Florida International University FIU Digital Commons

FCE LTER Journal Articles

FCE LTER

2007

Seasonal fish community variation in headwater mangrove creeks in the southwestern Everglades: an examination of their role as dry-down refuges

Jennifer S. Rehage Earth and Environment Department, Florida International University, rehagej@fiu.edu

William F. Loftus U.S. Geological Survey, Florida Integrated Science Center, Everglades National Park Field Station

Follow this and additional works at: http://digitalcommons.fiu.edu/fce_lter_journal_articles

Recommended Citation

Rehage, J.S., W.F. Loftus. 2007. Seasonal fish community variation in headwater mangrove creeks in the southwestern Everglades: an examination of their role as dry-down refuges. Bulletin of Marine Science 80(3): 625-645.

This material is based upon work supported by the National Science Foundation through the Florida Coastal Everglades Long-Term Ecological Research program under Cooperative Agreements #DBI-0620409 and #DEB-9910514. Any opinions, findings, conclusions, or recommendations expressed in the material are those of the author(s) and do not necessarily reflect the views of the National Science Foundation. This work is brought to you for free and open access by the FCE LTER at FIU Digital Commons. It has been accepted for inclusion in FCE LTER Journal Articles by an authorized administrator of FIU Digital Commons. For more information, please contact dcc@fiu.edu, jkrefft@fiu.edu.

SEASONAL FISH COMMUNITY VARIATION IN HEADWATER MANGROVE CREEKS IN THE SOUTHWESTERN EVERGLADES: AN EXAMINATION OF THEIR ROLE AS DRY-DOWN REFUGES

Jennifer S. Rehage and William F. Loftus

ABSTRACT

The connectivity between the fish community of estuarine mangroves and that of freshwater habitats upstream remains poorly understood. In the Florida Everglades, mangrove-lined creeks link freshwater marshes to estuarine habitats downstream and may act as dry-season refuges for freshwater fishes. We examined seasonal dynamics in the fish community of ecotonal creeks in the southwestern region of Everglades National Park, specifically Rookery Branch and the North and Watson rivers. Twelve low-order creeks were sampled via electrofishing, gill nets, and minnow traps during the wet season, transition period, and dry season in 2004-2005. Catches were greater in Rookery Branch than in the North and Watson rivers, particularly during the transition period. Community composition varied seasonally in Rookery Branch, and to a greater extent for the larger species, reflecting a pulse of freshwater taxa into creeks as marshes upstream dried periodically. The pulse was short-lived, a later sample showed substantial decreases in freshwater fish numbers. No evidence of a similar influx was seen in the North and Watson rivers, which drain shorter hydroperiod marshes and exhibit higher salinities. These results suggest that headwater creeks can serve as important dry-season refugia. Increased freshwater flow resulting from Everglades restoration may enhance this connectivity.

Biological connectivity between fish communities of mangrove regions and those of other marine and coastal habitats (e.g., coral reefs, seagrass beds, sand, and mudflats), although deserving further attention, has been explored in a number of recent studies (see reviews by Beck et al., 2001, Gillanders et al., 2003; Sheridan and Hays, 2003; Mason et al., 2005; Sheaves, 2005; Faunce and Serafy, 2006). The presence of mangroves along coastal areas enhances the richness, abundance, and biomass of fishes in marine habitats (e.g., coral reefs; Nagelkerken et al., 2002; Dorenbosch et al., 2004; Mumby et al., 2004). Mangroves provide nursery grounds for larval and juvenile marine fishes and crustaceans (Robertson and Duke, 1987; Laegdsgaard and Johnson, 1995; Nagelkerken et al., 2000) due to their high prey abundance (Robertson et al., 1988; Sheridan, 1997) and their role as a predation refuge (Primavera, 1998; Acosta and Butler, 1999). Juvenile survival may be enhanced in shallow mangrove habitats where structural complexity, shading, and turbidity are relatively high (Laegdsgaard and Johnson, 2001; Ellis and Bell, 2004). Mobile marine fishes use mangrove habitats transiently as foraging grounds (Blaber and Milton, 1990; Chong et al., 1990), reproductive grounds (Chaves and Bouchereau, 2000), or move in when environmental conditions in diel or seasonal cycles are favorable (e.g., at high tide or with increased salinity or temperature; Ley et al., 1999; Barletta et al., 2005).

By comparison, the connectivity between fish communities in mangrove regions and upstream freshwater habitats has received much less attention. A reason for this is that in many mangrove systems, the freshwater influence is small, and the contribution of freshwater fishes to the estuarine community is limited (Pinto and Punchihewa, 1996; Laroche et al., 1997; Kuo et al., 1999; Nordlie, 2003; Hindell and Jenkins, 2004). In the Greater Everglades Ecosystem, shallow vegetated freshwater marshes transition into an extensive region of tidal mangrove forests (up to 15 km in width, over 60,000 ha of mangroves), which dominates the landscape along the southwest Florida coast (Smith et al., 1994). At the ecotone, mangrove-lined creeks drain upland marshes into a network of interconnecting estuarine rivers, bays, and mangrove forests. The ecosystem is rainfall-driven, marked by strong seasonality (high rainfall in the summer and fall, low in the winter and spring), which greatly influences the spatial extent of inundation of freshwater marshes, as well as the salinity regime of this broad estuarine region (Gunderson and Loftus, 1993).

As in other estuarine systems, salinity levels play an important role in structuring the plant and animal communities of the Greater Everglades Ecosystem (Montague and Ley, 1993; Serafy et al., 1997; Ley et al., 1999; Lorenz, 1999; Faunce et al., 2004). Historically, large volumes of freshwater reached estuarine areas, particularly during the wet season (Fennema et al., 1994). Today, drainage, channelization, and impoundment of marshes have greatly diminished the freshwater inflow into estuarine areas, resulting in substantially higher and more variable salinity regimes (Smith et al., 1989; Montague and Ley, 1993; Light and Dineen, 1994; McIvor et al., 1994). Fish community response to the natural and the anthropogenically-derived variation in freshwater inflow and salinity has been relatively well-studied in the southern and eastern parts of the ecosystem, namely, Florida Bay and Biscayne Bay (Thayer et al., 1987; Montague and Ley, 1993; Serafy et al., 1997; Ley et al., 1999; Lorenz, 1999; Serafy et al., 2003; Faunce et al., 2004; Lorenz and Serafy, 2006); but remains understudied along the southwest region (but see Green et al., 2006), where the mangrove zone is substantially more extensive than in the southern and eastern parts (Smith et al., 1994). Mangrove creeks along this area also drain generally longer hydroperiod marshes than the southern and eastern regions (Fenema et al., 1994). These marshes (Shark Slough) support more diverse and abundant fish assemblages than southern marshes (Taylor Slough) (Trexler et al., 2001; Chick et al., 2004; Green et al., 2006); thus high connectivity between the mangrove and freshwater fish communities may be expected.

In this study, we examined variation in the fish community of headwater mangrove creeks in response to seasonal fluctuations in freshwater flow and salinity in the southwestern region of Everglades National Park (ENP). In particular, we explored the role of low-order, ecotonal mangrove creeks as dry-season refuges for freshwater fishes. As marsh water levels drop, fishes are forced into deeper habitats such as alligator holes, solution holes, canals (Kushlan, 1974; Nelson and Loftus, 1996; Chick et al., 2004; Kobza et al., 2004; Rehage and Trexler, 2006), and presumably headwater creeks. We sampled the fish community in the uppermost stretches of creeks, where habitat may be most suitable for freshwater species because of proximity to marshes and low salinity regimes. A secondary goal of this study was to compare sampling efficiency among gears. Sampling with electrofishing and gill nets targeted large fishes, whereas minnow traps targeted small fishes. Sampling focused on two regions: Rookery Branch (RB) and the North and Watson rivers (NW) (Fig. 1). Headwater creeks in the RB region link the main freshwater drainage of the southern Everglades (Shark Slough) to Tarpon Bay, and the Shark and Harney rivers. Creeks in the NW area are headwaters of the North and Watson rivers which flow into Whitewater Bay. In neither system have the fish communities in the oligohaline reaches received enough attention to describe their seasonal and long-term dynamics beyond surveys

that provided inventory data (Tabb and Manning, 1961; Odum, 1971; Loftus and Kushlan, 1987), despite their historical importance as a prey source for wading birds (Ogden, 1994) and their key role in the mangrove food web (Odum, 1971).

Methods

SITE DESCRIPTION.—We sampled the large and small fish community in the oligohaline to mesohaline headwater reaches of six creeks in the RB region and six creeks in the NW region (Fig. 1). RB sites included four creeks in the main stem of Rookery Branch, as well as Squawk and Otter creeks (RB7 and RB12, respectively). NW sites were located along three Watson River creeks and three North River creeks. Creeks in NW drain shorter hydroperiod marshes than RB creeks. By hydroperiod, we refer to the number of days the marsh is flooded in a yearly cycle. Marshes are typically considered dry if water levels drop below 5 cm, at which depth little standing water remains (Loftus and Eklund, 1994). According to data from hydrologic stations P35 and P38 (Fig. 1), over the past 20-yr period (1986–2006), the hydroperiod averages 332 d of flooding (\pm 8.3 d) in marshes upstream of RB creeks, and 305 d (\pm 13.5 d) in marshes upstream of NW creeks. Nutrient concentrations are similar between regions: relatively high nitrogen (approximately 1 mg L⁻¹) and low phosphorus concentrations (below 0.02 mg L⁻¹) are characteristic of both RB and NW waters (Levesque, 2004).

SAMPLING EFFORT.—All sampling was conducted in the main channel of headwater creeks and in the uppermost boat-accessible 600 m reach of each creek. Sampling included only first and second order creeks (Strahler, 1957). Creek shorelines were vegetated by riverine mangrove forests dominated by red mangrove, *Rhizophora mangle* Linnaeus, 1753 (Lugo and Snedaker, 1974). Creek depth at sampling locations averaged 1.37 m (\pm 0.03 m, n = 108); width averaged 10.8 m (\pm 1.0 m, n = 107). Sampling was conducted during November 2004, February 2005, and April 2005, corresponding to the wet season, the transition between wet and dry



Figure 1. Map of southwestern Everglades National Park showing location of headwater creeks included in this study. Twelve creeks (filled circles) were sampled: six in the North and Watson rivers (NR1-3 and WR4-6) and six in the Rookery Branch region (RB7-12). Location of four reference NPS hydrological stations is indicated by open triangles (see Figure 2 for data from these stations). Distance from sampling sites to stations varies. CN and NR stations are located in creek channels downstream of sites sampled (CN is 4200 m downstream from RB7 and NR is 900 m downstream from NR3). P35 and P38 are located in freshwater marshes upstream of creeks (P35 is 300 m from RB9 and P38 is 6300 m from NR2).



Figure 2. (A) Mean daily salinity and (B) water levels collected by the nearest four NPS monitoring stations to study headwater creeks. See Figure 1 for approximate location of stations in reference to sites. Bold lettering indicates sampling months. Dotted line indicates 5-cm water depth cutoff used for calculation of marsh hydroperiod (see text for explanation).

seasons (hereafter "transition"), and dry season. Daily marsh water level and creek salinity measurements were obtained from the nearest National Park Service (NPS) hydrologic stations (Figs. 1 and 2). While sampling, we measured salinity at creek sites with a YSI[®] 85 unit (Fig. 3).

LARGE-FISH SAMPLING.—Large fishes (55–750 mm standard length, SL) were sampled using a boat-mounted electrofishing unit (two-anode, one-cathode Smith-Root^{*} generator-powered pulsator 9.0 unit rated to a maximum salinity of 15). Electrofishing has been shown to be an effective method for sampling larger fishes in other Everglades habitats (Nelson and Loftus, 1996; Chick et al., 1999). At each creek, sampling was conducted in three 5-min (pedal time) bouts (three bouts × six creeks × two regions × three seasons = 108 electrofishing samples). For all bouts, electrofishing power was standardized to 1500 watts according to temperature and salinity conditions (Burkhardt and Gutreuter, 1995). On average, each bout sampled 122.6 m (\pm 2.8 m, n = 108) of creek shoreline. Bouts were distributed evenly over the 600-m segment of creek, so that each bout was considered an independent sampling unit. For each bout, we randomly selected a creek shoreline and made a single pass with the electrofish



Figure 3. Salinities over the three sampling seasons in the two study regions: the North and Watson rivers and Rookery Branch. Shown are means ± 1 standard error (SE).

ing boat. All fish captured were identified to species, measured to nearest mm SL, weighed to nearest g, and released after full recovery. Non-indigenous species were collected and brought to the laboratory for processing.

We sampled the upper 100-m reaches of each creek with two passive techniques—experimental gill nets and minnow traps. Gill nets are commonly used to monitor fish populations in a wide range of habitats, typically targeting highly mobile and large-bodied species (e.g., Hubert and O'Shea, 1992). Experimental gill nets have panels of several mesh sizes, thus reducing the potential for size or single-species selectivity (Argent and Kimmel, 2005). Nets were 38 m long, with six mesh sizes (25.4, 38.1, 50.8, 63.5, and 76.2 mm). One net was set in the upper 100 m of each creek (one gill net × six creeks × two regions × three seasons = 36 gill net samples). Logistic constrains prevented us from obtaining greater gill-net sample sizes that would be comparable to electrofishing sample sizes. To comply with NPS regulations, gill nets were set mid-channel, parallel to the direction of current flow, and for only 30-min periods. All fishes captured were identified, measured, and weighed in the field, then released.

SMALL-FISH SAMPLING.—Small fishes (< 50 mm SL) were sampled with 3-mm, metal-mesh minnow traps (25.4 mm opening) deployed unbaited, overnight along creek banks. Minnow traps are a commonly used and easily replicable sampling device, but it suffers from several sampling biases (Rozas and Minello, 1997), one of which is trap placement. Minnow traps are typically set on the substrate, where they are unlikely to be encountered by water-column or surface dwellers (Layman and Smith, 2001). In this study, we deployed minnow traps in pairs; one set on the substrate and a second suspended just beneath the water surface, secured to mangrove prop roots. In each creek, we deployed three pairs of traps during the November 2005 sampling event, but increased effort to five pairs for subsequent sampling events (November: six traps × six creeks × two regions = 72 samples; February and April: 10 traps × six creeks × two regions z two seasons = 240 samples; total sample size is 312). Fish captures from minnow traps were preserved in 10% formalin and brought to the laboratory for processing.

STATISTICAL ANALYSES.—We examined variation in the abundance of fishes among regions and creeks and as a function of season with nested, repeated-measures ANOVA or AN-COVA models. Season was the repeated measure in our analyses, and nesting allowed us to account for spatial variation among regions (RB and NW); creeks were nested within regions. Focal response variables included: CPUE for the large fishes caught in electrofishing (number 5-min⁻¹ pedal time) and gill nets (number 30-min⁻¹ soak time), CPUE for the small fishes caught in minnow traps (number 24 h⁻¹), and the proportion of CPUE that was freshwater in electrofishing and minnow trap samples (CPUE was too low in gill net samples). Species were classified as either marine, estuarine, or freshwater (Table 1) based upon their habitat occurrence (per Loftus and Kushlan, 1987; Loftus, 2000). Preliminary analyses examined seasonal and spatial variation in the number of species caught in all gears, but results were indistinguishable from analyses of CPUE; and thus, are not presented here.

A two-way ANOVA was used to examine seasonal and spatial variation in salinity levels. Salinity was used as a covariate in analyses of the large fish data; no salinity measurements were made at the time of minnow trap deployment. To better satisfy assumptions of parametric tests, CPUEs were ln (observed value + 1)-transformed and proportions were subject to angular transformations prior to analyses. Post-hoc pairwise comparisons were performed using Tukey-corrected contrasts. If salinity was a significant covariate, simple linear regressions were used to examine the relationship between response variables and salinity. All analyses were performed using Proc Mixed in SAS Version 9.1.3[°].

We used analyses of similarity (ANOSIM) based on Bray-Curtis similarity matrices to test for effects of region, season, and gear (electrofishing vs gill net, and top vs bottom minnow trap) on fish community structure (Clarke and Warwick, 2001). Dissimilarity matrices were constructed based on ln (observed value + 1)-transformed estimates of the relative abundance of all taxa in samples, except for the gear comparison of gillnets and electrofishing, where a presence/absence matrix was used. Analyses included 28 taxa from electrofishing samples, 10 taxa from gill nets, and 22 taxa from minnow traps (Table 1). We followed ANOSIM analyses with percentage of similarity analyses (SIMPER) to determine which taxa contributed most to groupings observed among samples. We constructed non-metric multi-dimensional scaling (NMDS) plots to illustrate dissimilarity among groups. In these plots, the distance between data points is proportional to the degree of similarity between samples. All community structure analyses were conducted using Primer[®] Version 5.2.9.

Results

FRESHWATER FLOW AND SALINITY.—Salinity levels near our study sites increased substantially with the yearly onset of the dry season as flows from upstream freshwater marshes decreased (Fig. 2). In 2005, these increases occurred earlier in NW creeks than in RB creeks (Fig. 2A). Maximum salinities were higher in the vicinity of our NW sites than near RB sites (29 and 19, respectively). Hydroperiod was shorter in marshes upstream of NW sites than in those upstream of RB sites. Marshes upstream of NW sites were flooded for 357 d in 2004–2005; whereas marshes upstream of RB creeks were flooded for 324 d (Fig. 2B). Marshes upstream of RB also dried more frequently, but for short periods of time. In contrast, marshes upstream of the North River dried less frequently, but once dry, remained dry for a longer period of time. At our study creeks, salinity varied both among sites and across seasons (ANOVA: significant season by region interaction, $F_{2,102} = 18.6$, P = 0.0001; Fig. 3). Salinities were comparable between regions during the wet season sample, but diverged as the dry season progressed, reaching 10 in NW but < 5 in RB (Tukey pairwise comparisons of RB vs NW, P < 0.0001 for both the transition period and dry season).

ESTIMATES OF ABUNDANCE.—Fish abundance in the oligohaline to mesohaline reaches of mangrove creeks, as estimated by electrofishing, gill net, and minnow trap CPUE, was consistently higher in RB creeks than in NW creeks (Figs. 4,5). CPUE varied as a function of season (significant season by region interactions for all three

							CPUI	[1]			
Species			Ele	ctrofis	hing		Gill ne	ts	M	nnow t	raps
Scientific name	Common name	Occurrence	8	F		8	F		M	F	D
Lepisosteus platyrhincus DeKay, 1842	Florida gar	FW, EF, E	∞	351	246	-	31	23			
Amia calva Linnaeus, 1766	bowfin	FW, EF	-	52	З						
Megalops atlanticus Valenciennes, 1847	Tarpon	EF, E, M					8				
Anguilla rostrata (Lesueur, 1817)	American eel	EF, E	2	9	0						
Notropis petersoni Fowler, 1942	coastal shiner	FW, EF	17	42						468	
Ariopsis felis (Linnaeus, 1766)	hardhead catfish	EF, E, M						1			
Esox americanus Gmelin, 1789	redfin pickerel	FW		0							
Opsanus beta (Goode and Bean, 1880)	gulf toadfish	E, M									0
Mugil cephalus Linnaeus, 1758	striped mullet	EF, E, M	6	56	19						
Menidia beryllina (Cope, 1867)	inland silverside	EF, E	٢		0						4
Strongylura notata (Poey, 1860)	Atlantic needlefish	EF, E, M			1						
Strongylura sp.	unidentified needlefish	EF, E, M		0	5						
Fundulus chrysotus (Günther, 1866)	golden topminnow	FW, EF								11	
Lucania goodei Jordan, 1880	bluefin killifish	FW, EF							15	1,016	119
Lucania parva (Baird and Girard, 1855)	rainwater killifish	EF, E, M							13	49	76
Gambusia holbrooki Girard, 1859	eastern mosquitofish	FW, EF, E, M							137	390	357
Heterandria formosa Agassiz, 1855	least killifish	FW, EF							0	25	44
Poecilia latipinna (Lesueur, 1821)	sailfin molly	FW, EF, E, M							21		
Cyprinodon variegatus Lacépède, 1803	sheepshead minnow	FW, EF, E, M									1
Centropomus undecimalis (Bloch, 1792)	Snook	EF, E, M	33	102	14		-	1			
Enneacanthus gloriosus (Holbrook, 1855)	bluespotted sunfish	FW							4	102	0
Lepomis gulosus (Cuvier, 1829)	Warmouth	FW, EF		9					1	1	
Lepomis macrochirus Rafinesque, 1819	Bluegill	FW, EF		-							

							CPUE				
Species			Ele	ctrofish	ing	Ŭ	Gill net	s	Minr	low tra	bs
Scientific name	Common name	Occurrence	×	F	D	M	F	D	M	F	D
Lepomis marginatus (Holbrook, 1855)	dollar sunfish	FW		213	5					216	11
Lepomis microlophus (Günther, 1859)	Redear	FW, EF	0	31	1						
Lepomis punctatus (Valenciennes, 1831)	spotted sunfish	FW, EF	5	38	2						
Lepomis sp.	unidentified sunfishes	FW, EF		8							
Micropterus salmoides (Lacépède, 1802)	largemouth bass	FW, EF	2	100	7		Ļ	1			
Caranx hippos (Linnaeus, 1766)	crevalle jack	EF, E, M			0			1			
Lutjanus griseus (Linnaeus, 1758)	gray snapper	EF, E, M	11								
Eucinostomus harengulus Goode and Bean, 1879	tidewater mojarra	EF, E, M	113	0	0				0		
Eugerres plumieri (Cuvier, 1830)	striped mojarra	EF, E, M	111	12	11	0		ю			
Archosargus probatocephalus (Walbaum, 1792)	Sheepshead	EF, E, M	0			0					
Cynoscion nebulosus (Cuvier, 1830)	spotted seatrout	E, M									1
Sciaenops ocellatus (Linnaeus, 1766)	Redfish	EF, E, M	5	9							
Elassoma evergladei Jordan, 1884	Everglades pygmy sunfish	FW, EF							0		
Cichlasoma urophthalmus (Günther, 1862) ^a	Mayan cichlid	FW, EF	S	21	16			1	0		
Oreochromis aureus (Steindachner, 1864) ^a	Blue tilapia	FW, EF		15	10		3	1			
Tilapia mariae Boulenger, 1899 ^a	spotted tilapia	FW, EF	3						б		
Gobiosoma bosc (Lacépède, 1800)	naked goby	EF, E							1	3	0
Lophogobius cyprinoides (Pallas, 1770)	crested goby	EF, E, M	17						4	-	
Microgobius gulosus (Girard, 1858)	clown goby	EF, E, M							З	0	4
Trinectes maculatus (Bloch and Schneider, 1801)	Hogchoker	EF, E, M	4							3	0
CPUE total by season			368	1,066	348	9	47	31	210 2,	287	646
Total number of samples				108			36			312	
Total CPUE by gear				1,782			84		Э,	143	
Total taxa				27			10			22	
^a Non-indigenous species											

632

Table 1. Continued.



Figure 4. (A) Electrofishing and (B) gill net catch-per-unit effort (CPUE) over the three sampling seasons in the two study regions. Shown are means ± 1 SE.

CPUEs, Table 2), but season had a different effect in the two sampled regions. Across sampling gears, CPUE was highest in RB samples during the transition period. CPUE increased four-fold in electrofishing samples, eight-fold in gill nets, and nine-fold in minnow traps between the wet and transition samples (electrofishing, P = 0.009; gill nets, P = 0.002; minnow traps, P = 0.0001). The abundance of large species, such as Florida gar, bowfin, snook, largemouth bass, Mayan cichlid, and sunfishes peaked in the transition period. Among the small fishes, catches of bluefin killifish, eastern mosquitofish, coastal shiners, and smaller-bodied sunfishes also peaked during the transition period (Table 1). RB CPUE decreased significantly, returning to wet-season levels, in the dry season for electrofishing and minnow traps, but not for gill nets (electrofishing, P = 0.003; minnow traps, P = 0.0001).

In NW creeks, electrofishing CPUE was highest in the wet season (wet vs dry, P = 0.03; wet vs transition, P = 0.01; Fig. 4A), whereas no seasonal variation was detected in minnow trap nor gill net CPUE (Figs. 4B, 5). There was a trend for electrofishing CPUE to be negatively related to salinity (Table 2). The relationship had a relatively better fit in NW than in RB creeks (NW, P = 0.0001, r^2 = 0.27; RB, P = 0.045, r^2 = 0.08; Fig. 6). We detected no relationship between salinity and gill net CPUE (Table 2).

All gears varied significantly among creeks within the two study regions, and this variation was affected by season (Table 2). In NW, electrofishing CPUE was higher in North River creeks than in Watson River creeks (P = 0.003), although CPUE in gill nets and minnow traps did not differ. Seasonally, electrofishing CPUE was higher in North River headwaters in the transition and dry-season samples (P = 0.06 and P = 0.0006, respectively), but not in the wet season. In RB, electrofishing CPUE was lower in Squawk Creek (RB 7, Fig. 1) than in other creeks, particularly in the transition sample (P < 0.05). Minnow trap and gill net CPUE were significantly higher in Otter Creek (RB 12, Fig. 1) than in Squawk Creek (P = 0.0006 and P = 0.02, respectively), while CPUE in other RB creeks was intermediate.



Figure 5. CPUE in minnow traps shown separately by trap placement: (A) top vs (B) bottom of water column) over the three sampling seasons and in the two study regions. Shown are means ± 1 SE.

LARGE-FISH COMMUNITY STRUCTURE.—The composition of electrofishing catches varied equally between regions and among seasons (Table 3). Community structure was similar between NR and RB creeks in the wet season, but diverged considerably during the transition and dry seasons (wet, R = 0.12, P = 0.14; transition, R = 0.54, P = 0.002; dry, R = 0.74, P = 0.002; Fig. 7A). This divergence can be explained by increases in the relative contribution of freshwater taxa to the creek community. The contribution of freshwater species to CPUE was comparable between regions in the wet season (5% in NW vs 20% in RB; P = 0.225), but differed significantly in later samples (Fig. 8A). NW catches remained < 10% freshwater, whereas in RB, 80%–90% of the catch was composed of freshwater taxa during the transition and dry seasons (transition, P = 0.0011; dry season, P = 0.0001). During these drier samples, Florida gar, largemouth bass, bowfin, Mayan cichlid, and several sunfish species were almost

Table 2. Summary of results of nested, repeated-measures	ANOVA and ANCOVAs testing the
effects of season, region, creek, placement (for minnow traps	only), and salinity on catch-per-unit-
effort (CPUE) from electrofishing, gill nets, and minnow traj	ps.

	Electr	ofishir	ng CPUE	Gill	l-net (CPUE	Minne	ow-trap	CPUE
Source of variation	df	F	Р	df	F	Р	df	F	Р
Season	2, 42	6.7	0.0031	2, 14	3.7	0.0502	2,200	26.9	0.0001
Region	1, 55	60.4	0.0001	1, 11	9.3	0.0113	1,220	46.8	0.0001
Placement							1, 112	2.6	0.1077
Season × region	2,45	14.5	0.0001	2, 16	4.1	0.0368	2,200	31.7	0.0001
Season × placement							2, 198	21	0.0001
Region × placement							1, 112	19.1	0.0001
Region × season × placement							2, 198	9.5	0.0001
Creek (region)	3, 38	8.4	0.0002	3,6	6.9	0.0202	4,140	4.8	0.0012
Creek (region) × season	6,37	2.3	0.0558	6,14	0.8	0.5973	6,201	4.1	0.0006
Salinity	1,41	3.8	0.058	1, 13	0.01	0.9453			



Figure 6. Estimates of large fish abundance (CPUE) in electrofishing samples (Log-transformed) plotted as a function of salinity. Separate least-squares regressions were fitted to the two regions: Rookery Branch (RB, solid line) and the North and Watson rivers (NW, dotted line).

exclusively caught in RB creeks. The relative abundance of snook was also higher in RB, whereas needlefishes and tidewater mojarras were exclusively caught in NW creeks. Composition of gill net samples was similar between regions during the wet season, but tended to differ in the dry season sample (wet, R = 0.29, P = 0.20; dry, R = 0.65, P = 0.10; Fig. 7B). Florida gar was dominant in RB gill net samples, whereas NW gill nets were dominated by a small number of striped mojarras.

SMALL-FISH COMMUNITY STRUCTURE.—Variation in the small community structure was higher between regions than among seasons (Table 3, Fig. 9A), yet the proportion of freshwater species in traps varied as a function of both season and region (Table 4). The contribution of freshwater species to the RB small fish fauna showed no seasonal variation, averaging 96% throughout the study (Fig. 8B). In NW, however, the contribution of freshwater species decreased significantly between the wet and dry seasons, from 24% to 2% (P = 0.024). Minnow trap CPUE in NW primarily contained estuarine species (rainwater killifish, tidewater mojarra, and clown goby), whereas the RB community primarily contained freshwater species (eastern mosquitofish, sailfin molly, bluefin killifish, least killifish, dollar sunfish, and bluespotted sunfish; Table 1).

GEAR COMPARISON.—Large fish catches averaged 16.5 fish 5-min⁻¹ bout in electrofishing samples, whereas gill nets only averaged 2.3 fish per 30-min set (Fig. 4). Gill nets failed to detect the marked seasonal variation in the numbers of large fresh-

Table 3. Su	mmary of ANOS	IM results testing v	ariation in fish-o	community structur	e as a function
of region, se	eason, and gear i	n electrofishing, gill	net, and minno	w trap samples.	

		Regi	on	Seas	on	Gear com	parison
Target community	Sampling method	Global R	Р	Global R	Р	Global R	Р
Large fishes (> 50 mm)	Electrofishing	0.446	0.001	0.416	0.001	0.279ª	0.002
Large fishes (> 50 mm)	Gill nets	0.511	0.019	0.275	0.012		
Large fishes (> 50 mm)	Minnow traps	0.641	0.001	0.1	0.001	0.135 ^b	0.002

^a Comparison of electrofishing and gill nets

^b Comparison of minnow trap placement: top vs bottom of water column



Figure 7. Two-dimensional non-metric MDS ordinations illustrating large fish (50–750 mm SL) community structure in (A) electrofishing samples, (B) gill net samples, and (C) gill net vs electrofishing samples based on Bray-Curtis similarities of log-transformed, standardized CPUE.

water fishes present in creeks seen in the electrofishing data, particularly in RB (Fig. 4A). Electrofishing and gill-net samples also differed significantly in composition, although this dissimilarity was less than that observed as a function of spatial or seasonal factors (Table 3, Fig. 7C). Florida gar was the most abundant species caught using both methods, but numbers caught by electrofishing were higher than those caught using gill nets. CPUE of snook, striped mullet, largemouth bass, mojarras, and largemouth bass were also higher in electrofishing samples than in gill nets.

In minnow traps, mean CPUE was similar between the top and bottom trap, but placement affected the magnitude of CPUE variation across seasons and between regions (Table 2). For instance, CPUE in RB doubled between the wet and transition



Figure 8. Average proportion of (A) electrofishing and (B) minnow trap CPUE composed of freshwater species over the three sampling seasons in the two study regions. Only species known to occur in freshwater marshes (listed as FW in Table 1) were included in these analyses. Shown are means ± 1 SE.

samples in the top trap, but increased by 50 times in the bottom trap (P = 0.0001 for both cases, Fig. 5). Large numbers of sunfishes, bluefin killifish, and coastal shiners accounted for this increase in the bottom trap (Table 1). In NW headwaters, seasonal variation in CPUE was detected only in the top trap. In spite of very low catches, CPUE increased between the transition period and the dry season (P = 0.0001; Fig. 5A). Dissimilarity between minnow trap samples as a function of trap placement was lower than the separation observed when comparing gill nets and electrofishing samples (Table 3, Fig. 9B). The contribution of freshwater species was higher in the top than in the bottom trap (80.6% and 54.2%, respectively; Table 4). Eastern mosquitofish and least killifish were more abundant in traps placed at the top of the water column, whereas bluefin killifish, rainwater killifish, clown gobies, and dollar sunfish were more abundant in traps placed at the bottom of the water column.

Table 4. Summary of nested, repeated-measures ANCOVA and ANOVA analyses testing variation in the proportion of CPUE that is composed of freshwater taxa (FW in Table 1) per sample as a function of region, season, and placement (for minnow traps only) in electrofishing and minnow trap samples.

		Pro	portion of fr	eshwater Cl	PUE	
	Electr	rofishing sa	mples	Minn	low trap sa	mples
Source of variation	df	F	Р	df	F	Р
Season	2, 61	5.8	0.0049	2, 129	5.7	0.0044
Region	1, 58	38.7	0.0001	1, 128	199.7	0.0001
Placement				1, 103	18.1	0.0001
Season \times region	2,63	16.9	0.0001	2,130	4.1	0.0191
Season \times placement				2, 129	5.1	0.007
Region × placement				1, 103	0.1	0.7619
Region \times season \times placement				2, 129	7.9	0.0006
Creek (region)	3, 29	3.4	0.0319	4, 105	0.9	0.4609
Creek (region) \times season	6, 53	1.1	0.3533	6, 115	1.1	0.3586
Salinity	1,61	0.04	0.8399			

A. Seasonal and drainage variation



B. Trap placement



Figure 9. Two-dimensional non-metric MDS ordinations illustrating variation in small fish community structure (< 5 cm SL): (A) among seasons and sites and (B) as a function of trap placement. MDS plots are based on Bray-Curtis similarities of standardized minnow traps CPUE.

DISCUSSION

Disturbance from recurrent, seasonal dry-down events has a strong structuring effect on freshwater fish communities inhabiting Everglades marshes (Kushlan, 1974; Loftus and Eklund, 1994; Trexler et al., 2001, 2005; Chick et al., 2004). In response to dry-down, fish move from marshes to deeper habitats such as alligator holes, solution holes, and canals (Nelson and Loftus, 1996; Trexler et al., 2001; Kobza et al., 2004; Rehage and Trexler, 2006). Thus, access to dry-season refuges is a key element underlying long-term population dynamics in freshwater fishes (DeAngelis et al., 1997). Our results indicate that mangrove creek headwaters in the southern part of the ecosystem can also serve as important dry-season refugia, particularly for large-bodied species, whose abundance is strongly limited by seasonal dry-down (Trexler et al., 2001, 2005; Chick et al., 2004). A pulse of freshwater fishes was detected in RB creeks in February as marshes upstream began to dry periodically, which resulted in marked seasonal variation in patterns of abundance and composition in RB headwaters. The pulse was composed of both predatory species, such as Florida

gar, bowfin, and centrarchids; and of the small cyprinodontoids, although some species (e.g., mosquitofish) appeared to reside in creek all-year around. No evidence of a similar pulse was noted in the North and Watson rivers, where the contribution of freshwater taxa to the community was consistently small (0%–24%) and showed lower seasonal variation.

Mangrove fish communities are highly variable in both short (tidal) and longer time scales (seasonal) because of pronounced environmental fluctuations (Kupschus and Tremain, 2001). Seasonal changes in the abundance and composition of tropical and subtropical fish communities have been reported in mangrove systems throughout the world, including Madagascar (Laroche et al., 1997), Brazil (Barletta et al., 2005), Australia (Loneragan et al., 1986), the Solomon Islands (Blaber and Milton, 1990), Taiwan (Lin and Shao, 1999), and Mexico (Yanez-Arancibia et al., 1988). Of these examples, mangrove creeks of the Caeté River estuary in Brazil (Barletta et al., 2005) exhibit the greatest freshwater inflow and may closely resemble the mangrove creeks at our Everglades sites. However, the directionality of seasonal variation in RB and NW creeks is opposite that of Caeté, where the influx of freshwater species occurs during the wet season when salinities are low. Seasonal community dynamics have also been shown in Everglades mangrove regions (Thayer et al., 1987; Ley et al., 1999; Lorenz et al., 1999; Faunce et al., 2004). In Florida Bay and Whitewater Bay, Thayer et al. (1987) reported increases in both fish numbers and biomass during the wet season. Our results from NW fit their findings. Fish abundances varied monthly in mangrove creeks of southeastern Florida Bay, with those of freshwater species increasing during February and March (Faunce et al., 2004), as seen in our RB sites.

Several factors may be responsible for the lack of a freshwater species influx in NW headwaters. Marshes upstream of NW creeks have shorter hydroperiods than those upstream of RB sites, and consequently may contain lower densities of fishes, particularly of the large species (Lorenz, 1999; Trexler et al., 2001, 2005; Chick et al., 2004). Freshwater fishes may be absent from NW creeks simply because the pool of potential marsh migrants is small. Secondly, salinity levels are higher in NW than RB headwaters, and may approach or exceed the physiological tolerances or preferences for some of the freshwater species such as centrarchids (Loftus and Kushlan, 1987). However, other marsh inhabitants exhibit high salinity tolerances (Lorenz and Serafy, 2006; Nordlie, 2006), and should find suitable salinity conditions in NW, despite the fact that there were rarely caught there.

Thirdly, the pattern of marsh dry-down differed between regions, and marshes upstream of NW remained flooded beyond our dry-season sample. A pulse of freshwater species could have possibly occurred later in the season, and would have been missed by our sampling. Other studies, however, suggest that marsh fishes move into deep-water refugia well in advance of low-water conditions (Chick et al., 2004; Re-hage and Trexler, 2006). Even in long-hydroperiod marshes that rarely dry, and where direct mortality due to dry-down conditions is unlikely, large-fish densities decrease significantly in the open marsh during the dry season and concentrate in deep-water refuges. Marsh water-levels upstream of NW were low (close to 5 cm); therefore, a pulse of migrants should have occurred by our April sample. Furthermore, salinities in later months exceeded 15 in NW, which is too stressful for many of the potential migrants. Another explanation may be a higher abundance of alternative dry-down refuges (e.g., solution and alligator holes) in marshes upstream of NW relative to RB. However, abiotic (high ammonia and low oxygen) and biotic (high predation) conditions in these alternative refuges are often stressful (Nelson and Loftus, 1996; Kobza et al., 2004), and could make these refuges less preferred relative to creeks. Lastly, small differences in local topography (e.g., the presence of berms along creeks) could limit fish movement in and out of creeks, perhaps to a greater extent in NW than RB sites (Green et al., 2006).

The pulse of freshwater species in RB occurred early in the dry season, and despite the fact that RB assemblages remained dominated by freshwater species in the later sample, their abundances decreased considerably. This decrease could be explained by a large-scale return of the freshwater taxa to marshes, if marshes had reflooded. However, this was unlikely in 2005 since water levels in upstream marshes at the time of our transition and dry season samples were relatively low and similar (11 and 13 cm, respectively). Alternatively, the increase in salinity between the transition and dry samples in RB (from < 1 to 5) could explain the decline in freshwater species through mortality or movement to more suitable salinity environments. An important source of mortality could also be predation. Mangrove habitats provide important foraging grounds for marine and estuarine piscivores (Blaber and Milton, 1990; Chong et al., 1990). Even though mangroves can provide a refuge from predation because of their high habitat complexity (Primavera, 1998; Acosta and Butler, 1999), the abundance of predators is not necessarily lower in these shallow coastal habitats (Sheaves, 2001). In RB creeks, piscine predators such as snook, Florida gar, largemouth bass, and bowfin were abundant early in the dry season and could account for the declines in the cyprinodontoids between February and April. Top predators, such as alligators, wading birds, and bull sharks could possibly account for the decreases in the abundance of the larger freshwater species. More extensive, paired sampling in creeks, marshes (including other dry-down refuges), and downstream portions of the estuary is needed to discriminate between these and other plausible explanations for the timing and extent of pulsing of freshwater taxa into headwater creeks.

Results from studies that rely on a single gear type to sample mangrove fishes may be restricted in their applicability because of gear selectivity (Rozas and Minello, 1997). CPUE in gill nets was appreciably lower than that for electrofishing, and catch composition differed between gears. This suggests that gill nets with the 30-min soak times used in our study do not provide a reliable index of abundance, nor detect seasonal variation in community structure, even if previous studies have shown that gill nets may be better at capturing certain aspects of target fish populations, such as size structure, relative to electrofishing (Colvin, 2002). Comparison of the smallfish CPUE showed that minnow-trap placement in the water column strongly affects catch numbers and species composition. This is likely explained by variation in microhabitat use among small-fish species; a factor that needs to be considered if sampling is targeted to multiple species.

The influx of freshwater species into RB headwater creeks may enhance estuarine fish abundance and richness, and should provide an important prey source for marine and estuarine piscine predators, as well as for avian predators. Interannual variation in drying patterns may create circumstances in which other creek headwater habitats (including NW) may serve as dry-season refugia. In other parts of the ecotone, factors such as high salinity and local topography could limit the connectivity between mangrove and upstream marsh habitats (i.e., Green et al., 2006). Ongoing restoration of the Greater Everglades ecosystem aimed at re-establishing historical freshwater flows (CERP, 1999), could greatly enhance this connectivity. Increased freshwater flows are expected to result in reduced salinities, prolonged pooling of freshwater, and a spatially-expanded and seasonally-extended oligohaline zone at that marsh-mangrove ecotone, including our study creeks (Davis et al., 2005). These conditions should make large portions of the mangrove region suitable for freshwater species. In northern parts of Florida Bay, increased freshwater flow has resulted in higher abundance and biomass of small-bodied freshwater taxa, and thus the recovery of the demersal forage fish community (Lorenz, 1999; Lorenz and Serafy, 2006). Similar effects could occur in our study area in southwestern Everglades, but overall responses of freshwater fishes are somewhat uncertain. Increased freshwater flow and decrease salinity are expected the influence multiple components of marine, estuarine, and freshwater food webs and how these interact over a complex and heterogeneous ecotonal landscape. Further research is needed to develop predictions and gain a better understanding of the net effects of hydrological restoration on the freshwater fish community of mangrove headwaters.

Acknowledgments

This study was funded by the Army Corps of Engineers (ACOE), Jacksonville District, as part of the Comprehensive Everglades Restoration Plan Monitoring and Assessment Program (Work Order 12) and supported by NSF grant No. DEB-9910514. We thank E. Kurzbach and K. Luisi of the ACOE, P. Telis, D. Elswick, and C. Fadeley of USGS, and J. Lorenz and J. Wolkowsky of Audubon of Florida for their support. We thank B. Zepp and P. Teague of ENP for their assistance with permitting and GIS. We are grateful to B. Dunker and B. Shamblin for their assistance with field work, and K. Dunker for her assistance with sampling processing.

LITERATURE CITED

- Acosta, C. A. and M. J. Butler IV. 1999. Adaptive strategies that reduce predation on spiny lobster postlarvae during onshore transport. Limnol. Oceanogr. 44: 494–501.
- Argent, D. G. and W. G. Kimmel. 2005. Efficiency and selectivity of gill nets for assessing fish community composition of large rivers. North Am. J. Fish. Manage. 25: 1315–1320.
- Barletta, M., A. Barletta-Bergan, U. Saint-Paul, and G. Hubold. 2005. The role of salinity in structuring the fish assemblages in a tropical estuary. J. Fish Biol. 66: 45–72.
- Beck, M. W., K. L. Heck Jr., K. W. Able, D. L. Childers, D. B. Eggleston, B. M. Gillanders, B.
- Halpern, C. G. Hays, K. Hoshino, T. J. Minello, R. J. Orth, P. F. Sheridan, and M. P. Weinstein. 2001. The identification, conservation and management of estuarine and marine nurseries for fish and invertebrates. BioScience 51: 633–641.
- Blaber, S. J. M. and D. A. Milton. 1990. Species composition, community structure and zoogeography of fishes of mangrove estuaries in the Solomon Islands. Mar. Biol. 105: 259–267.
- Burkhardt, R. W. and S. Gutreuter. 1995. Improving electrofishing catch consistency by standardizing power. North Am. J. Fish. Manage. 15: 375–381.
- Comprehensive Everglades Restoration Plan (CERP). 1999. U.S. Army Corps of Engineers and South Florida Water Management District. Jacksonville FL. Available from: http://www. evergladesplan.org/pub/restudy_eis.aspx.
- Chaves, P. and J. L. Bouchereau. 2000. Use of mangrove habitat for reproductive activity by the fish assemblage in the Guaratuba Bay, Brazil. Oceanol. Acta 23: 273–280.
- Chick, J. H., S. Coyne, and J. C. Trexler. 1999. Effectiveness of airboat electrofishing for sampling fishes in shallow vegetated habitats. North Am. J. Fish. Manage. 19: 957–967.
 - _____, C. R. Ruetz, and J. C. Trexler. 2004. Spatial scale and abundance patterns of large fish communities in freshwater marshes of the Florida Everglades. Wetlands 24: 652–664.

- Chong, V. C., A. Sasekumar, M. U. C. Leh, and R. D' Cruz. 1990. The fish and prawn communities of a Malaysian coastal mangrove system, with comparison to adjacent mud flats and inshore waters. Estuar. Coast. Shelf Sci. 31: 703–722.
- Clarke, K. R. and R. M. Warwick. 2001. Changes in marine communities: an approach to statistical analyses and interpretation, 2nd edition. National Environmental Research Council, Plymouth Marine Laboratory, Plymouth, United Kingdom. 124 p.
- Colvin, M. 2002. A comparison of gill netting and electrofishing as sampling techniques for white bass in Missouri's large reservoirs. North Am. J. Fish. Manage. 22: 690–702.
- Davis, S. M., D. L. Childers, J. J. Lorenz, H. R. Wanless, and T. E. Hopkins. 2005. A conceptualmodel of ecological interactions in the mangrove estuaries of the Florida Everglades. Wetlands 25: 832–842.
- DeAngelis, D. L., W. F. Loftus, J. C. Trexler, and R. E. Ulanowicz. 1997. Modeling fish dynamics in a hydrologically pulsed ecosystem. J. Aquatic Ecosyst. Stress and Recov. 6: 1–13.
- Dorenbosch, M., M. C. van Riel, I. Nagelkerken, and G. van der Velde. 2004. The relationship of reef fish densities to the proximity of mangrove and seagrass nurseries. Estuar. Coast. Shelf Sci. 60: 37–48.
- Ellis, W. L. and S. S. Bell. 2004. Conditional use of mangrove habitats by fishes: depth as a cue to avoid predators. Estuaries 27: 966–976.
- Faunce, C. H. and J. E. Serafy. 2006. Mangroves as fish habitat: 50 years of field studies. Mar. Ecol. Prog. Ser. 318: 1–18.
 - _____, ____, and J. J. Lorenz. 2004. Density-habitat relationships of mangrove
- creek fishes within the southeastern saline Everglades (USA), with reference to managed water releases. Wetland Ecol. Manag. 12: 377–394.
- Fennema, R. J., C. J. Neidrauer, R. A. Johnson, T. K. MacVicar, and W. A. Perkins. 1994. A computer model to simulate natural Everglades hydrology. Pages 249–289 in S. M. Davis and J. C. Ogden, eds. Everglades: the ecosystem and its restoration. St. Lucie Press, Delray Beach.
- Green, D. P. J., J. C. Trexler, J. J. Lorenz, C. C. McIvor, and T. Philippi. 2006. Spatial patterns of fish communities along two estuarine gradients in southern Florida. Hydrobiologia 569: 387–399.
- Gillanders, B. M., K. W. Able, J. A. Brown, D. B. Eggleston, and P. F. Sheridan. 2003. Evidence of connectivity between juvenile and adult habitats for mobile marine fauna: an important component of nurseries. Mar. Ecol. Prog. Ser. 247: 281–295.
- Gunderson, L. H. and W. F. Loftus. 1993. The Everglades. Pages 199–255 *in* W. H. Martin, S. G. Boyce, and A. C. Echternacht, eds. Biodiversity of the Southeastern United States. John Wiley and Sons, New York.
- Hindell, J. S. and G. P. Jenkins. 2004. Spatial and temporal variability in the assemblage structure of fishes associated with mangroves, *Avicennia marina*, and intertidal mudflats in temperate Australian embayments. Mar. Biol. 144: 385–395.
- Hubert, W. A. and D. T. O'Shea. 1992. Use of spatial resources by fishes in Grayrocks Reservoir, WY. J. Freshw. Ecol. 7: 219–225.
- Kobza, R. M, J. C. Trexler, W. F. Loftus, and S. A. Perry. 2004. Community structure of fishes inhabiting aquatic refuges in a threatened Karst wetland and its implications for ecosystem management. Biol. Conserv. 13: 898–911.
- Kuo, S. R., H. J. Lin, and K. T. Shao. 1999. Fish assemblages in the mangrove creeks of northern and southern Taiwan. Estuaries 22: 1004–1015.
- Kupschus, S. and D. Tremain. 2001. Associations between fish assemblages and environmental factors in nearshore habitats of a subtropical estuary. J. Fish Biol. 58: 1383–1403.
- Kushlan, J. A. 1974. Effects of a natural fish kill on the water quality, plankton, and fish population of a pond in the Big Cypress Swamp, Florida. Trans. Am. Fish. Soc. 103: 235–243.
- Laegdsgaard, P. and C. R. Johnson. 1995. Mangrove habitats as nurseries: unique assemblages of juvenile fish in subtropical mangroves in eastern Australia. Mar. Ecol. Prog. Ser. 126: 67–81.

_____ and _____. 2001. Why do juvenile fish utilize mangrove habitats? J. Exp. Mar. Biol. Ecol. 257: 229–253.

- Laroche, J., E. Baran, and N. B. Rasoanandrasana. 1997. Temporal patterns in fish assemblage of a semiarid mangrove zone in Madagascar. J. Fish. Biol. 51: 3–20.
- Layman, C. A. and D. E. Smith. 2001. Sampling bias of minnow traps in shallow aquatic habitats on the eastern shore of Virginia. Wetlands 21: 145–154.
- Levesque, V. A. 2004. Water flow and nutrient influx from five estuarine rives along the southwest coast of the Everglades National Park, Florida, 1997–2001. U.S. Geological Survey Scientific Investigations Report 2004-5142.
- Ley, J. A., C. C. McIvor, and C. L. Montague. 1999. Fishes in mangrove prop-root habitats of northeastern Florida Bay: distinct assemblages across an estuarine gradient. Estuar. Coast. Shelf Sci. 48: 701–723.
- Light, S. S. and J. W. Dineen. 1994. Water control in the Everglades: a historical perspective. Pages 47–84 *in* S. M. Davis and J. C. Ogden, eds. Everglades: the system and its restoration. St. Lucie Press, Delray Beach.
- Lin, H. J. and K. T. Shao. 1999. Seasonal and diel changes in a subtropical mangrove fish assemblage. Bull. Mar. Sci. 65: 775–794.
- Loftus, W. F. 2000. Inventory of fishes of Everglades National Park. Fla. Sci. 63: 27-47.
 - and A. M. Eklund. 1994. Long-term dynamics of an Everglades fish community. Pages 461–483 *in* S. M. Davis and J. C. Ogden, eds. Everglades: the system and its restoration. St. Lucie Press, Delray Beach.

- Loneragan, N. R., I. C. Potter, R. C. J. Lenanton, and N. Caputi. 1986. Spatial and seasonal differences in the fish fauna of the shallows in a large Australian estuary. Mar. Biol. 92: 575–586.
- Lorenz, J. J. 1999. The response of fishes to physiochemical changes in the mangroves of northeast Florida Bay. Estuaries 22: 500–517.

______ and J. E. Serafy. 2006. Subtropical wetland fish assemblages and changing salinity regimes: implications for Everglades restoration. Hydrobiologia 569: 401–422.

- Lugo, A. and S. Snedaker. 1974. The ecology of mangroves. Ann. Rev. Ecol. System. 5: 39-64.
- Mason, F. J., N. R. Loneragan, G. A. Skilleter, and S. R. Phinn. 2005. An evaluation of the evidence for linkages between mangroves and fisheries: a synthesis of the literature and identification of research directions. Oceanogr. Mar. Biol. Annu. Rev. 43: 485–515.
- McIvor, C. C., J. A. Ley, and R. D. Bjork. 1994. Changes in freshwater inflow from the Everglades to Florida Bay including effects on biota and biotic processes: a review. Pages 533–570 *in* S. M. Davis and J. C. Ogden, eds. Everglades: the system and its restoration. St. Lucie Press, Delray Beach.
- Montague, C. L. and J. A. Ley. 1993. A possible effect of salinity fluctuation on abundance of benthic vegetation and associated fauna in northeastern Florida Bay. Estuaries 16: 703– 717.
- Mumby, P. J., A. J. Edwards, J. E. Arlas Gonzales, K. C. Lindeman, P. G. Blackwell, A. Gall, M. I. Gorczynska, A. R. Harborne, C. L. Pescod, H. Renken, C. C. C. Wabnitz, and G. Llewellyn. 2004. Mangroves enhance the biomass of coral reef fish communities in the Caribbean. Nature 427: 533–536.
- Nagelkerken, I., G. van der Velde, M. W. Gorrissen, G. J. Meijer, T. van't Hof, and C. den Hartog. 2000. Importance of mangroves, seagrass beds, and shallow coral reef as a nursery for important coral reef fishes, using a visual census technique. Estuar. Coast. Shelf Sci. 51: 31–44.

, C. M. Roberts, G. van der Velde, M. Dorenbosch, M. C. van Riel, E. Cocheret de la Morinière, and P. H. Nienhuis. 2002. How important are mangrove and seagrass beds for coral-reef fish? The nursery hypothesis tested on an island scale. Mar. Ecol. Prog. Ser. 244: 299–305.

______ and J. A. Kushlan. 1987. Freshwater fishes of southern Florida. Bull. Fla. Mus. Nat. Hist. Biol. Sci. 31: 147–344.

- Nelson, C. M. and W. F. Loftus. 1996. Effects of high-water conditions on fish communities in Everglades alligator ponds. Pages 89–101 in T. V. Armentano, ed. Proc. 1996 conference: ecological assessment of the 1994–1995 high water conditions in the southern Everglades. Florida International University, Miami, FL.
- Nordlie, F. G. 2003. Fish communities of estuarine salt marshes of eastern North America, and comparisons with temperate estuaries of other continents. Rev. Fish Biol. Fish. 13: 281–325.

_____. 2006. Physicochemical environments and tolerances of cyprinodontoid fishes found in estuaries and salt marshes of eastern North America. 2006. Rev. Fish. Biol. Fish. 16: 51–106.

- Odum, W. E. 1971. Pathways of energy flow in a south Florida estuary. Sea Grant Tech. Bull. No. 7. Univ. of Miami Sea Grant Program, Miami, FL. 162 p.
- Ogden, J. C. 1994. A comparison of wading bird nesting colony dynamics (1931–1946 and 1974–1989) as an indication of ecosystem conditions in the southern Everglades. Pages 533–570 *in* S. M. Davis and J. C. Ogden, eds. Everglades: the system and its restoration. St. Lucie Press, Delray Beach.
- Pinto, L. and N. N. Punchihewa. 1996. Utilization of mangroves and seagrasses by fishes in the Negombo estuary, Sri Lanka. Mar. Biol. 126: 333–345.
- Primavera, J. H. 1998. Mangroves as nurseries: shrimp populations in mangrove and non-mangrove habitats. Estuar. Coast. Shelf Sci. 46: 457–464.
- Rehage, J. S. and J. C. Trexler. 2006. Assessing the net effect of anthropogenic disturbance on aquatic communities in wetlands: community structure relative to distance from canals. Hydrobiologia 569: 359–373.
- Robertson, A. I. and N. C. Duke. 1987. Mangroves as nursery sites: comparisons of the abundance and species composition of fish and crustaceans in mangroves and other nearshore habitats in tropical Australia. Mar. Biol. 96: 193–205.

______, P. Dixon, and P. A. Daniel. 1988. Zooplankton dynamics in mangrove and other nearshore habitats in tropical Australia. Mar. Ecol. Prog. Ser. 43: 139–150.

- Rozas, L. P. and T. J. Minello. 1997. Estimating densities of small fishes and decapod crustaceans in shallow estuarine habitats: A review of sampling design with focus on gear selection. Estuaries 20: 199–213.
- Serafy, J. E., C. H. Faunce, and J. J. Lorenz. 2003. Mangrove shoreline fishes of Biscayne Bay, Florida. Bull. Mar. Sci. 72: 161–180.

______, K. C. Lindeman, T. E. Hopkins, and J. S. Ault. 1997. Effects of freshwater canal discharge on fish assemblages in a subtropical bay: field and laboratory observations. Mar. Ecol. Prog. Ser. 160: 161–172.

Sheaves, M. J. 2001. Are there really few piscivorous fishes in shallow estuarine habitats? Mar. Ecol. Prog. Ser. 222: 272–290.

______. 2005. Nature and consequences of biological connectivity in mangrove systems. Mar. Ecol. Prog. Ser. 302: 293–305.

Sheridan, P. F. 1997. Benthos of adjacent mangrove, seagrass, and non-vegetated habitats in Rookery Bay, Florida, U.S.A. Estuar. Coast. Shelf Sci. 44: 455–469.

_____ and C. Hays. 2003. Are mangroves nursery habitat for transient fishes and decapods? Wetlands 23: 449–458.

Smith, T. J. III, M. B. Robblee, H. R. Wanless, and T. W. Doyle. 1994. Mangroves, hurricanes, and lighting strikes. Bioscience 44: 256–262.

, J. H. Hudson, M. B. Robblee, G. V. N. Powell, and P. J. Isdale. 1989. Freshwater flow from the Everglades to Florida Bay: a historical reconstruction based on fluorescent banding in the coral *Solenastrea bournoni*. Bull. Mar. Sci. 44: 274–282.

- Strahler, A. N. 1957. Quantitative analysis of watershed geomorphology. Amer. Geophys. Union T. 38: 913–920.
- Tabb, D. C. and R. B. Manning. 1961. Checklist of flora and fauna of north Florida Bay. Bull. Mar. Sci. 11: 552–640.

- Thayer, G. W., D. R. Colby, and W. F. Hettler. 1987. Utilization of the red mangrove prop root habitat by fishes in South Florida. Mar. Ecol. Prog. Ser. 35: 25–38.
- Trexler, J. C., W. F. Loftus, and S. Perry. 2005. Hydrological limitation of Everglades fish communities by a twenty-five year intervention study. Oecologia 145: 140–152.

, W. F. Loftus, C. F. Jordan, J. H. Chick, K. L. Kandl, T. C. McElroy, and O. L. Bass. 2001. Ecological scale and its implications for freshwater fishes in the Florida Everglades. Pages 153–181 *in* J. W. Porter and K. G. Porter, eds. The Everglades, Florida Bay, and coral reefs of the Florida Keys: An ecosystem sourcebook. CRC Press, Boca Raton.

Yañez-Arancibia, A., A. L. Lara-Domínguez, J. L. Rojas-Galaviz, P. Sánchez-Gil, J. W. Day, and C. J. Madden. 1988. Seasonal biomass and diversity of estuarine fishes coupled with tropical habitat heterogeneity (southern Gulf of Mexico). J. Fish. Biol. 33: 191–200.

ADDRESSES: (J.S.R.) Nova Southeastern University, Oceanographic Center, 8000 North Ocean Drive, Dania Beach, Florida 33004-3078. (W.F.L.) U.S. Geological Survey, Florida Integrated Science Center, Everglades National Park Field Station, 40001 State Road 9336, Florida 33034. CORRESPONDING AUTHOR: (J.S.R.) Email: <rehage@nova.edu>.

