

COMMUNICATIONS earth & environment

ARTICLE

<https://doi.org/10.1038/s43247-021-00192-w>

OPEN

Carbon dioxide fluxes increase from day to night across European streams

Katrin Attermeyer^{1,2,3}[✉], Joan Pere Casas-Ruiz^{4,5}, Thomas Fuss⁶, Ada Pastor^{4,5,34}, Sophie Cauvy-Fraunié⁷, Danny Sheath^{8,35}, Anna C. Nydahl¹, Alberto Doretto^{9,10}, Ana Paula Portela^{11,12}, Brian C. Doyle¹³, Nikolay Simov¹⁴, Catherine Gutmann Roberts⁸, Georg H. Niedrist¹⁵, Xisca Timoner^{4,5}, Vesela Evtimova¹⁶, Laura Barral-Fraga⁵, Tea Bašić^{8,36}, Joachim Audet^{17,37}, Anne Deininger^{18,38}, Georgina Busst⁸, Stefano Fenoglio^{10,19}, Núria Catalán^{4,5,39,40}, Elvira de Eyto²⁰, Francesca Pilotto^{18,41}, Jordi-René Mor^{4,21}, Juliana Monteiro²², David Fletcher⁸, Christian Noss²³, Miriam Colls^{4,5}, Magdalena Nagler²⁴, Liu Liu^{23,25}, Clara Romero González-Quijano²⁶, Ferran Romero^{4,5}, Nina Pansch²⁵, José L. J. Ledesma^{17,27,28}, Josephine Pegg^{8,29}, Marcus Klaus^{18,42}, Anna Freixa^{4,5}, Sonia Herrero Ortega²⁵, Clara Mendoza-Lera^{7,23}, Adam Bednařík^{30,43}, Jérémy A. Fonvielle²⁵, Peter J. Gilbert³¹, Lyubomir A. Kenderov³², Martin Rulík³⁰ & Pascal Bodmer^{23,33,44}

Globally, inland waters emit over 2 Pg of carbon per year as carbon dioxide, of which the majority originates from streams and rivers. Despite the global significance of fluvial carbon dioxide emissions, little is known about their diel dynamics. Here we present a large-scale assessment of day- and night-time carbon dioxide fluxes at the water-air interface across 34 European streams. We directly measured fluxes four times between October 2016 and July 2017 using drifting chambers. Median fluxes are 1.4 and 2.1 mmol m⁻² h⁻¹ at midday and midnight, respectively, with night fluxes exceeding those during the day by 39%. We attribute diel carbon dioxide flux variability mainly to changes in the water partial pressure of carbon dioxide. However, no consistent drivers could be identified across sites. Our findings highlight widespread day-night changes in fluvial carbon dioxide fluxes and suggest that the time of day greatly influences measured carbon dioxide fluxes across European streams.

A list of author affiliations appears at the end of the paper.

Land waters are important sources of atmospheric carbon dioxide (CO₂) partially offsetting the terrestrial carbon sink^{1,2}. Streams and rivers therein represent major CO₂ emitters³. Fluvial CO₂ fluxes are primarily controlled by the gas exchange velocity at the water–air interface (k) and the gradient between the water and atmospheric partial pressures of CO₂ ($p\text{CO}_2$)⁴. Both parameters are highly variable in space and time^{5,6}, causing uncertainty in the magnitude of regional and global fluvial CO₂ emissions².

The high spatiotemporal variability of k and water $p\text{CO}_2$ can be attributed to a complex interplay of underlying controls. While k in streams is mostly driven by water turbulence created by variations in flow and stream morphology⁷, the water $p\text{CO}_2$ is influenced by the degree of hydrological connectivity between the stream and the adjacent riparian soils⁸ as well as by in-stream processes (e.g., stream metabolism). The supply of CO₂ from external sources, such as soil water or groundwater, into streams, varies with reach and season^{5,9}. Furthermore, seasonal and diel changes in stream $p\text{CO}_2$ are attributed to stream metabolism driven by temperature and solar radiation^{10–13}. Ecosystem respiration, a source of CO₂ in the stream, takes place throughout the whole day, and gross primary production, a sink of CO₂, occurs only during daylight. Temperature and solar radiation also directly influence water $p\text{CO}_2$, the former by changing the solubility of the gas and the latter due to photomineralization¹⁴. However, questions remain regarding the magnitude and relative drivers of seasonal and diel fluctuations of CO₂ fluxes in streams.

Presently, most fluvial CO₂ emission values are derived from k estimates based on water velocity and stream channel slope and on water $p\text{CO}_2$ values indirectly calculated from alkalinity, pH, and temperature³. This approach fails to capture the high spatiotemporal variability observed for k and $p\text{CO}_2$ and therefore can provide imprecise estimates of CO₂ fluxes^{15,16}. Direct field observations provide the means to improve estimates and understanding of the drivers behind spatiotemporal variability, and thus the dynamics of CO₂ outgassing from running waters. However, besides mostly local studies that indirectly infer CO₂ fluxes from $p\text{CO}_2$ and k ^{11,12,17,18}, no direct measurements exist that compare day-time and night-time CO₂ fluxes from streams on a larger spatial scale.

The aim of this study was to assess the magnitude and drivers of stream CO₂ flux variations between day and night across European streams. We hypothesized that CO₂ fluxes would differ between day and night due to diel variations in terrestrial inorganic carbon inputs, in situ metabolism, and temperature. As higher temperatures and solar radiation may drive differences in $p\text{CO}_2$, we expected a higher difference between day-time and night-time fluxes with warmer temperatures and at lower latitudes. Hence, we measured day-time and night-time fluxes of CO₂ at four different periods throughout one year from 34 streams (Strahler stream orders from 1 to 6) in 11 countries across Europe following a standardized procedure. CO₂ fluxes were measured starting at midday (11 a.m. Greenwich Mean Time (GMT)) and midnight (11 p.m. GMT) with drifting flux chambers equipped with CO₂ sensors as described in Bastviken et al.¹⁹. In the majority of the European streams, we found increased CO₂ fluxes at the water–air interface in the night compared to the day with a median increase of 0.5 mmol m⁻² h⁻¹. Most of the observed CO₂ flux variability was explained by changes in $p\text{CO}_2$ from day to night with more pronounced changes at lower latitudes.

Results and discussion

Magnitude of CO₂ flux variation from day to night. Midday CO₂ fluxes at the water–air interface ranged from –2.7 (uptake)

to 19.9 mmol m⁻² h⁻¹ (emission) (1.4 [0.5, 3.1]; median [interquartile range (IQR)]; $n = 107$) and midnight fluxes ranged from –0.3 to 25.6 mmol m⁻² h⁻¹ (2.1 [0.9, 3.7]; $n = 107$) (Fig. 1a; Supplementary Table S3). Our measured fluxes are comparable to other studies conducted in temperate and boreal streams that used chambers^{20,21} or empirical models^{12,22,23}, although they were in the lower range of the numbers modeled in a study in the USA²³ (Supplementary Fig. S2). The lower numbers might be due to the lack of tributary inflows, large woody debris, and strong hydraulic jumps in the selected stream sections (Supplementary Sampling manual).

To assess stream CO₂ flux variations between day and night, we computed the difference of night-time minus day-time fluxes for each stream and sampling period, where positive numbers indicate an increase from day to night and vice versa (Fig. 1b). Differences in CO₂ fluxes amounted to 0.5 mmol m⁻² h⁻¹ [0.1, 1.4] ($n = 107$) across all sites and sampling periods, which is equivalent to a relative increase of 39% [4%, 100%] ($n = 101$; n reduced due to exclusion of relative comparisons to zero flux at day-time) (Fig. 2). Altogether, these results point towards a high relevance of night-time CO₂ fluxes as reported earlier for single pre-alpine streams¹², stream networks^{13,17} or rivers¹⁸, and in a recent compilation of diel CO₂ data from 66 streams worldwide²⁴. A rough annual extrapolation of fluxes from our study sites (Supplementary Methods) shows that the inclusion of night-time fluxes increases annual estimates of site-specific stream CO₂ emissions by 16% [6%; 25%] (Supplementary Table S4). Hence, our measurements and the simplified extrapolation of our data emphasize the need to collect and integrate night-time CO₂ flux data into sampling protocols as well as regional upscaling efforts.

Looking into the individual comparisons, we found 83 increases in median CO₂ fluxes from day to night with seven comparisons where the stream even switched from a sink to a source of CO₂ to the atmosphere (Supplementary Table S3). However, we also found four comparisons where median CO₂ fluxes at day and night were the same and 20 decreases in the night (Supplementary Table S3). These results and also other studies^{13,25,26} suggest that the direction and strength of diel $p\text{CO}_2$ pattern can be largely variable across space and time.

Diel CO₂ flux differences vary as a function of latitude and water temperature.

The diel differences in CO₂ fluxes were significantly negatively related to latitude (Table 1A), with substantial diel variation more likely at lower latitudes. Likewise, the interaction with latitude and the water temperature was significant (Table 1A), which might be explained by higher temperatures at lower latitudes during the sampling periods and higher solar radiation boosting in-stream primary production²⁷. This dataset is derived from only 34 streams distributed across different climate zones in Europe. However, to our knowledge, it is currently the largest study of its kind, using flux chambers to measure CO₂ fluxes, and compare those fluxes at day-time and night-time on such a spatial scale.

We found no significant differences in the magnitude of diel differences in CO₂ fluxes related to water temperature (Table 1A) using a linear mixed-effect model (LME). However, comparing the CO₂ fluxes at midday to midnight at the different sampling periods, we detected significant diel changes in CO₂ fluxes in October, January, and April (Fig. 1a). Contrary to our expectation that higher differences can be expected at higher temperatures, we did not detect significant changes from day to night in July (Fig. 1a), during which period the lowest changes in absolute numbers were recorded (0.3 mmol m⁻² h⁻¹; Fig. 1b). The highest differences of CO₂ fluxes from day to night were measured during April (1.1 mmol m⁻² h⁻¹), followed by January (0.5 mmol m⁻² h⁻¹)

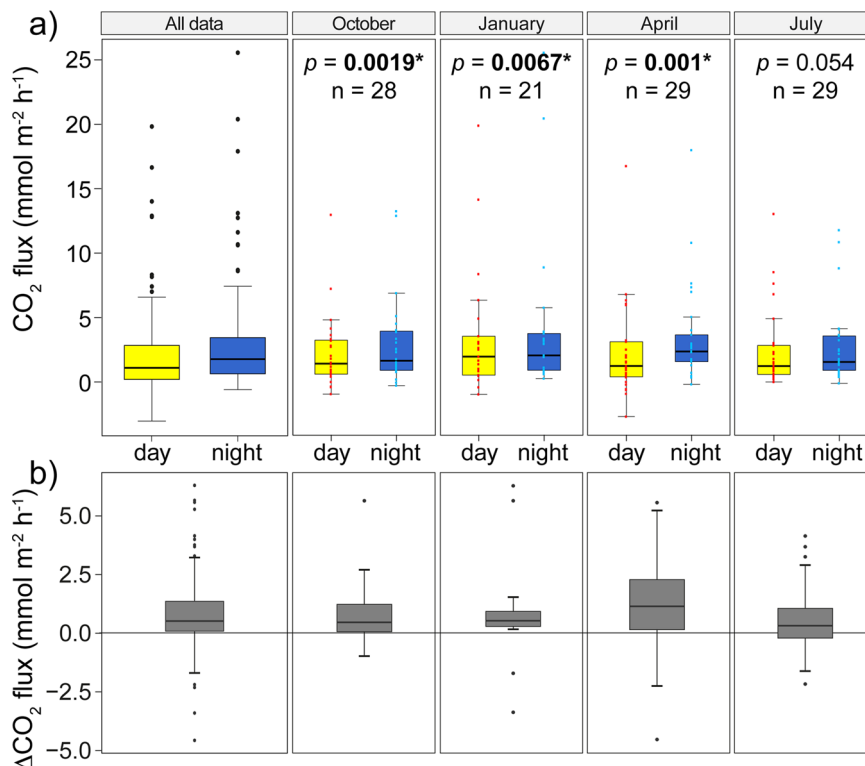


Fig. 1 Day-to-night changes of CO₂ fluxes at the water-air interface of the sampled European streams. Stream CO₂ fluxes (in mmol CO₂ m⁻² h⁻¹) at day-time (yellow) and night-time (blue) (**a**) and the calculated changes from night minus day (Δ CO₂ flux) (**b**) for all data and separately for each sampling period. In the sampling periods comparisons in (**a**), CO₂ fluxes for individual stream sites are indicated by red (day) and light blue (night) dots. The boxplots visualize the median of all stream sites (line), the first and third quartiles (hinges), the 1.5*inter-quartile ranges (whiskers), and the outliers outside the range of 1.5*inter-quartile ranges (black dots). On top of (**a**) are *p* values retrieved from paired comparisons of median CO₂ fluxes tested by Wilcoxon signed-rank tests and the sample size (*n*). Significant *p* values with *p* < 0.05 are in bold with an asterisk. The differences in the CO₂ fluxes (**b**) in mmol CO₂ m⁻² h⁻¹ from day to night are for October: 0.5 [0.1, 1.2]; January: 0.5 [0.3, 0.9]; April: 1.1 [0.1, 2.3]; July: 0.3 [-0.2, 1.1] (median [IQR]).

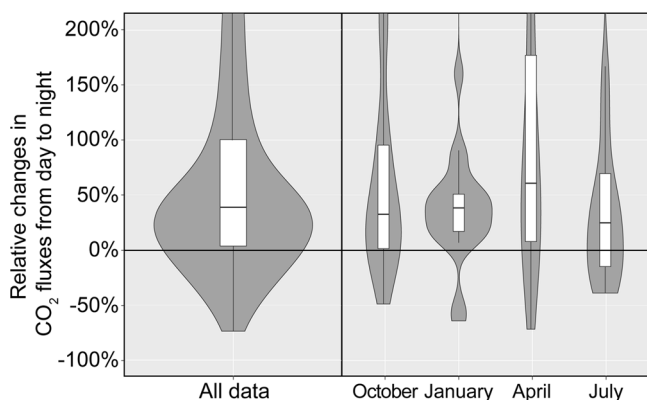


Fig. 2 Relative changes in CO₂ fluxes from day to night for all data together and for each sampling period. A positive value indicates an increase in CO₂ fluxes during the night and vice versa (expressed as a %-change of the daytime values). Outliers (>1.5*IQR) were excluded for illustration purposes as the large relative variation in these fluxes was due to minor absolute variation in fluxes close to zero. The median relative changes were positive throughout all sampling periods, ranging from 32% [0.6%, 95%] in October, 38% [16%, 50%] in January, 60% [7%, 177%] in April, to 24% [-16%, 69%] in July (median [IQR]; *n* = 26, 21, 28, and 26, respectively).

and October (0.5 mmol m⁻² h⁻¹). Lower day-night changes in July could be explained by increased riparian shading reducing photosynthesis^{28,29}. For example, reduced in-stream photosynthesis

in summer compared to spring has been shown for a subalpine stream network²⁹ or a temperate forested headwater stream²⁸. However, comparing the canopy cover of the streams and the differences in CO₂ fluxes from day to night (Supplementary Fig. S3h) revealed no clear pattern. A probable alternate explanation is that CO₂ production via photomineralization during the day counteracted a decrease via CO₂ fixation by photosynthesis³⁰ and diminished diel *p*CO₂ and ultimately CO₂ flux changes. This highlights the complex interplay between different light-dependent processes in streams influencing *p*CO₂ on a diel scale.

The importance of year-round measurements is highlighted by the January data set containing the second-highest diel CO₂ flux changes. European ice-free streams may be perceived “dormant” during these periods and representative CO₂ flux estimates are thus often missing³. Our January data showed a magnitude of flux compared to the rest of the year across the European streams as well as high diel variability in CO₂ fluxes (Fig. 1). This may be attributed in part to the latitudinal coverage of our study as we included streams from the boreal to the Mediterranean. For example, the water temperatures of the Spanish streams were still relatively high in winter with around 2.8–9.5 °C during the day whereas Swedish streams showed these temperatures in October and April. A study in the coterminous US looking into stream *p*CO₂ variability also reports varying strengths of diel *p*CO₂ variability, dependent on the investigated stream and time²⁵. Hence, diel *p*CO₂ and CO₂ flux variability can be large in streams of the northern hemisphere, stressing the need to unravel the site-specific drivers of and mechanisms behind these diel changes.

Table 1 Results of the linear mixed-effect models (LME).

Response variable	Fixed effect	χ^2 (1)	<i>p</i>	Sign
(A) Testing spatial and temporal hypotheses				
CO ₂ flux difference	Latitude	7.4207	0.006	–
from day to night	Water temperature (day)	0.0168	0.897	
	Water temperature (day)*latitude	4.9594	0.026	+
(B) Testing physical and biogeochemical drivers of CO₂ flux changes				
CO ₂ flux difference	Δ Water <i>p</i> CO ₂	4.9497	0.026	+
from day to night	Δ Gas transfer velocity <i>k</i>	0.5613	0.454	
(C) Testing biogeochemical drivers of <i>p</i>CO₂ changes				
<i>p</i> CO ₂ difference	Δ Water O ₂ concentration	7.9879	0.005	–
from day to night	Δ pH	0.0345	0.853	
	Δ Conductivity	0.0293	0.864	
	$\Delta T_w - T_a$ ^a (proxy for heat flux)	1.6720	0.196	
	Δ Water temperature	0.8731	0.350	
(D) Testing catchment and hydromorphological drivers of <i>p</i>CO₂ changes				
<i>p</i> CO ₂ difference	Day length	1.7244	0.189	
from day to night	Stream wetted width	0.3748	0.540	
	Discharge	3.4458	0.063	
	% forest	0.0950	0.758	
	Catchment area	2.3656	0.124	

^aHeat flux calculated as water temperature (T_w) minus air temperature (T_a).

(A) Marginal $R^2 = 0.12$, conditional $R^2 = 0.18$, sample size = 107.

(B) Marginal $R^2 = 0.08$, conditional $R^2 = 0.10$, sample size = 77.

(C) Marginal $R^2 = 0.13$, conditional $R^2 = 0.33$, sample size = 78.

(D) Marginal $R^2 = 0.11$, conditional $R^2 = 0.13$, sample size = 68.

The effects of latitude and water temperature during the day (A) and the effect of day-to-night differences of *p*CO₂ and the gas transfer velocity (Δ = night minus day values) (B) on the day-to-night difference of CO₂ fluxes were tested. Furthermore, the effect of day-to-night differences of physical and biogeochemical parameters (C) and the effect of catchment and hydromorphological related parameters (D) on the day-to-night differences of *p*CO₂ were evaluated. Stream ID was included as a random effect on the intercept. Significances of fixed effects were assessed with likelihood ratio tests with degrees of freedom = 1. The slope direction (sign) of the effect is indicated with – or + when significant. Significant *p* values < 0.05 are in bold.

Diel CO₂ flux variability driven by changes in water *p*CO₂. To understand the mechanisms behind the observed changes in CO₂ fluxes from day to night, we first selected the two primary controls of CO₂ fluxes at the water–air interface, i.e., the gas exchange velocity and water *p*CO₂ and explored the influence of these parameters on absolute CO₂ flux changes using an LME. The diel CO₂ flux variability in European streams could be mostly attributed to changes in water *p*CO₂ (Table 1B), whereas changes in the gas exchange velocity *k* appeared less important. In fact, we did not measure significant variations in *k* from day to night in our streams (Fig. 3; Supplementary Fig. S4h). Although diel variabilities of gas exchange velocities have been reported for CO₂ and other gases^{31,32}, the majority of the investigated streams in this study did not show those changes. The *p*CO₂ as a major driver of diel CO₂ flux variability was also identified by a global compilation of high-frequency CO₂ measurements²⁴. Consequently, if no major changes in physical drivers of gas exchanges occur that strongly affect the turbulence, such as heavy rain events, it is sufficient to focus on *p*CO₂ for assessing diel flux changes at the water–air interface.

In a second step, we tested the influence of biogeochemical parameters that vary on a diel scale on water *p*CO₂ day-to-night differences (Table 1C). This LME identified a link between the day-to-night changes in water *p*CO₂ and water dissolved O₂, with *p*CO₂ generally increasing and O₂ decreasing from day to night (Supplementary Fig. S4b, c). This potentially reflects a diel cycle

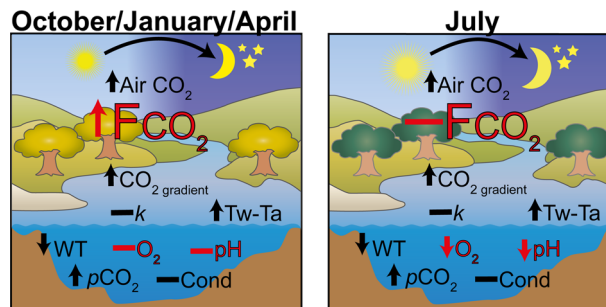


Fig. 3 Diel changes in CO₂ fluxes (FCO₂) and other physical and chemical parameters for October/January/April and July, respectively. The physical and chemical parameters comprise atmospheric CO₂ (Air CO₂), the differences of CO₂ concentrations in the water minus the air (CO₂ gradient), the water–air gas transfer velocity (*k*), the differences of temperatures in the water minus the air ($T_w - T_a$), the water temperature (WT), the oxygen concentration in the water (O₂), pH in the water, the partial pressure of CO₂ in the water (*p*CO₂), and conductivity (Cond). The arrows indicate significant increases (↑) or significant decreases (↓) from day to night and the line indicates no significant change (—) tested by a Wilcoxon signed-rank test (see Supplementary Fig. S4 for more information). The differences between the sampling periods October/January/April and July, respectively, detected in this European study are highlighted in red.

of CO₂ controlled by aquatic primary production and respiration (in-stream metabolism). Hence, even though in situ metabolism may play a minor role in determining the baseline *p*CO₂ and flux in smaller streams (mostly controlled by terrestrial inputs²³), our results suggest that metabolism can be an important driver of the diel fluctuations in CO₂ fluxes. Indeed, increased water *p*CO₂ during the night has been attributed to a decrease in CO₂ fixation by primary producers^{13,18,24}, although a recent study suggests that the adjacent groundwater can also show measurable but less pronounced diel *p*CO₂ variations³³. Previous research suggests that in situ mineralization of CO₂ should play a larger role in CO₂ dynamics in larger streams because they are less influenced by external CO₂ sources²³. Nevertheless, we did not find any trend in CO₂ flux day-to-night differences with stream width or discharge as a proxy for size (Supplementary Fig. S3c, f) or with stream order (Supplementary Fig. S5) although other studies suggest change over a size gradient^{23,34}. Furthermore, the LME testing hydromorphological and catchment variables on *p*CO₂ day-to-night differences (Table 1D) did not reveal significant relationships with either of these drivers. This could either be due to the fact that we missed the best proxy that determines day-to-night differences in *p*CO₂ in European streams or that there are no common drivers among the investigated streams. Large diel variability of CO₂ patterns within one Swedish stream²⁶ or among US headwater streams²⁵ have been described, which complicates the identification of general drivers. Hence, further research is needed to decipher the diel variability of the sources and dynamics of *p*CO₂ in streams and to understand the environmental, hydromorphological, and catchment drivers before their importance on a regional or global scale can be assessed.

In-stream metabolism with photosynthetic CO₂ fixation diminishing *p*CO₂ during the day may explain the increase in CO₂ fluxes from day to night, but cannot explain why in some instances we measured a lower CO₂ flux at night. Potential explanations for a lower night flux might include: (i) higher atmospheric CO₂ concentrations due to the absence of terrestrial CO₂ fixation during night and therefore a lower water–atmosphere *p*CO₂ gradient, (ii) photomineralization of

organic matter to CO₂ counteracting the CO₂ fixation by primary producers during day-time, and (iii) lower turbulence due to a decrease in stream discharge in the night. We found significant increases in atmospheric CO₂ close to the investigated streams at night. However, this was usually accompanied by concomitant increases in water *p*CO₂ and therefore did not translate into smaller CO₂ gradients between the water–air interface (Fig. 3; Supplementary Fig. S4b, e, i). Production of CO₂ due to photomineralization of dissolved organic carbon (DOC) could play a role in diel CO₂ dynamics in streams with high amounts of colored terrestrial organic matter³⁵. In the highly colored streams, diel CO₂ patterns can additionally be influenced by DOC shading diminishing benthic primary production³⁶. In October, we measured DOC concentrations in a subset of the investigated streams for another study³⁷ where an agricultural stream in Sweden and peatland-dominated streams in Great Britain had high DOC concentrations (>10 mg L⁻¹) whereas the median DOC was much lower with 2.6 mg L⁻¹³⁷. Due to the limited data, we could not test the effect of DOC on *p*CO₂ changes and we can neither confirm nor exclude that photomineralization might play a role for diel *p*CO₂ and consequently CO₂ flux variability in the studied streams. We did find, nonetheless, that the majority of the streams where CO₂ fluxes were lower during the night also had a lower gas transfer velocity (*k*₆₀₀), likely due to a slight decrease in stream discharge and therefore turbulence. Thus, while there was a general tendency of increased *p*CO₂ from day to night (only 4 out of 20 decreases in CO₂ fluxes from day to night showed a concomitant decrease in water *p*CO₂), individual streams at single time points seemed to experience diel fluctuations in discharge as described elsewhere³⁸. This can simultaneously reduce the gas exchange velocity of the stream and therefore cause lower night-time CO₂ fluxes. In this study, we only measured stream discharge during the day, and therefore the importance of this mechanism remains to be confirmed.

Maximum CO₂ flux differences might be even higher—limitations of the study design. For organizational reasons, the sampling scheme of this collaborative study was standardized to fixed times of measurements for the day and the night. All teams across Europe started their measurements at 11:00 (midday) and 23:00 GMT (midnight) during each sampling period, which has consequences for the magnitude of the observed diel variability of the CO₂ fluxes. The largest diel differences in stream *p*CO₂ have generally been detected at the end of the day compared to the end of the night^{12,18,39}. In an agricultural Swedish stream, diel maximum and minimum CO₂ concentrations were reached at 04:00 and 16:00 (GMT), respectively, during spring and early summer periods (late April to early July) where diel dynamics were most pronounced²⁶. In these scenarios, sampling midday and midnight, as conducted in this study, would be close to those maxima and minima as they can be reached already earlier during the day (see Supplementary Fig. S6 in May). However, the maxima and minima of diel CO₂ dynamics in streams can vary largely (see Supplementary Fig. S6 in October, April, July). In another example of German streams³⁹, the times of minima and maxima differ between streams and times, and the fixed time points chosen in this study would miss the maximum differences that can be observed (see Supplementary Fig. S7 in August). Hence, our estimates could be conservative as we compared fixed time points at midday and midnight. In general, CO₂ flux measurements in streams are highly sensitive towards the time of the day because diel minimum and maximum of *p*CO₂ can vary largely from month to month but also from day to day. As we found that the diel variability of *p*CO₂ was the major driver of diel CO₂ fluxes, we recommend future studies that plan to measure CO₂

fluxes directly with the chamber method, to additionally monitor the diel variability of *p*CO₂ with loggers at a high temporal resolution. This approach will provide the opportunity to estimate if the measurements are done during peak times or not.

While our results provide a first insight into the drivers of day-night differences in CO₂ fluxes, the high uncertainty in the models as well as the sometimes opposing patterns—increases and decreases from day to night in different streams and sampling periods—point towards different drivers varying on a temporal and spatial scale. We recommend that future study designs incorporate high-frequency CO₂ data together with biogeochemical variables from the stream (e.g., O₂) and the atmosphere (e.g., CO₂ or temperature)⁴⁰. Additionally, we recommend including radioactive or stable carbon isotope signatures to track potential sources of CO₂ and their changes in streams^{41,42} to better assess terrestrial–aquatic linkages. Linking temporal patterns of fluvial CO₂ fluxes with their drivers across large spatial scales is a path towards a more accurate understanding of their role in regional and global carbon cycles. Our results demonstrate that, in many streams across Europe, night-time CO₂ fluxes exceed day-time, resulting in a potential underestimation of global CO₂ emissions from inland waters if not considered. It is thus critical to account for the diel variability of fluvial CO₂ fluxes for accurate daily and annual estimates of CO₂ emissions from inland waters.

Methods

Sampling scheme. The project included 16 teams distributed across 11 European countries. Every team sampled one to three streams (Supplementary Table S1) every 3 months (October 2016/January 2017/April 2017/July 2017) within a time frame of 2 weeks throughout a whole year. These sampling periods roughly cover the seasons autumn/winter/spring/summer although, due to the large latitudinal coverage of the sampling sites, the seasons and their characteristics vary largely. In total, 34 stream sites (Supplementary Fig. S1) were visited each sampling period during the specified 2 weeks' time frame except for 11 streams in January that were frozen during the sampling weeks (Supplementary Table S3).

CO₂ fluxes were measured once every sampling period with drifting flux chambers equipped with CO₂ sensors. This method has proven to be a reliable and least biased direct measurement of CO₂ fluxes at the water–air interface in streams^{19,43}. CO₂ concentrations in the chamber headspace were logged every 30 s over a period of 5–10 min during each run, and CO₂ fluxes were calculated based on the rate of change over time in *p*CO₂ in the chamber headspace. At each stream, we measured CO₂ fluxes with the flux chamber (five times), *p*CO₂ in the atmosphere and water with the CO₂ sensors in the flux chamber (details described in Supplementary Methods), pH, temperature, conductivity, and oxygen in the water with a multiprobe (Supplementary Table S2). These measurements were started at 11:00 and 23:00 (GMT) and lasted approximately two hours and are referred to as midday and midnight throughout this article. Stream width, depth, canopy cover, and discharge were determined during the day (see Supplementary Sampling manual for details). In addition, the following information was collected for each stream once during the study: stream order, climate zone, catchment area until the endpoint of the investigated stream site and the percentage of coverage of different land use classes in this catchment area, and predominant geology (Supplementary Table S1).

Calculations of CO₂ fluxes and gas transfer velocity. Flux rates were obtained from the linear slopes of the *p*CO₂ in the chamber headspace over time and flux was accepted if the coefficient of determination (*R*²) of the slope was at least 0.65⁴⁴. An exception was made in cases where the slope was close to zero and the *p*CO₂ in the atmosphere and water (measured at the same time) were at equilibrium. These fluxes were set to zero. Final flux rates *F* (mmol CO₂ m⁻² h⁻¹) were calculated according to Eq. (1)⁴⁵:

$$F = S * 10^{-3} \frac{PV}{RTA} * 60 * 60, \quad (1)$$

where *S* is the slope (ppm s⁻¹), *P* is the *p*CO₂ in the atmosphere (atm), *V* is the volume (mL) of the drifting chamber, *R* is the gas constant (82.0562 mL atm K⁻¹ mol⁻¹), *T* is the chamber air temperature (K), *A* is the bottom area of the chamber (m²), and the last term is the conversion from seconds to hours. In this study, we followed the sign convention whereby positive values indicate a CO₂ flux from the stream to the atmosphere (source) and negative values indicate a flux from the atmosphere to the stream (sink). The magnitudes of variations between day-time and night-time measurements are additionally stated as percent increases, which

were computed by dividing the difference between the values at night minus day by the value at day and expressing the result as a percent change from day to night.

We used F (Eq. (1)) to calculate the gas transfer velocity (k in cm h^{-1}) by inverting the equation for Fick's law of gas diffusion, according to Eq. (2):

$$k = \frac{F}{kH(\text{CO}_{2,\text{water}} - \text{CO}_{2,\text{air}})} * 100, \quad (2)$$

where kH is Henry's constant (in $\text{mol L}^{-1} \text{atm}^{-1}$) adjusted for temperature⁴⁶.

For comparison of transfer velocities between sites and sampling periods and with the literature, k (Eq. (2)) was standardized to k_{600} (Eq. (3)):

$$k_{600} = k \left(\frac{600}{Sc} \right)^{-0.5} \quad (3)$$

where k is the transfer velocity at in situ temperature (T), Sc is the Schmidt number for in situ temperature T , the Schmidt number for 20 °C in freshwater is 600, and representing a hydrodynamic rough water surface typical in streams the exponent of -0.5 was chosen⁴⁷.

Statistical analyses. All statistical analyses were performed with median values of three to five floating chamber runs per day and night, respectively, using the statistical programming language R⁴⁸ (version 3.5.1). Samplings that generated less than three values for either day or night due to an R^2 of the slope <0.65 ⁴⁴ were excluded from further analysis reducing the number from 136 to 107 day–night comparisons. For our statistical tests, the alpha level was set to $\alpha = 0.05$. Significant differences between day-time and night-time measurements for each sampling period across all streams were tested with Wilcoxon signed-rank tests⁴⁹ where median day-time and night-time values for each stream site were paired (Fig. 1a). The same tests were conducted for the other biogeochemical variables measured at midday and midnight (see Fig. 3; Supplementary Fig. S4).

With a first linear mixed-effect model (LME) we tested the latitudinal and water temperature effect on CO_2 flux differences from day to night. A second LME was built to evaluate the two major drivers of CO_2 flux differences from day to night: $p\text{CO}_2$ and gas exchange velocity (k). A third LME was subsequently used to determine the biochemical factors potentially influencing the differences of the night-time minus day-time $p\text{CO}_2$, which was identified as the only significant driver in the second LME. Finally, a fourth LME was built to evaluate the effect of catchment and hydromorphological parameters on the day-to-night differences of $p\text{CO}_2$. For these tests, we used the “lmer” function of the R-package “lme4”⁵⁰ with maximum-likelihood estimation. Fixed effects for the LME with biogeochemical parameters for $p\text{CO}_2$ differences from day to night included absolute differences from day to night of oxygen concentration in the water, pH, conductivity, temperature gradient of atmosphere and water, and water temperature. Fixed effects for the LME with catchment and hydromorphological parameters included day length (i.e., sun hours from sunrise to sunset), stream wetted width, discharge, % forest of the catchment, and catchment area. These variables are mostly remotely available for streams. For the LMEs we included stream ID as a random effect allowing different intercepts for each stream to account for pseudoreplication (one data point per sampling period per stream) and z -scaled all fixed effects with the “scale” function before running the models. Statistical significances of fixed effects were assessed with likelihood ratio tests using the function “drop1”⁵¹. The respective LMEs were followed by a model validation, checking the residuals for normal distribution and homogeneity of variances⁵². A separation of the dataset to check if drivers between increases from day to night and decreases from day to night differ did not reveal acceptable models in terms of model validation (i.e., residuals were not normally distributed). Although our dataset provided a large spatial coverage on day–night differences in CO_2 fluxes in European streams, it did not have the statistical power to test for significant drivers separately for increases and decreases.

Data availability

The data that support the findings of this study are openly available in figshare at <https://doi.org/10.6084/m9.figshare.12717188>.

Code availability

This manuscript includes no code.

Received: 13 November 2020; Accepted: 21 May 2021;

Published online: 10 June 2021

References

- Butman, D. E. et al. Aquatic carbon cycling in the conterminous United States and implications for terrestrial carbon accounting. *Proc. Natl. Acad. Sci. USA* **113**, 58–63 (2016).
- Drake, T. W., Raymond, P. A. & Spencer, R. G. M. Terrestrial carbon inputs to inland waters: a current synthesis of estimates and uncertainty. *Limnol. Oceanogr. Lett.* **3**, 132–142 (2018).
- Raymond, P. A. et al. Global carbon dioxide emissions from inland waters. *Nature* **503**, 355–359 (2013).
- MacIntyre, S., Wanninkhof, R. & Chanton, J. P. Trace gas exchange in freshwater and coastal marine systems: flux across the air water interface. In *Methods in Ecology: Biogenic Trace Gases: Measuring Emissions from Soil and Water* (eds Matson, P. & Harriss, R.) 52–97 (Blackwell Publishing, 1995).
- Duvert, C., Butman, D. E., Marx, A., Ribolzi, O. & Hutley, L. B. CO_2 evasion along streams driven by groundwater inputs and geomorphic controls. *Nat. Geosci.* **11**, 813–818 (2018).
- Rocher-Ros, G., Sponseller, R. A., Lidberg, W., Mörth, C. & Giesler, R. Landscape process domains drive patterns of CO_2 evasion from river networks. *Limnol. Oceanogr. Lett.* **4**, 87–95 (2019).
- Hall, R. O. & Ulseth, A. J. Gas exchange in streams and rivers. *WIREs Water* e1391 (2019). <https://doi.org/10.1002/wat2.1391>
- Hope, D., Palmer, S. M., Billet, M. F. & Dawson, J. J. C. Variations in dissolved CO_2 and CH_4 in a first-order stream and catchment: an investigation of soil–stream linkages. *Hydrol. Process.* **18**, 3255–3275 (2004).
- Horgby, Å., Gómez-Gener, L., Escoffier, N. & Battin, T. J. Dynamics and potential drivers of CO_2 concentration and evasion across temporal scales in high-alpine streams. *Environ. Res. Lett.* **14**, 124082 (2019).
- Guasch, H., Armengol, J., Martí, E. & Sabater, S. Diurnal variation in dissolved oxygen and carbon dioxide in two low-order streams. *Water Res.* **32**, 1067–1074 (1998).
- Lynch, J. K., Beatty, C. M., Seidel, M. P., Jungst, L. J. & DeGrandpre, M. D. Controls of riverine CO_2 over an annual cycle determined using direct, high temporal resolution $p\text{CO}_2$ measurements. *J. Geophys. Res.* **115**, G03016 (2010).
- Peter, H. et al. Scales and drivers of temporal $p\text{CO}_2$ dynamics in an Alpine stream. *J. Geophys. Res. Biogeosci.* **119**, 1078–1091 (2014).
- Rocher-Ros, G., Sponseller, R. A., Bergström, A.-K., Myrsten, M. & Giesler, R. Stream metabolism controls diel patterns and evasion of CO_2 in Arctic streams. *Glob. Chang. Biol.* **00**, 1–14 (2019).
- Koehler, B., Landelius, T., Weyhenmeyer, G. A., Machida, N. & Tranvik, L. J. Sunlight-induced carbon dioxide emissions from inland waters. *Glob. Biogeochem. Cycles* **28**, 696–711 (2014).
- Golub, M., Desai, A. R., McKinley, G. A., Remcul, C. K. & Stanley, E. H. Large uncertainty in estimating $p\text{CO}_2$ from carbonate equilibria in lakes. *J. Geophys. Res. Biogeosci.* **122**, 2909–2924 (2017).
- Raymond, P. A. et al. Scaling the gas transfer velocity and hydraulic geometry in streams and small rivers. *Limnol. Oceanogr. Fluids Environ.* **2**, 41–53 (2012).
- Scheller, J., Singer, G. A., Ulseth, A. J., Hengsberger, S. & Battin, T. J. CO_2 evasion from a steep, high gradient stream network: importance of seasonal and diurnal variation in aquatic $p\text{CO}_2$ and gas transfer. *Limnol. Oceanogr.* **61**, 1826–1838 (2016).
- Reiman, J. H. & Xu, Y. J. Diel variability of $p\text{CO}_2$ and CO_2 outgassing from the lower Mississippi River: implications for riverine CO_2 outgassing estimation. *Water* **11**, 43 (2019).
- Bastviken, D., Sundgren, I., Natchimuthu, S., Reyier, H. & Gålfalk, M. Technical Note: cost-efficient approaches to measure carbon dioxide (CO_2) fluxes and concentrations in terrestrial and aquatic environments using mini loggers. *Biogeosciences* **12**, 3849–3859 (2015).
- Looman, A., Maher, D. T., Pendall, E., Bass, A. & Santos, I. R. The carbon dioxide evasion cycle of an intermittent first-order stream: contrasting water–air and soil–air exchange. *Biogeochemistry* **132**, 87–102 (2017).
- Crawford, J. T. et al. CO_2 and CH_4 emission from streams: patterns, controls, and regional significance. *Glob. Biogeochem. Cycles* **28**, 197–210 (2014).
- Teodoru, C. R., Del Giorgio, P. A., Prairie, Y. T. & Camire, M. Patterns in $p\text{CO}_2$ in boreal streams and rivers of northern Quebec, Canada. *Glob. Biogeochem. Cycles* **23**, GB2012 (2009).
- Hotchkiss, E. R. et al. Sources of and processes controlling CO_2 emissions change with the size of streams and rivers. *Nat. Geosci.* **8**, 696–699 (2015).
- Gómez-Gener, L. et al. Global carbon dioxide efflux from rivers enhanced by high nocturnal emissions. *Nat. Geosci.* 1–6 (2021). <https://doi.org/10.1038/s41561-021-00722-3>
- Crawford, J. T., Stanley, E. H., Dornblaser, M. M. & Striegl, R. G. CO_2 time series patterns in contrasting headwater streams of North America. *Aquat. Sci.* **79**, 473–486 (2016).
- Wallin, M. B., Audet, J., Peacock, M., Sahlée, E. & Winterdahl, M. Carbon dioxide dynamics in an agricultural headwater stream driven by hydrology and primary production. *Biogeosciences* **17**, 2487–2498 (2020).
- Demars, B. O. L. et al. Impact of warming on CO_2 emissions from streams countered by aquatic photosynthesis. *Nat. Geosci.* **9**, 758–761 (2016).

28. Roberts, B. J., Mulholland, P. J. & Hill, W. R. Multiple scales of temporal variability in ecosystem metabolism rates: results from 2 years of continuous monitoring in a forested headwater stream. *Ecosystems* **10**, 588–606 (2007).
29. Ulseth, A. J., Bertuzzo, E., Singer, G. A., Schelker, J. & Battin, T. J. Climate-induced changes in spring snowmelt impact ecosystem metabolism and carbon fluxes in an Alpine Stream Network. *Ecosystems* **21**, 373–390 (2018).
30. Cory, R. M., Ward, C. P., Crump, B. C. & Kling, G. W. Sunlight controls water column processing of carbon in arctic fresh waters. *Science (80-)* **345**, 925–928 (2014).
31. Tobias, C. R., Böhlke, J. K., Harvey, J. W. & Busenberg, E. A simple technique for continuous measurement of time-variable gas transfer in surface waters. *Limnol. Oceanogr. Methods* **7**, 185–195 (2009).
32. Berg, P. & Pace, M. L. Continuous measurement of air–water gas exchange by underwater eddy covariance. *Biogeosciences* **14**, 5595–5606 (2017).
33. Riml, J., Campeau, A., Bishop, K. & Wallin, M. B. Spectral decomposition reveals new perspectives on CO₂ concentration patterns and soil–stream linkages. *J. Geophys. Res. Biogeosci.* **124**, 3039–3056 (2019).
34. Liu, S. & Raymond, P. A. Hydrologic controls on pCO₂ and CO₂ efflux in US streams and rivers. *Limnol. Oceanogr. Lett.* **3**, 428–435 (2018).
35. Lindell, M. J., Granéli, H. W. & Bertilsson, S. Seasonal photoreactivity of dissolved organic matter from lakes with contrasting humic content. *Can. J. Fish. Aquat. Sci.* **57**, 875–885 (2000).
36. Ask, J., Karlsson, J., Persson, L. & Ask, P. Terrestrial organic matter and light penetration: effects on bacterial and primary production in lakes. *Limnol. Oceanogr.* **54**, 2034–2040 (2009).
37. Bravo, A. G. et al. The interplay between total mercury, methylmercury and dissolved organic matter in fluvial systems: a latitudinal study across Europe. *Water Res.* **144**, 172–182 (2018).
38. Schwab, M., Klaus, J., Pfister, L. & Weiler, M. Diel discharge cycles explained through viscosity fluctuations in riparian inflow. *Water Resour. Res.* **52**, 8744–8755 (2016).
39. Bodmer, P., Heinz, M., Pusch, M., Singer, G. & Premke, K. Carbon dynamics and their link to dissolved organic matter quality across contrasting stream ecosystems. *Sci. Total Environ.* **553**, 574–586 (2016).
40. Vachon, D. et al. Paired O₂–CO₂ measurements provide emergent insights into aquatic ecosystem function. *Limnol. Oceanogr. Lett.* **5**, 287–294 (2019).
41. Campeau, A. et al. Stable carbon isotopes reveal soil–stream DIC linkages in contrasting headwater catchments. *J. Geophys. Res. Biogeosci.* **123**, 149–167 (2018).
42. Campeau, A. et al. Current forest carbon fixation fuels stream CO₂ emissions. *Nat. Commun.* **10**, 1–9 (2019).
43. Lorke, A. et al. Technical note: drifting versus anchored flux chambers for measuring greenhouse gas emissions from running waters. *Biogeosciences* **12**, 7013–7024 (2015).
44. Tremblay, A., Varfalvy, L., Garneau, M. & Roehm, C. *Greenhouse Gas Emissions-Fluxes and Processes: Hydroelectric Reservoirs and Natural Environments* (Springer Science & Business Media, 2005).
45. Duc, N. T. et al. Automated flux chamber for investigating gas flux at water–air interfaces. *Environ. Sci. Technol.* **47**, 968–975 (2013).
46. Goldenfum, J. A. *GHG Measurement Guidelines for Freshwater Reservoirs: Derived From: the UNESCO/IHA Greenhouse Gas Emissions from Freshwater Reservoirs Research Project* (International Hydropower Association (IHA), 2010).
47. Jähne, B. et al. On the parameters influencing air–water gas exchange. *J. Geophys. Res. Ocean.* **92**, 1937–1949 (1987).
48. R Core Team. *R: A Language and Environment for Statistical Computing* (R Foundation for Statistical Computing, 2018).
49. Wilcoxon, F. Individual comparisons by ranking methods. *Biomet. Bull.* **1**, 80–83 (1945).
50. Bates, D., Maechler, M., Bolker, B. & Walker, S. Fitting linear mixed-effects models using lme4. *J. Stat. Softw.* **67**, 1–48 (2015).
51. Zuur, A., Ieno, E. N., Walker, N., Saveliev, A. A. & Smith, G. M. *Mixed Effects Models and Extensions in Ecology with R* (Springer Science & Business Media, 2009).
52. Zuur, A. F. & Ieno, E. N. A protocol for conducting and presenting results of regression-type analyses. *Methods Ecol. Evol.* **7**, 636–645 (2016).

Acknowledgements

We thank the initiators of the first Collaborative European Freshwater Science Project for Young Researchers, the European Federation of Freshwater Sciences (EFFS) board, the European Fresh and Young Researchers (EFYR) and the representatives of the Fresh

Blood for Fresh Water (FBFW) meetings. We also thank the seven national freshwater societies financing this project, namely the Iberian Association of Limnology (AIL; Spain and Portugal), Deutsche Gesellschaft für Limnologie e.V. (DGL; Germany), Swiss Society for Hydrology and Limnology (SGHL; Switzerland), Italian Association of Oceanography and Limnology (Italy), Freshwater Biological Association (FBA; United Kingdom), French Limnological Association (AFL; France), Austrian Limnological Society (SIL-Austria), as well as the Leibniz-Institute of Freshwater Ecology and Inland Fisheries for additional funds. Additional funding was awarded to J.P.C.-R. by a Juan de la Cierva postdoctoral grant from the Spanish Government (FJC2018-037791-I), to A.P.P. by a Ph.D. grant from the Fundação para a Ciência e Tecnologia (SFRH/BD/115030/2016), to B.C.D. by the Marine Institute’s Cullen Ph.D. fellowship (Grant No. CF/15/05), to N.C. by the European Union’s Horizon 2020 research and innovation program under the Marie Skłodowska-Curie grant agreement (No. 839709), to J.M. by FCT (Portuguese Science Foundation) through a Ph.D. grant (SFRH/BD/131924/2017), to J.P. by the DSI/NRF Research Chair in Inland Fisheries and Freshwater Ecology, to A.F. by the Juan de la Cierva postdoctoral grant from the Spanish Government (FJCI-2017-33171), and to C.M.-L. by the French National Agency for Water and Aquatic Environments (ONEMA, Action 13, “Colmatage, échanges nappe-rivière et processus biogéochimiques”). We acknowledge Luigi Naselli-Flores and Antonio Camacho for their encouragement and support during the project. We also thank David Bastviken, Ingrid Sundgren, and Thanh Duc Nguyen for the introduction to the logger and chamber construction and advice for measurements of CO₂ fluxes with the chamber, Vincent Fugère for his help in setting up the linear mixed-effect models, and Viktor Rosenberg for creating the map. Furthermore, we are very thankful to Rafael Marcé and Paul del Giorgio for their thoughtful comments on the manuscript and we thank three anonymous reviewers for constructive inputs that improved the manuscript. Open access funding provided by University of Vienna.

Author contributions

K.A. and P.B. conceived the study design, coordinated the project and contributed equally to this work; all authors collected and analyzed the field data and K.A. and P.B. gathered and performed the quality check of all data; K.A., P.B., and J.P.C.-R. co-wrote the paper with the help of M.K., G.H.N., and N.C. All authors (K.A., J.P.C.-R., T.F., A.P., S.C.-F., D.S., A.C.N., A.L.D., A.P.P., B.C.D., N.S., C.G.R., G.H.N., X.T., V.E., L.B.-F., T.B., J.A., An.D., G.B., S.F., N.C., E.d.E., F.P., J.-R.M., J.M., D.F., C.N., M.C., M.N., L.L., C.R. G.-Q., F.R., N.P., J.L.J.L., J.P., M.K., A.F., S.H.O., C.M.-L., A.B., J.A.F., P.J.G., L.A.K., M.R., P.B.) commented on the manuscript.

Competing interests

The authors declare no competing interests.

Additional information

Supplementary information The online version contains supplementary material available at <https://doi.org/10.1038/s43247-021-00192-w>.

Correspondence and requests for materials should be addressed to K.A.

Peer review information *Communications Earth & Environment* thanks the anonymous reviewers for their contribution to the peer review of this work. Primary Handling Editors: Joshua Dean and Joe Aslin.

Reprints and permission information is available at <http://www.nature.com/reprints>

Publisher’s note Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.



Open Access This article is licensed under a Creative Commons Attribution 4.0 International License, which permits use, sharing, adaptation, distribution and reproduction in any medium or format, as long as you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons license, and indicate if changes were made. The images or other third party material in this article are included in the article’s Creative Commons license, unless indicated otherwise in a credit line to the material. If material is not included in the article’s Creative Commons license and your intended use is not permitted by statutory regulation or exceeds the permitted use, you will need to obtain permission directly from the copyright holder. To view a copy of this license, visit <http://creativecommons.org/licenses/by/4.0/>.

© The Author(s) 2021

¹Limnology/Department of Ecology and Genetics, Uppsala University, Uppsala, Sweden. ²WasserCluster Lunz – Biologische Station, Lunz am See, Austria. ³Department of Functional and Evolutionary Ecology, University of Vienna, Vienna, Austria. ⁴Catalan Institute for Water Research (ICRA), Girona, Spain. ⁵University of Girona (UdG), Girona, Spain. ⁶Fluvial Ecosystem Ecology, Department of Ecology, University of Innsbruck, Innsbruck, Austria. ⁷INRAE, UR Riverly, Centre de Lyon-Villeurbanne, Villeurbanne, Cedex, France. ⁸Department of Life and Environmental Sciences, Bournemouth University, Poole, UK. ⁹Department of Sciences and Technological Innovation, University of Piemonte Orientale, Alessandria, Italy. ¹⁰ALPSTREAM – Alpine Stream Research Center, Ostana, Italy. ¹¹Research Centre in Biodiversity and Genetic Resources (CIBIO-InBIO), University of Porto, Vila do Conde, Portugal. ¹²Faculty of Sciences, University of Porto, Porto, Portugal. ¹³Centre for Freshwater and Environmental Studies, Dundalk Institute of Technology, Dundalk, Co Louth, Ireland. ¹⁴National Museum of Natural History, Bulgarian Academy of Sciences, Sofia, Bulgaria. ¹⁵River and Conservation Research, Department of Ecology, University of Innsbruck, Innsbruck, Austria. ¹⁶Department of Aquatic Ecosystems, Institute of Biodiversity and Ecosystem Research, Bulgarian Academy of Sciences, Sofia, Bulgaria. ¹⁷Department of Aquatic Sciences and Assessment, Swedish University of Agricultural Sciences, Uppsala, Sweden. ¹⁸Department of Ecology and Environmental Science, Umeå University, Umeå, Sweden. ¹⁹Department of Life Sciences and Systems Biology, University of Turin, Turin, Italy. ²⁰Marine Institute, Furnace, Newport, Co Mayo, Ireland. ²¹Department of Evolutionary Biology, Ecology and Environmental Sciences, Faculty of Biology, University of Barcelona (UB), Barcelona, Spain. ²²Centre for Ecology, Evolution and Environmental Changes (cE3c), Faculdade de Ciências, Universidade de Lisboa, Lisboa, Portugal. ²³Institute for Environmental Sciences, University of Koblenz-Landau, Landau, Germany. ²⁴Institute of Microbiology, University of Innsbruck, Innsbruck, Austria. ²⁵Experimental Limnology, Leibniz-Institute of Freshwater Ecology and Inland Fisheries (IGB), Stechlin, Germany. ²⁶Ecohydrology, Leibniz-Institute of Freshwater Ecology and Inland Fisheries (IGB), Berlin, Germany. ²⁷Center for Advanced Studies of Blanes, Spanish National Research Council, Blanes, Spain. ²⁸Institute of Geography and Geoecology, Karlsruhe Institute of Technology, Karlsruhe, Germany. ²⁹South African Institute for Aquatic Biodiversity, Makhanda, South Africa. ³⁰Department of Ecology and Environmental Sciences, Palacký University Olomouc, Olomouc, Czech Republic. ³¹Environmental Research Institute, University of Highlands and Islands (UHI), Thurso, Scotland, UK. ³²Department of General and Applied Hydrobiology, Sofia University “St. Kliment Ohridski”, Sofia, Bulgaria. ³³Chemical Analytics and Biogeochemistry, Leibniz-Institute of Freshwater Ecology and Inland Fisheries, Berlin, Germany. ³⁴Present address: Department of Biology, Aarhus University, Aarhus C, Denmark. ³⁵Present address: Institute of Global Health, Faculty of Medicine, University of Geneva, Campus Biotech, Geneva, Switzerland. ³⁶Present address: Centre for Environment, Fisheries and Aquaculture Science (Cefas), Lowestoft, Suffolk, UK. ³⁷Present address: Department of Bioscience, Aarhus University, Silkeborg, Denmark. ³⁸Present address: Norwegian Institute for Water Research, Oslo, Norway. ³⁹Present address: Laboratoire des Sciences du Climat et de l’Environnement (LSCE), CEA, CNRS, UVSQ, Gif-Sur-Yvette, France. ⁴⁰Present address: United States Geological Survey, Boulder, CO, USA. ⁴¹Present address: Environmental Archaeology Lab, Department of Historical, Philosophical and Religious studies, Umeå University, Umeå, Sweden. ⁴²Present address: Department of Forest Ecology and Management, Swedish University of Agricultural Sciences, Umeå, Sweden. ⁴³Present address: Global Change Research Institute of the Czech Academy of Sciences, Brno, Czech Republic. ⁴⁴Present address: Groupe de Recherche Interuniversitaire en Limnologie, Département des Sciences Biologiques, Université du Québec à Montréal, Montréal, Canada. ✉email: katrin.attermeyer@univie.ac.at