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Multi-Tissue Stable Isotope Analysis and Acoustic Telemetry Reveal Seasonal Variability in the Trophic Interactions of Juvenile Bull Sharks in a Coastal Estuary

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- 1 Multi-tissue stable isotope analysis and acoustic telemetry reveal seasonal variability in the
- 2 trophic interactions of juvenile bull sharks in a coastal estuary

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- 8 Running head: seasonal changes in bull shark diets
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12 Summary

- Understanding how natural and anthropogenic drivers affect extant food webs is critical
 to predicting the impacts of climate change and habitat alterations on ecosystem
 dynamics.
- In the Florida Everglades, seasonal reductions in freshwater flow and precipitation lead to
 annual migrations of aquatic taxa from marsh habitats to deep-water refugia in estuaries.
 The timing and intensity of freshwater reductions, however, will be modified by ongoing
 ecosystem restoration and predicted climate change.
- 3. Understanding the importance of seasonally pulsed resources to predators is critical to
 predicting the impacts of management and climate change on their populations. As with
 many large predators, however, it is difficult to determine to what extent predators like
 bull sharks (*Carcharhinus leucas*) in the coastal Everglades make use of prey pulses
 currently.

4. We used passive acoustic telemetry to determine whether shark movements responded to
the pulse of marsh prey. To investigate the possibility that sharks fed on marsh prey, we
modeled the predicted dynamics of stable isotope values in bull shark blood and plasma
under different assumptions of temporal variability in shark diets and physiological
dynamics of tissue turnover and isotopic discrimination.

5. Bull sharks increased their use of upstream channels during the late dry season, and
although our previous work shows long-term specialization in the diets of sharks, stable
isotope values suggested that some individuals adjusted their diets to take advantage of
prey entering the system from the marsh, and as such this may be an important resource
for the nursery.

35	6.	Restoration efforts are predicted to increase hydroperiods and marsh water levels, likely
36		shifting the timing, duration, and intensity of prey pulses, which could have negative
37		consequences for the bull shark population and/or induce shifts in behavior.
38	7.	Understanding the factors influencing the propensity to specialize or adopt more flexible
39		trophic interactions will be an important step in fully understanding the ecological role of
40		predators and how ecological roles may vary with environmental and anthropogenic
41		changes.
42		
43	Key w	ords: δ^{13} C, food webs, habitat use, movements, shark nursery, trophic ecology

45 Introduction

Coastal ecosystems experience considerable daily and seasonal variation in 46 environmental conditions (Lewis 2001; Kennish 2002). Also, they have been, and continue to 47 be, heavily influenced by human activities that have contributed to shifts in community 48 composition and have potentially altered the ecological roles of species (e.g. Cloern 2001; 49 50 Jackson et al. 2001; Parmesan & Yohe 2003). Within coastal ecosystems, predators serve important roles in controlling prey populations, linking disparate food webs, and transporting 51 biomass and nutrients across habitat boundaries (e.g. Bowen 1997; Darimont, Paquet & 52 53 Reimchen 2009). Thus, understanding how predators are affected by temporally variable and ephemeral food sources is important for understanding the trophic dynamics of a system. 54 However, this can be challenging, because predators are often highly mobile with relatively large 55 56 home ranges. Consequently, manipulative studies can be difficult to execute and/or lead to biased results. Data quantifying behavioral variability in response to natural variation in food 57 sources can provide valuable insight in the roles predators play. In addition, there is a growing 58 need to understand how both natural and anthropogenic factors influence variability in trophic 59 interactions to predict how they may affect the ecological roles of species and ultimately 60 61 ecosystem dynamics.

Seasonal changes in temperature, precipitation, and freshwater flow lead to noticeable
variation in the distribution, abundance, and behavior of many resident and migratory species in
the Florida Coastal Everglades (Chick, Ruetz & Trexler 2004; Ruetz, Trexler & Jordan 2005;
Rehage & Trexler 2006). Therefore, trophic interactions are likely to vary in space and time as
predators and prey move to stay within suitable environmental conditions or to take advantage of
seasonal pulses of prey. These seasonal pulses of prey occur in the coastal Everglades when

water levels in freshwater marshes drop and numerous aquatic taxa are forced into deep-water 68 channels (Rehage & Trexler 2006; Rehage & Loftus 2007). The magnitude and timing of these 69 pulses are likely to be affected by ecosystem restoration. Freshwater flow is predicted to 70 increase through freshwater marshes, likely reducing the duration and intensity of marsh dry-71 down (Sklar et al. 2001; Perry 2004; CERP 2006), and therefore the magnitude and timing of 72 73 resource pulses into creeks. Thus, understanding the value of this resource pulse in the trophic ecology of estuarine predators will be important for predicting the consequences of restoration 74 efforts within the ecosystem. 75

76 Stable isotope analysis has become a popular method used in ecological studies of food webs to investigate trophic interactions (reviewed by Layman et al. 2012). Because the materials 77 eaten by an animal are not immediately incorporated into its tissues, stable isotope values 78 79 provide dietary data over a previous timeframe based on the isotopic turnover rate of the sampled tissue(s) (Gannes, O'Brien & Martinez del Rio 1997; Post 2002; Martinez del Rio et al. 2009). 80 This lag time can provide a means to investigate the temporal variability in the diet of an 81 organism by serially sampling parts of metabolically inert tissues or comparing the isotopic 82 values of multiple tissues with different turnover rates (Bearhop et al. 2004). 83

Metabolically inert tissues, like vibrissae in California sea otters (*Enhydra lutris nereis*), provide dietary information about a particular time period or event in the life of an organism, and sequentially sampling inert tissues can provide a dietary record for an organism over its lifetime (Newsome et al. 2009). Unfortunately, many animals do not have easily accessible tissues that can be used for serial sampling. One alternative to sequentially sampling metabolically inert tissues is to sample metabolically active tissue(s) from animals over multiple time periods and quantify the variability in isotope values over time (Bearhop et al. 2004). This can be an effective method when individuals are easily recaptured (e.g. Drago et al. 2010). But many
ecosystems are open and animals can move across habitat boundaries, making it difficult to
regularly sample the same individuals over time, and limiting the use of this approach.

A third strategy to investigate temporal change in diets is to compare the isotopic values 94 of metabolically active tissues with significantly different turnover rates collected during one 95 sampling event. A tissue with a fast isotopic turnover rate like blood in Japanese quail (Coturnix 96 *japonica*, δ^{13} C half-life = 11.4 days), provides information on recent trophic interactions, while a 97 tissue with a slower turnover rate like bone collagen (δ^{13} C half-life = 173.3 days in *C. japonica*), 98 provides a view of the average trophic interactions over an extended time period (Hobson & 99 Clark 1992). If the isotope values of a fast turnover tissue are compared to the isotope values of 100 a slow turnover tissue, the difference in isotope values can provide insight into the temporal 101 102 variability of an organism's trophic interactions. Comparing the isotope values of multiple 103 metabolically active tissues, however, must be conducted carefully because the values of stable isotopes in the tissues of a consumer are not identical to that of their food due to isotopic 104 105 discrimination (Gannes, O'Brien & Martinez del Rio 1997), and different tissues from the same organism can have different discrimination factors (e.g. Vander Zanden & Rasmussen 2001; 106 Sweeting et al. 2007; Buchheister & Latour 2010). Thus, understanding the isotopic 107 discrimination values of the tissues being used is important when investigating temporal 108 variability in trophic interactions. 109

While stable isotope analysis provides useful information on the trophic interactions of animals, data from complimentary approaches strengthen inferences about the trophic ecology of individuals and populations. Acoustic telemetry, for example, provides data on the movements of tagged animals, and when paired with stable isotope analysis, can provide a powerful tool for

114 elucidating individual- and population-level patterns linking habitat use and diet (e.g.

Papastamatiou et al. 2010; Rosenblatt & Heithaus 2011; Speed et al. 2012). Here, we used a

116 combination of long-term, passive acoustic tracking, and stable isotopic analysis and modeling

using blood plasma (faster turnover tissue) and whole blood (slower turnover tissue) δ^{13} C values

to investigate whether juvenile bull sharks make use of seasonal prey pulses in the coastal

- 119 Everglades.
- 120

121 Methods

122 Study species and system

Bull sharks (Carcharhinus leucas; Müller & Henle 1839) inhabit coastal and estuarine 123 waters of the tropics and subtropics around the world, and use coastal estuaries as nurseries 124 125 during early years before moving into coastal ocean habitats (Wiley & Simpfendorfer 2007, Grubbs 2010). Bull sharks can travel between fresh and marine waters with minimal metabolic 126 costs, and young individuals can be found in salinities ranging from 0.2-41.7 parts per thousand 127 128 (Anderson et al. 2006; Steiner, Michel & O'Donnell 2007; Heupel & Simpfendorfer 2008). As a result, bull sharks can take advantage of a variety of prey types, including teleosts, crustaceans, 129 cephalopods, and other elasmobranchs in marine, brackish, and freshwater habitats (Snelson & 130 Williams 1981; Snelson, Mulligan & Williams 1984; O'Connell et al. 2007). 131 The Shark River Estuary of Everglades National Park, Florida, USA (Fig. 1) is primarily 132

a braided stream system lined by mangroves that extends more than 30 km upstream from the

134 Gulf of Mexico. The estuary serves as a nursery for juvenile bull sharks year-round, which are

found throughout the entire system (Wiley & Simpfendorfer 2007; Heithaus et al. 2009; Matich

136 & Heithaus 2012). Seasonal changes in precipitation and freshwater flow lead to noticeable

137 variation in the distribution, abundance, and behavior of many resident and migratory species in 138 the Florida Coastal Everglades, including the Shark River Estuary (Chick, Ruetz & Trexler 2004; Ruetz, Trexler & Jordan 2005; Rehage & Trexler 2006). Therefore, trophic interactions vary in 139 140 space and time annually as predators and prey move to stay within acceptable environmental conditions and/or to take advantage of seasonal pulses of prey. Seasonal pulses of freshwater 141 prey into mangrove-lined creeks in the upstream region of the Shark River Estuary occur when 142 water levels in freshwater marshes drop during the dry season (Rehage & Trexler 2006; Rehage 143 & Loftus 2007; Fig. 1), and teleost predators rely on this prey pulse as an important seasonal 144 145 component of their diets (Boucek & Rehage *in press*). Our previous work revealed that juvenile bull sharks have relatively high levels of individual dietary specialization in the Shark River 146 Estuary (Matich, Heithaus & Layman 2011). Yet, stable isotope analysis revealed that some 147 148 individuals (ca. 13%) exhibit temporal variability in their trophic interactions, possibly driven by use of this seasonal pulse of marsh prey. 149

Marsh water levels serve as a seasonal indicator for when taxa migrate from the marsh 150 151 into deep-water refuges. For the purposes of our analyses, we used water level data from United States Geological Survey water station 252820080505400 Everglades National Park 152 (N25°28'20", W80°50'54"; Fig. 1) adjacent to our study system. When marsh water elevation 153 drops below 10 cm in depth in reference to elevation, the marsh becomes unsuitable for large 154 aquatic taxa (> 8 cm), which are forced to seek out deep-water habitat. As such, the dry season, 155 156 in reference to water levels, occurs when marsh water elevations are less than 10 cm and the wet season occurs when water levels are greater than 10 cm. These thresholds have been used in 157 studies of movements of Everglades marsh taxa into estuarine creeks (e.g. Chick, Ruetz & 158 159 Trexler 2004; Rehage & Loftus 2007; Parkos, Ruetz & Trexler 2011). Because the abundance of

marsh prey within mangrove-lined creeks changes considerably within seasons (Rehage &
Loftus 2007), we further divided each season into sub-seasons (i.e. early and late dry seasons,
and early and late wet seasons). During our study, the late wet season of 2008/2009 ended on 29
Feb 2009, and the early dry season was from 1 Mar to 13 Apr 2009. The late dry season was
from 14 Apr to 28 May 2009, the early wet season was from 29 May to 16 Oct 2009, and the late
wet season began 17 Oct 2009.

Within the confines of the Shark River Estuary, there are two isotopically distinct food 166 webs - freshwater/estuarine ($\delta^{13}C < -25\%$) and marine ($\delta^{13}C > -19\%$; Frv & Smith 2002; Chasar 167 et al. 2005; Williams & Trexler 2006; Fig. 2). Marsh taxa that enter the estuary during the dry 168 season prey pulse have more depleted δ^{13} C values (mean ± SE = -30.5 ± 0.5%) than resident 169 freshwater/estuarine taxa (mean \pm SE = -28.0 \pm 0.5%; Matich & Boucek unpublished data). 170 These differences in the δ^{13} C values of potential prey species enabled us to investigate seasonal 171 shifts in bull shark diets between prey with different basal carbon sources, and the potential use 172 of the freshwater prey pulse, by quantifying temporal variability in the δ^{13} C values of bull shark 173 174 tissues.

175

176 Field Sampling

Bull sharks were captured from Oct 2008 to Mar 2012 on ~500 m longlines, fitted with 40-55 14/0 or 15/0 Mustad tuna circle hooks baited with mullet (*Mugil* sp.) and attached to ~2 m of 400 kg monofilament line (see Heithaus et al. 2009 for further details of sampling equipment). Captured sharks were processed alongside the sampling vessel, or within a water-filled, aerated cooler on board. Shark total length was measured to the nearest centimeter. An 18 gauge needle was used to collect 4 mL of blood from the caudal vein. During collection, 3 mL of blood was 183 placed into BD Vacutainer blood collection vials with neither additives nor interior coating, and 184 then immediately separated into its components, including plasma, using a centrifuge spun for one minute at 3000 rpm. The remaining 1 mL of blood was retained in its original composition 185 186 (whole blood, "blood" hereafter). Based on several lab studies, plasma has an isotopic half-life of ~32 days in elasmobranchs (Kim et al. 2012), and likely serves as a short-term diet indicator 187 for juvenile bull sharks, while blood has an isotopic half-life of ~61 days (MacNeil, Drouillard & 188 Fisk 2006), and likely serves as a longer-term diet indicator for juvenile bull sharks. As such, 189 when more dynamic plasma δ^{13} C values are compared to more stable blood δ^{13} C values they can 190 be used to study short-term changes in the diets of sharks, and provide diet information over the 191 time-frame juvenile bull sharks may respond to the pulse of marsh prey into the Shark River 192 Estuary. Importantly, such inter-tissue comparisons are useful even if tissues do not reach full 193 194 dietary equilibrium (i.e. four half lives), because they can provide data on the direction (i.e. an increase of decrease in δ^{13} C depleted prey) and magnitude of dietary change. Tissue samples 195 were put on ice and frozen before laboratory preparations. All samples were dried and 196 197 homogenized prior to stable isotopic analysis.

Muscle tissue was collected from known estuarine (*Lutjanus griseus*, *Mugil cephalus*) 198 and marsh teleosts (Lepomis marginatus, L. microlophus, L. punctatus) that may serve as prey 199 200 for juvenile bull sharks (based on gape size of sharks, size of teleosts, and stomach contents analysis of juvenile bull sharks in other systems: Snelson & Williams 1981; Snelson, Mulligan & 201 Williams 1984; O'Connell et al. 2007). Samples were collected during ongoing community 202 level surveys using electrofishing, which occurred during the bull shark study (see Rehage & 203 Loftus 2007 for description of sampling method). Samples were frozen until being dried and 204 homogenized in the lab. Stable isotope data from teleosts provided the framework for the 205

206 difference in δ^{13} C values of estuarine and freshwater prey for stable isotope diet change models 207 (see below).

Passive acoustic tracking was used to quantify the movement patterns of individual bull 208 209 sharks to assess their use of upstream areas of the estuary in response to the marsh prey pulse. From Oct 2008 to May 2009, sharks caught in excellent condition (swimming strongly upon 210 capture; n = 23) were surgically fitted with a Vemco V16-4H transmitter (Vemco, Halifax, NS). 211 Transmitters were set to emit a unique series of pulses for each shark at a random interval 212 between 30-90 sec (mean emission interval = 60 sec). Movements of acoustically tagged sharks 213 were tracked within an array of 43 Vemco VR2 and VR2W acoustic receivers (Fig. 1) from Oct 214 2008 to Nov 2009. In situ measurements revealed mean detection ranges of receivers were ~500 215 m (see Rosenblatt & Heithaus 2011 for detection ranges of individual receivers). Each receiver 216 217 was attached to a PVC pipe set in a 10 kg cement anchor. Data from receivers were downloaded every 3-4 months for the duration of the study, and batteries were replaced as needed. 218

219

220 *Stable isotope analysis*

All shark samples were analyzed at the Florida International University Stable Isotope 221 Facility (29 blood samples and 30 plasma samples) or the Yale Earth System Center for Stable 222 Isotopic Studies (61 blood samples and 60 plasma samples). Lipid extraction was not performed 223 because C:N ratios (mean blood = 2.63 ± 0.25 SD; mean plasma = 2.03 ± 0.26 SD) were below 224 225 those suggested for extraction or mathematical correction (3.5; Post et al. 2007). To verify analytical consistency, we randomly selected samples to be analyzed at both Florida 226 International University and Yale University, for which the variation between resulting δ^{13} C 227 values and δ^{15} N values were 0.13% \pm 0.20 SE. The standard deviations of standards run for 228

Yale were 0.14‰ for δ¹³C and 0.22‰ for δ¹⁵N, and 0.29‰ for δ¹³C and 0.24‰ for δ¹⁵N for
Florida International. All teleost samples were analyzed at the Florida International University
Stable Isotope Facility.

232

233 *Quantitative Analysis*

234 <u>Acoustic tracking</u>

We quantified the proportion of days each tagged shark was detected in the upstream 235 region based on receiver detections of tagged sharks (Fig. 1). We predicted that if sharks fed 236 237 from the prey pulse, they would have increased their use of the upstream region where freshwater taxa enter the system early in the dry season, and decreased their use of the upstream 238 region later in the dry season when the abundance of marsh prey decreased (Rehage & Loftus 239 240 2007). Sharks were only used for analysis if they were present within the array for > 3 months, and were within the array during the 2009 dry season when the marsh prey pulse was expected to 241 enter the estuary (1 Mar to 28 May). We used a random effects GLMM to test the effect of 242 243 month on the average proportion of days individual sharks were detected by upstream receivers, with individual as a random effect, and used a Post hoc Tukey's test to test for significant 244 245 differences across months. We used linear regression to examine the relationship between marsh water level and the proportion of sharks detected per day by upstream receivers during the dry 246 season. Finally, based on the movements of marsh taxa during the dry season, we used t-tests to 247 investigate if there was a significant change in the use of the upstream region by sharks when 248 water elevations were ≤ 0 cm, between 0-5 cm, and between 0-10 cm to develop predictions for 249 our diet change models (see below). Investigating shark habitat use in relation to these water 250 251 depths allowed us to examine if sharks changed their movement behavior in response to the entry

of larger marsh taxa (> 8 cm; enter the estuary from the marsh when water levels drop below 10
cm) or smaller marsh taxa (< 8 cm; enter the estuary from the marsh when water levels drop
below 5 cm) (Chick, Ruetz & Trexler 2004; Rehage & Loftus 2007; Trexler & Goss 2009;
Parkos, Ruetz & Trexler 2011).

256

257 Discrimination difference between blood and plasma

Studies quantifying isotopic discrimination values in sharks are limited. Hussey et al. 258 (2010) investigated discrimination in captive sand tiger (*Carcharias taurus*; n = 3) and lemon 259 sharks (*Negaprion brevirostris*; n = 1), however the only tissue they analyzed that could 260 collected without lethal sampling was muscle tissue, which has a long isotopic half-life (~98 261 days, MacNeil, Drouillard & Fisk 2006) and was not useful for our study. Both Kim et al. 262 263 (2012) and Malpica-Cruz et al. (2012) investigated isotopic discrimination in captive leopard sharks (*Triakis semifasciata*; n = 6, n = 16, respectively). Kim et al. (2012) calculated Δ^{13} C 264 plasma-blood values between 0.5-0.9%. Despite sampling a variety of tissues, including blood, 265 266 muscle, and fin tissue, Malpica-Cruz et al. (2012) did not collect plasma, and therefore data from this study was not useful for our analyses. 267

Tissue-specific incorporation of stable isotopes can be affected by variability in environmental conditions, and can vary between species (reviewed by Vander Zanden & Rasmussen 2001; Crawford, McDonald & Bearhop 2008; Newsome, Clementz & Koch 2010). Therefore, we used linear regression to estimate the inter-tissue discrimination difference between blood and plasma (Δ^{13} C plasma-blood) in juvenile bull sharks, and compared this to values calculated by Kim et al. (2012). To do so, we plotted paired blood and plasma δ^{13} C values from 90 juvenile bull sharks against one another, and performed linear regression to

quantify the relationship between δ^{13} C values (see Matich, Heithaus & Layman 2010 for further details). To test whether differences between δ^{13} C values of blood and plasma varied across the observed range of δ^{13} C values, we also used a t-test to determine if the slope of the best-fit-line was different from 1:1. We would expect deviation from a slope of 1:1 if δ^{13} C discrimination varied with δ^{13} C values of shark diets.

280

281 <u>Temporal change in diet</u>

In addition to providing an estimate of Δ^{13} C plasma-blood, the regression plot of blood 282 and plasma δ^{13} C values described above also provided data that could be used to investigate 283 dietary variability in sharks. Data points above the best fit line indicate plasma δ^{13} C values more 284 enriched (i.e. less negative) than predicted by the model, and data points below the regression 285 line indicate plasma δ^{13} C values more depleted (i.e. more negative) than predicted by the model 286 (Fig. 3). In elasmobranchs, plasma has a faster isotopic turnover rate (half-life = 32 days, Kim et 287 al. 2012) than blood (half-life = 61 days, MacNeil, Drouillard & Fisk 2006), and therefore 288 289 dietary changes made by bull sharks should be detected by plasma isotope values considerably faster than blood isotope values. As such, plasma δ^{13} C values more than 0.4% enriched above 290 δ^{13} C values of blood (calculated Δ^{13} C plasma-blood - See Results) indicate a recent change to 291 either more marine food web-based diets or a change from feeding on marsh taxa to resident 292 estuarine taxa. Plasma values more depleted than 0.4% above blood indicate either a recent 293 change to more estuarine food web-based diets or a change from feeding on resident estuarine 294 taxa to migratory marsh taxa (Fig. 2). Thus, we plotted the residuals from the regression analysis 295 of blood and plasma δ^{13} C against shark capture date [day of year (DOY)] to investigate temporal 296 297 variability in the diets of bull sharks and to elucidate their potential use of the freshwater prey

298 pulse during the dry season. We then evaluated the effectiveness of using a piecewise function 299 to describe the data against using the best fit line from linear or non-linear least squares regression (Toms & Lesperance 2003). We selected breakpoints at which δ^{13} C residuals 300 301 exhibited a notable change (DOY 128, 150, 163, 175, 213) and quantified the coefficient of determination for each model. We selected the piecewise model with the highest coefficient of 302 determination and compared it to the coefficient of determination for linear and non-linear least 303 squares fits to determine if it was significantly higher in order to choose the best overall model 304 (Fisher 1921). To gain insight into general patterns of bull shark dietary changes in response to 305 the prey pulse, we used ANOVA to test the effect of season on δ^{13} C residual values. Post hoc 306 Tukey's tests were used to test for significant differences across seasons. 307

Despite having isotope data from 2008-2012, we only used data from sharks caught from 308 309 Oct 2008 to Dec 2009 because an extreme weather event in Jan 2010 significantly reduced the number of juvenile bull sharks in the estuary and changed the population structure (Matich & 310 Heithaus 2012). It also possibly affected the community composition, and thereby trophic 311 312 dynamics, of the ecosystem (Rehage et al. 2010). Additionally, we did not have acoustic tracking data for sharks after 2009 due to this extreme weather event, and therefore could not 313 314 investigate the correlations between marsh water levels, shark movements, and stable isotope values. Future studies investigating interannual variation in shark trophic interactions and 315 movements, however, would provide additional insights. 316

Due to the slow isotopic turnover rates of tissues in elasmobranchs (MacNeil, Drouillard & Fisk 2006; Logan & Lutcavage 2010; Kim et al. 2012) and the potential for maternal diets to be reflected in the tissues of newborns sharks (McMeans, Olin & Benz 2009; Matich, Heithaus & Layman 2010; Vaudo, Matich & Heithaus 2010), isotope values of bull sharks may not be

321 indicative of their current diet for individuals less than 90 days old (Belicka et al. 2012). 322 Because bull sharks in the Shark River Estuary are likely born at 60-70 cm TL (based on captures of neonate individuals; Heithaus et al. 2009; Matich & Heithaus 2012) between May 323 324 and August (based on the presence of umbilical scars; Curtis, Adams & Burgess 2011), and grow 10-20 cm/year (based on recaptured individuals; Neer, Thompson & Carlson 2005), we only 325 included tissues from individuals that were greater than 84 cm total length (at least one year of 326 age) and individuals less than 85 cm TL that were caught between December and April with 327 closed umbilical scars (at least 90 days old). 328

To determine if bull sharks changed their diets during the freshwater pulse, we developed 329 a series of theoretical models to predict the differences in plasma and blood δ^{13} C values to 330 determine if plasma had recently become more enriched or depleted in δ^{13} C in response to a 331 332 dietary change (Fig. 4). We modeled six plausible dietary shifts. These included 1) estuarine prey \rightarrow marsh prey (E \rightarrow F); 2) marine + estuarine prey \rightarrow estuarine + marsh prey (M+E \rightarrow E+F); 333 3) marine + estuarine prey \rightarrow marsh prey (M+E \rightarrow F); 4) marine prey \rightarrow estuarine prey (M \rightarrow E); 334 5) marine prey \rightarrow estuarine + marsh prey (M \rightarrow E+F); and 6) marine prey \rightarrow marsh prey (M \rightarrow F) 335 (Fig. 4b; Table 1). We used turnover data from MacNeil, Drouillard & Fisk (2006) (blood half-336 life = 61 days) and Kim et al. (2012) (plasma half-life = 32 days) to predict the rate of change in 337 blood and plasma isotopes based on the differences in δ^{13} C values of prev items from different 338 food webs (mean $\delta^{13}C \pm SE$; marine = -14.1 ± 0.2%, estuarine = -28.0 ± 0.5%, and marsh = -339 $30.5 \pm 0.5\%$). The models assumed that sharks would change their diets in response to marsh 340 prey entering the system, with the timing of the modeled change based on the movements of bull 341 sharks (see Results). Thus, the model assumed that shark diets changed when marsh water levels 342 343 were ≤ 0 cm, which corresponds to the time when sharks significantly increased their use of the

upstream region [31 Mar 2009 (DOY 90)]. A second diet switch, to a diet similar to that before
the prey pulse, was modeled to occur when water levels rose above 0 cm [3 Jun 2009 (DOY
154)]. During predicted periods of dietary equilibrium (wet season), we assumed that blood and

- 347 plasma values would differ by our calculated Δ^{13} C plasma-blood (0.4%); see Results).

Differences in δ^{15} N values of marine, estuarine, and marsh taxa bull sharks may have fed 348 upon (mean $\delta^{13}C \pm SE$; marine = 8.8 ± 0.5%, estuarine = 9.0 ± 0.5%, and marsh = 9.1 ± 0.3%) 349 did not provide the same resolution as differences in δ^{13} C values for taxa from each food web. 350 Therefore, we only modeled changes in δ^{13} C, rather than both δ^{13} C and δ^{15} N. Stable isotope 351 mixing models have become a popular analytical tool to investigate the trophic interactions of 352 animals (reviewed by Layman et al. 2012), but mixing models do not provide adequate output to 353 investigate temporal variability in the diets of individuals without repeated sampling, which is 354 355 often difficult for highly mobile species, such as sharks. Our modeling approach, however, enabled us to quantify variability in the diets of each sampled shark in response to the freshwater 356 prey pulse, and therefore we chose not to employ a mixing model. 357 358 Despite recent lab studies quantifying the turnover rates and discrimination values of

blood and plasma stable isotopes in elasmobranchs (MacNeil, Drouillard & Fisk 2006; Logan & 359 Lutcavage 2010; Kim et al. 2012; Malpica-Cruz et al. 2012), these processes can vary among 360 similar species (reviewed by Vander Zanden & Rasmussen 2001; Crawford, McDonald & 361 Bearhop 2008; Newsome, Clementz & Koch 2010). To investigate whether our estimates of 362 discrimination and turnover rates might affect the performance of our models, we tested 363 additional models in which we varied blood and plasma isotopic half-lives and Δ^{13} C plasma-364 blood. We created models with the half-lives of blood and plasma decreased to half of published 365 366 values (31 and 16 days, respectively) and increased to twice published values (122 days and 64

367 days, respectively; MacNeil, Drouillard & Fisk 2006; Kim et al. 2012) (Fig. 5a). We also 368 created models with Δ^{13} C plasma-blood of 0.9%, 0.7%, and 0.2%, representing a range of Δ^{13} C 369 plasma-blood values across the calculated values of Kim et al. (2012) (Fig. 5b). As such, we 370 created six different models (each diet change scenario; see above) for 12 different treatments of 371 isotopic half-life and Δ^{13} C plasma-blood.

Because a piecewise function best described δ^{13} C residuals across time (see Results), we 372 used piecewise linear regression with the same breakpoint as the true δ^{13} C residuals and DOY 373 model (DOY = 169) to investigate the relationship between the predicted difference in δ^{13} C 374 values (from theoretical models) and DOY. Because regression plots of predicted and true 375 differences in δ^{13} C values produced best fit lines with the same correlation coefficients and f-376 values for each diet change model across each isotopic half-life and Δ^{13} C plasma-blood, we 377 could not use traditional model selection. We therefore compared the best fit lines of the 378 theoretical models to that of the model for true δ^{13} C residuals and DOY. This approach allowed 379 us to qualitatively select the best model(s) describing if and how bull sharks changed their 380 trophic interactions in response to the prev pulse, and how isotopic half-life and Δ^{13} C plasma-381 blood affected model selection. Criteria for qualitatively selecting the best theoretical models 382 included 1) slopes of the piecewise functions with the same direction (positive or negative) as the 383 model for true δ^{13} C residuals and DOY; 2) slopes not significantly different from that of true 384 δ^{13} C residuals and DOY (t-test); and 3) piecewise functions with the closest mean distance to the 385 true δ^{13} C residuals and DOY regression lines. ANOVA was used to test the effects of model, 386 isotopic half-life, and Δ^{13} C plasma-blood on mean distance from the true δ^{13} C residuals and 387 DOY piecewise function for theoretical models that passed the first two criteria. Post-hoc 388

Tukey's tests were used to test for significant differences across these factors. All statisticalanalyses were conducted in JMP 6.0.0.

391

392 **Results**

From 2008 to 2012, we captured 90 juvenile bull sharks. Twenty-three individuals 393 between 71-142 cm total length (mean TL \pm SD = 102 \pm 22 cm) had acoustic transmitters 394 surgically implanted in them, and were tracked from 10 Oct 2008 to 30 Nov 2009 for a total of 395 5343 tracking days. Three individuals were not present for > 3 months within the system, and 396 therefore were not included in movement analyses. Shark detections by upstream receivers 397 varied by month, and were highest in April and May (DOY 91-151; $R^2 = 0.59$, p < 0.01; Fig. 6). 398 During the dry season, there was a negative correlation between shark use of the upstream region 399 and marsh water levels (DOY 60-148; $R^2 = 0.52$, p < 0.01; Fig 6), and the proportion of sharks 400 401 detected in the upstream region was significantly higher when water levels were ≤ 0 cm (mean \pm $SD = 0.38 \pm 0.14$) compared to > 0-5 (0.12 \pm 0.07) and > 0-10 cm (0.10 \pm 0.06) (t = 6.09, p < 402 403 0.01; t = 8.54, p < 0.01, respectively; Fig. 6).

Thirty-nine of the sampled sharks (n = 17 males, 22 females) captured from Oct 2008 to Dec 2009 and ranging in size from 75-182 cm TL (mean TL \pm SD = 116.5 \pm 28.3 cm) were used to investigate seasonal shifts in shark diets relative to the 2009 pulse of prey from the marsh. Sharks had blood and plasma δ^{13} C values that ranged from -17.5% to -26.5% (mean δ^{13} C \pm SD = -22.9 \pm 2.4%) and -17.8% to -25.3% (mean δ^{13} C \pm SD = -22.4 \pm 2.3%), respectively, and blood and plasma δ^{15} N values that ranged from 10.5% to 12.8% (mean δ^{15} N \pm SD = 11.6 \pm 0.5%) and 9.9% to 12.4% (mean δ^{15} N \pm SD = 11.5 \pm 0.6%), respectively (Fig. 2). There was a significant, positive relationship between blood and plasma δ^{13} C values with a high coefficient of determination (Fig. 3; R² = 0.81, p < 0.01), and the slope of the best fit line was not different from one (slope = 0.84, t₍₉₀₎ = 0.18). This suggests the mean difference between blood and plasma δ^{13} C values (plasma was 0.4‰ greater than blood) was consistent across the δ^{13} C range of the sampled sharks.

The δ^{13} C residuals of bull sharks caught in 2008-2009 varied significantly with DOY and capture season. A piecewise function with a breakpoint at DOY 169 was significantly better than a polynomial fit or linear fit for the relationship between DOY and δ^{13} C residuals (z-score = 5.48, p < 0.01; Fig. 7), with the magnitude of the slope for the first section (DOY 0-169) more than three times greater than the magnitude of the second section (DOY 169-365). Mean δ^{13} C residuals decreased significantly between the early dry season and the late dry season, and then increased from the late dry season to the early wet season (Fig. 8).

Changing the parameters of the models (isotopic half-life and Δ^{13} C plasma-blood) 423 changed their predictions of δ^{13} C residuals. As the duration of isotopic half-life increased (i.e. 424 from 0.5 half-lives to 2 half-lives), models predicted an increase in the duration of time δ^{13} C 425 residuals were in a state of change in response to diet shifts, and as Δ^{13} C plasma-blood increased 426 (i.e from 0.2-0.9%), models predicted greater positive δ^{13} C residuals during non-pulse periods 427 and smaller negative δ^{13} C residuals during the prey pulse (Fig. 5). ANOVA revealed that model 428 (F = 10.26, p < 0.01) and Δ^{13} C plasma-blood (F = 14.08, p < 0.01) were significant factors 429 explaining variability in mean distance between piecewise functions of theoretical models and 430 the model of true δ^{13} C residuals and DOY (Appendix 1). Models with Δ^{13} C plasma-blood = 431 0.7% and 0.9% had significantly lower mean distances from the true δ^{13} C residuals and DOY 432 piecewise function than discrimination differences of 0.2% and 0.4% (Fig. 9a), and models with 433

434	Δ^{13} C plasma-blood = 0.9% were the best fit (mean distance \pm SE = 0.5 \pm 0.1%, 0.4 \pm 0.1% for
435	Δ^{13} C plasma-blood = 0.7% and 0.9%, respectively). Models 2 and 3 (marine + estuarine prey
436	\rightarrow estuarine + freshwater prey, and marine + estuarine prey \rightarrow freshwater prey, respectively) had
437	the lowest mean distances from the δ^{13} C residuals and DOY piecewise function (Fig. 9b), with
438	model 2 having the best fit (distance \pm SE = 0.4 \pm 0.1%, 0.5 \pm 0.1% for model 2 and model 3,
439	respectively). This suggests that model 2 at Δ^{13} C plasma-blood = 0.9% was the best model for
440	predicting changes in δ^{13} C values of blood and plasma, (Table 2, Appendix 1). Parameter
441	estimates for isotopic half-life did not affect model performance ($F = 0.90$, $p = 0.42$).

442

443 Discussion

Seasonal resource pulses are important components of annual energy budgets for many 444 species (reviewed by Otsfeld & Keesing 2000, Yang et al. 2008). For example, brown bears 445 (Ursus arctos) in North American Pacific riparian ecosystems rely on predictable annual pulses 446 of spawning salmon to sustain their biomass levels for overwinter hibernation (Naimen et al. 447 448 2002, Helfield & Naimen 2006). Along the South African coastline, sardines (Sardinops sagax) 449 make annual migrations into nearshore areas and serve as an important pulse of food for marine mammals, birds, bony fishes, and elasmobranchs (Dudley & Cliff 2010; O'Donoghue, Drapeau 450 & Peddemors 2010). Within the Shark River Estuary, the influx of marsh taxa into upstream 451 channels comprises a considerable proportion of the annual energy budget of teleost predators in 452 453 the ecotone region (e.g. Amia calva, Centropomus undecimalis, Micropterus salmoides; Rehage & Loftus 2007; Boucek & Rehage *in press*), suggesting this resource pulse is likely a seasonally 454 important component of estuarine food webs within the ecosystem. Our study shows that 455 456 numerous juvenile bull sharks move upstream to take advantage of this influx of marsh prey,

despite relatively high levels of individual specialization within the population found in ourprevious work (Matich, Heithaus & Layman 2011).

Previously, we found that juvenile bull sharks in the Shark River Estuary show 459 considerable inter-individual variation in trophic interactions, and many individuals (ca. 57%) 460 showed relatively high degrees of specialization on one type of resource pool (i.e. marine food 461 webs vs freshwater/estuarine food webs; Matich, Heithaus & Layman 2011). Our results from 462 this study suggest the trophic interactions of some sharks in the estuary (i.e. those identified 463 previously as specialists) are flexible, at least during the dry season when marsh taxa enter the 464 system and provide an additional food source. Blood and plasma δ^{13} C values (mean \pm SD = -465 $22.9 \pm 2.4\%$ and $-22.4 \pm 2.3\%$, respectively) suggest that many bull sharks fed on marine and 466 freshwater/estuarine prey throughout the year, and during the wet and early dry seasons, sharks 467 had δ^{13} C residuals (plasma-blood) similar to our predictions attributed to Δ^{13} C plasma-blood (*ca.* 468 0.3-0.9‰), suggesting they had relatively stable diets. Yet, sharks had significantly lower $\delta^{13}C$ 469 residuals during the late dry season (mean \pm SE = -0.5 \pm 0.4%), and model selection predicted 470 471 diet switches from marine and estuarine prey to estuarine and marsh prey during the marsh prey pulse with a relatively rapid return to the previous diet at the terminus of the prev pulse when 472 marsh prey were depleted, suggesting bull sharks fed on this seasonal resource pulse from 473 freshwater marshes despite many individuals specializing on other resources outside this time 474 period (Matich, Heithaus & Layman 2011). 475

Individual specialization has been hypothesized as a means to reduce intraspecific
competition, especially in ecosystems with limited resources (reviewed by Bolnick et al. 2003).
The Shark River Estuary is an oligotrophic system, and limited food abundance may be a driver
of individual specialization in juvenile bull sharks (Matich, Heithaus & Layman 2011), however

480 the additional suite of resources from the marsh during the prey pulse may relax intraspecific 481 competition for food. Apparently similar to bull sharks in the Shark River Estuary, specialist bluegill sunfishes (Lepomis macrochirus) were more successful foragers than generalists, but 482 483 individual specialists exhibited flexibility in their habitat use and switched foraging behaviors when preferred resources became depleted (Werner, Mittlebach & Hall 1981). When preferred 484 485 prey were scarce, silver perch (Bidyanus bidyanus) in aquaculture ponds switched from specializing on *Daphnia* to specializing on calanoids and insects, suggesting individual 486 specialization may be a flexible trait in some populations (Warburton, Retif & Hume 1998). If 487 488 bull shark trophic specialization is driven by density dependent processes, then the influx of marsh taxa into the system may considerably increase the availability of food, and reduce the 489 need for sharks to have specialized diets when the prey pulse enters the estuary (Svanback & 490 491 Persson 2004).

492

493 *Drivers of bull shark behavior*

494 Previous studies in the Everglades suggest that large marsh taxa (> 8 cm TL) vacate the marsh in search of deep water refugia early in the dry season when water levels drop below 10 495 496 cm in reference to elevation, and small marsh taxa (< 8 cm TL) enter the estuary later when water levels drop below 5 cm (Rehage & Loftus 2007; Trexler & Goss 2009). Thus, we 497 predicted bull sharks would use the upstream region of the estuary early in the dry season to take 498 499 advantage of all marsh taxa entering the system. Yet, bull sharks began using upstream areas heavily later than we predicted, which may be due to several reasons. Because many sharks in 500 the estuary are less than three years old (Heithaus et al. 2009; Matich & Heithaus 2012), a lack of 501 502 foraging experience may hinder their ability to detect when marsh prey are available (e.g.

Werner and Giliam 1984). Interannual variation in timing and magnitude of the prey pulse due
to variation in precipitation and freshwater flow (Boucek & Rehage *personal communication*),
may further reduce the ability of bull sharks to detect the start of this event. Unfortunately, our
data set will not currently allow us to test these hypotheses.

Prey preference may also play a role in the timing of the bull sharks' responses to the 507 prey pulse (Lanszki & Sallai 2006; Hawlena & Perez-Mellado 2009). If bull sharks preferred to 508 eat large mesopredators like bass or bowfin, we would have expected them to use the upstream 509 region earlier than observed, and their diets and isotope values would have changed accordingly. 510 511 Instead, bull sharks did not significantly increase their use of the upstream region of the estuary until marsh water levels dropped below 0 cm, when all aquatic taxa have vacated the marsh. 512 Thus, bull sharks may wait until the overall abundance of marsh taxa of all sizes in the system is 513 relatively high, or they may be targeting smaller prey that arrive in the estuary later. Shark $\delta^{15}N$ 514 values suggest that bull sharks likely targeted smaller prey from the marsh. Plasma and blood 515 δ^{15} N values of bull sharks caught during the dry season (mean \pm SE = 11.8 \pm 0.1% and 11.9 \pm 516 0.2%, respectively) were comparable to δ^{15} N values of muscle tissue of other large aquatic 517 predators like snook (mean \pm SE = 11.3 \pm 0.3%) and bass (mean \pm SE =10.93 \pm 0.14%) that are 518 known to feed on small marsh taxa. Therefore, bull sharks likely compete for with these large 519 mesopredators for small prey that decline in abundance as the dry season progresses rather than 520 consuming them (Boucek & Rehage *in press*). Comparison of δ^{15} N values must be made 521 cautiously, however. For example, muscle tissue in elasmobranchs has a slow turnover rate 522 (half-life = 98 days, MacNeil, Drouillard & Fisk 2006), and thus we may not expect to detect 523 large seasonal changes in δ^{15} N values. Additionally, δ^{15} N turnover and discrimination rates may 524 525 vary with diet quality, trophic pathway, metabolic activity, and body size (reviewed by Vander

Zanden & Rasmussen 2001; Martinez del Rio et al. 2009; Hussey et al. 2012). Future studies
incorporating stomach content analysis and fatty acid analysis should help further elucidate the
importance of resource pulses to bull shark diets as well as intraspecific variation in the use of
these resources.

530

531 *Alternative explanations*

Alternative explanations are unlikely to account for observed temporal variation in 532 habitat use and δ^{13} C values of sharks within the estuary. For example, shifts in habitat use by 533 sharks could be driven by upstream movements of preferred prey (e.g. Ford et al. 1998; Rolstad, 534 Loken & Rolstad 2000). Yet, if sharks were feeding on the same prey year round, δ^{13} C residuals 535 would be expected to remain similar during the year or exhibit longer lag-times if the prey of 536 537 sharks had moved upstream to feed on the marsh prey pulse (i.e. the time for preferred prey to integrate marsh prey into tissues which would then be integrated into shark tissues). 538 Increased use of the upstream area by bull sharks when marsh water levels were ≤ 0 cm 539 540 may have been driven by changes in environmental conditions. Juvenile bull sharks in other estuaries modify their space use in accordance with changes in salinity (e.g. Heupel & 541 Simpfendorfer 2008; Froeschke, Stunz & Wildhaber 2010). Thus, bull sharks may have 542 increased their use of the upstream region of the estuary during the dry season when salinities in 543 areas further downstream increased and became higher than sharks preferred. However, salinity 544 remains relatively low in the upstream region year-round (Heithaus et al. 2009, Rosenblatt & 545

Heithaus 2011) and bull sharks are found in all areas of the estuary in all seasons (Matich &

547 Heithaus 2012), suggesting physical factors are unlikely to be driving the significant increase in

the use of the upstream area when marsh water levels are ≤ 0 cm.

549 Alterations in metabolic processes in response to environmental change can cause variability in stable isotope values (Kelly 2000; McCutchan et al. 2003; Vanderklift & Ponsard 550 551 2003). Although bull sharks experience seasonal changes in salinity that may lead to changes in stable isotope values of tissues (Heithaus et al. 2009; Rosenblatt & Heithaus 2011), daily and 552 weekly changes in salinity within the estuary would be expected to buffer a detectable change in 553 isotope values attributed to osmoregulatory processes. Additionally, changing the isotopic half-554 lives and discrimination differences of our theoretical models did not affect the performance of 555 our models or model selection (models 2 and 3 were the best models for all permutations), 556 557 suggesting changes in metabolic processes attributed to environmental variability are unlikely to have produced the trends in δ^{13} C observed during our study. As such, our results do indeed 558 suggest sharks changed their diets during the dry season in response to the prey pulse, which 559 560 may be a seasonally important source of nutrients and energy as observed in other predators within the system (Boucek & Rehage in press). 561

562

563 Conclusions

Stable isotope analysis is an attractive tool for ecologists because it can provide a time-564 565 integrated view of trophic interactions (Bearhop et al. 2004). While stable isotopes often provide only course information with regard to prey identity (reviewed by Gannes, O'Brien & Martinez 566 del Rio 1997; Post 2002), employing this tool with complimentary approaches can be used to 567 568 elucidate patterns and drivers of variability in trophic interactions and make predictions about how future conditions may lead to changes in food webs. Here we used a combination of stable 569 isotope analysis, acoustic telemetry, and predictive modeling to elucidate changes in bull shark 570 571 behavior in response to a resource pulse of taxa from adjacent marshland. Isotope data suggest

sharks increased the proportion marsh prey in their diets during the dry season, and movement
data show that bull sharks increased their use of areas adjacent to freshwater marshes during this
time. Annual variability in hydrology and planned changes in Everglades water management,
however, may affect the importance of marsh taxa in the diets of bull sharks during the dry
season.

Restoration efforts are planned to increase freshwater flow through the Everglades 577 (CERP 2006), which will likely affect marsh water elevations (Obeysekera et al. 2011) and, in 578 turn, the onset and duration of this resource pulse and the total biomass entering the Shark River 579 580 Estuary. Increased freshwater flow and higher water levels in the marsh may lead to increased productivity, but may diminish the magnitude of the resource pulses into estuarine creeks, which 581 582 could have negative consequences for the bull shark population and/or affect their ecological role 583 within the ecosystem. If marsh taxa are not available within the estuary during the dry season, prey availability within the upper areas of the estuary may decrease and force bull sharks to 584 increase their use of marine resources. This potential dietary shift may lead to decreased survival 585 586 of young sharks, because downstream areas where marine taxa are most abundant are the riskiest habitats for small sharks to forage in due to high predation risk from large sharks (Heithaus et al. 587 2009; Matich, Heithaus & Layman 2011). However, this shift in behavior may lead to upstream 588 nutrient transport if juvenile sharks forage in areas where marine taxa are prominent, but rest 589 upstream where large sharks are rarely found (Rosenblatt & Heithaus 2011). Additionally, if 590 591 marsh taxa are not available to bull sharks, levels of individual specialization may further increase in the face of increased intraspecific competition (Matich, Heithaus & Layman 2011). 592 Understanding how environmental variability currently affects the magnitude and timing of this 593 594 pulse and the effects it has on aquatic communities is important for making predictions about

how changes in freshwater flow may alter slough communities in the Everglades. Using 595 596 modeling approaches such as the one developed for this study can advance our understanding of temporal variation in trophic interactions, and provide predictions about how changes in the 597 environment should affect food webs. Further research investigating the importance of resource 598 pulses and disturbance regimes on the trophic dynamics of systems should increase our 599 600 understanding of how predicted environmental changes due to natural and anthropogenic drivers may cause important ecological changes, and affect the role of predators within their respective 601 602 ecosystems.

603

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618 Capture data and isotopic data are available upon request through the FCE LTER website:619 http://fce.lternet.edu/.

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Tables

- Table 1: Predicted δ^{13} C values (in ‰) of prey in bull shark diets during periods of dietary equilibrium (wet season) and during dietary change (attributed to the freshwater prey pulse) used to predict δ^{13} C residuals for the theoretical models. M = marine prey, E = estuarine prey, F =
- marsh prey entering channels during marsh dry down.

Model	δ^{13} C at equilibrium	δ^{13} C at prey pulse	
1 (E→F)	-28.0	-30.5	
2 (M+E→E+F)	-21.0	-29.3	
$3 (M+E \rightarrow F)$	-21.0	-30.5	
4 (M→E)	-14.1	-28.0	
5 (M → E+F)	-14.1	-29.3	
6 (M → F)	-14.1	-30.5	

- Table 2: Mean distances \pm SE (in %) between actual δ^{13} C residuals of bull shark blood and
- plasma isotope values, and those predicted by theoretical models for each tissue-specific
- discrimination difference between plasma and blood (Δ^{13} C plasma-blood with plasma more
- 856 enriched for each scenario) to determine the best fit model(s) for sharks' diet change during the
- freshwater prey pulse. Smaller distances indicate a better fit. M = marine prey, E = estuarine
- 858 prey, F = marsh prey entering channels during marsh dry down.

	o C Discrimination difference				
Model	0.2%	0.4%	0.7%	0.9%0	
1 (E → F)	$0.7 \pm < 0.1$	0.6 ± 0.1	0.5 ± 0.1	0.5 ± 0.1	
$2 (M+E \rightarrow E+F)$	$0.7 \pm < 0.1$	$0.5 \pm < 0.1$	$0.2 \pm < 0.1$	$0.1 \pm < 0.1$	
$3 (M+E \rightarrow F)$	$0.8 \pm < 0.1$	$0.5 \pm < 0.1$	$0.3 \pm < 0.1$	$0.2 \pm < 0.1$	
4 (M→E)	1.0 ± 0.1	0.8 ± 0.1	0.6 ± 0.1	0.5 ± 0.2	
5 (M → E+F)	1.0 ± 0.1	0.8 ± 0.1	0.7 ± 0.1	0.6 ± 0.2	
6 (M → F)	1.1 ± 0.1	0.9 ± 0.1	0.8 ± 0.2	0.7 ± 0.2	

 δ^{13} C Discrimination difference

860 **Figure captions:**

Figure 1: The study occurred in the Shark River Estuary, Florida, USA. The star represents

United States Geological Survey water station 252820080505400, which was used to define
 seasons. Locations of acoustic receivers are indicated by white circles, and the white rectangle
 encompasses the upstream region where freshwater prey enter the estuary during marsh dry
 down.

- Figure 2: Mean isotope values for producers and consumers in the Shark River Estuary and
 adjacent marine waters from Fry & Smith (2002), Chasar et al. (2005), Williams & Trexler
 (2006), and our own sampling. Producers and consumers from the freshwater/estuarine food
 web are black, those from the marine food web are gray, and migratory marsh taxa are white.
 Producers are pluses (+), primary consumers are triangles (▲), secondary consumers are squares
- 871 (\blacksquare), tertiary consumers are circles (\bullet), and bull sharks (blood isotope values) are diamonds (\bullet).
- Figure 3: Linear regression of paired blood and plasma δ^{13} C values. The mean difference
- between blood and plasma δ^{13} C values (0.4‰) serves as an approximation for the difference in
- δ^{13} C discrimination between blood and plasma in bull sharks. The location of data points relative
- to the regression line provides insights into whether an individual's diet has become more

enriched in δ^{13} C or more depleted in δ^{13} C than predicted by differences in discrimination factors

- of blood and plasma. Open circles are data from sharks caught in 2008-2009, and closed
- diamonds are data from sharks caught in 2010-2012. Only sharks caught in 2008-2009 were
- 879 considered for temporal analysis.
- Figure 4: Model predictions for changes in a) δ^{13} C values of plasma and blood and b) δ^{13} C
- residuals if bull sharks switched to using freshwater prey during the dry season. If bull shark
- diets consist of resident estuarine taxa and are at equilibrium during the wet season, mean blood
- δ^{13} C values should be -28.0‰ and mean plasma δ^{13} C values should be -27.6‰. When marsh taxa enter the estuary during the dry season, if bull sharks switch to feeding on marsh taxa
- taxa enter the estuary during the dry season, if bull sharks switch to feeding on marsh taxa (Model 1), plasma δ^{13} C values will become more depleted faster than blood δ^{13} C values because
- plasma δ^{13} C turnover (~32 day half-life) is faster in elasmobranchs than blood δ^{13} C turnover (~
- 61 day half-life). In this scenario, differences between plasma and blood δ^{13} C values are
- predicted to switch from being positive to negative on DOY 112 and then revert to being positive
- on DOY 162 after marsh taxa have become depleted and bull sharks return to feeding on
- 890 estuarine taxa. Note that in b) the inconsistencies at the ends of the diet switch periods (near 133
- B91 DOY 148 and 200) are attributed to the different rates of change in plasma and blood δ^{13} C
- 892 (plasma approaches it asymptote much sooner than blood).
- Figure 5: Effects of changing a) isotopic half-life at Δ^{13} C plasma-blood = 0.4‰ and b) Δ^{13} C plasma-blood at one half-life on predictions of diet-change model 2.
- 895
- Figure 6: Relationship between marsh water elevation (gray line) at United States Geological
- 897 Survey water station 252820080505400 and the proportion of sharks detected by upstream
- 898 receivers per day (black line) from 10 Oct 2008 (DOY 284) to 31 Nov 2009 (DOY 335).

- Figure 7: Temporal variation in δ^{13} C residuals. The black lines are predicted residuals based on a
- 900 piecewise function and the gray dashed line is the predicted change in the difference between
- 901 plasma and blood δ^{13} C values for model 2 (marine + estuarine prey \rightarrow estuarine + freshwater
- prey attributed) at one isotopic half-life and Δ^{13} C plasma-blood = 0.9% $_{o}$, which was the best
- 903 model for predicting changes in δ^{13} C residuals. Model selection was not influenced by
- 904 assumptions about isotopic half life or Δ^{13} C.

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Figure 8: Seasonal variation in mean \delta^{13}C residuals. Error bars are ± SE, and bars with different
letters are significantly different based on post hoc Tukey's tests.
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- 907 Figure 9: Mean differences between actual δ^{13} C residuals and those predicted by a) all models at
- 908 each δ^{13} C discrimination difference between tissues and b) across each model for all
- 909 discrimination differences and half-lives. Data are means and error bars are ± SE, and bars with
- 910 different letters are significantly different based on post hoc Tukey's tests.

































