

# Effect of the purification treatment on the valorization of natural cellulosic residues as fillers in PHB-based composites for short shelf life applications

Estefanía Lidón Sánchez-Safont<sup>1</sup>, Abdulaziz Aldureid<sup>1</sup>, José María Lagarón<sup>2</sup>, José Gámez-Pérez<sup>1</sup>, Luis Cabedo<sup>1\*</sup>

1. Polymers and Advanced Materials Group (PIMA), Universitat Jaume I (UJI), Av. de Vicent Sos Baynat s/n, 12071 Castelló, Spain
2. bNovel Materials and Nanotechnology Group, Institute of Agrochemistry and Food Technology (IATA), Spanish National Research Council (CSIC), Calle Catedrático Agustín Escardino Benlloch 7, 46980 Paterna, Spain

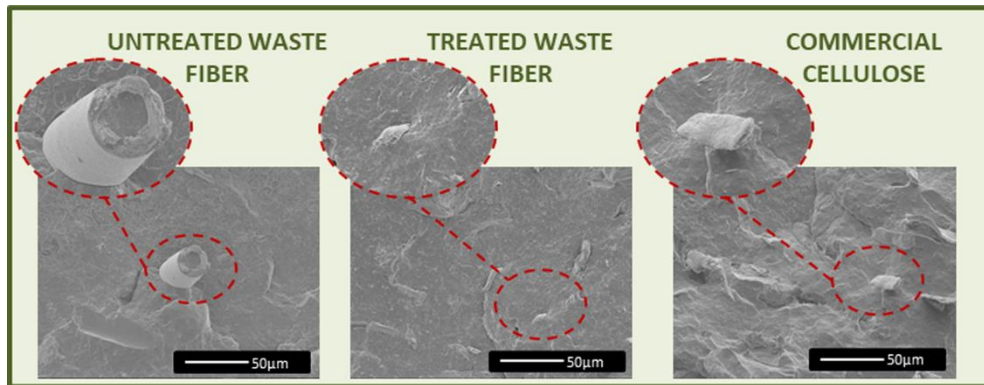
## Abstract

In this work the effect of a combined NaOH+ peracetic acid (PAA) purification treatment on the valorization of almond shell (AS) and rice husk (RH) lignocellulosic residues as fillers in PHB-based composites for short shelf life applications has been studied. The efficiency of the treatment at removing the non cellulosic components of the fibers has been evaluated by SEM, FTIR, WAXS and TGA taking a commercial cellulose as reference. The influence of the untreated and treated fibers on the morphology, thermal, crystallization, tensile properties, fracture toughness and dynamo mechanical behavior of the PHB/fiber composites has been studied. The treatment has demonstrated its ability at removing the lignin, hemicelluloses and waxes allowing the obtention of fibers with relative crystallinity, thermal stability and composition similar to the commercial cellulose. The different agro-food based lignocellulosic residues used resulted in two suitable reinforcing fillers for a PHB matrix showing the composites prepared with the treated fibers better thermal and mechanical performance with respect to those prepared with the untreated ones.

## Keywords

PHB, natural fiber, cellulose, biocomposite, waste valorization

## Graphical abstract



## State of novelty

The manuscript explores the valorization of agrocellulosic wastes for the obtention of low cost biocomposites for short shelf life applications.

### 1. Introduction

The growing concern for the environment and the increasingly restrictive environmental directives results in an urgent need to find sustainable alternatives to conventional plastics. Particularly, in short shelf life applications or single-use products (e.g. in packaging or tableware articles) biodegradable or compostable materials pose as especially appropriate candidates. Within this context, biopolymers are called to be the ideal substitutes to the so-called “commodities” [1]. Concretely, polyhydroxyalkanoates (PHAs) have aroused great interest in these applicability fields and good proof of this is the large number of related publications that exist in literature [2–5].

In case of polyhydroxybutyrate (PHB) the main reasons that make it attractive for these type of applications are its bio-based origin, full biodegradability and compostability, mechanical properties close to polypropylene in terms of stiffness and high barrier properties [6], comparable or even superior to PET [7]. On the other hand, in spite of the extent research done, PHB still presenting some shortcomings that handicap its industrial applicability. Among them, its still high production cost, low elongation-at-break, low toughness and tear resistance mainly due to its high crystallinity [8]. Moreover, PHB undergo an important embrittlement over time caused by secondary crystallization and physical ageing [9]. PHB also present a narrow

1 processing window and is very sensitive to thermal degradation [10]. Altogether, these problems  
2 limit the application of PHB to packaging applications.  
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4 In the last decades, the development of natural fiber based polymer composites has become  
5 one of the research topics of greater tendency in the field of polymeric materials, and could be  
6 considered as an attractive approach to overcome some of the above-mentioned shortcomings  
7 of PHB [11–14]. Cellulosic fibers are cheap and abundant in nature and have, in general, high  
8 elastic modulus and relatively high thermal stability (at polymer processing temperatures).  
9 These characteristics make them appropriate for their use as reinforcing materials in polymeric  
10 matrices, enhancing the mechanical performance and reducing the final costs of the eventual  
11 composite [15–17] while decreasing the overall environmental impact of the product [18] Being  
12 cellulose the main component of plants, its use as reinforcing material allows the valorization of  
13 vegetal residues, thus promoting the circular economy while reducing the environmental impact  
14 of plastics. In this sense, the use of lignocellulosic residues from the agro-food industry as fillers  
15 for compostable polymer composite applications is an excellent circular approach to valorization  
16 of a residue, while enhancing the quality of the final compost.  
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28 Nevertheless, due to the natural origin of the cellulose fibers, the main features of the fibers  
29 (such as morphology, crystallinity, composition, purity, etc...) highly depend on the source [19]  
30 the extraction and purification method, the application of surface modifications or chemical  
31 treatments [14, 20]. In this sense, the influence of the source of the fiber on the overall  
32 properties of a polymer composite requires to be assessed for each particular fiber.  
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38 In a previous work the valorization of lignocellulosic fibers obtained from almond shell (AS) and  
39 rice husk (RH) as fillers in PHB based composites obtained by melt mixing has been assessed  
40 [21]. That work focused on the influence of fiber morphology, type and content on the  
41 mechanical, thermal, barrier properties, compostability and thermoforming ability of the  
42 PHB/fiber composites. Although some differences were found, mainly related with the  
43 morphology of the fibers, both AS and RH have demonstrate their suitability as reinforcing  
44 materials for the studied matrix, and PHB/AS composites showed promising thermoformability.  
45 However, the barrier properties and thermal stability of the composites was considerably  
46 compromised.  
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55 Taking into account these preliminary results, some questions are still present. On the one hand,  
56 the effect of the use of untreated fibers on the PHB properties has not been evaluated in the  
57 previous study. On the other hand, an assessment of the effectiveness of a fiber treatment, in  
58 terms of whether the treatment of the fiber could yield to an improvement of the performance  
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1 of the composite without a severe increase in the cost and environmental impact of the so-  
2 obtained material, would be relevant. In particular, a study covering how the treatment  
3 influences the overall mechanical, thermal and crystallization behavior of the composites. And  
4 finally, what is the effect of the fibers incorporation on the embrittlement over time of PHB.  
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7 Aiming at answering these questions from an industrial point of view, in these work, PHB/fiber  
8 composites containing 10 phr of AS or RH fibers without and with a combined NaOH + peracetic  
9 acid (PAA) treatment has been prepared by twin screw extrusion. A commercial cellulose grade  
10 (TC90) has been used as reference material. Peracetic acid treatment was selected because of it  
11 has higher selectivity in the removal of lignin and other non-cellulosic components than other  
12 methods such as chlorite/acetic acid while producing less severe impacts in cellulose structure,  
13 as reported by Kumar et al. [22]. In addition, according to Zhao et al. [23] a combined NaOH +  
14 PAA leads to high pulping yields and the fibers obtained possess superior mechanical properties  
15 and brightness compared to other usual pulping methods such as Kraft, while being a chlorine-  
16 free method is more environmentally friendly.  
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26 The influence of the untreated and treated fibers on morphology, thermal, crystallization,  
27 tensile properties, fracture toughness and dynamo mechanical behavior of the PHB/fiber  
28 composites has been studied. The effect of the fibers on the ageing and secondary crystallization  
29 of PHB has been also assessed.  
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## 35 **2. Experimental**

### 36 ***2.1. Materials***

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42 Commercial Poly(3-hydroxybutyrate) grade (P309) was purchased from Biomer® (Krailling,  
43 Germany) in pellet form. Micronized almond shell (AS) was kindly supplied by Unió Corporació  
44 Alimentària (Reus, Spain). Rice husk (RH) by-product from the rice production process was kindly  
45 provided by Herba Ingredients (Valencia, Spain). Purified alpha-cellulose fiber grade with an  
46 alpha-cellulose content >99.5% (TC90) was purchased from CreaFill Fibers Corp. (US). Sodium  
47 hydroxide (NaOH, 98%), hydrogen peroxide (H<sub>2</sub>O<sub>2</sub>, 30%), glacial acetic acid (CH<sub>3</sub>COOH, 99%) and  
48 sulphuric acid (H<sub>2</sub>SO<sub>4</sub>, 98%) were purchased from Sigma Aldrich.  
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## ***2.2. Fiber treatment***

Almond Shell (AS) and Rice Husk (RH) fibers were subjected to a two-stage purification treatment in order to remove the major part of impurities and non-cellulosic components such as waxes, lignin and hemicelluloses.

Prior to purification treatment micronized Almond Shell was sieved through a 140  $\mu\text{m}$  mesh and native rice husk was grinded in a mechanical knife mill and then sieved in 140  $\mu\text{m}$  mesh. Grinded and sieved AS and RH powders were subjected to the two-stage purification treatment. The first stage consisted in an alkaline attack with NaOH. The fibers were soaked in 5% (wt/v) NaOH solution for 2h at 80°C under vigorous stirring maintaining a fibre:liquid ratio of 1:20. This treatment was applied twice. The second stage consisted in an oxidative attack with peracetic acid (PAA). Alkaline pretreated fibers were soaked into peracetic acid with a fiber:liquid ratio of 1:20 under vigorous stirring at 80°C for 4h (until whitening of suspension was visually detected). The peracetic acid was prepared by the mixing of 30%(v/v) hydrogen peroxide and acetic acid in the reaction medium with a volume ratio of 3:1 at room temperature. 1% (w/w) of sulfuric acid was added as catalyzer to ensure a chemical shift in the direction of the products. This procedure was adapted from literature [23, 24]. After each stage the fibers were filtered and washed repeatedly in distilled water until neutral pH was reached. The purified powder was dried at 60°C for at least 24 hours and grinded again and sieved through a 140  $\mu\text{m}$  mesh.

Untreated Almond Shell and Rice Husk fibers were named as U\_AS and U\_RH, and treated ones as T\_AS and T\_RH, respectively.

## ***2.3. Samples preparation***

Before extrusion, the PHB used in this study was dried according to the manufacturer instructions in a Piovan dehumidifying drier and the commercial cellulose (TC90) and the untreated (U\_AS, U\_RH) and treated (T\_AS, T\_RH) fibers were dried at 100°C in oven for at least 2h. Composites containing 10 phr (referred to 100% wt. unit PHB) fiber loading were prepared in a twin-screw co-rotating extruder from DUPRA SL (Castalla, Spain) with L/D ratio of 24 and a diameter of 2.5 cm, with a temperature profile of 165/170/175/180 °C (from the hopper to the extruder die) at a speed of 40 rpm. The PHB pellets and the fibers were manually dry-mixed in zip-bags before extrusion and fed to the extruder hopper. The extruded material was pelletized and dried at in a Piovan. Compression moulded samples were obtained in a hot-plate press (Carver Inc. E 43201) at 180°C. Bars of 50x12.5x3.5 mm for dynamo-mechanical analysis tests,

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films of 0.4 nominal thickness for uniaxial mechanical tests and plates of 2.4 mm nominal thickness for Linear Elastic Fracture Mechanics (LEFM) tests were obtained. The different formulations studied were named as PHB/X, where X corresponds to the fiber type: TC90, U\_AS, U\_RH, T\_AS and T\_RH.

## 2.4. Characterization

The morphology of the untreated and treated AS and RH fibers, the commercial cellulose (TC90) and the PHB/fiber composites was examined by scanning electron microscopy (SEM) using a high-resolution field-emission JEOL 7001F microscope. The aspect ratio and the average particle size of the different fibers were estimated using the image analysis software Fiji® (the number of particles measured was never below 250). Composite films were cryofractured after immersion in liquid nitrogen to avoid plastic deformation. The fibers and composites were coated by sputtering with a thin layer of platinum for the SEM analysis. The elemental analysis of RH fibers was conducted by energy dispersion X ray microanalysis (EDX).

The crystallinity of the untreated and treated AS and RH fibers and the TC90 was assessed by Wide angle X-ray scattering (WAXS) using a Bruker AXS D4 Endeavor diffractometer. Radial scans of intensity versus scattering angle ( $2\theta$ ) were recorded at room temperature in the range of 2 to  $40^\circ(2\theta)$  (step size =  $0.02^\circ(2\theta)$ , scanning rate = 4 s/step) with filtered  $\text{CuK}\alpha$  radiation ( $\lambda = 1.54$  A), an operating voltage of 40kV, and a filament current of 40 mA. The crystallinity was evaluated according to Seagal et al. [25] method, where the crystallinity index  $C_i$  can be determined with the following equation (1):

$$C_i(\%) = \frac{I_{002} - I_{am}}{I_{002}} \times 100 \quad (1)$$

where  $I_{002}$  is the maximum intensity of the (002) lattice diffraction peak of cellulose located at a diffraction angle of around  $22^\circ(2\theta)$  and  $I_{am}$  is the intensity scattered by the amorphous phase of the sample measured at around  $18^\circ(2\theta)$ .

Fourier infrared (FT-IR) spectra of the untreated and treated AS and RH fibers and the TC90 were recorded by a Jasco FT/IR-6200 equipped with an attenuated total reflection (ATR) accessory in the range of  $400\text{-}4000\text{ cm}^{-1}$  in transmission mode.

Thermogravimetric analysis (TGA) of the untreated and treated AS and RH fibers, the TC90, as well as the neat PHB and its composites, was performed with a TG-STDA Mettler Toledo model TGA/STDA851e/LF/1600 analyzer. The samples with an initial mass of typically about 15 mg were

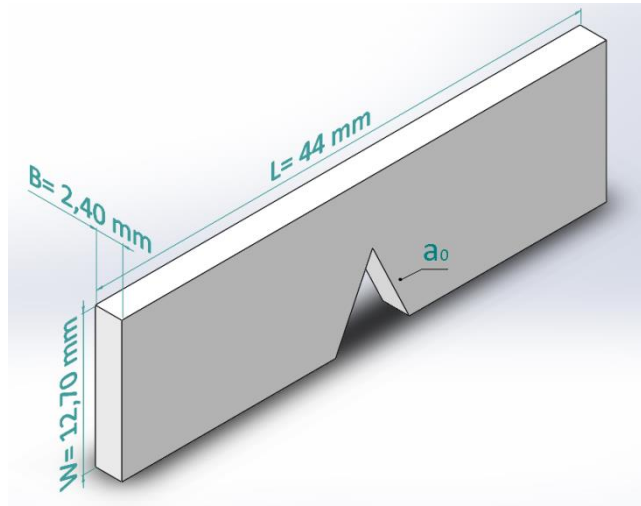
1 heated from 50 to 600°C at a heating rate of 10°C/min under nitrogen flow. The thermal stability  
2 of both the fibers and the PHB/fiber composites was evaluated and the residue at 600°C was  
3 determined. For the composites, the initial decomposition temperature ( $T_{5\%}$ , temperature at 5%  
4 weight loss) and the maximum decomposition rate temperature ( $T_d$ ) were determined from the  
5 weight loss curve and the maximum value of weight loss derivative respectively.  
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9 The mechanical characterization of neat PHB and the composites was performed by tensile tests  
10 according to the ASTM D638 standard, using dumbbell 400  $\mu\text{m}$ -thick samples (Type IV) die-cut  
11 from the hot pressed films. Tensile tests were conducted in a universal testing machine  
12 (Shimatzu AGS-X 500N) at room temperature with a cross-head speed of 10 mm/min. The  
13 samples were tested immediately after be processed (0 days) and after 15 days to explore the  
14 effect of ageing and secondary crystallization on their mechanical performance [9]. All the  
15 samples were stored in a vacuum desiccator at ambient temperature until be tested.  
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19 Linear Elastic Fracture Mechanic (LEFM) was applied using single edge notched bend (SENB)  
20 specimens, as depicted in Fig. 1. The samples were prepared by cutting the hot-pressed plates  
21 (nominal dimensions: 2.4 x 12.7 x 44 mm). Notches were produced centrally on the narrowest  
22 side of the prismatic bars, using a 45° V notch-broaching tool. Prior to testing, the notches were  
23 sharpened with a single cut from a razor blade (razor pushing) in order to get an initial sharp  
24 crack, as indicated in the European Structural and Integrity Society (ESIS) protocols for KIC  
25 determination [26]. The depth of such sharpening,  $\Delta a$ , was measured using a binocular  
26 microscope on each fractured specimen, in order to determine the real value of the crack ( $a_0 +$   
27  $\Delta a$ ).  $\Delta a$  values were around 0.2 mm in all specimens. The tests were performed using a universal  
28 testing machine Shimatzu AGS-X equipped with a 5 kN load cell. The crosshead speed of the  
29 test was 10 mm/min. The notch depth (i.e. initial crack length),  $a_0$  of SENB specimens was set to  
30 7 mm and the tests were conducted at room temperature. The fracture toughness ( $K_{IC}$ ) was  
31 calculated by the ESIS protocol [26].  
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$$34 \quad K_{IC} = f \frac{P_C}{B W^{1/2}} \quad (2)$$

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36 where  $P_C$  is the maximum load point. B and W are thickness and width of the specimens. The  
37 geometrical correction factor, f, a function of  $a/W$  where a is the initial crack length (i.e.  $a_0 + \Delta a$ ),  
38 was obtained from the ESIS protocol [26].  
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**Figure 1. Scheme of SENB specimens**

Dynamic mechanical analysis (DMA) experiments were conducted on hot pressed sample platelets (55 × 12.5 × 3.5 mm) in an AR G2 oscillatory rheometer (TA Instruments, New Castle, DE) equipped with a clamp system for solid samples (torsion mode). Samples were heated from -20 °C to melting temperature with a heating rate of 2 °C/min at a constant frequency of 1 Hz. The maximum deformation ( $\gamma$ ) was set to 0.1%.

Differential scanning calorimetry (DSC) experiments were conducted on a DSC2 (Mettler Toledo) with an intracooler (Julabo FT900) calibrated with an Indium standard before use. The samples weighing typically 6 mg were first heated from -20°C to 200°C at 10°C/min and kept for 5 min to erase thermal history, followed by cooling to -20°C and heating to 200°C at 10°C/min. The samples were tested at 0 days and after 100 days of ageing at room temperature. Melting temperature ( $T_m$ ) and enthalpy ( $\Delta H_m$ ) were calculated from the first and second heating scans and crystallization temperature ( $T_c$ ) and enthalpy ( $\Delta H_c$ ) from the cooling scan. The crystallinity ( $X_c$ ) of the PHB phase of the composites was determined by applying the following expression [27]:

$$X_c(\%) = \frac{\Delta H_m}{w \cdot \Delta H_m^0} \times 100 \quad (3)$$

where  $\Delta H_m$  (J/g) is the melting enthalpy of the polymer matrix,  $\Delta H_m^0$  is the melting enthalpy of 100% crystalline PHB (perfect crystal) (146 J/g) [9] and  $w$  is the polymer weight fraction of PHB in the blend.



## 3. Results and discussion

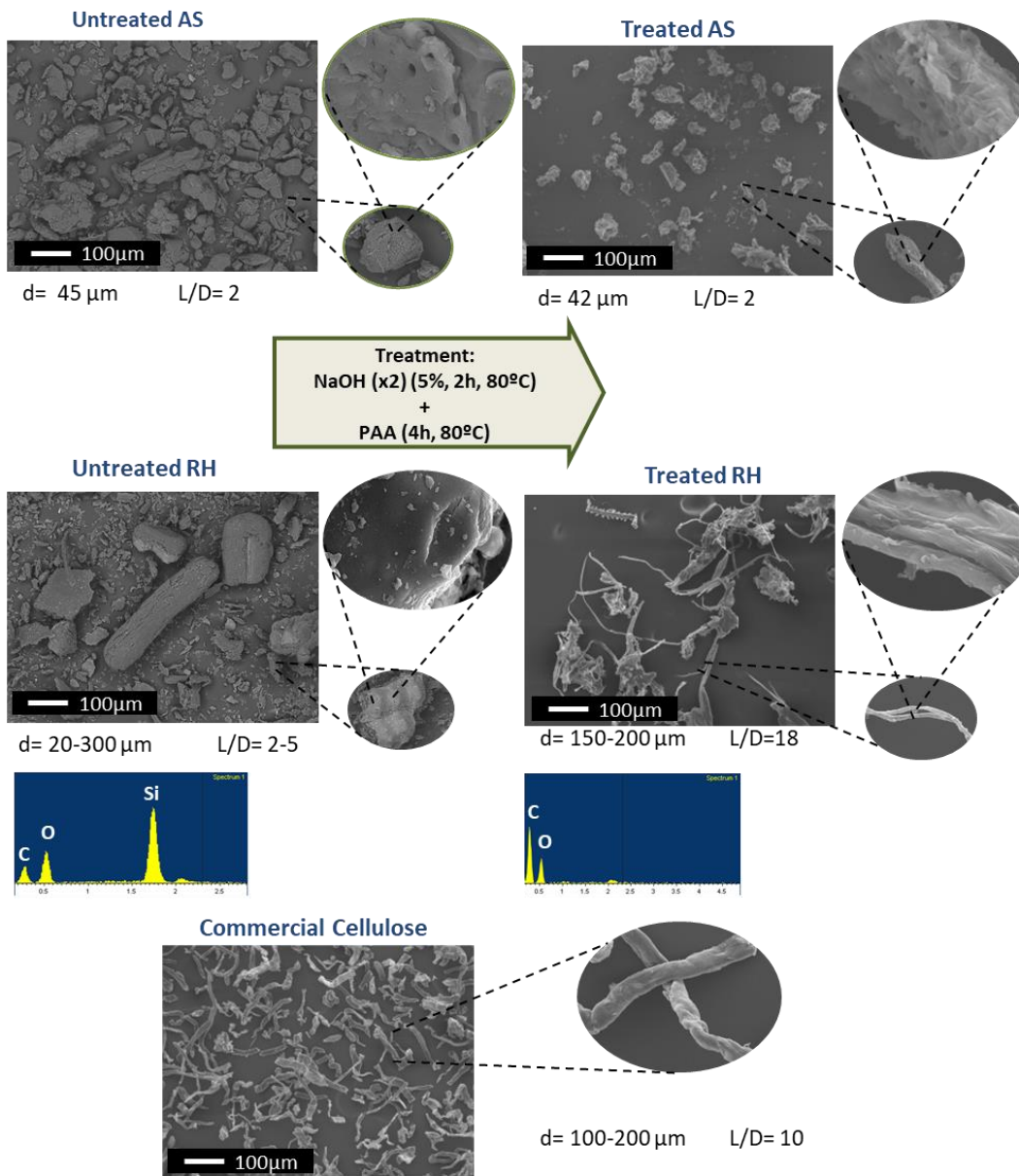
### 3.1. Fiber characterization

SEM micrographs of the untreated and treated AS and RH fibers and commercial cellulose, the estimated aspect ratio and average particle size of the different fibers and the EDX microanalyses of RH fibers are shown in Figure 2.

Untreated Almond Shell (U\_AS) powder was composed by irregular particles and aggregates with an average particle size of about 45  $\mu\text{m}$  and an estimated aspect ratio of around 2. The particles present a porous surface with the presence of small pinholes. The combined NaOH and PAA treatment didn't significantly change the morphology of AS fibers. Only a slight increase in the surface roughness can be observed in T\_AS particles with respect to U\_AS; however, the average size is of about 42  $\mu\text{m}$  and the aspect ratio is of c.a.2. Hence, the treatment has no apparent effect on the morphology of the fibers.

The U\_RH powder is formed by particles of different morphologies (rod-shaped, junks and fibers) whose particle sizes vary from a few microns up to 300  $\mu\text{m}$  in length. With close examination a smooth surface of the particles can be observed, probably related with the presence of waxes as it will be further discussed. The EDX microanalysis reveals the presence of silica with an estimated content around 14%, this value is in accordance with the ones reported in literature for this type of fibers [28]. In this case, the combined NaOH and PAA treatment had a big impact on the morphology of RH fibers. After treatment the particles present fibrillary morphology with more homogeneous particle size distribution with an average particle size ranging from 150 to 200  $\mu\text{m}$  in length and high aspect ratio, estimated at about 18. The particles surface presents a rough topography. This finding suggests the removal of waxes and other non-cellulosic components by the treatment. The EDX microanalysis didn't reveal the presence of silica, evidencing the effectiveness of the treatment in removing this component.

The commercial cellulose used in this study as a reference material has a fibrillary morphology. The fibers present varying lengths up to 200  $\mu\text{m}$ . In all cases, the diameter is contained within the 10-20  $\mu\text{m}$  range, thus rendering an average aspect ratio of about 10.



**Figure 2. SEM micrographs of the untreated and treated RH and AS fibers, the commercial cellulose (TC90), and EDX microanalysis of RH fibers**

The crystallinity index ( $C_i$ ) of the U\_AS, U\_RH, T\_AS, T\_RH and TC90 fibers was determined by WAXS according to Seagal et al. [25] (equation 1). WAXS patterns of the studied fibers are shown in Figure 3.

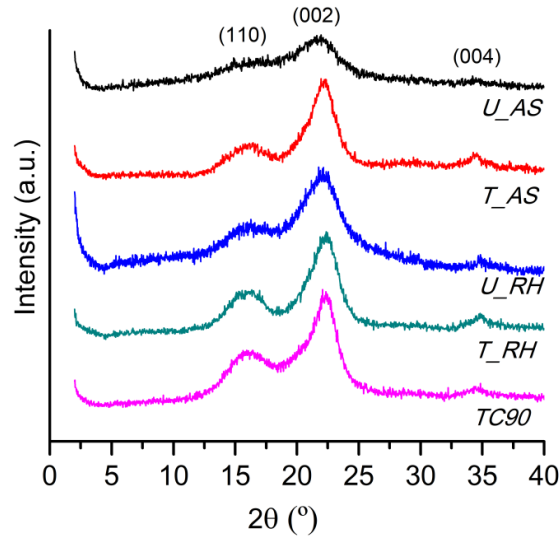


Figure 3. WAXS patterns of TC90 and untreated and treated AS and RH fibers

All the fibers present a characteristic X-ray diffraction pattern of a semicrystalline material with an amorphous broad hump and different crystalline peaks. Three peaks located at  $16^\circ$ ,  $22^\circ$  and  $34^\circ$  can be observed in all cases, which are characteristics of the (110), (002) and (004) reflections of type I cellulose lattice [29, 30]. When assessing the crystallinity of the untreated AS and RH, the values of crystalline index ( $C_i$ ) (according to eq. (1)) of 40 and 48 %, respectively. After treatment their crystallinity indexes were increased to 69 and 70%. These values are comparable to that corresponding to commercial cellulose, TC90, that presents a  $C_i$  of 63 % and are coherent with those reported in literature for these types of fibers [31, 32]. The increase in crystallinity after treatment is indicative of the elimination of amorphous components present in the fibers (mainly lignin and hemicelluloses) [30, 33].

FTIR was employed in this work in order to verify the effectiveness of the chemical treatment in removing the non-cellulosic components of fibers, such as lignin, hemicelluloses, pectin or waxes. The FTIR spectra of untreated and treated AS and RH fibers and the commercial cellulose (TC90), as reference, are represented in Figure 4.

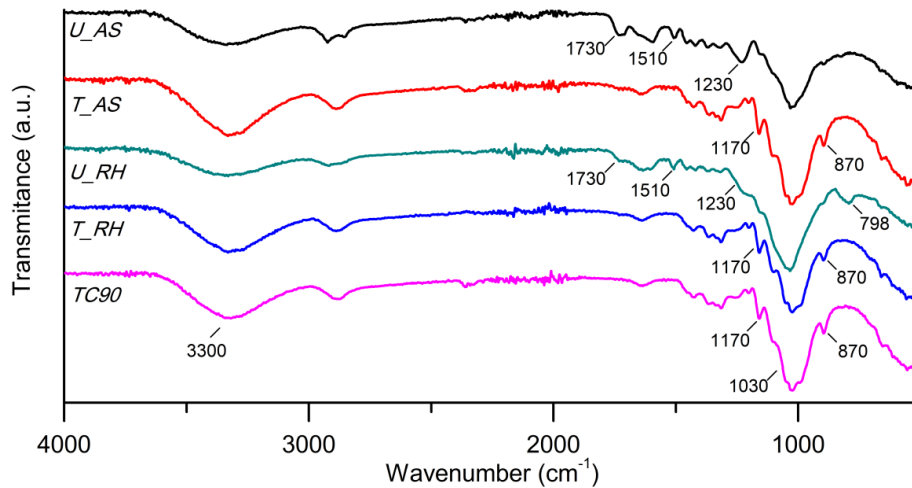
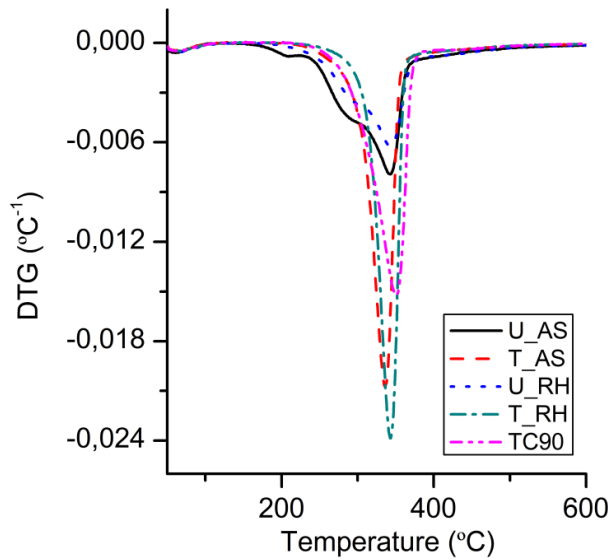
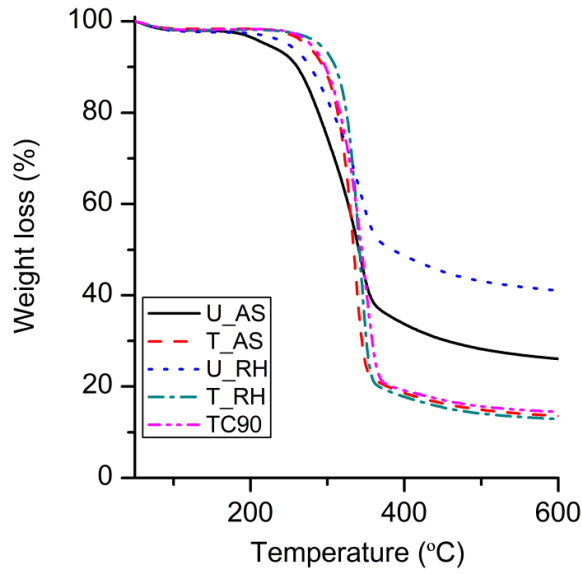


Figure 4. FTIR spectrograms of TC90 and untreated and treated AS and RH fibers

Untreated AS and RH fibers present the characteristic bands of lignin, hemicelluloses, waxes and pectin. The band at 1730  $\text{cm}^{-1}$  is ascribed to either acetyl and carbonyl groups from carboxylic ester linkages in hemicelluloses and lignin [34, 35], or to pectin, wax and natural fats [36, 37], the band at 1510  $\text{cm}^{-1}$  is due to the C=C stretching of the aromatic rings present in the lignin structure [38, 39], and the band in the 1200-1250  $\text{cm}^{-1}$  spectral region is ascribed to -C-O stretching vibration of the acetyl groups in lignin and hemicelluloses [35, 39]. All these bands are not detected in the spectra corresponding to treated AS and RH fibers and commercial cellulose, thus indicating the successful removal of these components through the treatment. The band at 798  $\text{cm}^{-1}$  present in the U\_RH corresponding to the silica vibrations [34] also disappears in T\_RH spectrum confirming the results obtained by EDX microanalysis, as it has been discussed in SEM results. Additionally, the intensity of the broad band around 3300  $\text{cm}^{-1}$  is increased in T\_AS and T\_RH spectra with respect to untreated fibers and it starts to appear a shoulder on this band. According to Ndazi et al. [34] and Mariano et al. [24] the increased intensity of this band and the shoulder are indicative of the presence of more free -OH reactive groups on the fibers surface. Also, the characteristic bands of vibrational C-O-C, C-O and C-H strain of cellulose [35, 39] at 1170  $\text{cm}^{-1}$ , 1030  $\text{cm}^{-1}$  and 896  $\text{cm}^{-1}$  show better resolution and higher intensity in spectra corresponding to treated and commercial cellulose fibers than in the spectra corresponding to untreated ones, also confirming the purification of the fibers by the treatment.

The thermal stability of the untreated and treated AS and RH fibers and the TC90 was studied by TGA. The weight loss and first derivative (DTG) curves are shown in Figure 5.



**Figure 5. Weight loss (a) and DTG (b) curves corresponding to TC90 and untreated and treated AS and RH fibers**

As shown in Figure 5 the thermal degradation of the untreated fibers takes place in a broad range of temperatures with different weight loss steps related to the thermal degradation of its main components (hemicelluloses, cellulose and lignin). The decomposition of hemicelluloses primarily occurs between 220–315 °C. Cellulose decomposes at higher temperatures (around 345 °C), whilst lignin decomposition happens slowly over the whole temperature range studied [30, 40]. In addition, U\_AS and U\_RH present a residue at 600°C of 26 and 41%, respectively. The higher residues with respect to treated fibers are related with their high content of lignin and, in case of RH, also by the silica present in its composition. On the other hand, the thermal

1 degradation of TC90 and the treated fibers takes place in a single step with the highest weight  
2 loss rate at, approximately, 345°C corresponding to cellulose degradation and a residue of 12%,  
3 confirming the successfully removal of the non-cellulosic components through the treatment. In  
4 addition, after treatment, the onset degradation temperature of the AS and RH fibers was  
5 displaced about 40°C to higher temperatures thus indicating that their thermal stability were  
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11 In a previous work we used a combined NaOH+H<sub>2</sub>O<sub>2</sub> purification treatment for AS and RH fibers  
12 [21]. Attending to the results obtained by means of the treatment applied in here (combined  
13 NaOH+ PAA), it seems to be more efficient in removing the non cellulosic components, thus  
14 yielding to fibers with higher purity grade and relative crystallinity indexes.  
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### 20 ***3.2. Composites characterization***

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23 To evaluate the particle distribution and the fiber/matrix interactions, the morphology of the  
24 cryofractured surfaces of PHB/fiber composites was studied by SEM. The micrographs of the  
25 composites with TC90, U\_AS, T\_AS, U\_RH and T\_RH fibers are presented in figure 6.  
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29 In general, the composites prepared with the studied fibers (U\_AS, T\_AS, U\_RH and T\_RH)  
30 present a particle distribution similar to that corresponding to the commercial cellulose (TC90).  
31 In all cases the fibers appear individualized and homogeneously distributed within the polymeric  
32 matrix indicating that an effective blending was achieved. However, some differences can be  
33 detected among the different fibers regarding to the fiber-matrix interactions. For the untreated  
34 fibers, pull-out effect (marked with an arrow) can be observed, whilst this is not evident in the  
35 composites prepared with the treated fibers and the TC90. This phenomenon could be related  
36 to the presence of waxes and impurities on the untreated fiber surface, together with a more  
37 limited –OH reactive groups exposed. These waxes and other non-cellulosic components could  
38 form a weak layer that hinders the fiber/matrix interaction. In the micrograph corresponding to  
39 PHB/U\_RH it can be observed a fiber with a very smooth surface as a result of the presence of  
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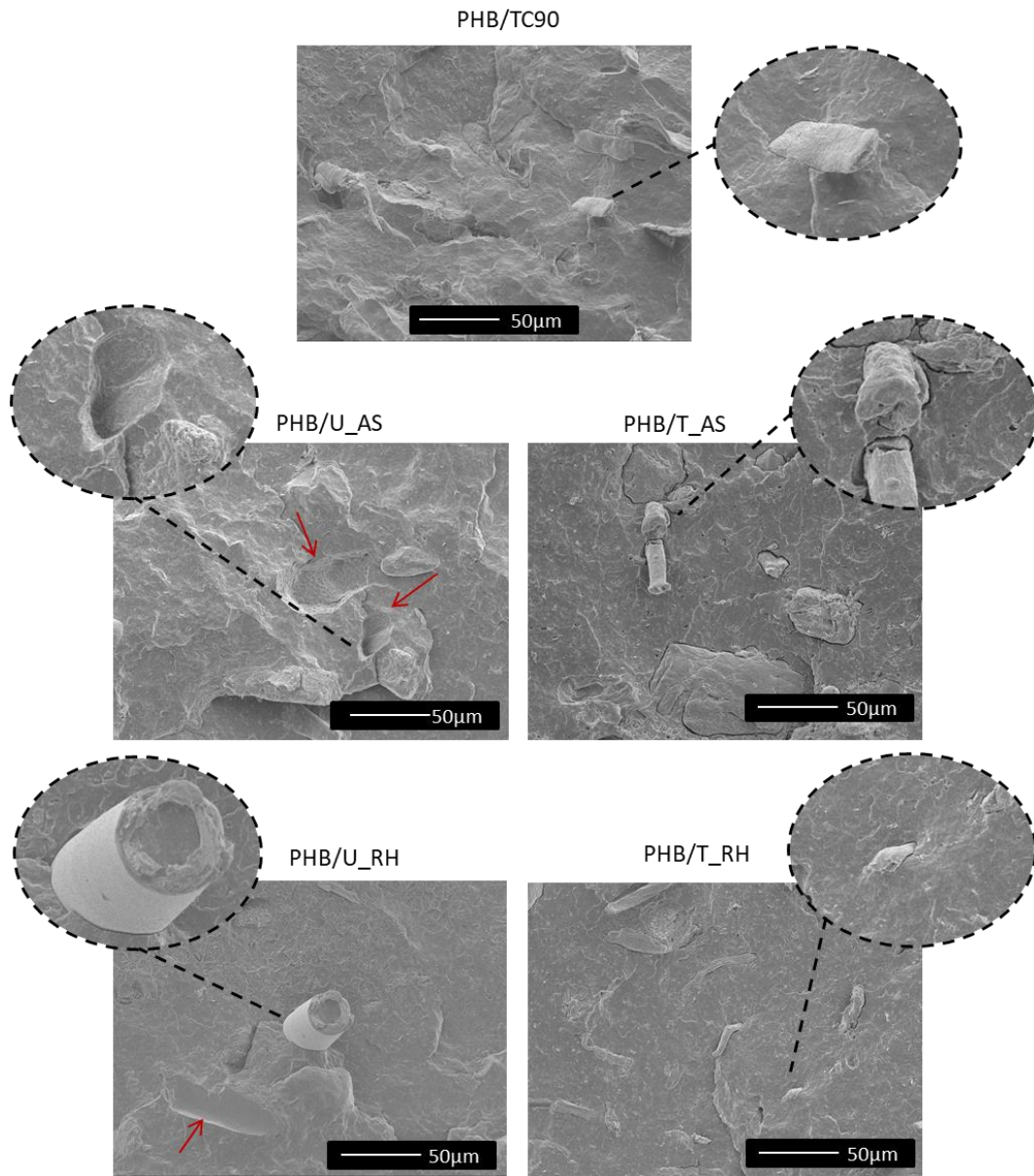


Figure 6. SEM micrographs of PHB/TC90, PHB/U\_AS, PHB/T\_AS, PHB/U\_RH and PHB/T\_RH composites

these non-cellulosic components (probably waxes). In case of the composites containing the T\_AS, T\_RH and TC90 the presence of some broken fibers that seem to be well embedded into the PHB matrix suggesting certain enhanced fiber-matrix interaction with respect to untreated fibers. However, some small gaps between the fibers and the polymeric matrix can be also detected, especially in PHB/T\_AS composites. These gaps may have been induced during the specimen preparation by means of immersion in liquid nitrogen and subsequent brittle fracture.

The mechanical properties of the composites were evaluated by uniaxial tensile tests up to break. The effect of the intrinsic embrittlement of PHB with time due to secondary crystallization and physical ageing [9] on the mechanical performance of PHB/composites was also evaluated.

Tensile modulus of elasticity, tensile strength at yield and elongation at break of the composites at 0 days and after 15 days of ageing at room conditions are represented in Figure 7 a, b and c, respectively.

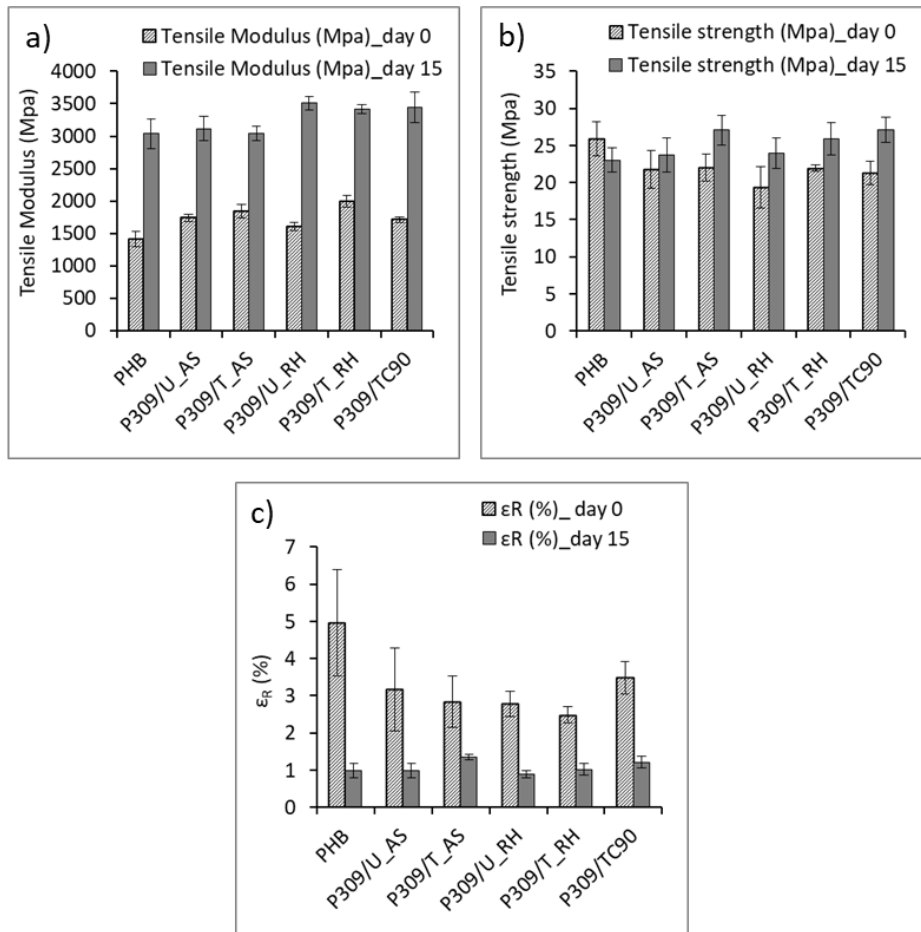


Figure 7. Tensile modulus (a), Tensile strength (b) and elongation at break (c) of neat PHB and the composites.

The mechanical behavior of neat PHB and PHB/fiber composites is not the same at 0 days and after 15 days of ageing. At 0 days the mechanical behavior is dominated by the fiber/matrix interactions whilst at 15 days the behavior is more dependent on the secondary crystallization and physical ageing.

As shown in Figure 6 at 0 days PHB presents a tensile modulus of 1400 MPa, a tensile strength of 26 MPa and an elongation at break of 5%. The incorporation of the fibers leads in all cases to clear improvement of the elastic modulus due as a result of their reinforcing nature; while a slight reduction of tensile strength and elongation at break can be also detected, as is typical in short fiber reinforced composites with a brittle polymer matrix. At certain level of stress, the fibers act as flaws (i.e. stress concentrators) leading to a premature failure and, consequently, a reduction in tensile strength and elongation at break [41].



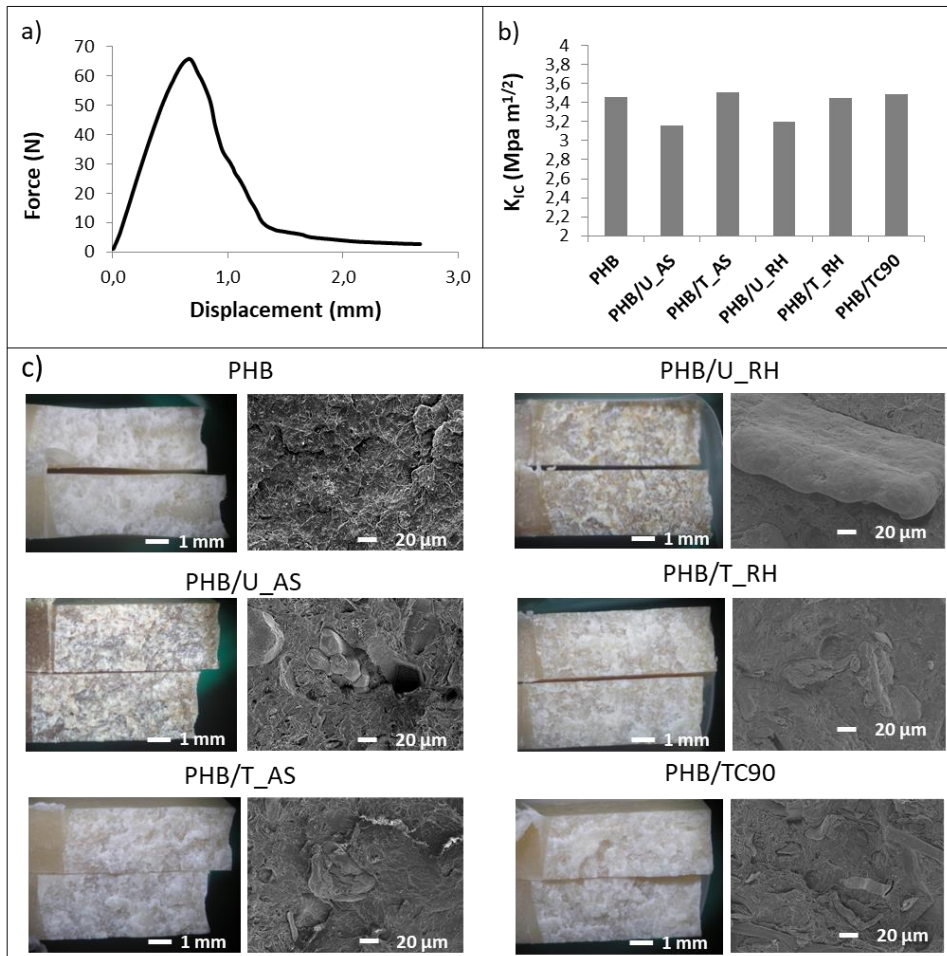
1 Most noticeable increase in tensile modulus is found for the treated fibers. In particular, the  
2 tensile modulus of PHB/T\_RH composites was increased of about 40% with respect to neat PHB  
3 and 17% compared to the commercial cellulose. The higher reinforcing effect of treated fibers  
4 with respect to untreated ones could be attributed to the increased crystallinity and purity of  
5 the fibers, together with a higher presence of hydroxyl groups on the surface leading to a better  
6 interaction by hydrogen bonds with the polymer carbonyl groups [42–44].  
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11 After 15 days of ageing at room conditions, a clear embrittlement of the PHB matrix is evidenced.  
12 This behavior is attributed in literature to a secondary crystallization and physical ageing  
13 (reduction of the mobility of amorphous phase and constriction of rigid amorphous regions) [45,  
14 46]. In neat PHB ageing produces an increase in modulus and a decrease in elongation at break  
15 (Fig. 7). For the composites the increase in tensile modulus is also observed, while the tensile  
16 strength and the elongation at break are slightly superior to those corresponding to neat PHB.  
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23 The fiber treatment does not seem to have any influence on the elastic modulus of the  
24 composites after ageing. Nevertheless the tensile strength is slightly higher for the composites  
25 prepared with the treated fibers with respect to the untreated ones. This could be due to a less  
26 fiber/matrix detachment owing to a better fiber adhesion.. In addition, the presence of  
27 impurities and waxes on the untreated fiber surfaces could promote the fiber/matrix  
28 detachment by a weak layer mechanism as it has been discussed in SEM results, as pull-outs  
29 have been detected (Fig 6). In any case, all the compositions present fragile behavior.  
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### 39 LEFM

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41 To assess the influence of the fiber type on the PHB fracture behavior three point bending tests  
42 at slow speed (10 mm/min) were performed and the results were analyzed following the  
43 fracture mechanics approach. The behavior was in all cases sufficiently linear to allow the  
44 application of the linear elastic fracture mechanics (LEFM) to characterize the fracture  
45 toughness ( $K_{IC}$ ) according with ESIS protocol [26]. The results are summarized in Figure 8: a  
46 representative force-displacement curve are represented in Figure 8a, the  $K_{IC}$  values of the  
47 materials are shown in Figure 8b and representative photographs and SEM micrographs of the  
48 fractured specimens are depicted in Figure 8c.  
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**Figure 8. LEFM results: representative force-displacement curve (a), K<sub>IC</sub> values of neat PHB and the composites (b), and representative SEM micrographs of the fractured specimens (c)**

According to the results presented in Figure 8b, the composites containing the treated fibers and the commercial cellulose (PHB/T\_AS, PHB/T\_RH and PHB/TC90) present K<sub>IC</sub> values similar to neat PHB. A slight decrease in fracture toughness is observed for the composites containing the untreated fibers (PHB/U\_AS and PHB/U\_RH). In any case, the differences in K<sub>IC</sub> values are below 10%.

As shown in Figure 8c, the matrix presents a fracture morphology with microcavities and unstable crack propagation and even deviation of the transverse plane to the application of force which implies high concentration of elastic energy attributable to a high crystallinity (see DSC Table 1) and little fraction of tie molecules. Nevertheless, the fracture surface also shows some small crests that are indicative of certain localized plastic deformation of the matrix that is coherent at temperatures above T<sub>g</sub>. On the one hand, the introduction of cellulose should suppose an increase in rigidity and toughness, but, on the other hand, the fibers act as crack initiators or stress concentrators. For untreated fibers the reinforcing effect is lower and the stress concentration is higher (due to minor fiber /matrix interaction). As shown in Figure 8c, in

PHB/U\_AS and PHB/U\_RH the detachment of the fibers is evident supporting the hypothesis of a “weak layer mechanism” failure caused by the presence of impurities and waxes on the fiber surface.

### DMA

In addition to mechanical and fracture toughness characterization dynamic mechanical analysis (DMA) was performed in order to assess the effect of temperature on the mechanical behavior of the PHB and the composites. The storage modulus ( $G'$ ) and the Tan delta ( $\text{Tan } \delta$ ) curves as a function of temperature are shown in Figures 9a and 9b, respectively.

As it was expected, the  $G'$  of PHB, which is a measure of the stiffness of a viscoelastic material, decreases as the temperature increases due to the softening of the sample at higher temperatures (Fig.9a). A slight increase in  $G'$  is observed in filled compositions that is related with the increased rigidity of the samples because of the reinforcing effect of the fibers [43, 47] Nevertheless, the differences in  $G'$  among the different compositions are quite small.

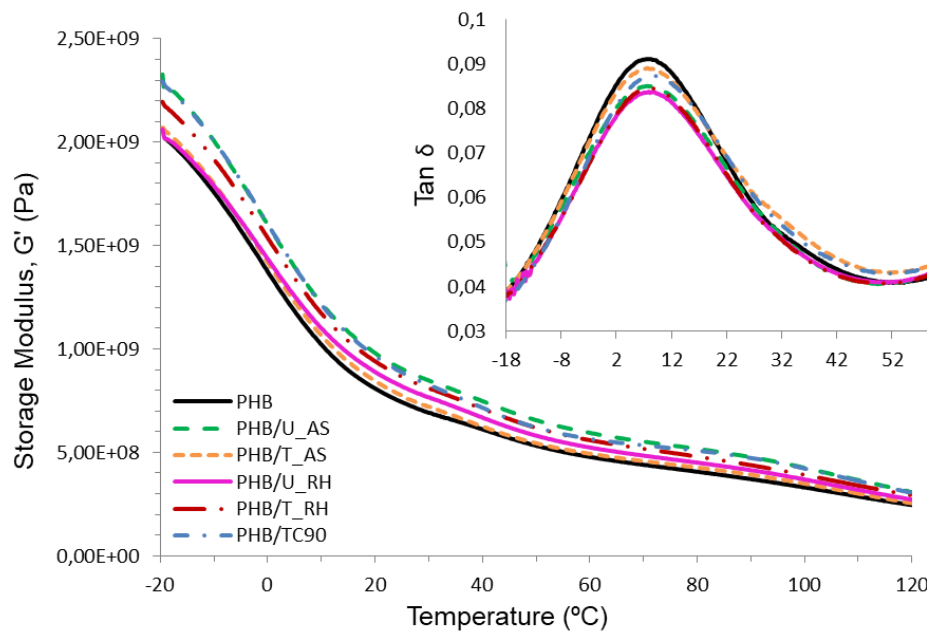


Figure 9. Storage modulus ( $G'$ ) and Tan  $\delta$  (inset) evolution with temperature of neat PHB and the composites

Tan  $\delta$  represents the ratio of the viscous and elastic components of the modulus and is a measure of the energy dissipated as heat during the dynamic test [48]. The Tan  $\delta$  curve presents a single event before melting for the studied range, being this event the  $\alpha$  transition, generally

1 attributed to the glass transition ( $T_g$ ) in semicrystalline polymers [49]. The  $T_g$  value for neat PHB  
 2 is around 8°C, while fiber addition does not have any effect on the glass transition temperature.  
 3 The only noticeable difference is a slight reduction in peak height for the composites with  
 4 respect to neat PHB. According to Wong et al. [50], a decrease in the value of the  $T_g$  peak can  
 5 be attributed to a major number of polymer chains with restricted movement (i.e. does not  
 6 undergo this transition). For the samples studied, this decrease could be ascribed to the  
 7 presence of the fibers.  
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### 10 DSC

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 12 The influence of the fiber type in crystallization and melting behavior of PHB has been studied  
 13 by DSC measurements. DSC measurements were performed at 0 days and after 100 days of  
 14 ageing at room conditions in order to study the effect of the fibers on the secondary  
 15 crystallization of PHB. The main thermal parameters, obtained from the first heating scans, the  
 16 cooling scan and the second heating scan are summarized in Table 1.  
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27 **Table 1. DSC parameters of neat PHB and the composites**

	1st heating scan					
	0 days			100 days		
	$T_m$ (°C)	$\Delta H_m$ (J/g)	$X_c$ (%)	$T_m$ (°C)	$\Delta H_m$ (J/g)	$X_c$ (%)
PHB	175,1	72,0	49,3	169,6	85,8	58,7
PHB/U_AS	174,0	71,9	54,2	171,8	74,0	55,8
PHB/T_AS	171,6	60,0	45,2	172,1	72,0	54,2
PHB/U_RH	172,1	70,8	53,3	172,1	73,5	55,4
PHB/T_RH	175,0	68,5	51,6	172,8	73,4	55,3
PHB/TC90	172,8	73,7	55,5	172,7	73,0	55,0
	cooling scan		2nd heating scan			
	$T_c$ (°C)	$\Delta H_c$ (J/g)	$T_m$ (°C)	$\Delta H_m$ (J/g)	$X_c$ (%)	
PHB	117,2	90,6	169,6	93,9	64,3	
PHB/U_AS	118,2	85,0	168,3	86,3	65,0	
PHB/T_AS	117,7	82,6	169,7	86,3	65,0	
PHB/U_RH	115,9	81,8	168,5	85,0	64,1	
PHB/T_RH	116,8	83,7	169,9	86,3	65,0	
PHB/TC90	117,1	82,2	168,8	84,1	63,4	

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 53 Regarding the results obtained from the first heating scans, with the processing conditions  
 54 applied, at initial times after samples be processed (0days) neat PHB present lower values of  
 55 crystallinity than the composites, except for U\_AS. However, after 100 days of ageing the  
 56 crystallinity of PHB increases about 20% due to secondary crystallization whilst the crystallinity  
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1 of composites increases on average less than 10% achieving lower final crystallinity than PHB.  
2 This behavior can be attributed to hindered motion of the polymer chains because of the  
3 presence of the fibers as it has been reported in literature for PHA/fiber composites [51].  
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6 With respect to the fiber type or treatment, no remarkable differences have been found in  
7 crystallization and melting behavior. After full erasure of the thermal history, at low heating and  
8 cooling rates (such as current DSC conditions), thus allowing full crystallization to take place, all  
9 the samples present comparable values of enthalpy, crystallization and melting temperatures  
10 obtained from the cooling and second heating scans.  
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### 14 TGA

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16 The thermal stability of neat PHB and the composites was evaluated by thermogravimetric  
17 analysis (TGA). The weight loss as a function of temperature and their first derivative (DTG)  
18 curves are shown in Figure 10. The onset degradation temperature (T5%), the maximum  
19 degradation temperature (Td) and the residue at 600°C are summarized in Table 2.  
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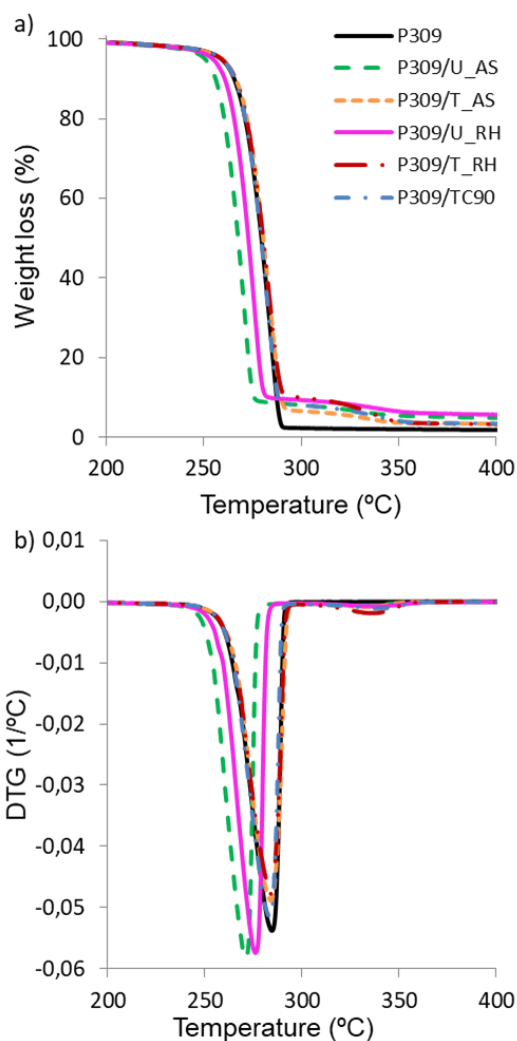


Figure 10. Weight loss (a) and DTG (b) curves corresponding to neat PHB and the composites.

Table 2. Thermal parameters obtained from TGA for neat PHB and the composites

	$T_{5\%}$ (°C)	$T_d$ (°C)	residue (600°C)
<b>PHB</b>	259	284	1,74
<b>PHB/U_AS</b>	251	271	4,27
<b>PHB/T_AS</b>	259	284	2,8
<b>PHB/U_RH</b>	255	276	5,08
<b>PHB/T_RH</b>	259	284	2,9
<b>PHB/TC90</b>	259	283	3,11

Thermal degradation of PHB takes place abruptly in a single weight loss step by a random chain scission mechanism as it has been widely reported [52]. The  $T_{5\%}$  and  $T_d$  values for neat PHB are 259°C and 284 °C, respectively, and the residue at 600°C is close to 2%. On the other hand, the

1 thermal degradation of the composites takes place in two steps. The second step, at  
2 approximately 345°C, corresponds to the degradation of the cellulosic fibers. The treatment of  
3 the fibers has a clear positive influence on the thermal stability of the PHB. As shown in Table 2,  
4 for the composites containing the untreated fibers, both  $T_{5\%}$  and  $T_d$  values are reduced with  
5 respect to neat PHB. This reduction is attributed to the presence of non-cellulosic components  
6 that starts their thermal degradation at lower temperatures than pure cellulose as discussed in  
7 fiber characterization section (see Figure 5). On the contrary, the incorporation of treated fibers  
8 does not affect the thermal stability of PHB (similarly to commercial cellulose). With respect to  
9 residue values at 600°C, they are coherent with the fiber content and type, that is, the values  
10 are proportional to the fiber content and the higher values correspond to the untreated fibers  
11 according with the residues reported in fiber characterization section (Figure 5).  
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## 21 **4. Conclusions**

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25 A combined NaOH+PAA treatment was applied to AS and RH fibers in order to remove the non-  
26 cellulosic components of the fibers and improve their surface characteristics to be used as  
27 reinforcing materials in PHB based composites taking as reference a commercial cellulose.  
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31 The combined NaOH + PAA treatment has demonstrated its effectiveness in the removal of lignin  
32 and other non-cellulosic components such as hemicelluloses and waxes based on FTIR, WAXS  
33 and TGA results. The resultant fibers present comparable relative crystallinity indexes, chemical  
34 composition and thermal stability to the commercial cellulose. In addition, treated fibers present  
35 more reactive free -OH groups in their surface with respect to untreated ones as it has been  
36 observed by FTIR. Nevertheless, some differences were found regarding to their morphology.  
37 Treated RH presents fibrillary morphology with high aspect ratio, whilst treated AS presents  
38 irregular shape with high surface roughness. In both cases the treatment leads to a surface  
39 roughening with respect to untreated ones.  
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48 In general, all the fibers have a reinforcing effect on the PHB matrix as it has been deduced by  
49 the increased elastic modulus and storage modulus of the composites with respect to neat PHB.  
50 However, best results were found for treated fibers that allows overall performance similar to  
51 commercially purified cellulose. In case of untreated fibers, some detachment of the fibers has  
52 been observed in SEM (thus revealing poor adhesion on the interface) probably due to the  
53 presence of waxes and other impurities on their surface. For these composites, the tensile  
54 strength, elongation at break and  $K_{Ic}$  values are lower than those corresponding to the  
55 composites containing the treated fibers and TC90. In addition, the thermal stability of the  
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composites with untreated fibers was slightly reduced with respect to neat PHB and the composites with treated fibers and commercial cellulose due to the presence of the non-cellulosic components.

The effect of the fibers on the mechanical properties and crystallinity over time of the composites were also evaluated. The secondary crystallization and the physical ageing of PHB conduce to a drastic embrittlement of the matrix over time. As it has been observed by DSC the presence of the fibers partly hinder the development of crystallinity of PHB during ageing.

The incorporation of the fibers to the PHB does not seem to affect negatively to the mechanical behavior of the matrix once the aging has taken place, since the brittleness of the system is governed by the intrinsic fragility of the PHB matrix.

To conclude, a sustainable straightforward purification treatment has been applied to two different agro-food based lignocellulosic residues resulting in two suitable reinforcing fillers for a PHB matrix. An assessment of the environmental impact and the cost of applying this treatment to the abovementioned residues, with respect to the use of a commercially purified cellulose would determine whether its use is convenient for a particular application, attending to the fact that there are no main technical differences for its use as fillers in PHB matrix.

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## References

1. Lambert, S., Wagner, M.: Environmental performance of bio-based and biodegradable plastics: The road ahead. *Chem. Soc. Rev.* 46, 6855–6871 (2017). <https://doi.org/10.1039/c7cs00149e>
2. Możejko-Ciesielska, J., Kiewisz, R.: Bacterial polyhydroxyalkanoates: Still fabulous? *Microbiol. Res.* 192, 271–282 (2016). <https://doi.org/10.1016/j.micres.2016.07.010>
3. Singh, M., Kumar, P., Ray, S., Kalia, V.C.: Challenges and Opportunities for Customizing Polyhydroxyalkanoates. *Indian J. Microbiol.* 55, 235–249 (2015). <https://doi.org/10.1007/s12088-015-0528-6>
4. Wang, Y., Yin, J., Chen, G.Q.: Polyhydroxyalkanoates, challenges and opportunities. *Curr. Opin. Biotechnol.* 30, 59–65 (2014). <https://doi.org/10.1016/j.copbio.2014.06.001>
5. Anjum, A., Zuber, M., Zia, K.M., Noreen, A., Anjum, M.N., Tabasum, S.: Microbial production of polyhydroxyalkanoates (PHAs) and its copolymers: A review of recent advancements. *Int. J. Biol. Macromol.* 89, 161–174 (2016). <https://doi.org/10.1016/j.ijbiomac.2016.04.069>
6. Laycock, B., Halley, P., Pratt, S., Werker, A., Lant, P.: The chemomechanical properties of microbial polyhydroxyalkanoates. *Prog. Polym. Sci.* 38, 536–583 (2013). <https://doi.org/10.1016/j.progpolymsci.2012.06.003>
7. Cava, D., Giménez, E., Gavara, R., Lagaron, J.M.: Comparative Performance and Barrier Properties of Biodegradable Thermoplastics and Nanobiocomposites versus PET for Food Packaging Applications. *J. Plast. Film Sheeting.* 22, 265–274 (2006). <https://doi.org/10.1177/8756087906071354>
8. Bugnicourt, E., Cinelli, P., Lazzeri, A., Alvarez, V.: Polyhydroxyalkanoate (PHA): Review of synthesis, characteristics, processing and potential applications in packaging. *Express Polym. Lett.* 8, 791–808 (2014). <https://doi.org/10.3144/expresspolymlett.2014.82>
9. Corre, Y.-M., Bruzaud, S., Audic, J.-L., Grohens, Y.: Morphology and functional properties of commercial polyhydroxyalkanoates: A comprehensive and comparative study. *Polym. Test.* 31, 226–235 (2012). <https://doi.org/10.1016/j.polymertesting.2011.11.002>
10. Liu, Q.-S., Zhu, M.-F., Wu, W.-H., Qin, Z.-Y.: Reducing the formation of six-membered ring ester during thermal degradation of biodegradable PHBV to enhance its thermal stability. *Polym. Degrad. Stab.* 94, 18–24 (2009). <https://doi.org/10.1016/j.polymdegradstab.2008.10.016>
11. Mohanty, A.K., Vivekanandhan, S., Pin, J.M., Misra, M.: Composites from renewable and sustainable resources: Challenges and innovations. *Science (80-. )*. 362, 536–542 (2018). <https://doi.org/10.1126/science.aat9072>
12. Väisänen, T., Haapala, A., Lappalainen, R., Tomppo, L.: Utilization of agricultural

and forest industry waste and residues in natural fiber-polymer composites: A review. *Waste Manag.* 54, 62–73 (2016). <https://doi.org/10.1016/j.wasman.2016.04.037>

13. Berthet, M.-A., Angellier-Coussy, H., Guillard, V., Gontard, N.: Vegetal fiber-based biocomposites: Which stakes for food packaging applications? *J. Appl. Polym. Sci.* 133, (2016). <https://doi.org/10.1002/app.42528>
14. Pereira, P.H.F., Rosa, M. de F., Cioffi, M.O.H., Benini, K.C.C. de C., Milanese, A.C., Voorwald, H.J.C., Mulinari, D.R.: Vegetal fibers in polymeric composites: a review. *Polímeros.* 25, 9–22 (2015). <https://doi.org/10.1590/0104-1428.1722>
15. Loureiro, N.C., Esteves, J.L., Viana, J.C., Ghosh, S.: Development of polyhydroxyalkanoates/poly(lactic acid) composites reinforced with cellulosic fibers. *Compos. Part B Eng.* 60, 603–611 (2014). <https://doi.org/10.1016/j.compositesb.2014.01.001>
16. Georgiopoulos, P., Christopoulos, A., Koutsoumpis, S., Kontou, E.: The effect of surface treatment on the performance of flax/biodegradable composites. *Compos. Part B Eng.* 106, 88–98 (2016). <https://doi.org/10.1016/j.compositesb.2016.09.027>
17. Satyanarayana, K.G., Arizaga, G.G.C., Wypych, F.: Biodegradable composites based on lignocellulosic fibers—An overview. *Prog. Polym. Sci.* 34, 982–1021 (2009). <https://doi.org/10.1016/j.progpolymsci.2008.12.002>
18. Basalp, D., Tihminlioglu, F., Sofuoglu, S.C., Inal, F., Sofuoglu, A.: Utilization of Municipal Plastic and Wood Waste in Industrial Manufacturing of Wood Plastic Composites. *Waste and Biomass Valorization.* 1–12 (2020). <https://doi.org/10.1007/s12649-020-00986-7>
19. Hejna, A., Sulyman, M., Przybysz, M., Saeb, M.R., Klein, M., Formela, K.: On the Correlation of Lignocellulosic Filler Composition with the Performance Properties of Poly( $\epsilon$ -Caprolactone) Based Biocomposites. *Waste and Biomass Valorization.* 11, 1467–1479 (2018). <https://doi.org/10.1007/s12649-018-0485-5>
20. Picard, M.C., Rodriguez-Uribe, A., Thimmanagari, M., Misra, M., Mohanty, A.K.: Sustainable Biocomposites from Poly(butylene succinate) and Apple Pomace: A Study on Compatibilization Performance. *Waste and Biomass Valorization.* 1–13 (2019). <https://doi.org/10.1007/s12649-019-00591-3>
21. Sánchez-Safont, E.L., Aldureid, A., Lagarón, J.M., Gámez-Pérez, J., Cabedo, L.: Biocomposites of different lignocellulosic wastes for sustainable food packaging applications. *Compos. Part B Eng.* 145, 215–225 (2018). <https://doi.org/10.1016/j.compositesb.2018.03.037>
22. Kumar, R., Hu, F., Hubbell, C.A., Ragauskas, A.J., Wyman, C.E.: Comparison of laboratory delignification methods, their selectivity, and impacts on physiochemical characteristics of cellulosic biomass. *Bioresour. Technol.* 130, 372–381 (2013). <https://doi.org/10.1016/j.biortech.2012.12.028>
23. Zhao, X., van der Heide, E., Zhang, T., Liu, D.: Delignification of sugarcane bagasse with alkali and peracetic acid and characterization of the pulp. *BioResources.* 5, 1565–1580 (2010)

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65
24. Mariano, M., Cercená, R., Soldi, V.: Thermal characterization of cellulose nanocrystals isolated from sisal fibers using acid hydrolysis. *Ind. Crops Prod.* 94, 454–462 (2016). <https://doi.org/10.1016/j.indcrop.2016.09.011>
25. Segal, L., Creely, J.J., Martin, A.E., Conrad, C.M.: An Empirical Method for Estimating the Degree of Crystallinity of Native Cellulose Using the X-Ray Diffractometer. *Text. Res. J.* 29, 786–794 (1959). <https://doi.org/10.1177/004051755902901003>
26. Brown, R.: Fracture Mechanics Testing Methods for Polymers Adhesives and composites. *Polym. Test.* 21, 363 (2002). [https://doi.org/10.1016/S0142-9418\(01\)00080-0](https://doi.org/10.1016/S0142-9418(01)00080-0)
27. Carli, L.N., Crespo, J.S., Mauler, R.S.: PHBV nanocomposites based on organomodified montmorillonite and halloysite: The effect of clay type on the morphology and thermal and mechanical properties. *Compos. Part A Appl. Sci. Manuf.* 42, 1601–1608 (2011). <https://doi.org/10.1016/j.compositesa.2011.07.007>
28. Arjmandi, R., Hassan, A., Majeed, K., Zakaria, Z.: Rice Husk Filled Polymer Composites. *Int. J. Polym. Sci.* 2015, (2015). <https://doi.org/10.1155/2015/501471>
29. Khiari, R., Mhenni, M.F., Belgacem, M.N., Mauret, E.: Valorisation of Vegetal Wastes as a Source of Cellulose and Cellulose Derivatives. *J. Polym. Environ.* 19, 80–89 (2011). <https://doi.org/10.1007/s10924-010-0207-y>
30. Paschoal, G.B., Muller, C.M.O., Carvalho, G.M., Tischer, C.A., Mali, S.: ISOLATION AND CHARACTERIZATION OF NANOFIBRILLATED CELLULOSE FROM OAT HULLS. *Quim. Nova.* 38, 478–482 (2015). <https://doi.org/10.5935/0100-4042.20150029>
31. Battegazzore, D., Bocchini, S., Alongi, J., Frache, A., Marino, F.: Cellulose extracted from rice husk as filler for poly(lactic acid): preparation and characterization. *Cellulose.* 21, 1813–1821 (2014). <https://doi.org/10.1007/s10570-014-0207-5>
32. Urruzola, I., Robles, E., Serrano, L., Labidi, J.: Nanopaper from almond (*Prunus dulcis*) shell. *Cellulose.* 21, 1619–1629 (2014). <https://doi.org/10.1007/s10570-014-0238-y>
33. Duan, L., Yu, W., Li, Z.: Analysis of structural changes in jute fibers after peracetic acid treatment. *J. Eng. Fiber. Fabr.* 12, 33–42 (2017). <https://doi.org/10.1177/155892501701200104>
34. Ndazi, B.S., Karlsson, S., Tesha, J. V., Nyahumwa, C.W.: Chemical and physical modifications of rice husks for use as composite panels. *Compos. Part A Appl. Sci. Manuf.* 38, 925–935 (2007). <https://doi.org/10.1016/j.compositesa.2006.07.004>
35. El Mechtali, F.Z., Essabir, H., Nekhlaoui, S., Bensalah, M.O., Jawaid, M., Bouhfid, R., Quaiss, A.: Mechanical and Thermal Properties of Polypropylene Reinforced with Almond Shells Particles: Impact of Chemical Treatments. *J. Bionic Eng.* 12, 483–494 (2015). [https://doi.org/10.1016/S1672-6529\(14\)60139-6](https://doi.org/10.1016/S1672-6529(14)60139-6)
36. Chanda, A.K., Hazra, A., Praveen Kumar, M., Neogi, S., Neogi, S.: Chemical treatments of rice husk filler and jute fiber for the use in green composites. *Fibers*

Polym. 16, 902–910 (2015). <https://doi.org/10.1007/s12221-015-0902-3>

37. Johar, N., Ahmad, I., Dufresne, A.: Extraction , preparation and characterization of cellulose fibres and nanocrystals from rice husk. *Ind. Crop. Prod.* 37, 93–99 (2012). <https://doi.org/10.1016/j.indcrop.2011.12.016>
38. Paulo, J., Oliveira, D., Pinheiro, G., Oliveira, K., Lisie, S., El, M., Silveira, G., Renato, A., Dias, G., Zavareze, R.: Cellulose fibers extracted from rice and oat husks and their application in hydrogel. *Food Chem.* 221, 153–160 (2017). <https://doi.org/10.1016/j.foodchem.2016.10.048>
39. Rosa, S.M.L., Rehman, N., Miranda, M.I.G. De, Nachtigall, S.M.B., Bica, C.I.D.: Chlorine-free extraction of cellulose from rice husk and whisker isolation. *Carbohydr. Polym.* 87, 1131–1138 (2012). <https://doi.org/10.1016/j.carbpol.2011.08.084>
40. Yang, H., Yan, R., Chen, H., Lee, D.H., Zheng, C.: Characteristics of hemicellulose, cellulose and lignin pyrolysis. *Fuel.* 86, 1781–1788 (2007). <https://doi.org/10.1016/j.fuel.2006.12.013>
41. Tobergte, D.R., Curtis, S.: *Biocomposites: Desing and Mechanical Performance.* Elsevier Inc. (2013)
42. Michael A. Gunning, Luke M. Geever , John A. Killion , John G. Lyonsa, C.L.H.: Mechanical and biodegradation performance of short natural fibre polyhydroxybutyrate composites. *Polym. Test.* 32, 1603–1611 (2013)
43. Torres-Tello, E. V., Robledo-Ortíz, J.R., González-García, Y., Pérez-Fonseca, A.A., Jasso-Gastinel, C.F., Mendizábal, E.: Effect of agave fiber content in the thermal and mechanical properties of green composites based on polyhydroxybutyrate or poly(hydroxybutyrate-co-hydroxyvalerate). *Ind. Crops Prod.* 99, 117–125 (2017). <https://doi.org/10.1016/j.indcrop.2017.01.035>
44. Fei, B., Chen, C., Chen, S., Peng, S., Zhuang, Y., An, Y., Dong, L.: Crosslinking of poly[(3-hydroxybutyrate)-co-(3-hydroxyvalerate)] using dicumyl peroxide as initiator. *Polym. Int.* 53, 937–943 (2004). <https://doi.org/10.1002/pi.1477>
45. Srubar, W. V., Wright, Z.C., Tsui, A., Michel, A.T., Billington, S.L., Frank, C.W.: Characterizing the effects of ambient aging on the mechanical and physical properties of two commercially available bacterial thermoplastics. In: *Polymer Degradation and Stability.* pp. 1922–1929 (2012)
46. Esposito, A., Delpouve, N., Causin, V., Dhotel, A., Delbreilh, L., Dargent, E.: From a Three-Phase Model to a Continuous Description of Molecular Mobility in Semicrystalline Poly(hydroxybutyrate-co-hydroxyvalerate). *Macromolecules.* 49, 4850–4861 (2016). <https://doi.org/10.1021/acs.macromol.6b00384>
47. Sánchez-Safont, E.L., González-Ausejo, J., Gámez-Pérez, J., Lagarón, J.M., Cabedo, L.: Poly(3-Hydroxybutyrate-co-3-Hydroxyvalerate)/Purified Cellulose Fiber Composites by Melt Blending: Characterization and Degradation in Composting Conditions. *J. Renew. Mater.* 4, 123–132 (2016). <https://doi.org/10.7569/JRM.2015.634127>
48. Turi, E.A.: *Thermal characterization of polymeric materials.* Academic Press

(1981)

- 1  
2 49. Saba, N., Jawaid, M., Alothman, O.Y., Paridah, M.T.: A review on dynamic  
3 mechanical properties of natural fibre reinforced polymer composites. *Constr.*  
4 *Build. Mater.* 106, 149–159 (2016).  
5 <https://doi.org/10.1016/j.conbuildmat.2015.12.075>  
6
- 7  
8 50. Wong, S., Shanks, R., Hodzic, A.: Interfacial improvements in poly(3-  
9 hydroxybutyrate)-flax fibre composites with hydrogen bonding additives.  
10 *Compos. Sci. Technol.* 64, 1321–1330 (2004).  
11 <https://doi.org/10.1016/j.compscitech.2003.10.012>  
12
- 13  
14 51. Wu, C.S.: Preparation and Characterization of Polyhydroxyalkanoate Bioplastic-  
15 Based Green Renewable Composites from Rice Husk. *J. Polym. Environ.* 22, 384–  
16 392 (2014). <https://doi.org/10.1007/s10924-014-0662-y>  
17
- 18  
19 52. Grassie, N., Murray, E.J., Holmes, P.A.: The thermal degradation of poly(-(d)- $\beta$ -  
20 hydroxybutyric acid): Part 2-Changes in molecular weight. *Polym. Degrad. Stab.*  
21 6, 95–103 (1984). [https://doi.org/10.1016/0141-3910\(84\)90075-2](https://doi.org/10.1016/0141-3910(84)90075-2)  
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