536	6.791887
0.09	1.064	613.6301	4.970404
0.095	0.606	349.4923	3.154168
0.1	0.29	167.2488	1.672488
0.125	0.0356	20.53123	0.320801
0.15	0.00245	1.412964	0.031792
0.175	0.00028	0.161482	0.004945
0.2	0.00004	0.023069	0.000923

Table B-2. Beta DPK of ¹⁷⁷Lu

Appendix C Calculation of Gamma DPK of '"Lu

Tables C-1 and C-2 present the results of the logarithmic linear interpolation that was done to obtain the DPK values for gamma emissions of ¹⁷⁷Lu with energies of .208 MeV and 0.113 MeV for a given distance d. The interpolation was done using the values provided in the tables by Luxton and Jozsef (1999). Table C-3 provides the final DPK of ¹⁷⁷Lu obtain by summing up the DPK for both energies multiplied by the fraction of emissions according to equation 11 (Chapter 3).

d (cm)	0.200	0.250	0.208
	MeV	MeV	MeV
0.05	0.2852	0.3648	0.296657
0.1	0.28	0.3629	0.291863
0.2	0.2736	0.357	0.285499
0.24	0.2743	0.3545	0.285791
0.3	0.2693	0.3626	0.282428
04	0.2779	0.3699	0.290911
0.5	0.2719	0.361	0 284515
0.6	0.2803	0.3598	0.291725
0.8	0.2811	0.363	0.292838
0.9	0.2861	0 3648	0.297443
	0 286	0.364	0.297251
1.11	0 2883	0.3688	0.299886
$1.2\,$	0.2875	0.3745	ი 299922
15	0.2925	0.3706	0 303788
2	0.2985	0.3763	0.30977

Table C-1. Dose to water per gamma ray emission from a point source (nGycm²/Bq-hr) for **gamma energy 0.208 MeV (11.0%)**

	0.100	0.125	0.113
d (cm)	MeV	MeV	MeV
0.05	0.126	0.1559	0.140753
0.1	0.1231	0.1514	0.137085
0.2	0.1226	0.1519	0.137052
0.24	0.1206	0.1584	0.138969
03	0.1226	0.1563	0.139102
0.4	0.1244	0.161	0.142254
0.5	0.1223	0.1602	0.140731
0.6	0.1344	0.1574	0.145906
0.8	0.1313	0.1671	0.148838
0.9	0.1344	0.1675	0.150702
1.0	0.1347	0.1674	0.150817
1.11	0.1367	0.1708	0.153484
1.2	0.1371	0.175	0.155653
1.5	0.1418	0.1786	0.159876
2.0	0.152	0.1823	0.167068

Table C-2. Dose to water per gamma ray emission from a point source (nGycm²/Bq-hr) for **gamma energy 0.113 MeV (6.4%)**

Table C-3. Gamma DPK of 17 Lu

	DPK.
d (cm)	$(nGy-cm^2/Bq-hr)$
0.05	0.041818
0.1	0.041053
0.2	0 040348
0.24	0.040503
0.3	0 040139
0.4	0.041279
0.5	0.040474
0.6	0.041603
0.8	0.041914
0.9	0.042542
1	0.042528
1.11	0.04299
1.2	0.043133
15	0 043831
2	0.044953

APPENDIX D Stent Coordinates

The following table provides the set of Cartesian coordinates of the stent geometry that was used as the radiation source in this project. The stent at 15.5 mm in length consists of four repeating segments that are joined together to obtain a stent of this specific desired length. Table C.1 shows the set of (x,y,z) coordinates for this segment, which are equivalent to one fourth of the total length of the stent.

Table D-1 Stent Coordinates

APPENDIX E **Code for Dose Calculation of a Radioactive Stent with One Radioisotope**

Part A. represents the code for the calculation of the dose surrounding a stent coated with a pure beta emitter such as $3^{2}P$. Part B shows the code of a beta and gamma emitter such as 177 Lu. Note that the final dose is given by the sum of the dose given by both beta and gamma DPK. Both beta and gamma components are multiplied by their respective attenuation factors.

A.

- % DOSE CALCULATION ALGORITHM FOR RADIOACTIVE STENTS
- % DOSE POINT KERNEL CONVOLUTION
- % Author: Maria Alejandra Caceres
- % Date: May 1, 2003

clear

close all

% Input Source Points

% Source points are the coordinates defining the geometry of the stent and are this algorithm's INPUT

 $\%$ The coordinates are Cartesian coordinates (x,y,z) and are in a Microsoft Excel format

% Can include option to select only several slices for a certain radioisotope

tic

xlsread source; source=ans; $xs = source(:,1);$ $ys = source(:,2);$ $zs=source(:,3);$

% Target points are defined in the First Quadrant Only (for computation efficiency purposes).

% The assumption therefore is that the dose is symmetrical **in all** quadrants.

% Loop to find x and y target points

 $a=1$;

for $z=12:0.2:12$; for *y=0:0.2:5;* for x=2.1026:0.2:5.1026; $target(a, :, :,)=[a x y z];$ **a=a+1;**

```
end
     end
  end
 xt=target(:,2);
 yt=target(:,3);
 zt=target(:,4);
  % Dose calculation
  % P-32 tl/2= 14.28 days(342.72 hours)
  % lambda=\ln 2/t(1/2)% Initial Activity = 37 kBq = 1 microCi
  nsource=size(xs);
  ntarget=size(xt);
  sumdose=zeros(ntarget); % allocate empty matrix to store dose values
  for I=1:ntarget;
    for j=1:nsource;
       dis=sqrt((xt(i)-xs(j))^2+(yt(i)-ys(j))^2+(zt(i)-zs(j))^2);
       if dis <= 6.68
          DPK=((-0.0161. *(dis.^5))+(0.0866. *(dis.^4))+(1.8508.*(dis.^3))-
(15.253. *(dis.^2))+(9.889. *dis)+104.6)./dis.^2;
          'warning off MATLAB:divideByZero'
          sumdose(i)=sumdose(i)+DPK;
       end
     end
  end
  dose=sumdose.*(0.37/(0.00202*1840)); %unit conversion factor
```

```
DOSE=[xt yt zt dose];
```
% PLOT **for dose along stent axis for different depths**

B.

% DOSE CALCULATION ALGORITHM FOR RADIOACTIVE STENTS Lu-177 **% DOSE** POINT KERNEL CONVOLUTION for both BETA and GAMMA **energies**

% Author: **Maria Alejandra Caceres**

% Date: June 1, 2003

clear

close all % Input Source Points

% Source points are the coordinates defining the geometry of the stent and is this algorithm's INPUT

% The coordinates are Cartesian coordinates (x,y,z) and are in a Microsoft Excel format

% Can include option to select only several slices for a certain radioisotope

tic

```
xlsread source2;
source2=ans;
xs = source2(:, 1);ys = source2(:,2);zs=source2(:,3);xlsread lu177;
```
 $lu177=ans;$ $r=lu177(:,1);$ dpk=lu177(:,2);

% Target Points

% Target points are defined in the First Quadrant Only (for computation efficiency purposes)

% The assumption therefore is that the dose is symmetrical in all quadrants.

```
% Loop to find x and y target points
```

```
a=1;
```

```
for z=0;
  for y=0;
```

```
for x=2.10407 7 5 :0.1: 7 .
10407 7 5 ;
```

```
target(a, \ldots, \ldots)=[a x y z];
```

```
a=a+1;
```
end

end

end

```
xt = target(:,2);
```

```
yt = target(:,3);
```

```
zt=target(:,4);
```
- % Dose calculation using Lu-177
- % Lu-177 t1/2= 6.734 days=161.6160 hours
- % lambda=ln 2/t(1/2)=0.693/161.6160=0.0043
- % Initial Activity = 37 kBq = 1 microCi
- % # of struts= 1840
- $%$ unit conversion factor, $k=0.37$

```
nsource=size(xs);
ntarget=size(xt);
```

```
sumdose1=zeros(ntarget); % allocate empty matrix to store dose values
```

```
%BETA DPK
 for i=1 : ntarget;
    for i=1: nsource;
      dis1=sqrt((xt(i)-xs(j))^2+(yt(i)-ys(j))^2+(zt(i)-zs(j))^2);if dis1 \leq 2.0DPKB=interp1(r,dpk,disl,linear');'warning off MATLAB:divideByZero';
         DPKB=DPKB./dis1.^2;
         sumdose1(i)=sumdose1(i)+DPKB;
       end
    end
  end
  beta=sumdosel.*(0.37/(0.0043*1840))*0.28; %unit conversion factor
  nsource=size(xs);
  ntarget=size(xt);
  sumdose2=zeros(ntarget); % allocate empty matrix to store dose values
  %GAMMA DPK
  for i=1: ntarget;
    for j=1:nsource;
       dis2=sqrt((xt(i)-xs(j))^2+(yt(i)-ys(j))^2+(zt(i)-zs(j))^2);
       dis2 = dis2./10;DPKG = ((0.007.*(dis2.^6))-(0.0447.*(dis2.^5))+(0.1107.*(dis2.^4))-
(0.1377. * (dis2.^3))+(0.0896. * (dis2.^2))-(0.0253. * (dis2))+0.0428)./dis2.^2;
               'warning off MATLAB : divideByZero';
       sumdose2(i)=sumdose2(i)+DPKG;
     end
  end
  gamma=sumdose2.*(0.37/(0.0043*1840))*0.991; %unit conversion factor
  dose=beta+gamma;
  DOSE=[xt yt zt dose];
  doserate=dose.*(0.0043);
  RE=1+doserate./((1.386+0.0043)*0.08);
  BED=dose. *RE;
  table=[dose BED];
```
% PLOT for dose along stent axis for different depths

I=zt(1:26: size(zt)); m=dose(1:26: size (xt));n=dose(2:26: size (xt));o=dose(3 :26:size (xt));p=dose(6:26:size (xt)); q=dose(11:26:size (xt)); plot(l,m,'b-', l,n,'r-', l,o,'m-', l,p,'y-', l,q,'g-') grid on xlabel('Z(mm)') ylabel('Dose (cGy/uCi)') title ('Dose along stent axis') legend('O.1mm', '0.3 mm', '0.5 mm', '1.0 mm', '2.0 mm')

APPENDIX F Code for Dose Calculation of a Radioactive Stent with Two Radioisotopes

% DOSE CALCULATION ALGORITHM FOR HYRBID (More than one radioisotope) RADIOACTIVE STENT

% DOSE POINT KERNEL CONVOLUTION

% Author: Maria Alejandra Caceres

% Date: May 21, 2003

clear

close all

% Input Source Points

% Source points are the coordinates defining the geometry of the stent and is this algorithm's INPUT

 $\%$ The coordinates are Cartesian coordinates (x,y,z) and are in a Microsoft Excel format

% Can include option to select only several slices for a certain radioisotope

tic

xlsread source;

source=ans;

% Selection of slices of source for a specific radioisotope

%Selection of slices corresponding to the body of the stent

 $s1=source(461:1380,...,c)$;

%Selection of slices corresponding to the stent edges

 $s2=[source(1:460,:,:,:);source(1381:1840,:,:,:)]$;

 $xsl=sl(.1);$ $\text{ys1} = \text{sl}(.2);$ $zsl=sl(.3);$ $xs2=s2(:,1);$ $ys2= s2(:,2);$ $zs2= s2(:,3);$

% Target Points

% Target points are defined in the First Quadrant Only (for computation efficiency purposes)

% The assumption therefore is that the dose is symmetrical in all quadrants.

```
% Loop to find x, y, z target points
a=1;
for z=-10:0.2:10;
   for y=0:0.2:5;
      for x=2.1026:0.2:5.1026;
        target(a, \ldots)=[a x y z];
        a=a+1;
      end
   end
end
```

```
xt = target(:,2);yt = target(:,3);zt=target(:,4);%Dose Point Kernel Convolution
  ns1=size(xs1):
  ntarget=size(xt);
  sumdose1=zeros(ntarget); % allocate empty matrix to store dose values
  %Isotope in body of the stent (P-32)
  for i=1:ntarget;
     for j=1:ns1;
        dis I =sqrt((xt(i)-xs 1(j))^2+(yt(i)-ys1(j))^2+(zt(i)-zs1 (j))^2);
        if dis1 < 6.5DPK1=((0.009.*(dist.^6))-(0.2008.*(dist.^5))+(1.5189.*(dist.^4))-
(3.2879.*(dis1.^{3}))-(6.8498.*(dis1.^{2}))+(4.7765.*dis1)+105.12)./dis1.^2;
  %'warning off MATLAB :divideByZero'
          sumdose1(i)=sumdose1(i)+DPK1;
        end
     end
  end
  r = sumdose1.*(0.2);ns2=size(xs2);ntarget=size(xt);
  sumdose2=zeros(ntarget);
  %Isotope in stent edges (Y-90)
     for i=1: ntarget;
        for k=1:ns2;
          dis2=sqrt((xt(i)-xs2(k))^2+(yt(i)-ys2(k))^2+(zt(i)-zs2(k))^2);
          if dis2 < 8.0DPK2= ((0.001.*(dis2.^{6}))-(0.0331.*(dis2.^{5}))+(0.4201.*(dis2.^{4}))-
(2.1246. *(dis2.^3))+(2.6294. *(dis2.^2))-(7.8638. *dis2)+101.4)./dis2.^2;
               sumdose2(i)=sumdose2(i)+DPK2;
          end
        end
     end
     q=sumdose2.*(3.72*10^{\circ}–2);sumdose=r+q;
   DOSE=[xt yt zt sumdose];
```
%Plot 1: Dose along stent axis at different radial distances from stent edge figure;

```
c=zt(1:416:size(xt)); b=sumdose(1:416: size (sumdose));d=sumdose(2:416: size (sumdose));e=sumdose(3:416: size (sumdose));
                                      (sumdose));f=sumdose(6:416:size (sumdose));
g=r(11:416:size (sumdose));
   plot(c,b,'b-', c,d,'r-', c,e,'m-', c,f,'y-', c,g,'g-')
   grid on
   xlabel('Z(mm)')
   ylabel('Dose (cGy)')
   title ('Dose along stent axis')
   legend('0.1mm', '0.3 mm', '0.5 mm', '1.0 mm', '2.0 mm')
   %Plot 2: Dose as a function of radial distance calculated at three
   %stent planes: a)z=0, b)z=4 c)z=8figure;
   z1=0.1:0.2:3.1;a=sumdose(20801:20816);
   b=sumdose(29121:29136);
   c=sumdose(3 7025:37040);
   plot(zl,a,'b-s', zl,b,'r-.', zl,c,'g-x')
   grid on
   xlabel('distance (mm)')
   ylabel('Dose (cGy)')
   title('Dose as a function of radial distance at different z planes')
   legend ('z=0 mm', 'z=4 mm', 'z=8 mm')
```
APPENDIX G **Code for Calculation of Biological Equivalent Dose (BED) for Radioactive Stents with One Radioisotope**

- **% Radiobiological Model for P-32 stent**
- **% Author: Maria Alejandra Caceres**
- **% Date: May 25, 2003**
- **% Repair half-time is 0.5 hours** (30 min.)
- **% alpha/beta ratio is 8 Gy**
- **% lambda is 0.00202 h"1**
- **% repair half-life of tissue u=1.38 h-1**

clear

close all tic

load dose;

```
%Dose rate is equal to total dose x decay constant (hours)
doserate=dose. *(0.00202);
% Relative effectiveness per unit dose (RE)
RE=1 +doserate./((1.386+0.00202)*0.08);
```

```
%Calculation of Biological Equivalent Dose
BED=dose. *RE;
table=[dose BED];
```
toc t=toc

APPENDIX H **Code for Calculation of Biological Equivalent Dose (BED) for Radioactive Stents with Two Radioisotopes**

- **% Radiobiological Model for stent containing two radioisotopes**
- % Author: **Maria Alejandra Caceres**
- **% Date:** May 25, 2003
- **% Repair half-time is 0.5 hours (30 min.)**
- **% alpha/beta ratio is 8 Gy** (.08 cGy)

clear

close all tic

load dose;

%Dose rate is equal to total dose x decay constant of radioisotope(hours) $r=DOSE(:,4);$ **%a=dose from isotope1 (P-32) %b=dose from isotope2** (Lu-177) %L1=decay **constant of P-32 %L2=decay constant** of Lu-177 %P-32 **doserate=a. *(0.00202);** %Lu-177 **doseratel=b.** *(0.00428); **u=1.386** L1=0.00202 L2=0.00428

%Relative effectiveness per unit dose RE=1 +(0.08 *((doseratel .^2./(L 1. ***(L[+u)))+(doserate2.^2./(L2. *(L2+u)))+(2*(dosera tel. *doserate2.** *(L1+L2+2. *u))./((L1+L2). *(L1+u). ***(L2+u)))./((doseratel ./L1)+(dose** rate2./L2)))

```
%Calculation for Biologically Equivalent Dose
BED=r. *RE;
table=[r BED];
```
toc t=toc