

Northumbria Research Link

Citation: Rupasinghe, Vasantha, Sekhlon-Loodu, Satvir, Mantso, Theodora and Panagiotidis, Mihalis (2016) Phytochemicals in regulating fatty acid β -oxidation: Potential underlying mechanisms and their involvement in obesity and weight loss. *Pharmacology & Therapeutics*, 165. pp. 153-163. ISSN 0163-7258

Published by: Elsevier

URL: <http://dx.doi.org/10.1016/j.pharmthera.2016.06.005>
<<http://dx.doi.org/10.1016/j.pharmthera.2016.06.005>>

This version was downloaded from Northumbria Research Link:
<http://nrl.northumbria.ac.uk/28085/>

Northumbria University has developed Northumbria Research Link (NRL) to enable users to access the University's research output. Copyright © and moral rights for items on NRL are retained by the individual author(s) and/or other copyright owners. Single copies of full items can be reproduced, displayed or performed, and given to third parties in any format or medium for personal research or study, educational, or not-for-profit purposes without prior permission or charge, provided the authors, title and full bibliographic details are given, as well as a hyperlink and/or URL to the original metadata page. The content must not be changed in any way. Full items must not be sold commercially in any format or medium without formal permission of the copyright holder. The full policy is available online: <http://nrl.northumbria.ac.uk/policies.html>

This document may differ from the final, published version of the research and has been made available online in accordance with publisher policies. To read and/or cite from the published version of the research, please visit the publisher's website (a subscription may be required.)

www.northumbria.ac.uk/nrl



A review:

Phytochemicals in regulating fatty acid β -oxidation: Potential underlying mechanisms and their involvement in obesity and weight loss

H.P. Vasantha Rupasinghe^{1,2*}, Satvir Sekhon-Loodu¹, Theodora Mantso³, Mihalis I. Panayiotidis³

¹Department of Environmental Sciences, Faculty of Agriculture, Dalhousie University, P.O. Box 550, Truro, Nova Scotia B2N 5E3, Canada;

²Department of Pathology, Faculty of Medicine, Dalhousie University, Halifax, Nova Scotia B3H 4R2, Canada;

³Heriot-Watt University, School of Life Sciences John Muir Building, Riccarton Campus, Edinburgh, EH14 4AS, Scotland, UK.

*To whom correspondence should be addressed:

H.P. Vasantha Rupasinghe

Department of Environmental Sciences

Faculty of Agriculture, Dalhousie University

Truro, NS B2N 5E3, Canada

Tel: +1 902 893 6623

Fax: +1 902 893 1404

E-mail: vrupasinghe@dal.ca.

Abstract

Excessive accumulation of fat as the result of more energy intake and less energy expenditure is known as obesity. Lipids are essential components in the human body and are vital for maintaining homeostasis, physiological as well as cellular metabolism. Fatty acid synthesis and catabolism (by fatty acid oxidation) are normal part of basic fuel metabolism in animals. Fatty acids are degraded in the mitochondria by a biochemical process called β -oxidation in which two-carbon fragments are produced in each cycle. The increase in fatty acid oxidation is positively correlated with body mass index. Although healthy life style, avoiding Western diet, dieting and strenuous exercise are commonly used methods to lose weight, they are not considered a permanent solution in addition to risk attenuation of in basal metabolic rate (BMR). Pharmacotherapy offers benefits of weight loss by altering the satiety and lowering absorption of fat from the food; however, its side effects may outweigh the benefits of weight loss. Alternatively, dietary phytochemicals and natural health products offer great potential as an efficient weight loss strategy by modulating lipid metabolism and/or increasing BMR and thermogenesis. Specifically, polyphenols such as citrus flavonoids, green tea epigallocatechin gallate, resveratrol, capsaicin and curcumin, have been reported to increase lipolysis and induce fatty acid β -oxidation through modulation of hormone sensitive lipase, acetyl-coA carboxylase, carnitine acyl transferase and peroxisome proliferator-activated receptor gamma coactivator-1. In this review article, we discuss selected phytochemicals in relation to their integrated functionalities and specific mechanisms for weight loss.

Key words: Lipid metabolism, obesity, weight loss, beta-oxidation, phytochemicals, epigenetics

Abbreviations

Acetyl-CoA Carboxylase, ACC

Acyl CoA Synthetase, ACS

5' Adenosine Monophosphate-Activated Protein Kinase, AMPK

Activated or phosphorylated AMPK, AMPK-P

Body Mass Index, BMI

Carnitine Palmitoyl Transferase, CPT

Carnitine Palmitoyl Transferase-1, CPT-1

Carnitine Palmitoyl Transferase-1B, CPT-1B

Citrate Lyase, CL

Cyclic Adenosine Monophosphate, cAMP

DNA Methyltransferases, DNMTs

Fatty Acid, FA

Fatty Acid Oxidation, FAO

Fatty Acid Synthase, FAS

Glycerol-3-Phosphate Acyl Transferase-1, GPAT-1

Histone Acetyltransferases, HATs

Histone Deacetylases, HDACs

Hormone Sensitive Lipase, HSL

Hydroxycitric Acid, HCA

Lipoprotein Lipase, LPL

Mitochondrial Electron Transport Chain, ETC

Mitochondrial Uncoupling Protein-2, UCP-2

Peroxisome Proliferator-Activated Receptor-Gamma, PPAR- γ

Peroxisome Proliferator-Activated Receptor δ , PPAR δ

Raspberry Ketone, RK

Respiratory Quotient, RQ

Small Non-Coding RNAs, miRNAs

Triacylglycerol, TAG

Tricarboxylic Acid, TCA

Table of Contents

1. Introduction
2. β -oxidation and its regulation
3. Metabolic understanding of obesity
4. Possible mechanisms of weight loss
5. Phytochemicals stimulating β -oxidation
 - 5.1 Epigenetic properties of phytochemicals
 - 5.2 Classes of phytochemicals
 - 5.2.1 Green tea catechins
 - 5.2.2 Resveratrol
 - 5.2.3 Capsaicinoids
 - 5.2.4 Citrus flavonoids
 - 5.2.5 Piperine
 - 5.2.6 Anthocyanins
 - 5.2.7 Curcumin
 - 5.2.8 Raspberry ketones
 - 5.2.9 Cocoa polyphenols
 - 5.2.10 Soyabean phytochemicals
 - 5.2.11 Hydroxycitric acid
6. Conclusions
7. Acknowledgements
8. References

1 **1. Introduction**

2 The prevalence of obesity has progressively increased over the past 30 years worldwide
3 especially in the Western countries. Obesity is a condition characterized by accumulation of
4 excessive body fat. It is classified by body mass index (BMI) [a ratio of body weight (in kg) to
5 height (in meter squared)] in a way where individuals with a value over 30 are considered obese
6 (Witkamp, 2011; Bessesen, 2008). Obesity is an alarming indicator of onset of metabolic disorder
7 which is a cluster of health complications including hypertension, type 2 diabetes and
8 cardiovascular disease (Faulds et al., 2012; Salas-Salvado et al., 2011). Therefore, apart from
9 personal interest, the treatment of obesity is of clinical significance.

10 Weight management is a commonly recommended approach which is based on lifestyle
11 modifications including dieting, increased physical activity, exercise, etc. However, physical
12 exercise and dieting is often a difficult routine to maintain for lifetime and the results can be
13 disappointing in long term. At present, the combination therapy of reducing calorie intake,
14 increased energy expenditure and pharmacotherapy is becoming more popular. To this end, several
15 drugs such as Fenfluramine, R-Fenfluramine, Temin, Sibutramine, Orlistat, Qsymia, and Belviq
16 have been approved, by FDA, towards weight management aid. However, four of these were
17 removed later on due to their adverse health effects (WHO, 2000). In addition, all current weight
18 management drugs in the market have high cost as well as potential side effects thus causing
19 dissatisfaction to the consumers. Finally, gastric surgery has had the most effective approach in
20 severely obese to show long term effects.

21 Despite the progress in weight management strategy in recent years, obesity still poses a
22 serious challenge for the scientific community (WHO, 2000; González-Castejón and Rodríguez-
23 Casado, 2011). Therefore, there is considerable demand to explore natural therapies in developing

24 an alternative, safer and effective strategy. For this reason, a variety of natural phytochemicals
25 have been explored for their ability to increase fatty acid oxidation, fat absorption and suppress
26 appetite control. This review article will focus on most recent evidence on those phytochemicals
27 that potentially increase fatty acid β -oxidation in relation to weight loss.

28

29 **2. Fatty acid β -oxidation and its regulation**

30 Fats are stored in our body as triacylglycerols (TAG) which are hydrolyzed into free fatty
31 acids and glycerol by lipases as the first step of lipid catabolism. The fatty acid β -oxidation
32 pathway consists of multistep reactions which oxidizes fatty acids by degrading two carbons at a
33 time (Fig. 1). It takes place in mitochondria and peroxisomes, in eukaryotes, and it is a major
34 source of energy supply by providing more energy as compared to equivalent amount of glucose.
35 In the peroxisomes, long-chain fatty acids are converted to acyl CoA which cannot diffuse across
36 the inner mitochondrial membrane to be utilized for the fatty acid β -oxidation pathway. Therefore,
37 a transport system is required, called the carnitine shuttle system, catalyzed by carnitine
38 acyltransferase-1 or carnitine palmitoyltransferase-1 (CPT-1). While in cytosol, fatty acyl CoA is
39 converted into acylcarnitine (by carnitine acyltransferase I) which enters the mitochondrial matrix
40 and fatty acyl CoA is regenerated by a reaction catalyzed by carnitine acyltransferase II (Horton
41 et al., 2006). **Beta-oxidation is catalyzed by the sequential action of four enzyme families: acyl
42 CoA dehydrogenase (E1), enoyl CoA hydratase (E2), 3-hydroxy acyl CoA dehydrogenase (E3),
43 and 3-ketoacyl CoA thiolase (E4) (Fig. 1).**

44 Acetyl-CoA carboxylase (ACC) plays as central element both in fatty acid β -oxidation and
45 fatty acid biosynthesis. ACC catalyzes the carboxylation of acetyl-CoA producing malonyl-CoA,
46 which can be used by fatty acid synthase for fatty acid biosynthesis. As malonyl-CoA is the

47 substrate for fatty acid biosynthesis, malonyl-CoA is also a direct inhibitor of mitochondrial fatty
48 acid uptake as well as inhibition of CPT-1. 5' Adenosine monophosphate-activated protein kinase
49 (AMPK) regulates fatty acid metabolism by phosphorylation-induced inhibition of ACC activity
50 and eventually stimulate fatty acid β -oxidation and down-regulate fatty acid biosynthesis (Fig. 1)
51 (Lopaschuk et al., 2010).

52

53 **3. Metabolic understanding of obesity**

54 Cellular energy is produced from energy sources in the mitochondria. The major two
55 sources of energy in a human body are carbohydrates and fatty acids. The body produces energy
56 in the form of ATP by oxidation of carbohydrates, fats and proteins through tricarboxylic acid
57 (TCA) cycle; and by fatty acid oxidation through β -oxidation. The body derives energy for its
58 cellular processes by breaking down ATP to ADP and AMP. Under normal conditions, more ATP
59 is produced through β -oxidation of fatty acids in the mitochondria as compared to carbohydrates.
60 The first requirement in fatty acid β -oxidation is the presence of fatty acyl-CoA and its transport
61 into the mitochondria facilitated by CPT-1 (a rate-limiting step for β -oxidation) (McGarry et al.,
62 1983; Eaton et al., 2001). Malonyl-CoA (a precursor of fatty acid synthesis) is a competitive
63 inhibitor of CPT-1 meaning that when an energy level is high, it prevents fatty acid oxidation
64 whereas when energy level is low, malonyl and acetyl CoA levels fall and consequently β -
65 oxidation is induced by the activation of CPT-1 (Zammit et al., 1999). Therefore, the enzymes
66 CPT-1 and fatty acid synthase (FAS) directly regulate catabolism and anabolism of fatty acids
67 (Ronnett et al., 2005). In addition, glucose oxidation directly inhibits fatty acid oxidation in a
68 manner characterized by an insulin dependent response where a high glucose level (after a meal)
69 is regulated by insulin thus facilitating glucose uptake in the cells and consequently inhibiting

70 lipolysis and β -oxidation. Furthermore, low circulating levels of glucose and increased energy
71 demand can both stimulate cellular fatty acid β -oxidation pathway (Smith, 1994).

72 Obesity can lead to impaired cellular metabolism including dependence on glucose
73 oxidation (for ATP production) and decrease in fatty acid oxidation, thus leading to more fat
74 deposition in skeletal muscles, hepatocytes and other cells (Rogge, 2009). The reduced fatty acid
75 oxidation can be marked relative to the respiratory quotient (RQ). This way, when energy is
76 produced from fats (by β -oxidation) more oxygen is consumed and the RQ is low (e.g. 0.7) and
77 alternatively, when carbohydrates are the main source of ATP generation in the body, less oxygen
78 is consumed and the RQ is high (e.g. 1.0). Obese individuals have been reported to have high RQ
79 values, indicating low fat oxidation and thus more dependence on glucose than lean individuals
80 (Filozof et al., 2000; Simoneau et al., 1999). Therefore, reduced fatty acid oxidation is considered
81 as a risk factor for the development of obesity. Other studies indicate that obese individuals have
82 reduced CPT-1 activity, which impairs the flow of fatty acid transfer to mitochondria and hereby
83 reduce β -oxidation, suggesting that fatty acids cannot be oxidized even after lipolysis if CPT-1 is
84 not activated (Simoneau et al., 1999; Rogge 2009).

85

86 **4. Possible mechanisms of weight loss**

- 87 i. One of the popular approach of weight loss is through appetite control. The food urge and
88 satiety is controlled by serotonin, histamine, dopamine and their receptors. Sibutramine is
89 an anti-obesity drug which functions as appetite suppressant; however, coupled with
90 various side effects such as dry mouth, constipation and insomnia (Tziomalos et al., 2009).
- 91 ii. Stimulated energy expenditure can be used to reduce body weight by induction of non-
92 shivering thermogenesis. Thermogenesis is mainly regulated by leakage of protons

93 generated in oxidative phosphorylation, bypassing ATP generation and activating UCP-1
94 which thereby, dissipates energy as heat (Kumar et al., 1999). UCP-1 is expressed in
95 mitochondria-rich brown adipose tissue. Likewise, UCP-3 also mediates thermogenesis
96 regulated by the thyroid hormone, β -adrenergic receptor agonist and leptin (Gong et al.,
97 2000). The function of UCP family was demonstrated in a mice study, where the mice
98 over-expressing UCP-1, UCP-2 and UCP-3 were resistant to diet-induced obesity;
99 however, they were susceptible to cold due to the lack of thermogenesis (Arsenijevic et al.,
100 2000, Gong et al., 2000).

101 iii. Adipocytes increase in size and differentiate when fat storage increases under obesity.
102 Thus, the compounds that inhibit adipocyte differentiation and induce apoptosis in mature
103 adipocytes can be considered as potentially promising anti-obesity agents (Kim et al., 2006;
104 Yun, 2010).

105 iv. Many pharmaceutical drugs stimulate triacylglycerol hydrolysis and release fatty acids.
106 Lipolysis diminishes storage fat (leading to dyslipidemia) thus, intervening the β -
107 adrenergic receptor agonist is required to oxidize the released fatty acids (Langin, 2006).

108 v. In lipid metabolism, peroxisome proliferator-activated receptor gamma (PPAR- γ) and 5'
109 adenosine monophosphate-activated protein kinase (AMPK) play crucial roles. PPAR- γ is
110 a transcriptional factor (mediating gene expression) predominately expressed in adipose
111 tissue that stimulates adipose differentiation. Therefore, PPAR- γ agonists can ameliorate
112 dyslipidemia, as well as improve adiposity and insulin resistance (Cornalius et al., 1994).

113 vi. AMPK is an enzyme which regulates the target proteins controlling metabolism. AMPK
114 activation regulates glucose transport and fatty acid oxidation. Increase in AMPK in muscle
115 stimulates CPT-1 production and eventually increases fatty acid oxidation (Lee et al.,

116 2005). Activation of AMPK by exercise and fuel deprivation (AMP:ATP ratio) have led to
117 studies of the effects of AMPK on lipid metabolism, obesity and metabolic syndrome–
118 related diseases (O’Neill, 2013).

119 vii. One of the most promising approaches to weight management is the decrease in fat
120 absorption. In gastrointestinal tract, before fat gets absorbed, it is subjected to the action of
121 pancreatic lipase with its inhibition being a clinically approved strategy for controlling
122 obesity. One such drug compound is Orlistat; however, also associated with certain side
123 effects like oily spotting, liquid stools, abdominal cramps, etc. (Chaput et al., 2007).

124 viii. SIRT1 and SIRT3 belong to the sirtuin family of the silent information regulator 2
125 enzymes which have been found to regulate insulin secretion as well as lipid metabolism.
126 SIRT1 plays an important role in regulation of obesity during fasting and feeding
127 (Chalkiadaki and Guarente, 2012; Guarente, 2006). Its major role is played in hepatic fatty
128 acid metabolism, at various steps such as activation of the AMPK/LKB1 pathway thus
129 facilitating fatty acid oxidation (Hou et al., 2008). The specific action of SIRT1 in
130 regulating PPAR- α was demonstrated in mice studies when hepatocyte specific deletion of
131 the SIRT1 gene led to decreased rate of fatty acid oxidation (Purushotham et al., 2009). On
132 the other hand, SIRT3 directly regulates hydroxyacyl-CoA dehydrogenase, acyl-CoA
133 dehydrogenases and deacetylates as well as activates acyl-CoA synthetase short-chain
134 hereby, modulating β -oxidation (Hallows et al., 2011, Hirschev et al., 2010, Hallows et al.,
135 2006).

136 **5. Phytochemicals stimulating fatty acid β -oxidation**

137 Nutritional supplements have been claimed to increase energy metabolism, reduce fat
138 absorption, increase fat oxidation all of which thereby increase weight loss and consequently

139 described popularly as fat burners (Jeukendrup and Randall, 2011). The majority of the ingredients
140 used in these nutritional supplements are from plant origin and commonly referred as
141 phytochemicals. These phytonutrients are secondary metabolites produced by plants and play a
142 central role in defensive mechanism against stress, pathogens, herbivores and disease conditions.
143 Phytochemicals are divided into polyphenols, alkaloids and isoprenoids on the basis of their basic
144 structure and biosynthesis (Table 1). The list of phytochemicals capable of facilitating weight loss
145 by reducing appetite suppressants and/or fat absorption is still on-growing; however, not all of
146 them regulate fatty acid β -oxidation. Thus, it is within the scope of this review article to focus on
147 those phytochemicals capable of influencing the β -oxidation pathway (Table 2).

148

149 **5.1 Epigenetic properties of phytochemicals**

150 Over the past few decades, there is a growing interest in investigating and understanding
151 the beneficial properties of phytochemicals. A number of studies have revealed that the presence
152 of phytochemicals is responsible for exerting a plethora of different biological effects such as
153 antioxidant, anti-inflammatory, anti-aging, anti-proliferative, etc. To this end, after their isolation
154 and characterisation, there is a continuously increasing trend towards promoting their utilization
155 in various fields of biology and medicine such as drug design, disease therapy, cosmeceuticals,
156 nutrition/dietetics, etc. (Su et al., 2013, Szarc del Szic et al., 2015). In recent years, emerging
157 reports have provided evidence that phytochemicals can exert their advantageous effects by
158 targeting epigenetic mechanisms via regulation of specific epigenetic components such as DNA
159 methyltransferases (DNMTs), histone deacetylases (HDACs), histone acetyltransferases (HATs)
160 and small non-coding RNAs (miRNAs) (Guo et al., 2015, Shankar et al., 2013). Epigenetic
161 modifications are defined as reversible and heritable alterations in gene expression without

162 changes in the DNA sequence. The most common types are DNA methylation as well as histone
163 acetylation, deacetylation and methylation all of which are capable for modulating gene
164 expression. In addition, miRNAs have been implicated in several cellular processes while at the
165 same time they have been shown to regulate gene expression (Sharma et al., 2010).

166 Current research reports have outlined that there is a relation between epigenetic
167 modifications and metabolic disorders like obesity. More specifically, evidence from a recent
168 report showed that there are different methylation patterns of genes implicated in fatty acid β -
169 oxidation (FAO) in samples obtained from lean and severely obese women in response to lipid
170 exposure. According to the results, there was an immediate induction of genes participating in
171 FAO in response to lipid exposure among lean women whereas this was not observed in the case
172 of the severely obese ones. The mRNA levels of peroxisome proliferator-activated receptor δ
173 (PPAR- δ ; a molecule participating in FAO) were found to be differentially regulated in the case
174 of severe obesity, a fact that was attributed to different methylation patterns of the gene (Maples
175 et al., 2015a). Moreover, data from a similar study demonstrated that the expression of carnitine
176 palmitoyltransferase 1B (CPT-1B; a protein responsible for transferring the long-chain fatty acids
177 across the outer mitochondrial membrane) was reduced in skeletal muscle cells isolated from
178 severely obese women in contrast to lean women following lipid exposure. The observed
179 differential expression of CPT-1B, in obese women, was due to alterations in DNA methylation,
180 histone acetylation and transcription factor binding (Maples et al., 2015b). As a consequence, it is
181 logical that the link between epigenetic modifications and obesity could be influenced by
182 phytochemicals (given their ability to modulate key epigenetic processes); however, such link is
183 purely speculative and yet to be established.

184

185 5.2 Classes of phytochemicals

186 5.2.1 Green tea catechins

187 Health benefits of regular consumption of green tea are mostly attributed to the large
188 amount of catechins, polyphenols of flava-3-ol sub-family of flavonoids. Unlike black tea, green
189 tea manufacturing preserves high amount of epicatechin, epigallocatechine, epicatechin-3-gallate
190 and epigallocatechine-3-gallate (EGCG) as it is prepared from non-oxidized and non-fermented
191 leaves. Catechins are considered to inhibit catechol-*O*-methyltransferase which is responsible for
192 breaking down norepinephrine and thereby stimulate fat oxidation (Borchardt, 1975). The hepatic
193 fatty acid oxidation and ATP production directly influence appetite by influencing appetite
194 regulating centers of the brain. Green tea catechins can control appetite as a result of up-regulation
195 of hepatic fat oxidation and ATP generation (Friedman 2007; Kamphuis et al., 2003).

196 Green tea catechins have been supported as fat burning phytochemicals in various animal
197 studies; however, the clinical evidence is still lacking behind in confirming these findings. For
198 instance, results from a study using obese C57BL/6J mice showed that green tea-EGCG had the
199 ability to induce body mass reduction while it also caused changes in the mRNA expression levels
200 of PPAR- γ , C/EBP- α , SREBP-1c, aP2, LPL and FAS all of which decreased in white adipose
201 tissue. On the other hand, the mRNA levels of CPT-1, UCP2, HSL and ATGL increased (Lee et
202 al., 2009). Additionally, another study associated the supplementation of green tea catechins in the
203 diet of lactating maternal rats with the increase of mRNA expression levels of DNMT1, DNMT3a,
204 SIRT1 and SIRT2 in the kidneys of their three week old offspring. This, in turn, supports the
205 correlation between maternal levels of catechins and increased levels of enzymes (in newborn
206 female offspring) capable of influencing epigenetic marks capable of potentially regulating energy
207 metabolism (Sun et al., 2013). In a recent study, addition of 1% green tea to high-fat diet of mice

208 has reduced mass of adipose tissue and TAG, glucose, insulin, and leptin levels of blood (Lee et
209 al., 2015). It was postulated that green tea has the ability of modulation of abnormal fatty acid β -
210 oxidation caused by high-fat diet.

211 Beneficial effects of combined flavonoids on diet-induced obesity have been demonstrated.
212 Recently, supplementation of flavonoids from green tea combined with cocoa, coffee and *Garcinia*
213 has shown to stimulate lipid metabolism in high-energy diet-induced obese rats, which is
214 attributable to fat mobilization from adipose tissue (Huang et al., 2016). A recent review on dietary
215 polyphenols and obesity also confirmed that green tea catechins (especially EGCG), resveratrol
216 and curcumin all exert anti-obesity properties (Wang et al., 2014).

217

218 5.2.2 Resveratrol

219 Resveratrol (3,4,5-trihydroxystilbene) is a naturally occurring stilbene sub-group of
220 polyphenol in grapes, red wine and some berries (Freemont, 2000). It has been studied for its
221 involvement in regulating fatty acid β -oxidation in relation to preventing degradation of
222 intracellular cyclic adenosine monophosphate (cAMP) through inhibition of cAMP
223 phosphodiesterase enzymes which in turn activate the AMPK enzyme (Park et al., 2012a, Chung,
224 2012a,b), which consequently activates mitochondrial biogenesis and function by activating PGC-
225 1α (Wu et al., 1999). A recent review has also revealed that anti-obesity activity of resveratrol
226 could also be through down-regulation of PPAR- γ , CCAAT-enhancer-binding protein (C/EBP α),
227 and sterol regulatory element binding protein 1c (SREBP-1c) (Aguirre et al., 2014). In an another
228 resveratrol supplementation (0.02% of diet) study conducted using ApoE-deficient mice,
229 resveratrol exerts not only anti-obesity and hypolipidemic effects, but also protective effects for
230 the liver and aorta through the modulation of lipid metabolism in liver and white adipose tissue

231 (Jeon et al., 2014). In addition, resveratrol has been proposed as a natural SIRT-1 activator which
232 can also further activate PGC-1 α (Lagouge et al., 2006). Moreover, in several animal studies, the
233 supplementation of resveratrol resulted in a remarkable increase of AMPK activity (Baur et al.,
234 2006; Shang et al., 2008b; Rivera et al., 2009). Exposure to resveratrol has reported to increase
235 fatty acid β -oxidation in CPT-II and very long chain acyl CoA dehydrogenase deficient cultured
236 patient fibroblast model (Aires et al., 2014). In another study, resveratrol increased fatty acid β -
237 oxidation by inhibiting the production of malonyl-CoA (Szkudelska and Szkudelski, 2010).

238 Animal studies have demonstrated the role of resveratrol in energy expenditure in a way
239 where animals were capable of surviving cold longer with supplementation of high doses of
240 resveratrol than untreated ones (Lagouge et al., 2006). Similar observations were recorded after
241 one year of treatment with resveratrol (200 mg/kg/day) where such treatment was found to increase
242 basal metabolic rate and total daily energy expenditure in the non-human primate *Microcebus*
243 *murinus* (Dal-Pan et al., 2010; Dal-Pan et al., 2011). These studies further strengthen the capacity
244 of resveratrol to enhance energy expenditure and potentially promote weight loss. Only recently,
245 resveratrol has become the subject of intense research as being a phytochemical associated with a
246 great range of health promoting properties. For this reason, it has attracted the attention of the
247 nutraceutical industry as it is consumed by two-thirds of consumers taken dietary nutritional
248 supplements (Block et al., 2007).

249 Finally, according to the findings from a recent report, the combined administration of
250 resveratrol and pterostilbene (in rats) resulted in preventing the up-regulation of the FAS gene
251 (induced in response to a high fat and sucrose contained diet) while pterostilbene was also
252 demonstrated to be responsible for the differential methylation pattern of the gene as well (Gracia
253 et al., 2014).

254

255 **5.2.3 Capsaicinoids**

256 Red hot chillies or peppers are a commonly used spice in food worldwide. Capsaicinoid is
257 the class of pungent polyphenol derivative compound in red chillies. The genus capsicum includes
258 more than 200 varieties and concentration of capsaicin also varies (0 - 13,000 mg/kg) (Kozukue et
259 al., 2005). Capsaicin (N-[(4-hydroxy-3-methoxyphenyl)-methyl]-8-methyl-6-nonamide) is the
260 pungent compound which has been reported to increase thermogenesis and secretion of
261 catecholamines from adrenal medulla (Watanbe et al., 1987) which stimulate adrenergic receptors
262 in liver and adipose tissue resulting in lipolysis and energy expenditure (Diepvens and Westerterp,
263 2007). A meal containing red pepper instantaneously increases energy expenditure in humans
264 (Yoshioka et al., 1995). Although supplementation with capsaicinoids has been reported to
265 increase BMR in human subjects by increasing fat oxidation (Lejeune et al., 2003) they cannot be
266 generally consumed in high dosages due to their strong pungency and nociceptive stimulus.
267 However, capsiate (a non-pungent capsaicinoid analogue derived from *Capsicum annuum* L.; CH-
268 19 Sweet) has been reported to increase body temperature and increase mRNA and protein levels
269 of uncoupling proteins (UCPs) of brown adipose tissues in a two-week mice study (Masuda et al.,
270 2003). The thermogenic effects of capsaicin are attributed to its structure and not its pungency
271 (Ohnuki et al., 2001a). Similarly, there was an increase in catecholamine and free fatty acid levels
272 together with a decrease in triacylglycerol levels resulting in elevated levels of fat oxidation in
273 mice after supplementation with a single oral dosage of capsiate (Ohnuki et al., 2001b; Haramizu
274 et al., 2006).

275 CPT-1 α is the rate-limiting enzyme in mitochondrial β -oxidation and a target for reducing
276 body fat (McGarry and Brown, 1997). On the other hand, UCP-2 is a mitochondrial proton

277 transporter and has been suggested to influence body temperature, energy expenditure and fat mass
278 (Rousset et al., 2004). Capsaicin is reported to stimulate lipolysis by mediating CPT-1 α and UCP-
279 2 in adipocytes (Lee et al., 2011). Capsaicin-induced thermogenesis is proposed to function
280 through stimulation of β -adrenergic receptors as various studies have demonstrated decreased
281 thermogenesis after administration of β -adrenergic blockers (Kawada, 1986). In a combination
282 study of capsaicin with green tea, a decrease in fat mass was observed (Tsi et al., 2003); however,
283 long term supplementation of capsaicin is found to be more effective in weight loss (Lejeune et
284 al., 2003). CH-13 Sweet has similar capsaicin structure and is more suitable for long term use by
285 maintaining effectiveness and without pungency (Reinbach et al., 2009). Furthermore, only
286 recently, another study has demonstrated a different approach to minimize the pungency of
287 capsaicin (and thus increase the possibility of its long term use) by preparing chitosan
288 microspheres (Tan et al., 2014). Chitosan is a polysaccharide extracted from crab shells and it aids
289 to overcome strong pungent taste and smell of capsaicin. Finally, the findings of a study using rats
290 demonstrated that the administration of capsiate resulted in a reduced abdominal fat volume and
291 body weight gain, which were associated with the differential gene expression levels of UCP3 and
292 more specifically a reduction in its mRNA levels (Faraut et al., 2009).

293

294 **5.2.4 Citrus flavonoids**

295 These are a class of polyphenols found in citrus. Naringin and hesperidin belong to
296 flavanone sub-group and nobiletin and tangeretin to *O*-polymethoxylated flavones. In animal
297 studies, supplementation of naringin (3%) and nobiletin (0.3%) with high fat diet has demonstrated
298 increase in fatty acid oxidation by up-regulating CPT-1 α production (Mulvihill et al., 2009;
299 Mulvihill et al., 2011; Jung et al., 2006). Interestingly, dietary supplementation of citrus peel

300 flavonoid extract (rich in nobiletin, tangeretin, rutin and hesperidin) to high-fat diet-induced obese
301 C57BL/6 mice reversed the suppressed activities of AMPK and ACC (Kang et al., 2012). In mature
302 3T3-L1 adipocytes, the citrus peel flavonoid extract increased AMPK and ACC phosphorylation
303 and also enhanced lipolysis by phosphorylation of cAMP-dependent protein kinase (PKA) and
304 hormones-sensitive lipase (HSL) (Kang et al., 2012).

305 Apart from the citrus flavonoids, the alkaloid synephrine is also a bioactive component of
306 bitter orange (*Citrus aurantium*). The fruit is also often used for herbal medicine as appetite
307 suppressant. Bitter orange alkaloids act as adrenergic agonists with octapamine and synephrine
308 being similar to epinephrine and norepinephrine, respectively. Para-synephrine has properties of
309 both α -adrenergic and β -adrenergic agonists and is also known as oxedrine. Its anti-obesity effects
310 are proposed to be due to its action on β 3-receptors and increased thermogenesis leading to β -
311 oxidation (Arbo et al., 2008). On the other hand, in a rat study, bitter orange extracts with 4-6%
312 synephrine decreased body weight after 7 days (Calapai et al., 1999). However, there is a little
313 evidence reported the ability of synephrine for weight loss in human.

314 Finally, according to a research report, naringenin (a grapefruit flavonoid) is capable of
315 regulating the activation of PPAR α and PPAR γ , while it is also responsible for the induction of
316 several genes in fatty acid oxidation including CYP4A11, ACOX, UCP1 and ApoAI in
317 hepatocytes (Goldwasser et al., 2010). Data from a similar study showed that the administration
318 of naringenin, in rats, led to a differential regulation of the expression levels of PPAR α , CPT1 and
319 UCP2, with the up-regulation of PPAR α consequently resulting in the increase of CPT1 and UCP2
320 expression levels (Cho et al., 2011). As well, Huong et al. (2006) has also demonstrated that
321 naringenin (1% of the diet) increases hepatic fatty acid oxidation through up-regulation of gene
322 expression of enzymes involved in peroxisomal β -oxidation in mice.

323

324 **5.2.5 Piperine**

325 This is a pungent lipophilic alkaloid found in black pepper which is prepared from ground
326 unripe berries from the plant *Piper nigrum* Linn. Piperine has been found to increase
327 catecholamine secretion (particularly epinephrine) from the adrenal medulla in rats. These effects
328 are similar to capsaicin but not as much potent. This effect can be described as a sympathetic
329 nervous system (SNS)-mediated thermogenesis given that it is diminished after administration of
330 cholinergic blockers (Kawada et al., 1984). Finally, piperine was shown to inhibit the
331 differentiation of 3T3-L1 cells to adipocytes as it induced the down-regulation of PPAR γ , SREPB-
332 1c and C/EBP β and thus implying its potentially beneficial use in the treatment of metabolic
333 disorders (Park et al., 2012b).

334

335 **5.2.6 Anthocyanins**

336 These are well-known phytochemicals for their antioxidant effects. Apart from dietary
337 antioxidants, they also have several biological activities including anti-convulsant, anti-
338 carcinogenic, anti-atherosclerotic, anti-diabetic and anti-inflammatory thus reducing the risk of
339 disease and in particular coronary heart disease (Drenska et al., 1989; Satue-Gracia et al., 1997;
340 Wang et al., 1999; Koide et al., 1997; Sancho et al., 2012). Recently, studies have documented the
341 role of anthocyanins as anti-obesity agents (Tsuda et al., 2003; Jayaprakasam et al., 2006; Matsui
342 et al., 2001; Kwon et al., 2007). Anthocyanins are water-soluble compounds widely found in fruits
343 and vegetables and responsible for most of the red, purple, and blue colors exhibited by flowers,
344 fruits, and other plant tissues (Castañeda et al., 2009). In the last decade, anthocyanins from purple
345 corn (*Zea mays L.*), blood orange (*Citrus sinensis L.*Osbeck), strawberries (*Fragaria ananassa*),

346 blueberries (*Vaccinium angustifolium*), blackberries (*Rubus* species) and mulberry (*Morus*
347 *australis* P.) have been reported to exhibit anti-obesity effects in various *in vivo* studies (Tsuda et
348 al., 2003; Titta et al., 2010; Prior et al., 2008; Prior et al., 2012; Kaume et al., 2012; Wu et al.,
349 2013a).

350 In addition, other studies have suggested that treatment with anthocyanin induced ACC
351 phosphorylation and increased mitochondrial fatty acid oxidation via the allosteric regulation of
352 CPT-1, which catalyses the entry of long-chain fatty acyl-CoA into mitochondria in HepG2 cells.
353 Therefore, a decrease in malonyl CoA levels is directly responsible for increase in CPT-1
354 expression, leading to fatty acid oxidation (Hurley et al., 2005). On the other hand, AMPK
355 regulates the enzymes of lipid metabolism and also directs fatty acid both in oxidative and
356 biosynthetic pathways in the liver (Kahn et al., 2005). AMPK knockdown failed to stimulate
357 AMPK and reduce hepatocellular lipid accumulation. Thus, the possible mechanism of
358 anthocyanin-induced fatty acid oxidation is via AMPK directed inhibition of ACC and FAS which
359 are two key downstream regulators of AMPK in the control of lipid metabolism. A recent study,
360 in mice, has demonstrated down-regulation of CPT-1 gene expression after supplementation with
361 anthocyanin-rich blueberry and mulberry juices, indicating an anthocyanin-induced stimulation of
362 fatty acid oxidation while inhibiting fatty acid synthesis (Wu et al., 2013b). In addition, evidence
363 from a current report further supports the ability of anthocyanins to differentially regulate various
364 genes participating in fatty acid oxidation (e.g. PPAR- α , PPAR- δ , UCP-2, UCP-3, mitochondrial
365 transcription factor A) as their mRNA levels were considerably increased when C57BL/6J mice
366 were fed a high in fat and cholesterol diet with a polyphenol-rich blackcurrant extract (Benn et al.,
367 2014).

368

369 5.2.7 Curcumin

370 This is the main bioactive polyphenol (hydroxycinnamic acid derivative) present in the
371 rhizome of turmeric (*Curcuma longa*) which is commonly used as dietary spice and food color in
372 Asian countries. In addition, it has been found to regulate signal transduction and gene expression
373 apart of its anti-inflammatory and antioxidant properties and thus of potential benefit in disease
374 prevention and therapy (Ohara et al., 2009; Zingg et al., 2013). Furthermore, an animal study has
375 demonstrated that curcumin reduced the body weight gain in high fat fed mice without altering
376 food intake in addition of influencing energy metabolism and fatty acid β -oxidation in adipocytes,
377 through AMPK (Ejaz et al., 2009). Likewise, curcumin facilitated β -oxidation in *in vitro*
378 experiments (by up-regulation of CPT-1) and reduced lipid biosynthesis (by down-regulation of
379 glycerol-3-phosphate acyl transferase-1; GPAT-1 and acyl-CoA carboxylase) (Ejaz et al., 2009).
380 Finally, another suggested mechanism of fatty acid oxidation, by curcumin, can be explained in
381 terms of an increase in mitochondrial biogenesis by activation of PGC-1 (Chung et al., 2012a,b;
382 Zingg et al., 2013).

383 On another note, curcumin i) inhibits lipogenic enzymes in liver, ii) stimulates lipid
384 mobilization from adipose tissue by activating HSL, iii) inhibits fatty acid synthase (FAS) activity
385 and iv) activates fatty acid β -oxidation (Prakash and Srinivasan, 2012; Zhao et al., 2011; Jang et
386 al., 2008). In particular, curcumin has been shown to specifically down-regulate FAS leading to
387 an effective decrease in fat storage. Thus, there is substantial evidence to suggest that curcumin is
388 effective in inhibiting lipid synthesis and storage as well as stimulating fatty acid degradation
389 (Smith, 1994). To this end, data from a recent *in vitro* study demonstrated that curcumin was able
390 to reduce the mRNA levels of DNMT3B suggesting its ability to affect epigenetic mechanisms

391 thus leading to altered gene expression (Jiang et al., 2015), a fact that might account for curcumin's
392 observed beneficial effects in weight loss and activation of fatty acid degradation.

393

394 **5.2.8 Raspberry ketons**

395 Raspberry ketones [4-(4-hydroxyphenyl) butan-2-one; RK] are major phenolic acid
396 derivative compounds present in red raspberry (*Rubus idaeus*) and are responsible for the sweet
397 aroma of raspberries. Like other berries, raspberry has also been reported to have significant
398 biological effects (Ravai, 1996). RKs have similar structures with capsaicin and synephrine, which
399 are known for their active role in obesity and lipid metabolism (Harada et al., 2008).

400 RKs' supplementation inhibits body weight gain in high fat-fed rats as they are unable to
401 bind beta-adrenergic receptors and do not trigger lipolysis in the absence of norepinephrines.
402 Therefore, RKs can stimulate norepinephrine-induced lipolysis by facilitating the translocation of
403 HSL from the cytosol to the lipid droplets in the fat cells in addition to increasing fat oxidation
404 and energy expenditure by stimulating thermogenesis (Morimoto et al., 2005). In another study,
405 treatment with 10 μ M of RK induced lipolysis, fat oxidation and increased the adiponectin levels
406 in cultured 3T3-L1 pre-adipocytes all of which led to decreased fat mass in adipocytes and
407 potentially have a key role in body weight regulation (Park, 2010). According to the literature,
408 administration of adiponectin increases fat oxidation in obese mice circulating free fatty acid levels
409 by enhancing skeletal muscle fat oxidation (Wolf, 2003; Mullen et al., 2007). The other suggested
410 mechanism of RK regulated fat loss can be through reversing leptin resistance and elevating
411 PPAR- α (Meng et al., 2008; Wang et al., 2012). Leptin is a hormone secreted by adipocytes which
412 stimulate fatty acid oxidation by induction of AMPK (Monokoshi et al., 2002).

413

414 **5.2.9 Cocoa polyphenols**

415 Cocoa is a major ingredient of chocolate and it is derived from the beans of the *Theobroma*
416 *cacao* (Baba et al., 2000). The cocoa beans consist of approximately 6-8% polyphenols (by weight)
417 with their presence contributing to dark chocolate being a rich source of antioxidants. These are
418 predominantly flavonoids and mainly epicatechin, catechin and proanthocyanidins with a small
419 amount of quercetin also present (Manach et al., 2004; Andres-Lacueva et al., 2008). Polyphenol-
420 rich cocoa extracts possess many bioactivities including anti-hyperlipidemic (Hamed et al., 2008),
421 anti-diabetic (Grassi et al., 2005), antioxidant (Galleano et al., 2009), in addition to improving
422 cognitive and visual performance (Field et al., 2011) and boosting the immune system (Katz et al.,
423 2011).

424 Genistein, which is the main isoflavone in cocoa extract, directly interacts with PPAR- α
425 and PPAR- γ and functions as an activator for stimulating fatty acid catabolism (Kim et al., 2004a;
426 Kim et al., 2004b). Furthermore, activation of PPAR- α is reported to stimulate the expression of
427 β -oxidation genes, including CPT-1, ACO and UCP3. Adiponectin expression also increases with
428 the activation of PPAR- γ (Maeda et al., 2001) in addition to activating the AMPK pathway which
429 regulates glucose and lipid metabolism (Arts and Hollman, 2005; Kurlandsky and Stote, 2006).
430 Finally, cocoa polyphenols have been reported to increase plasma adiponectin levels and also
431 increase thermogenesis through activation of the AMPK pathway and specifically via up-
432 regulation of UCPs which are involved in facilitating thermogenesis and energy expenditure
433 (Yamashita et al., 2012; Corti et al., 2009).

434 Even though various studies have mentioned different types of cocoa flavonoids, it is not
435 evident yet which phytochemicals are efficacious for exerting their anti-obesity (Farhat et al.,
436 2014). Nogueira et al., (2011) reported that the supplementation of 2 mg/kg/day of cocoa-derived

437 epicatechin stimulated fat oxidation whereas in another study, supplementation with a dose of
438 cocoa (containing 18.4 mg epicatechin and 380 mg of polyphenols and equivalent to 40 g/day in
439 humans) exhibited anti-obesity effects in mice (Gu et al., 2014). Moreover, the weight reducing
440 effects of dark chocolate can be partially attributed to caffeine which is present in significant
441 amount (Stark et al., 2006; Zheng et al., 2004). Findings from a very recent study demonstrates
442 that cocoa polyphenol administration in the diet of Sprague-Dawley rats resulted in the
443 differentially regulated expression of genes implicated in lipid metabolism in mesenteric white
444 adipose tissue, as the mRNA levels of several lipolysis enzymes were found to be increased (Ali
445 et al., 2015). Finally, further studies support the ability of cocoa polyphenols to affect DNA
446 methylation patterns of peripheral leukocytes in subjects with cardiovascular risk factors including
447 obesity (Crescenti et al., 2013).

448

449 **5.2.10 Soybean phytochemicals**

450 Soybeans (*Glycine max*) are consumed mainly as a source of protein, besides being also rich
451 in micronutrients such as isoflavones, phytate, soyasaponins, phytosterol, vitamins and minerals
452 (Rupasinghe et al., 2003). They are known to be the richest source of isoflavones in food (Cederroth
453 and Nef, 2009) whereas soya-derived phytoestrogens (non-steroidal plant-derived compounds
454 which can bind estrogen receptors and thus mimic estrogen) have been shown to exert beneficial
455 effects in cardiovascular disease, diabetes, osteoporosis and prostate cancer (Setchell, 1998; Tham
456 et al., 1998; Sacks et al., 2006; Kuiper et al., 1997). Soybean isoflavones have been the subject of
457 intense research and thus shown to exert estrogenic effects hence influencing glucose and lipid
458 metabolism (Velasquez and Bhathena, 2007). Various animal studies have demonstrated that a soya-
459 rich diet significantly reduces fat accumulation (Bu et al., 2005; Lephart et al., 2004) and increases

460 energy expenditure and locomotor activity by utilizing lipid resources (Cederroth et al., 2007;
461 Cederroth et al., 2008). In a study of high-fat diet-induced obesity in C57BL/6 mice,
462 supplementation of fermented black soybean has significantly lowered the body and liver weight
463 and the levels of blood glucose, total cholesterol and leptin (Oh et al., 2014). Similarly, when high
464 fat-diet is supplemented with 0.15% of kaempferol glycosides isolated from soybean leaves, body
465 and adipose tissue weight and blood TAG of C57BL/6J mice were reduced significantly.
466 Furthermore, expression of genes of PPAR- γ and SREBP-1c was also reduced by the diet
467 supplementation of soybean flavonoids (Zang et al., 2015).

468 Although the anti-obesity effect of soya isoflavones is well-evident, the exact mechanism
469 remains unclear. To this end, suggested mechanisms include correlation of decrease of adiposity
470 with increase in AMPK and ACC activation (Hwang et al., 2005) along with increased lipolysis
471 through inhibition of cAMP phosphodiesterases (Szkudelska et al., 2000). The up-regulation of
472 AMPK, PPAR- γ co-activator-1 α (PGC-1 α) and PPAR- α resulted in increased β -oxidation and
473 energy expenditure (Cederroth et al., 2007; Cederroth et al., 2008). Although speculative, it may be
474 that such gene up-regulation is the result of the induction of epigenetic mechanisms as recently, a
475 study utilizing monkeys showed the presence of epigenetic alterations (by means of altered DNA
476 methylation patterns) induced when a high in fat content of a soy-based diet was changed to one
477 without soy (Howard et al., 2011).

478

479 **5.2.11 Hydroxycitric acid**

480 Hydroxycitric acid (HCA), an organic acid, is one of widely known supplements for anti-
481 obesity and weight management. *G. cambogia* extract is a commercially available and richest source
482 of HCA that contributes to anti-obesity mainly by suppressing appetite (Leonhardt et al., 2002),

483 inhibiting *de novo* lipogenesis (Kovacs et al., 2006) and increasing fat oxidation (Preuss et al. 2004).
484 In addition, another suggested mechanism is reduction of the acetyl-CoA by HCA and thus,
485 eventually inhibiting lipogenesis by regulating the availability of precursors for fatty acid and
486 cholesterol biosynthesis (Chuah et al., 2013). In addition, a study by Ishihara et al., (2000) conducted
487 in mice suggested that chronic HCA administration increased fatty acid oxidation during a 3-week
488 experimental period. Moreover, another study determined increase in HCA-induced fatty acid
489 oxidation by means of measuring urinary concentration of fatty acid oxidation by-products (Preuss
490 et al., 2004).

491 Although, currently there are no reports supporting the contribution of HCA in regulating
492 the expression of genes involved in FAO, a recent study showed that cambogin (a compound from
493 the *Garcinia genus*) was responsible for inducing epigenetic changes (via an increase in the
494 trimethylation of histone H3K9) in a different experimental setting and in order to exhibit its anti-
495 proliferative effects in various human breast cancer cell lines (Shen et al., 2015). Nevertheless, to
496 this end, other compounds of *G. cambogia*, like HCA, might also exhibit an ability to induce
497 epigenetic alterations in the context of anti-obesity and consequently management of weight control.
498

499 **6. Conclusions**

500 There is a vast majority of numerous phytochemicals being the subject of intense research
501 as potentially efficient dietary agents for the management of weight control. However, only some
502 of them are directly involved in weight reduction by stimulation of fatty acid β -oxidation. To this
503 end, the phytochemicals enlisted in this review have demonstrated the capacity for weight loss in
504 both cell-based assays and pre-clinical studies. Even though the clinical evidence is very limited,
505 these plant-based compounds have been traditionally used for their anti-obesity benefits without any

506 toxicity or health hazard concerns. Most of these phytochemicals, apart from their weight loss
507 properties, also have other additional health benefits including anti-inflammatory, antioxidant and
508 other biological functions. Overall, the benefit of weight loss leads to reduction in fat mass, decrease
509 in inflammation and further reduction of the risk of developing metabolic disease. In addition, it
510 should be noted that there is also evidence to support a role of phytochemicals in regulating the
511 differential expression of various genes, implicated in various cellular pathways through epigenetic
512 mechanisms. Although the current evidence is substantially speculative, the fatty acid β -oxidation
513 pathway can be one such target pathway the significance of which is of extreme importance given
514 its relevance to weight loss and the overall management of weight control.

515

516 **7. Acknowledgements**

517 This work was supported, in part, by a Heriot Watt University PhD Studentship (Theodora
518 Mantso) and by Canada Research Chair program funds (H.P.V. Rupasinghe).

8. References

- 1) Ali, F., Ismail, A., Esa, N.M., Pei, C.P. (2015). Transcriptomics expression analysis to unveil the molecular mechanisms underlying the cocoa polyphenol treatment in diet-induced obesity rats. *Genomics*, *105*, 23-30.
- 2) Aguirre, L., Fernandez-Quintela, A., Arias, N., Portillo, M.P. (2014). Resveratrol: Anti-obesity mechanisms of action. *Molecules*, *19*, 18632–18655.
- 3) Aires, V., Delmas, D., Le Bachelier, C., Latruffe, N., Latruffe, N., Schlemmer, D. et al. (2014). Stilbenes and resveratrol metabolites improve mitochondrial fatty acid oxidation defects in human fibroblasts. *Orphanet J Rare Dis*, *9*, 79-85.
- 4) Andres-Lacueva, C., Monagas, M., Khan, N., Izquierdo- Pulido, M., et al. (2008). Flavanol and flavonol contents of cocoa powder products: influence of the manufacturing process. *J Agric Food Chem*, *56*, 3111-3117.
- 5) Arbo, M.D., Lartentis, E.R., Linck, V.M., Aboy, A.L., Pimentel, A.L., Henriques, A.T., et al. (2008). *Food Chem Toxicol*, *46*, 2770-2775.
- 6) Arsenijevic, D., Onuma, H., Pecqueur, C., Raimbault, S., Manning, B.S., et al. (2000). Disruption of the uncoupling protein-2 gene in mice reveals a role in immunity and reactive oxygen species production. *Nat Genet*, *26*, 435-439.
- 7) Arts, I. C. W., Hollman, P. C. H. (2005). Polyphenols and disease risk in epidemiologic studies. *Am J Clin Nutr*, *81*, S317–S325.
- 8) Baba, S., Osakabe, N., Natsume, M., Yasuda, A., et al. (2000). Cocoa powder enhances the level of antioxidative activity in rat plasma. *Br J Nutr*, *84*, 673-680.

- 9) Baur, J.A., Pearson, K.J., Price, N.L., Jamieson, N.L., Lerin, C., Kalra, A., et al. (2006). Resveratrol improves health and survival of mice on high-calorie diet. *Nature*, *444*, 337-342.
- 10) Benn, T., Kim, B., Park, Y.K., Wegner, C.J., Harness, E., Nam, T.G., et al. (2014). Polyphenol-rich blackcurrant extract prevents inflammation in diet-induced obese mice. *J Nutr Biochem*, *25*, 1019-1025.
- 11) Bessesen, D.H. (2008). Update on obesity. *J Clin Endocrinol Metab*, *93*, 2027-2034.
- 12) Block, G., Jensen, C. D., Norkus, E. P., Dalvi, T. B., et al. (2007). Usage patterns, health, and nutritional status of long-term multiple dietary supplement users: a cross-sectional study. *Nutr J*, *6*, 30-41.
- 13) Borchardt, R.T. (1975). Affinity Labeling of Catechol O-Methyltransferase by the Oxidation Products of 6-Hydroxydopamine. *Mol Pharmacol*, *11*, 436-449.
- 14) Bu, L., Setchell, K.D., Lephart, E.D. (2005). Influences of dietary soy isoflavones on metabolism but not nociception and stress hormone responses in ovariectomized female rats. *Reprod Biol Endocrinol*, *3*, 58-64.
- 15) Calapai, G., Firenzuoli, F., Saitta, A., Squadrito, F., Arlotta, M.R., Costantino, G., et al. (1999). *Fitoterapia*, *70*, 586-592.
- 16) Castañeda, A., Pacheco, H.M.L., Páez, H.M.E., Rodríguez, J.A., Galán, C.A. (2009). Chemical studies of anthocyanins: a review. *Food Chem*, *113*, 859-871.
- 17) Cederroth, C.R., Vinciguerra, M., Kuhne, F., Madani, R., Doerge, D.R., Visser, T.J., Foti, et al. (2007). A phytoestrogen-rich diet increases energy expenditure and decreases adiposity in mice. *Environ Health Perspect*, *115*, 1467-1473.

- 18) Cederroth, C.R., Vinciguerra, M., Gjinovci, A., Kuhne, F., Klein, M., Cederroth, M., Caille, et al. (2008). Dietary phytoestrogens activate AMP-activated protein kinase with improvement in lipid and glucose metabolism. *Diabetes*, 57, 1176-1185.
- 19) Cederroth, C.R., Nef, S. (2009). Soy, phytoestrogens and metabolism: a review. *Mol Cell Endocrinol*, 304, 30-42.
- 20) Chalkiadaki, A., Guarente, L. (2012). High-fat diet triggers inflammation induced cleavage of SIRT1 in adipose tissue to promote metabolic dysfunction. *Cell Metab*, 16, 180-188.
- 21) Chaput, J-P., St-Pierre, S., Tremblay, A. (2007). Currently Available Drugs for the Treatment of Obesity: Sibutramine and Orlistat. *Mini Reviews Med Chem*, 7, 3-10.
- 22) Cho, K.W., Kim, Y.O., Andrade, J.E., Burgess, J.R., Kim, Y.C. (2011). Dietary naringenin increases hepatic peroxisome proliferators-activated receptor alpha protein expression and decreases plasma triglyceride and adiposity in rats. *Eur J Nutr*, 50, 81-88.
- 23) Chuah L.O., Ho, W.Y., Beh, B.K., Yeap, S.K. (2013). Updates on antiobesity effect of Garcinia Origin (-)-HCA. *Evid Based Complement Alternat Med*, 2013, 751658.
- 24) Chung, J.H. (2012a). Using PDE inhibitors to harness the benefits of calorie restriction: lessons from resveratrol. *Aging*, 4, 144-145.
- 25) Chung, J.H., Manganiello, V., Dyck, J.R. (2012b) Resveratrol as a calorie restriction mimetic: therapeutic implications. *Trends Cell Biol*, 22, 546-554.
- 26) Corti, R., Flammer, A.J., Hollenberg, N.K., Lüscher, T.F. (2009). Cocoa and cardiovascular health. *Circulation*, 119, 1433-1441.
- 27) Cornelius, P., MacDougald, O.A., Lane, M.D. (1994). Regulation of adipocyte development. *Annu Rev Nutr*, 14, 99-129.

- 28) Crescenti, A., Sola, R., Valls, R.M., Caimari, A., Del Bas, J.M., Anguera, A., et al. (2013). Cocoa consumption alters the global DNA methylation of peripheral leukocytes in humans with cardiovascular disease risk factors: A randomized controlled trial. *PLoS One*, 8, e65744.
- 29) Dal-Pan, A., Blanc, S., Aujard, F., (2010). Resveratrol suppresses body mass gain in a seasonal non-human primate model of obesity. *BMC Physiol*, 10, 11-21.
- 30) Dal-Pan, A., Terrien, J., Pifferi, F., Botalla, R. et al. (2011). Caloric restriction or resveratrol supplementation and ageing in a non-human primate: first-year outcome of the RESTRIKAL study in *Microcebus murinus*. *Age (Dordr.)*. 33, 15-31.
- 31) Diepvens, K., Westerterp, K.R., Westerterp-Plantenga, M.S. (2007). Obesity and thermogenesis related to the consumption of caffeine, ephedrine, capsaicin, and green tea. *Am J Physiol Regul Integr Comp Physiol*, 292, 77-85.
- 32) Drenska, D., Bantutova, I., Oveharov, R. (1989). Anticonvulsant effect of anthocyanins and antioxidants. *Fomatsiya (Sofia)*, 30, 33-40.
- 33) Eaton, S., Bartlett, K., Quant, P.A. (2001). Carnitine palmitoyltransferase I and the control of beta-oxidation in heart mitochondria. *Biochem Biophys Res Commun*, 285, 537-539.
- 34) Ejaz, A., Wu, D., Kwan, P., Meydani, M. (2009). Curcumin inhibits adipogenesis in 3T3-L1 adipocytes and angiogenesis and obesity in C57/BL mice. *J Nutr*, 139, 919-925.
- 35) Faraut, B., Giannesini, B., Matarazzo, V., Le Fur, Y., Rougon, G., Cozzone, P.J., et al. (2009). Capsiate administration results in an uncoupling protein-3 downregulation, an enhanced muscle oxidative capacity and a decreased abdominal fat content *in vivo*. *Int J Obes (Lond)*, 33, 1348-1355.

- 36) Farhat, G., Drummond, S., Fyfe, L., Al-Dujaili, E.A.S. (2014). Dark Chocolate: An Obesity Paradox or a Culprit for Weight Gain? *Phytother Res*, 28, 791-797.
- 37) Faulds, M.H., Dahlman, W.K. (2012). Metabolic diseases and cancer risk. *Curr Opin Oncol*, 24, 58-61.
- 38) Field, D.T., Williams, C.M., Butler, L.T. (2011). Consumption of cocoa flavanols results in an acute improvement in visual and cognitive functions. *Physiol Behav*, 103, 255-260.
- 39) Filozof, C.M., Murua, C., Sanchez, M.P., Brailovsky, C., Perman, M., Gonzalez, C.D., et al. (2000). Low plasma leptin concentration and low rates of fat oxidation in weight-stable post-obese subjects. *Obesity Research*, 8, 205-210.
- 40) Friedman, M.I. (2007). Obesity and the hepatic control of feeding behavior. *Drug News Perspect* 20, 573-578.
- 41) Galleano, M., Oteiza, P., Fraga, C. (2009). Cocoa, chocolate and cardiovascular disease. *J Cardiovasc Pharmacol*, 54, 483-490.
- 42) Goldwasser, J., Cohen, P.Y., Yang, E., Balaguer, P., Yarmush, M.L., Nahmias, Y. (2010). Transcriptional regulation of human and rat hepatic lipid metabolism by the grapefruit flavonoid naringenin: role of PPARalpha, PPARgamma and LXRA. *PLoS One*, 5, e12399.
- 43) Gong, D.W., Monemdjou, S., Gavrilova, O., Leon, L.R., Marcus-Samuels, B., et al. (2000). Lack of obesity and normal response to fasting and thyroid hormone in mice lacking uncoupling protein-3. *J Biol Chem*, 275, 16251-16257.
- 44) González-Castejón, M., & Rodríguez-Casado, A. (2011). Dietary phytochemicals and their potential effects on obesity: a review. *Pharmacol Research*, 64, 438-455.

- 45) Gracia, A., Elcoroaristizabal, X., Fernandez-Quintela, A., Miranda, J., Bediaga, N.G., et al. (2014). Fatty acid synthase methylation levels in adipose tissue: effects of an obesogenic diet and phenol compounds. *Genes Nutr*, 9, 411-420.
- 46) Grassi, D., Lippi, C., Necozione, S., et al. (2005). Short-term administration of dark chocolate is followed by a significant increase in insulin sensitivity and a decrease in blood pressure in healthy persons. *Am J Clin Nutr*, 81, 611-614.
- 47) Gu, Y., Yu, S., Lambert, J. (2014). Dietary cocoa ameliorates obesity-related inflammation in high in high fat-fed mice. *Eur J Nutr*, 53, 149-58.
- 48) Guarente, L. (2006). Sirtuins as potential targets for metabolic syndrome. *Nature*, 444, 868-874.
- 49) Guo, Y., Su, Z.Y., Kong, A.T. (2015). Current perspectives on epigenetic modifications by dietary chemopreventive and herbal phytochemicals. *Curr Pharmacol Rep*, 1, 245-257.
- 50) Hallows, W.C., Lee, S., Denu, J.M. (2006). Sirtuins deacetylate and activate mammalian acetyl-CoA synthetases. *Proc Natl Acad Sci USA*, 103, 10230-10235.
- 51) Hallows, W.C., Yu, W., Smith, B.C., Devires, M.K., Ellinger, J.J., et al. (2011). Sirt3 promotes the urea cycle and fatty acid oxidation during dietary restriction. *Mol Cell*, 41, 139-149.
- 52) Hamed, M.S., Gambert, S., Bliden, K.P. et al. (2008). Dark chocolate effect on platelet activity, C-reactive protein and lipid profile: a pilot study. *South Med J*, 101, 1203-1208.
- 53) Harada, N., Okajima, K., Narimatsu, N., Kurihara, H., Nakagata, N. (2008). Effect of topical application of raspberry ketone on dermal production of insulin- like growth factor-I in mice and on hair growth and skin elasticity in humans. *Growth Horm IGF Res*, 18, 335-344.

- 54) Haramizu, S., Mizunoya, W., Masuda, Y., Ohnuki, K., Watanabe, T., Yazawa, S., Fushiki, T. (2006). Capsiate, a nonpungent capsaicin analog, increases endurance swimming capacity of mice by stimulation of vanilloid receptors. *Biosci Biotechnol Biochem*, 70, 774-781.
- 55) Huang, C.C., Tung, Y.T., Huang, W.C., Chen, Y.M., Hsu, Y.J., Hsu, M.C. (2016). Beneficial effects of cocoa, coffee, green tea, and garcinia complex supplement on diet induced obesity in rats. *BMC Complement Altern Med*, 16, 100-110.
- 56) Huong, D.T.T., Takahashi, Y., Ide, T. (2006). Activity and mRNA levels of enzymes involved in hepatic fatty acid oxidation in mice fed citrus flavonoids. *Nutrition*, 22, 546-552.
- 57) Hirschey, M.D., Shimazu, T., Goetzman, E., Jing, E., Schwer, B., et al. (2010). SIRT3 regulates mitochondrial fatty-acid oxidation by reversible enzyme deacetylation. *Nature*, 464, 121-125.
- 58) Horton, R.A., Moran, L.A., Scrimgeour, G., Perry, M., Rawn, D. (2006). Principles of Biochemistry, 4th, Fourth Edition. Pearson, Glenview, IL, USA.
- 59) Hou, X., Xu, S., Maitland-Toolan, K.A., Sato, K., Jiang, B., et al. (2008). SIRT1 regulates hepatocyte lipid metabolism through activating AMP activated protein kinase. *J Biol Chem*, 283, 20015-20026.
- 60) Howard, T.D., Ho, S.M., Zhang, L., Chen, J., Cui, W., Slager, R., Gray, S., Hawkins, G.A., Medvedovic, M., Wagner, J.D. (2011). Epigenetic changes with dietary soy in cynomolgus monkeys. *PLoS One*, 6, e26791.

- 61) Hurley, R.L., Anderson, K.A., Franzone, J.M., Kemp, B.E., Means, A.R., Witters, L.A. (2005). The Ca²⁺/calmodulin-dependent protein kinase kinases are AMP-activated protein kinase kinases. *J Biol Chem*, 280, 29060-29066.
- 62) Hwang, J.T., Park, I.J., Shin, J.I., Lee, Y.K., Lee, S.K., Baik, H.W., et al. (2005). Genistein, EGCG, and capsaicin inhibit adipocyte differentiation process via activating AMP-activated protein kinase. *Biochem Biophys Res Commun*, 338, 694-649.
- 63) Ishihara, K., Oyaizu, S., Onuki, K., Lim, K., Fushiki, T. (2000). Chronic (-)-hydroxycitrate administration spares carbohydrate utilization and promotes lipid oxidation during exercise in mice. *J Nutr*, 130, 2990-2995.
- 64) Jang, E.M., Choi, M.S., Jung, U.J., et al. (2008). Beneficial effects of curcumin on hyperlipidemia and insulin resistance in high-fatfed hamsters. *Metabolism*, 57, 1576-1583.
- 65) Jayaprakasam, B., Olson, L.K., Schutzki, R.E., Tai, M.H., Nair, M.G. (2006). Amelioration of obesity and glucose intolerance in high-fat-fed C57BL/6 mice by anthocyanins and ursolic acid in Cornelian cherry (*Cornus mas*). *J Agric Food Chem*, 11, 243-248.
- 66) Jeon, S.M., Lee, S.A., Choi, M.S. (2014). Antiobesity and vasoprotective effects of resveratrol in apoE-deficient mice. *J Med Food*, 17, 310-316.
- 67) Jeukendrup, A.E., Randell, R. (2011). Fat burners: nutrition supplements that increase fat metabolism. *Obesity Reviews*, 12, 841-851.
- 68) Jiang, A., Wang, X., Shan, X., Li, Y., Wang, P., Jiang, P., Feng, Q. (2015). Curcumin reactivates silenced tumor suppressor gene RARbeta by reducing DNA methylation. *Phytother Res*, 29, 1237-1245.

- 69) Jung, U.J., Lee, M.K., Park, Y.B., Kang, M.A., Choi, M.S. (2006). Effect of citrus flavonoids on lipid metabolism and glucose-regulating enzyme mRNA levels in type-2 diabetic mice. *Int J Biochem Cell Biol*, 38, 1134-1145.
- 70) Kahn, B.B., Alquier, T., Carling, D., Hardie, D.G. (2005). AMP-activated protein kinase: ancient energy gauge provides clues to modern understanding of metabolism. *Cell Metab*, 1, 15-25.
- 71) Kamphuis, M.M., Mela, D.J., Westerterp-Plantenga, M.S. (2003). Diacylglycerols affect substrate oxidation and appetite in humans. *Am J Clin Nutr*, 77, 1133-1139.
- 72) Kang, S.I., Shin, H.S., Kim, H.M., Hong, Y.S., Yoon, S.A., Kang, S.W. et al. (2012). Immature citrus sunki peel extract exhibits antiobesity effects by β -oxidation and lipolysis in high-fat diet-induced obese mice. *Biol Pharm Bull*, 35, 223–230.
- 73) Katz, D., Doughty, K., Ather, A. (2011). Cocoa and chocolate in human health and disease. *Antiox & Redox Signal*, 15, 2779-2811.
- 74) Kaume, L., Gilbert, W.C., Brownmiller, C., Howard, L.R., Devareddy, L. (2012). Cyanidin 3-O- β -D-glucoside-rich blackberries modulate hepatic gene expression and anti-obesity effects in ovariectomized rats. *J Funct Foods*, 4, 480-488.
- 75) Kawada, T., Watanabe, T., Takaishi, T., Tanaka, T., Iwai, K. (1986). Capsaicin- induced beta-adrenergic action on energy metabolism in rats: influence of capsaicin on oxygen consumption, the respiratory quotient, and substrate utilization. *Proc Soc Exp Biol Med*, 183, 250-256.
- 76) Kawada, T., Kawada, T., Suzuki, T., Takahashi, M., Iwai, K. (1984). Gastrointestinal absorption and metabolism of capsaicin and dihydrocapsaicin in rats. *Toxicol Appl Pharmacol*, 72, 449-456.

- 77) Kim, H.K., Nelson-Dooley, C., Della-Fera, M.A., Yang, J.Y., Zhang, W., Duan, J., et al. (2006). Genistein decreases food intake, body weight, and fat pad weight and causes adipose tissue apoptosis in ovariectomized female mice. *J Nutr*, *136*, 409-414.
- 78) Kim, S., Shin, H-J., Kim, S.Y., Kim, J.H., et al. (2004a). Genistein enhances expression of genes involved in fatty acid catabolism through activation of PPAR. *Mol Cell Endocrinol*, *220*, 51-58.
- 79) Kim, S., Sohn, I., Lee, Y.S., Lee, Y.S. (2004b). Hepatic gene expression profiles are altered by genistein supplementation in mice with diet-induced obesity. *J Nutr*, *135*, 33-41.
- 80) Koide, T., Hashimoto, Y., Kamei, H., Kojima, T., Hasegawa, M., Terabe, K. (1997). Antitumor effect of anthocyanin fractions extracted from red soybeans and red beans *in vitro* and *in vivo*. *Cancer Biother Radiopharm*, *12*, 277-280.
- 81) Kovacs, E.M.R., Westerterp-Plantenga, M.S. (2006). Effects of (-)-hydroxycitrate on net fat synthesis as *de novo* lipogenesis. *Physiology & Behavior*, *88*, 371-381.
- 82) Kozukue, N., Han, J-S., Kozukue, E., Lee, S-J., Kim, J-A., Lee, K-R., Levin, C-E., Friedman, M. (2005). Analysis of eight capsaicinoids in peppers and pepper-containing foods by high-performance liquid chromatography and liquid chromatography-mass spectrometry. *J Agric Food Chem*, *53*, 9172-9181.
- 83) Kuiper, G.G., Carlsson, B., Grandien, K., Enmark, E., Haggblad, J., Nilsson, S., et al. (1997). Comparison of the ligand binding specificity and transcript tissue distribution of estrogen receptors alpha and beta. *Endocrinology*, *138*, 863-870.
- 84) Kumar, M.V., Sunvold, G.D., Scarpace, P.J. (1999). Dietary vitamin A supplementation in rats: suppression of leptin and induction of UCP1 mRNA. *J Lipid Res*. *40*, 824-829.

- 85) Kurlandsky, S.B., Stote, K.S. (2006). Cardioprotective effects of chocolate and almond consumption in healthy women. *Nutr Res*, 26, 509-516.
- 86) Lagogue, M., Argmann, C., Gerhart-Hines, Z., Meziane, H., Lerin. C., Daussin, F., et al. (2006). Resveratrol improves mitochondrial function and protects against metabolic disease by activating SIRT1 and PGC-1alpha. *Cell*, 127, 1109-1122.
- 87) Langin, D. (2006). Control of fatty acid and glycerol release in adipose tissue lipolysis. *C R Biologies*. 329, 598-607.
- 88) Lee, W.J., Koh, E.H., Won, J.C., Kim, M-S., Park, J-Y. et al. (2005). Obesity: The role of hypothalamic AMP-activated protein kinase in body weight regulation. *Internatl. J. Biochem. Cell Biol*. 37, 2254-2259.
- 89) Lee, M.S., Kim, C.T., Kim, Y. (2009). Green tea (-)-epigallocatechin-3-gallate reduces body weight with regulation of multiple genes expression in adipose tissue of diet-induced obese mice. *Ann Nutr Metab*, 54, 151-157.
- 90) Lee, M. S., Kim, C. T., Kim, I. H., Kim, Y. (2011). Effects of capsaicin on lipid catabolism in 3T3-L1 adipocytes. *Phytotherapy Research*, 25, 935-939.
- 91) Lee, L.S., Choi, J.H., Sung, M.J., Hur, J.Y., Hur, H.J., Park, J.D. et al. (2015). Green tea changes serum and liver metabolomic profiles in mice with high-fat diet-induced obesity. *Mol Nutr Food Res*, 59, 784-794.
- 92) Lejeune, M.P., Kovacs, E.M., Westerterp-Plantenga, M.S. (2003). Effect of capsaicin on substrate oxidation and weight maintenance after modest body-weight loss in human subjects. *Br J Nutr*, 90, 651-659.

- 93) Leonhardt, M., Langhans, W. (2002). Hydroxycitrate has long-term effects on feeding behavior, body weight regain and metabolism after body weight loss in male rats. *J Nutr*, *132*, 1977-1982.
- 94) Lephart, E.D., Setchell, K.D., Handa, R.J., Lund, T.D. (2004). Behavioral effects of endocrine-disrupting substances: phytoestrogens. *Ilar J*, *45*, 443-454.
- 95) Lopaschuk, G.D., Ussher, J.R., Folmes, C.D., Jaswal, J.S., Stanley, W.C. (2010). Myocardial fatty acid metabolism in health and disease. *Physiol Rev*, *90*, 207-258.
- 96) Maeda, N., Takahashi, M., Funahashi, T., Kihara, S. et al. (2001). PPAR ligands increase expression and plasma concentrations of adiponectin, an adipose-derived protein. *Diabetes*, *50*, 2094-2098.
- 97) Manach, C., Scalbert, A., Morand, C., Remesy, C. et al. (2004). Polyphenols: food sources and bioavailability. *Am J Clin Nutr*, *79*, 727-747.
- 98) Maples, J.M., Brault, J.J., Shewchuk, B.M., Witczak, C.A., Zou, K., Rowland, N., et al. (2015a). Lipid exposure elicits differential responses in gene expression and DNA methylation in primary human skeletal muscle cells from severely obese women. *Physiol Genomics*, *47*, 139-46.
- 99) Maples, J.M., Brault, J.J., Witczak, C.A., Park, S., Hubal, M.J., Weber, T.M., et al. (2015b). Differential epigenetic and transcriptional response of the skeletal muscle carnitine palmitoyltransferase 1B (CPT1B) gene to lipid exposure with obesity. *Am J Physiol Endocrinol Metab*, *309*, E345-356.
- 100) Masuda, Y., Haramizu, S., Oki, K., Ohnuki, K., Watanabe, T., Yazawa, S., et al. (2003). Upregulation of uncoupling proteins by oral administration of capsiate, a nonpungent capsaicin analog. *J Appl Physiol*, *95*, 2408-2415.

- 101) Matsui, T., Ueda, T., Oki, T., Sugita, K., Terahara, N., Matsumoto, K. (2001). Alpha-glucosidase inhibitory action of natural acylated anthocyanins: Survey of natural pigments with potent inhibitory activity. *J Agric Food Chem*, 49, 1948-1951.
- 102) McGarry, J.D., Brown, N.F. (1997). The mitochondrial carnitine palmitoyltransferase system: from concept to molecular analysis. *Eur J Biochem*, 244, 1-14.
- 103) McGarry, J.D., Mills, S.E., Long, C.S., Foster, D.W. (1983). Observations on the affinity for carnitine, and malonyl-CoA sensitivity, of carnitine palmitoyltransferase I in animal and human tissues. *Biochem J*, 214, 21-28.
- 104) Meng, X.J., Zhou, Y., Liu, X., Zheng F. (2008). Experimental study on the anti-obesity action of simple obesity in rats by raspberry ketone. *Food Industry*, 1, 1-3.
- 105) Minokoshi, Y., Kim, Y.B., Peroni, O.D., Fryer, L.G., Müller, C., Carling, D., et al. (2002). Leptin stimulates fatty-acid oxidation by activating AMP-activated protein kinase. *Nature*, 17, 339-343.
- 106) Morimoto, C., Satoh, Y., Hara, M., Inoue, S., Tsujita, T., Okuda, H. (2005). Anti-obese action of raspberry ketone. *Life Sci*, 27, 194-204.
- 107) Mullen, K.L., Smith, A.C., Junkin, K.A., Dyck, D.J. (2007). Globular adiponectin resistance develops independently of impaired insulin-stimulated glucose transport in soleus muscle from high-fat-fed rats. *Am J Physiol Endocrinol Metab*, 293, E83-E90.
- 108) Mulvihill, E.E., Allister, E.M., Sutherland, B.G., et al. (2009). Naringenin prevents dyslipidemia, apolipoprotein B overproduction, and hyperinsulinemia in LDL receptor-null mice with diet-induced insulin resistance. *Diabetes*, 58, 2198-2210.

- 109) Mulvihill, E.E., Assini, J.M., Lee, J.K., et al. (2011). Nobiletin attenuates VLDL overproduction, dyslipidemia, and atherosclerosis in mice with diet-induced insulin resistance. *Diabetes*, *60*, 1446-1457.
- 110) Nogueira, L., Ramirez-Sanchez, I., Perkins, G., et al. (2011). (-)-Epicatechin enhances fatigue resistance and oxidative capacity in mouse muscle. *J Physiol*, *589*, 4615-4631.
- 111) Oh, H.G., Kang, Y.R., Lee, H.Y., Kim, J.H., Shin, E.H., Lee, B.G. et al. (2014). Ameliorative effects of *Monascus pilosus* - fermented black soybean (*Glycine max* L. Merrill) on high-fat diet-induced obesity. *J Med Food*, *17*, 972–978.
- 112) Ohara, K., Uchida, A., Nagasaka, R., Ushio, H., Ohshima, T. (2009). The effects of hydroxycinnamic acid derivatives on adiponectin secretion. *Phytomedicine*, *16*, 130-137.
- 113) Ohnuki, K., Haramizu, S., Oki, K., Watanabe, T., Yazawa, S., Fushiki, T. (2001a) Administration of capsiate, a non-pungent capsaicin analog, promotes energy metabolism and suppresses body fat accumulation in mice. *Biosci Biotechnol Biochem*, *65*, 2735-2740.
- 114) Ohnuki, K., Niwa, S., Maeda, S., Inoue, N., Yazawa, S., Fushiki, T. (2001b). CH-19 sweet, a non-pungent cultivar of red pepper, increased body temperature and oxygen consumption in humans. *Biosci Biotechnol Biochem*, *65*, 2033-2036.
- 115) O'Neill, H. M. (2013). AMPK and exercise: glucose uptake and insulin sensitivity. *Diabetes Metabol J*, *37*, 1-21.
- 116) Park, K.S. (2010). Raspberry ketone increases both lipolysis and fatty acid oxidation in 3T3-L1 adipocytes. *Planta Med*, *76*, 1654-1658.

- 117) Park, S.J., Ahmad, F., Philp, A., Baar, K., Williams, T., Luo, H., et al. (2012). Resveratrol ameliorates aging-related metabolic phenotypes by inhibiting cAMP phosphodiesterases. *Cell*, 148, 421-433.
- 118) Park, U.H., Jeong, H.S., Jo, E.Y., Park, T., Yoon, S.K., Kim, E.J., et al. (2012). Piperine, a component of black pepper, inhibits adipogenesis by antagonizing PPARgamma activity in 3T3-L1 cells. *J Agric Food Chem*, 60, 3853-3860.
- 119) Prakash, U.N., Srinivasan, K. (2012). Fat digestion and absorption in spice-pretreated rats. *J Sci Food Agric*, 92, 503-510.
- 120) Preuss, H.G., Bagchi, D., Bagchi, M., Rao, C.V.S., Dey, D.K., Satyanarayana, S. (2004). Effects of a natural extract of (-)-hydroxycitric acid (HCA-SX) and a combination of HCA-SX plus niacin-bound chromium and *Gymnema sylvestre* extract on weight loss. *Diabetes, Obesity & Metabolism*, 6, 171-180.
- 121) Prior, R.L., Wilkes, E.S.R., Rogers, T., Khanal, R.C., Wu, X., et al. (2012). Purified blueberry anthocyanins and blueberry juice alter development of obesity in mice fed an obesogenic high-fat diet. *J Agric Food Chem*, 58, 3970-3976.
- 122) Prior, R.L., Wu, X., Gu, L., Hager, T.J., Hager, A., et al. (2008). Whole berries versus berry anthocyanins: interactions with dietary fat levels in the C57BL/6J mouse model of obesity. *J Agric Food Chem*, 56, 647-653.
- 123) Purushotham, A., Schug, T.T., Xu, Q., Surapureddi, S., Guo, X., et al. (2009). Hepatocyte-specific deletion of SIRT1 alters fatty acid metabolism and results in hepatic steatosis and inflammation. *Cell Metab*, 9, 327-338.
- 124) Ravai, M. (1996). Quality characteristics of raspberries and blackberries. *Cereal Foods World*, 41, 773-775.

- 125) Reinbach, H.C., Smeets, A., Martinussen, T., Moller, P., Westerterp-Plantenga, M.S. (2009). Effects of capsaicin, green tea and CH-19 sweet pepper on appetite and energy intake in humans in negative and positive energy balance. *Clin Nutr*, 28, 260-265.
- 126) Rivera, L., Morón, R., Zarzuelo, A., Galisteo, M. (2009). Long-term resveratrol administration reduces metabolic disturbances and lowers blood pressure in obese Zucker rats. *Biochem Pharmacol*, 77, 1053-1063.
- 127) Rogge, M.M. (2009). The role of impaired mitochondrial lipid oxidation in obesity. *Biol Res Nurs*, 10, 356-373.
- 128) Ronnett, G.V., Kim, E.K., Landree, L.E., Tu, Y. (2005). Fatty acid metabolism as a target for obesity treatment. *Physiol Behav*, 19, 25-35.
- 129) Rousset, S., Alves-Guerra, M.C., Mozo, J., et al. (2004). The biology of mitochondrial uncoupling proteins. *Diabetes*, 53, S130-S135.
- 130) Rupasinghe, H.P.V., Jackson, C-J. C., Poysa, V., Berardo, C.D., Bewley, J.D., and Jenkinson, J. (2003). Soyasapogenol A and B distribution in soybean (*Glycine max* L. Merr.) in relation to seed physiology, genetic variability, and growing location. *J Agric Food Chem*, 51, 5888-5894.
- 131) Sacks, F.M., Lichtenstein, A., Van Horn, L., Harris, W., Kris-Etherton, P., Winston, M. (2006). Soy protein, isoflavones, and cardiovascular health: an American Heart Association Science Advisory for professionals from the Nutrition Committee. *Circulation*, 113, 1034-1044.
- 132) Salas-Salvado, J., Martinez-Gonzalez, M.A., Bullo, M., Ros, E. (2011). The role of diet in the prevention of type 2 diabetes. *Nutr Metab Cardiovasc Dis*, 21, B32-48.

- 133) Sancho, R.A.S., Pastore, G.M. (2012). Evaluation of the effects of anthocyanins in type 2 diabetes. *Food Res Int*, 46, 378-386.
- 134) Satue-Gracia, M.T., Heinonen, M., Frankel, E.N. (1997). Anthocyanins as antioxidants on human low-density lipoprotein and lecithinliposome systems. *J Agric Food Chem*, 45, 3362-3367.
- 135) Setchell, K.D. (1998). Phytoestrogens: the biochemistry, physiology, and implications for human health of soy isoflavones. *Am J Clin Nutr*, 68, 1333S-1346S.
- 136) Shang, J., Chen, L.L., Xiao, F.X., Sun, H., Ding, H.C., Xiao, H. (2008b). Resveratrol improves non-alcoholic fatty liver disease by activating AMP-activated protein kinase. *Acta Pharmacol Sin*, 29, 698-706.
- 137) Shankar, S., Kumar, D., Srivastava, R.K. (2013). Epigenetic modifications by dietary phytochemicals: implications for personalized nutrition. *Pharmacol Ther*, 138, 1-17.
- 138) Sharma, .S, Kelly, T.K., Jones, P.A. (2010). Epigenetics in cancer. *Carcinogenesis*, 31, 27-36.
- 139) Shen, K., Xie, J., Wang, H., Zhang, H., Yu, M., Lu, F., et al. (2015). Cambogin induces caspase-independent apoptosis through the ROS/JNK pathway and epigenetic regulation in breast cancer cells. *Mol Cancer Ther*, 14, 1738-1749
- 140) Simoneau, J.A., Veerkamp, J.H., Turcotte, L.P., Kelley, D.E. (1999). Markers of capacity to utilize fatty acids in human skeletal muscle: Relation to insulin resistance and obesity and effects of weight loss. *FASEB J*, 13, 2051-2060.
- 141) Smith, S. (1994). The animal fatty acid synthase: one gene, one polypeptide, seven enzymes. *FASEB J*, 8, 1248-1259.

- 142) Stark, T., Bareuther, S., Hofmann, T. (2006). Molecular definition of the taste of roasted cocoa nibs (*Theobroma cacao*) by means of quantitative studies and sensory experiments. *J Agric Food Chem*, 54, 5530-5539.
- 143) Su, Z.Y., Shu, L., Khor, T.O., Lee, J.H., Fuentes, F., Kong, A.N. (2013). A perspective on dietary phytochemicals and cancer chemoprevention: oxidative stress, nrf2, and epigenomics. *Top Curr Chem*, 329, 133-162.
- 144) Sun, Y., Mukai, Y., Tanaka, M., Saito, T., Sato, S., Kurasaki, M. (2013). Green tea extract increases mRNA expression of enzymes which influence epigenetic marks in newborn female offspring from undernourished pregnant mother. *PLoS One*, 8, e74559.
- 145) Szkudelska, K., Szkudelski, T. (2010). Resveratrol, obesity and diabetes. *Eur J Pharmacol*, 635, 1-8.
- 146) Szkudelska, K., Nogowski, L., Szkudelski, T. (2000). Genistein affects lipogenesis and lipolysis in isolated rat adipocytes. *J Steroid Biochem Mol Biol*, 75, 265-271.
- 147) Szarc vel Szic, K., Declerck, K., Vidakovic, M., Vanden Berghe, W. (2015). From inflammaging to healthy aging by dietary lifestyle choices: is epigenetics the key to personalized nutrition? *Clin Epigenetics*, 7, 33-39.
- 148) Tan, S., Bing Gao, B. Tao, Y., Guo, J., Su, Z.Q. (2014). Antiobese effects of Capsaicin–Chitosan Microsphere (CCMS) in obese rats induced by high fat diet. *J Agric Food Chem*, 62, 1866-1874.
- 149) Tham, D.M., Gardner, C.D., Haskell, W.L. (1998). Clinical review 97: potential health benefits of dietary phytoestrogens: a review of the clinical, epidemiological and mechanistic evidence. *J Clin Endocrinol. Metab*, 83, 2223-2235.

- 150) Titta, L., Trinei, M., Stendardo, M., Berniakovich, I., Petroni, I., et al. (2010). Blood orange juice inhibits fat accumulation in mice. *Int J Obes*, 34, 578-588.
- 151) Tsuda, T., Horio, F., Uchida, K., Aoki, H., Osawa, T. (2003). Dietary cyaniding 3-O-D-glucoside rich purple corn color prevents obesity and ameliorates hyperglycemia in mice. *J Nutr*, 133, 2125-2130.
- 152) Tziomalos, K., Krassas, G.E., Tzotzas, T. (2009). The use of sibutramine in the management of obesity and related disorders: an update. *Vascular Health Rick Managet*. 5, 441-452.
- 153) Velasquez, M.T. and Bhathena, S.J. (2007). Role of dietary soy protein in obesity. *Int J Med Sci*, 4, 72-82.
- 154) Wang, H., Nair, M.G., Strasburg, G.M., Chang, Y.C., Booren, A.M., Gray, J.I., DeWitt, D.L. (1999). Antioxidant and anti-inflammatory activities of anthocyanins and their aglycon, cyanidin, from tart cherries. *J Nat Prod*, 62, 294-296.
- 155) Wang, L., Meng, X., Zhang, F. (2012). Raspberry ketone protects rats fed high-fat diets against nonalcoholic steatohepatitis. *J Med Food*, 15, 495-503.
- 156) Wang, S., Moustaid-Moussa, N., Chen, L., Mo, H., Shastri, A., Su, R. et al. (2014). Novel insights of dietary polyphenols and obesity. *J Nutr Biochem*, 25, 1–18.
- 157) Watanabe, T., Kawada, T., Yamamoto, M., Iwai, K. (1987). Capsaicin, a pungent principle of hot red pepper, evokes catecholamine secretion from the adrenal medulla of anesthetized rats. *Biochem Biophys Res Commun*, 142, 259-264.
- 158) WHO (2000). Obesity: Preventing and Managing the Global Epidemic; World Health Organization: Geneva, Switzerland. 894, 1-252.

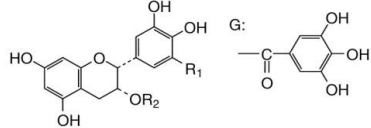
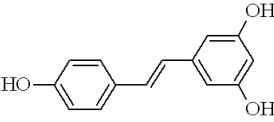
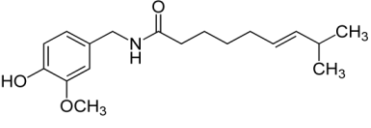
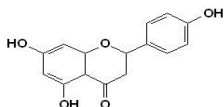
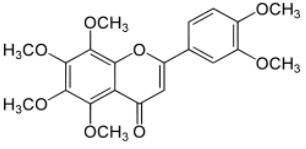
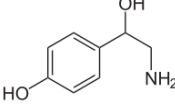
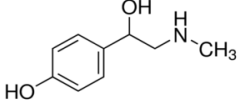
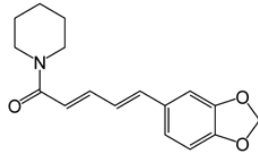
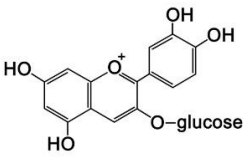
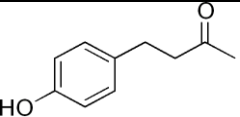
- 159) Witkamp, R.F. (2011). Current and future drug targets in weight management. *Pharm Res*, 28, 1792-818.
- 160) Wolf, G. (2003). Adiponectin: a regulator of energy homeostasis. *Nutr Rev*, 61, 290-292.
- 161) Wu, T., Qi, X., Liu, Y., Guo, J., Zhu, R., et al. (2013a). Dietary supplementation with purified mulberry (*Morus australis* Poir) anthocyanins suppresses body weight gain in high fat diet fed C57BL/6 mice. *Food Chem*, 141, 482-487.
- 162) Wu, T., Tang, Q., Gao, Z., Yu, Z., Song, H., Zheng, X., et al. (2013b). Blueberry and mulberry juice prevent obesity development in C57BL/6 mice. *PLoS One*, e77585. doi: 10.1371/journal.pone.0077585.
- 163) Wu, Z., Puigserver, P., Andersson, U., Zhang, C., Adelman, G., Mootha, V., et al. (1999). Mechanisms controlling mitochondrial biogenesis and respiration through the thermogenic coactivator PGC-1. *Cell*, 98, 115-124.
- 164) Yamashita, Y., Okabe, M., Natsume, M., Ashida, H. (2012). Prevention mechanisms of glucose intolerance and obesity by cacao liquor procyanidin extract in high-fat diet-fed C57BL/6 mice. *Arch Biochem Biophys*, 527, 95-104.
- 165) Yoshioka, K., Lim, K., Kikuzato, S., Kiyonaga, A., Tanaka, H. et al. (1995). Effects of red-pepper diet on the energy metabolism in men. *J Nutr Sci Vitaminology* 41, 647-656.
- 166) Yun, J.W. (2010). Possible anti-obesity therapeutics from nature – A review. *Phytochem*, 71, 1625-1641.
- 167) Zammit, V.A. (1999). The malonyl-CoA-long-chain acyl-CoA axis in the maintenance of mammalian cell function. *Biochem J*, 343, 505-515.

- 168) Zang, Y., Zhang, L., Igarashi, K., Yu, C. (2015). The anti-obesity and anti-diabetic effects of kaempferol glycosides from unripe soybean leaves in high-fat-diet mice. *Food Funct*, 6, 834–841.
- 169) Zhao, J., Sun, X.B., Ye, F., Tian, W.X. (2011). Suppression of fatty acid synthase, differentiation and lipid accumulation in adipocytes by curcumin. *Mol Cell Biochem*, 351, 19-28.
- 170) Zheng, G., Sayama, K., Okubo, T., et al. (2004). Anti-obesity effects of three major components of green tea, catechins, caffeine and theanine, in mice. *In vivo*, 18, 55-62.
- 171) Zingg, J.M., Hasan, S.T., Meydani, M. (2013). Molecular mechanisms of hypolipidemic effects of curcumin. *Biofactors*, 39, 101-21.

Table 1: Classification of major phytochemicals

Phytochemicals	Examples
Polyphenols	Anthocyanins, flavonols, catechins, isoflavonoids, flavones, flavanones, stilbenes, phenolic acids, capsaicinoids, curcuminoids
Alkaloids	Caffeine, nicotine, piperine
Isoprenoids	Beta-carotene, lycopene, essential oils

Table 2: Structure of phytochemicals and their sources

Phytochemicals	Structure	Source	Effect on β -oxidation
Catechin Epicatechin Epigallocatechin		Green tea, Cocoa	catechol- <i>O</i> -methyltransferase ↓
Resveratrol (3,4,5-trihydroxystilbene)		Grapes, Red wine	SIRT1, AMPK ↑ Malonyl CoA ↓
Capsaicin (N-[(4-hydroxy-3-methoxyphenyl)-methyl]-8-methyl-6-nonamide)		Red chillies	β -adrenergic receptors CPT-1, UCP-2 ↑
Naringin		Citrus	CPT-1 α ↑
Nobiletin		Citrus	CPT-1 α ↑
Octapamine		Bitter orange	β -adrenergic agonist
Synephrine		Bitter orange	β -adrenergic agonist
Piperine		Black pepper	SNS-mediated thermogenesis ↑
Anthocyanins		Purple corn, Blueberry, Strawberry, Bitter orange Pomegranat	CPT-1, AMPK ↑
Raspberry ketones (4-(4-hydroxyphenyl)butan-2-one)		Raspberry	PPAR- α , AMPK ↑ HSL

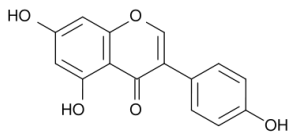
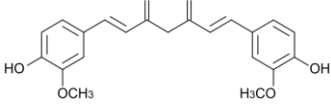
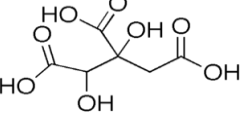
Genistein isoflavones		Soyabeans, Cocoa	AMPK, adiponectin, PPAR- α , CPT \uparrow
Curcumin		Turmeric	CPT-1 \uparrow FAS \downarrow
(-)Hydroxycitric Acid		<i>Garcinia cambogia</i>	Acetyl-CoA \downarrow

Figure Legends

Figure 1: Key elements involved in the regulation of fatty acid β -oxidation at various steps.

ACC, acetyl CoA carboxylase; ACS, acyl CoA synthetase; AMPK-P, phosphorylated AMP-activated protein kinase; CL, citrate lyase; CPT, carnitine palmitoyl transferase; ETC, mitochondrial electron transport chain; E1, acyl CoA dehydrogenase; E2, enoyl CoA hydratase; E3, 3-hydroxy acyl CoA dehydrogenase; E4, 3-ketoacyl CoA thiolase; FA, fatty acid; FAS, fatty acid synthase; HSL, hormone-sensitive lipase; PPAR- δ , peroxisome proliferator-activated receptor δ ; TAG, triacylglycerol; TCA, tricarboxylic acid; UCP-2, mitochondrial uncoupling protein-2.

Figure 1

