

Title	Non-steady-state hematopoiesis regulated by the C/EBP transcription factor.
Author(s)	Hirai, Hideyo; Yokota, Asumi; Tamura, Akihiro; Sato, Atsushi; Maekawa, Taira
Citation	Cancer science (2015), 106(7): 797-802
Issue Date	2015-07
URL	http://hdl.handle.net/2433/215155
Right	© 2015 The Authors. Cancer Science published by Wiley Publishing Asia Pty Ltd on behalf of Japanese Cancer Association.; This is an open access article under the terms of the Creative Commons Attribution-NonCommercial License, which permits use, distribution and reproduction in any medium, provided the original work is properly cited and is not used for commercial purposes.
Type	Journal Article
Textversion	publisher

Review Article

Non-steady-state hematopoiesis regulated by the C/EBP β transcription factor

Hideyo Hirai, Asumi Yokota, Akihiro Tamura, Atsushi Sato and Taira Maekawa

Department of Transfusion Medicine and Cell Therapy, Kyoto University Hospital, Kyoto, Japan

Key words

Cancer, C/EBP β , emergency, hematological malignancy, steady-state

Correspondence

Hideyo Hirai, Department of Transfusion Medicine and Cell Therapy, Kyoto University Hospital, 54 Kawahara-cho, Shogo-in, Sakyo-ku, Kyoto 606-8507, Japan.
Tel: +81-75-751-3630; Fax: +81-75-751-4283;
E-mail: hhirai@kuhp.kyoto-u.ac.jp

Funding Information

Ministry of Education, Culture, Sports, Science and Technology of Japan; Ministry of Health, Labor and Welfare of Japan; National Cancer Center Research and Development Fund; Takeda Science Foundation; Princess Takamatsu Cancer Research Fund; Kobayashi Foundation for Cancer Research.

Received March 23, 2015; Revised April 26, 2015;
Accepted April 27, 2015

Cancer Sci 106 (2015) 797–802

doi: 10.1111/cas.12690

Steady-state hematopoiesis responds to extracellular stimuli to meet changing demands and also to pathologically altered intracellular signaling. Granulocyte production increases following infection or in response to cytokine stimulation, and activation of the CCAAT/enhancer-binding protein β (C/EBP β) transcription factor is required for such stress-induced granulopoiesis, whereas C/EBP α plays a critical role in maintaining steady-state granulopoiesis. Different roles of these C/EBP transcription factors in different modes of hematopoiesis are evolutionally conserved from zebrafish to humans. In addition to reactions against infections, C/EBP β is responsible for cancer-driven myelopoiesis, which promotes cancer progression, at least in part, by abrogating the immune response in the cancer microenvironment. The BCR–ABL fusion protein activates emergency-specific pathway of granulopoiesis by upregulating C/EBP β . This in turn causes chronic phase chronic myeloid leukemia, which is characterized by myeloid expansion. The C/EBP β transcription factor also plays a role in other hematological malignancies of both myeloid and lymphoid lineage origin. Thus, elucidation of the upstream and downstream networks surrounding C/EBP β will lead to the development of novel therapeutic strategies for diseases mediated by non-steady-state hematopoiesis.

Transcription Factor CCAAT/Enhancer Binding Protein β

C CAAT/Enhancer Binding Protein β (C/EBP β) belongs to the C/EBP leucine zipper domain-containing family of transcription factors (Fig. 1).^(1,2) This intronless gene product binds to certain genomic regulatory regions either as a homodimer or as a heterodimer with other molecules, including other members of the C/EBP family. In addition to direct DNA binding, C/EBP β cooperates with the switch/sucrose non-fermentable complex to regulate gene expression through chromatin remodeling.⁽³⁾ It induces or represses the expression of target genes and, ultimately, regulates the proliferation, differentiation, metabolism, and survival of many different cell types.⁽¹⁾

The expression and function of C/EBP β are regulated in a complex way during transcription, translation, post-translational modification, and protein–protein interactions.^(4–8) Notably, alternative translation through the use of different initiation codons generates three different isoforms of C/EBP β : liver-enriched activating protein* (LAP* or full-length), liver-enriched activating protein (LAP), and liver-enriched inhibitory protein (LIP) (Fig. 1).⁽⁶⁾ Both LAP* and LAP are transcriptional activators, whereas LIP (which is the shortest isoform and lacks transactivation domains but retains DNA binding and dimerization domains) acts as a repressor or a dominant negative

inhibitor of other C/EBP family transcription factors.⁽⁹⁾ The ratio of these isoforms is regulated by different signaling events and has a significant impact on the overall function of C/EBP β .^(10,11)

Within the hematopoietic system, C/EBP β is expressed at high levels by monocytes and macrophages, and regulates genes involved in immune and inflammatory responses.^(12–16) In addition, we found that C/EBP β plays a crucial role in hematopoiesis, especially under stress conditions.^(17–19) Here, we discuss the role of this transcription factor in non-steady-state hematopoiesis, including the emergency response to infection and cancer, and in hematological malignancies.

Modes of Hematopoiesis

Hematopoiesis is a continuous process that supplies an organism with all blood cells over its lifetime. To avoid either an excess or lack of any specific type of blood cell, hematopoiesis must be tightly regulated according to demand. During steady-state conditions, the constant production of mature blood cells is maintained by fine-tuning the proliferation and differentiation of hematopoietic precursors in both a cell-intrinsic and a cell-extrinsic manner. By contrast, in emergency situations such as infection or bleeding, large numbers

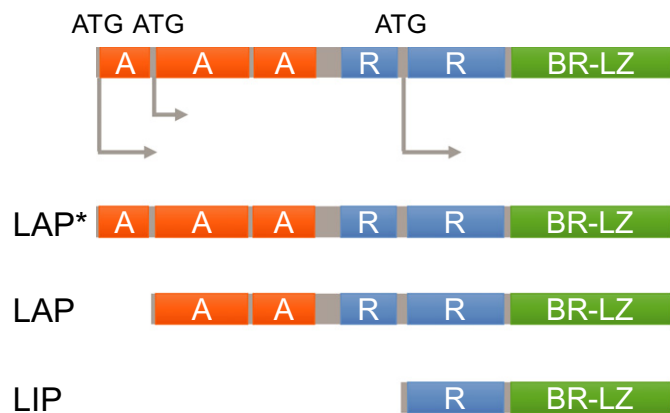


Fig. 1. Schematic illustration of the C/EBP β transcription factor and its isoforms. A, transactivating domain; BR-LZ, basic region–leucine zipper domain; LAP*, liver-enriched activating protein*; LAP, liver-enriched activating protein; LIP, liver-enriched inhibitory protein; R, repression domain.

of functionally mature cells are required. These increased demands must be met by an immediate increase in the production or release of specific cell types (Fig. 2).⁽²⁰⁾ These physiological non-steady-state responses are triggered by external stimuli and are resolved when the activating signals cease. In addition to responses to infection or bleeding, hematopoietic stress can be elicited by various kinds of pro-inflammatory disease, including cancer and autoimmune diseases.^(21–24) At the molecular level, steady-state-specific regulatory mechanisms are thought to be modulated in response to external stimuli. The shift from steady-state to emergency hematopoiesis and vice versa is a continuous process, and the extent of the shift is dependent on the type, strength, and duration of the stimuli and/or signals.^(20,25) It is difficult to determine clear boundaries between steady-state and emergency hemato-

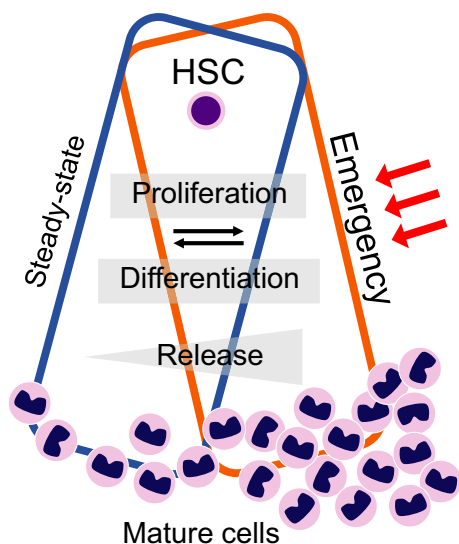


Fig. 2. Modes of hematopoiesis. The hematopoietic system in the bone marrow supplies mature blood cells on demand. Under both steady-state and emergency conditions, overlapping and distinct signals ensure the adequate production and release of mature cells. Red arrows indicate extracellular stimuli, including infection and cancer. HSC, hematopoietic stem cells.

poiesis, partly because the shift between these modes may be an extension of the fine-tuning of steady-state hematopoiesis mediated by co-operation between steady-state-specific and emergency-specific signals. Recent studies report the involvement of inflammatory signals in both the development and ageing of hematopoiesis.^(26–28) Therefore, it is necessary to identify the emergency-specific signals if we are to fully understand the mechanisms that fine-tune hematopoiesis in general.

Emergency Granulopoiesis

Neutrophilic granulocytes (granulocytes) are recruited to the frontline of infection, where they expel their granular contents to fight microbes.^(20,25) In the bone marrow, hematopoietic stem cells give rise to mature granulocytes through successive intermediates, such as common myeloid progenitors and granulocyte–macrophage progenitors.⁽²⁹⁾ Granulocytes have an extremely short half-life; therefore, they must be produced continuously in the bone marrow, stored, and supplied to the periphery. As is the cases with other hematopoietic lineages, either an excess or a lack of granulocytes is harmful to the host; therefore, granulopoiesis must be tightly regulated according to demand. It is well known that C/EBP α plays critical roles in granulopoiesis. In *Cebpa*-deficient mice, transition from common myeloid progenitors to granulocyte–macrophage progenitors is completely abrogated and no granulocytes are present under steady-state conditions.^(17,30,31) Overexpression of C/EBP α represses the proliferation of leukemic cells and induces their differentiation into granulocytes.^(17,32) Collectively, these findings suggest that C/EBP α is the master regulator of steady-state granulopoiesis.

While searching for the regulatory mechanisms involved in emergency granulopoiesis, we found that granulopoiesis can be induced by cytokines in the absence of C/EBP α . This suggests the existence of a C/EBP α -independent pathway of granulopoiesis under emergency conditions.⁽¹⁷⁾ Interestingly, all members of the C/EBP family, except C/EBP β , were downregulated in response to cytokine stimulation. Cytokine- or infection-induced enhancement of granulopoiesis is impaired in *Cebpb* knockout mice, and the C/EBP α -independent pathway of granulopoiesis is significantly attenuated by inhibiting C/EBP β .⁽¹⁷⁾ By contrast, C/EBP β is not necessary for steady-state granulopoiesis. These results clearly suggest that C/EBP β is required for stress-induced granulopoiesis; indeed, this requirement has been verified in other mouse models and in a zebrafish model.^(33–35) Both C/EBP β and C/EBP α share many common target molecules, including genes associated with granulocytic differentiation.⁽³⁶⁾ By contrast, they show a differing ability to regulate the cell cycle. C/EBP α strongly inhibits the cell cycle through direct or indirect interactions with cell cycle regulators,^(37–39) whereas C/EBP β has a less inhibitory effect.^(17,40) These differences might be the reason for the selective requirement of C/EBP α and C/EBP β for steady-state and emergency granulopoiesis, respectively. As the transition from steady-state to emergency granulopoiesis (or vice versa) is a continuous process, C/EBP α or C/EBP β might collaborate with each other to ensure an adequate supply of granulocytes by fine-tuning the proliferation and differentiation of granulocyte precursors (Fig. 3). Furthermore, we also found that CEBP β is required by early granulocyte precursors under emergency conditions; we are currently investigating the role of CEBP β in regulating hematopoietic stem cells.⁽¹⁸⁾

Role of C/EBPs in the Pathophysiology of Severe Congenital Neutropenia

Severe congenital neutropenia (SCN) is an inherited condition characterized by severe neutropenia in the peripheral blood (<500/ μ L) and by arrest of myeloid precursor maturation at the promyelocyte/myelocyte stage in the bone marrow, resulting in increased vulnerability to bacterial and fungal infections.^(41,42) The majority of patients with SCN respond to treatment with recombinant granulocyte-colony stimulating factor, which increases the neutrophil count and reduces both the frequency and severity of infections. Patients with SCN harbor mutations in diverse genes. These heterogeneous genetic alterations reflect the complex mechanisms governing the homeostasis of neutrophils.⁽⁴²⁾ Establishing induced pluripotent stem cells from SCN cells in combination with an *in vitro* differentiation system will further our understanding of both the pathogenesis of this disease and the physiological regulation of granulopoiesis.^(43,44) The majority of SCN patients harbor mutations in *ELANE* and *HAX1* (approximately 60% and 10%, respectively).⁽⁴⁵⁾ Recently, the lymphoid enhancer-binding factor 1 (LEF-1) transcription factor was identified as a common factor responsible for defective granulopoiesis in SCN patients with mutations in *ELANE* (ELA2) or *HAX1*.⁽⁴⁶⁾ LEF-1 regulates C/EBP α during granulopoiesis. Both the expression and function of LEF-1 and C/EBP α are severely reduced in myeloid precursors in SCN patients with *ELANE* or *HAX1* mutations, and the reduction in C/EBP α (the master regulator of steady-state granulopoiesis) might be a critical mechanism underlying neutropenia in SCN.^(46,47) Maturation arrest can be overcome by treatment with granulocyte-colony stimulating factor, presumably because it activates the C/EBP β -mediated pathway of granulopoiesis, which is thought to be intact in SCN patients.⁽⁴⁸⁾ The new insights into the pathophysiology of SCN suggest that the different roles of C/EBP α and C/EBP β during granulopoiesis may also be true in humans.

Cancer-associated Myeloipoesis

Cancer progression, including tumor growth, invasion, and metastasis, cannot be achieved by tumor cells alone; it requires the appropriate microenvironment.^(49,50) Accumulating

evidence suggests that myeloid cells are major components of the cancer microenvironment.^(21–23) Indeed, there is a strong association between increased numbers of macrophages or neutrophils in cancer tissues and poor patient survival.^(51,52) Thus, these myeloid cells can be good candidate therapeutic targets.

Tumor cells, or other stromal cells, in the microenvironment produce a variety of growth factors and chemokines, which then recruit myeloid cells from the bone marrow or reservoir tissues.^(21–23) Therefore, the mode of hematopoiesis is altered in the presence of cancer, and hematopoietic systems release a variety of myeloid cells into the cancer microenvironment. Such cells include monocytes, macrophages (tumor-associated macrophages), dendritic cells, neutrophils (tumor-associated neutrophils), and eosinophils. Recent studies by ourselves and others identified fibrocytes as important constituents of the cancer microenvironment.^(53–55) In such microenvironments, myeloid cells support cancer progression by secreting growth factors and promoting angiogenesis and/or tissue remodeling. In addition, it is widely accepted that a special subset of myeloid cells, called myeloid-derived suppressor cells (MDSCs), are induced by tumor-induced factors and are responsible for immune dysfunction.^(22,56) MDSCs in mice are classified as either monocytic or granulocytic based on their surface expression of Ly6C and Ly6G, respectively. Upregulated in the bone marrow of tumor-bearing hosts, C/EBP β regulates the expression of enzymes such as arginase and inducible nitric oxide synthase, both of which are required for the lymphocyte-inhibitory activities of MDSCs (Fig. 4a).⁽⁵⁷⁾ Accordingly, in tumor-bearing mice, both the emergence and the immunosuppressive function of MDSCs are severely abrogated in the absence of C/EBP β , resulting in attenuated tumor spread.⁽⁵⁷⁾ A similar relationship between C/EBP β and cancer-driven myeloipoesis is observed in humans.⁽⁵⁸⁾ These findings suggest that C/EBP β plays a critically important role in cancer-induced inflammation; thus, C/EBP β may be a therapeutic target for regulating the cancer microenvironment. Further studies should examine the roles of C/EBP β in generating or regulating the function of MDSCs in other diseases.

Role of CEBP β in Chronic Myeloid Leukemia

Chronic phase chronic myeloid leukemia (CP-CML) is characterized by a massive expansion of myeloid cells.⁽⁵⁹⁾ In sharp contrast to acute myeloid leukemia (AML) with leukemic hiatus, both myeloid progenitors and mature granulocytes accumulate in the bone marrow, peripheral blood, and spleen in CP-CML. The myeloid expansion in CP-CML is attributed to the BCR-ABL fusion protein, which arises from a translocation between chromosomes 9 and 22.⁽⁵⁹⁾ The leukocytosis observed in patients with infections, severe burns, or cancer is sometimes referred to as a “leukemoid” reaction because of the marked increase in the number of myeloid cells with a “left shift” in the shape of the nucleus. The resemblance between leukemoid reactions and CP-CML prompted us to examine whether BCR-ABL might use the emergency-specific pathway of granulopoiesis. Therefore, we investigated the role of C/EBP β in CP-CML (Fig. 4b). BCR-ABL upregulates C/EBP β , at least in part, by activating signal transducer and activator of transcription 5.⁽¹⁹⁾ Myeloid differentiation and proliferation (induced by BCR-ABL) are significantly impaired in *Cebpb*-deficient bone marrow cells both *in vitro* and *in vivo*.⁽¹⁹⁾ Interestingly, higher numbers of *Cebpb*-deficient leukemic stem cells were maintained after serial transplantation than wild-type leukemic stem cells in this mouse model.⁽¹⁹⁾ These results sug-

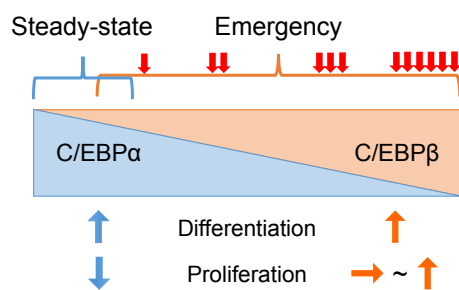


Fig. 3. Role of C/EBP transcription factors in steady-state and emergency granulopoiesis. Activation of the C/EBP β transcription factor is required to generate increased numbers of granulocytes under emergency conditions, such as severe infection or cytokine exposure, whereas C/EBP α plays a critical role under steady-state conditions. C/EBP β and C/EBP α share many common target molecules, including genes associated with granulocytic differentiation. By contrast, they show a differing ability to regulate the cell cycle. C/EBP α strongly inhibits the cell cycle, whereas C/EBP β has a less inhibitory effect. Red arrows indicate extracellular stimuli that activate C/EBP β .

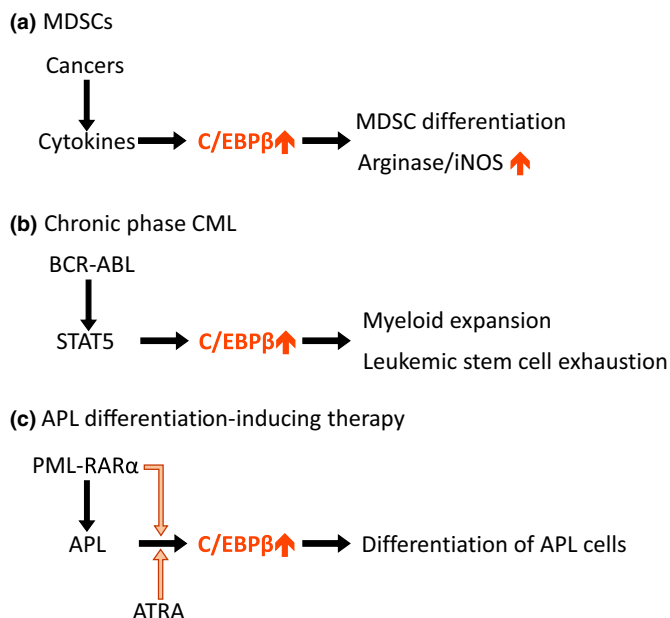


Fig. 4. Involvement of C/EBP β in non-infectious modes of hematopoiesis. (a) C/EBP β is upregulated in the bone marrow of tumor-bearing hosts. C/EBP β regulates the differentiation of myeloid-derived myeloid suppressor cells (MDSCs) and the expression of enzymes such as arginase and inducible nitric oxide synthase (iNOS), both of which are required for the lymphocyte-inhibitory activities of MDSCs. (b) In chronic phase chronic myeloid leukemia (CML), C/EBP β is activated by signal transducer and activator of transcription 5 (STAT5), which is located downstream of BCR-ABL. C/EBP β is involved in BCR-ABL-mediated myeloid expansion and leukemic stem cell exhaustion in chronic phase CML. (c) Acute promyelocytic leukemia (APL) is characterized by a promyelocytic leukemia-retinoic acid receptor α (PML-RAR α)-mediated differentiation block at the promyelocyte stage. During the processes of differentiation-inducing therapy using all trans retinoic acid (ATRA), C/EBP β is upregulated in the presence of PML-RAR α and increases the number of neutrophils derived from APL cells by promoting their proliferation and differentiation.

gest that C/EBP β is involved in BCR-ABL-mediated myeloid expansion and leukemic stem cell exhaustion in CP-CML. Consistent with our observations, C/EBP β is markedly upregulated in a pluripotent hematopoietic cell line transduced with BCR-ABL.⁽⁶⁰⁾ By contrast, downregulation of C/EBP β is associated with progression of CML toward a blast crisis.⁽⁶¹⁾ Changes in the BCR-ABL-mediated regulation of C/EBP β during the progression of CML may be a consequence of

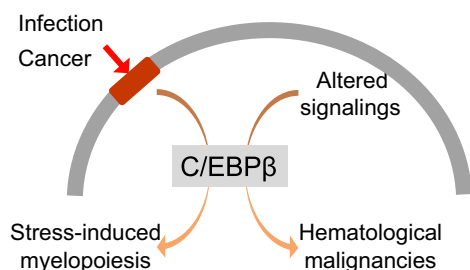


Fig. 5. Role of C/EBP β and non-steady-state hematopoiesis. Different types of cell-extrinsic stress, including infections and cancer, activate C/EBP β to increase the supply of functionally mature myeloid cells or myeloid-derived suppressor cells. Dysregulation of C/EBP β is observed in some hematological malignancies, resulting in maintenance or progression of disease.

genetic or epigenetic changes. Isoforms of C/EBP β involved in the pathogenesis of CML remain to be identified. Further identification of the molecular mechanisms underlying the regulation of C/EBP β and C/EBP β -mediated leukemic stem cell exhaustion might lead to novel therapeutic strategies for eradicating CML stem cells.

Role of CEBP β in other Hematological Malignancies

Hematological malignancies are the consequence of dysregulated differentiation and/or proliferation; therefore, they can be regarded as a form of pathologically induced non-steady-state hematopoiesis. Because C/EBP α promotes neutrophilic differentiation and inhibits the cell cycle, many cases of AML are associated with recurrent mutations in, or dysregulation of, C/EBP α .⁽⁶²⁻⁶⁴⁾ By contrast, no recurrent mutations in C/EBP β have been identified in AML,⁽⁶⁵⁾ possibly reflecting the fact that this transcription factor is required for emergency-specific responses. However, C/EBP β plays a role in the pathogenesis of many hematological malignancies. In AML, LIP (the shortest isoform of C/EBP β) collaborates with a proto-oncogene, *Evi1*, to induce leukemia in a mouse bone marrow transplantation model.⁽⁶⁶⁾ The same isoform is induced by signaling downstream of internal tandem duplication of fms-like tyrosine kinase 3, thereby supporting the proliferation of blasts.⁽⁶⁷⁾ These findings suggest that regulating the amount or the ratio of C/EBP β isoforms might be a common pathway that is abrogated during the development of AML.

Acute promyelocytic leukemia (APL) is a subtype of AML characterized by a promyelocytic leukemia-retinoic acid receptor α (PML-RAR α)-mediated differentiation block at the promyelocyte stage, which occurs (at least in part) through an impairment in C/EBP α function.⁽⁶⁸⁾ This block is reversed by all-trans retinoic acid (ATRA), which is used as frontline therapy for APL.⁽⁶⁹⁾ After the start of ATRA treatment, mature neutrophil-like cells originate from leukemic promyelocytes and their numbers increase in the bone marrow and peripheral blood of responder APL cases. During this process of differentiation-inducing therapy, C/EBP β is upregulated in the presence of PML-RAR α and increases the number of neutrophils derived from APL cells by promoting their proliferation and differentiation (Fig. 4c).⁽⁷⁰⁾ In other words, C/EBP β is an ATRA-dependent PML-RAR α target gene in APL cells.

It is clear that C/EBP β regulates not only myeloid hematopoiesis, but also bone marrow B lymphopoiesis, in both a cell-intrinsic and cell-extrinsic manner.^(71,72) One study examined the contribution of C/EBP β to the development of lymphoid neoplasias in cases with acute B-cell precursor leukemia and identified recurrent translocations in C/EBP β , which resulted in the upregulation of C/EBP β .⁽⁷³⁾

Anaplastic large cell lymphoma (ALCL) is a subset of non-Hodgkin's lymphoma characterized by unique cell morphology and expression of CD30.⁽⁷⁴⁾ In ALCL cells, the anaplastic lymphoma kinase (*ALK*) gene is frequently fused to the nucleophosmin (*NPM*) gene, and the resulting ALK activity is the central driver for the survival of ALCL cells. Recently, C/EBP β was identified as a downstream target of ALK-mediated signaling.⁽⁷⁴⁾ C/EBP β is upregulated in the presence of activated ALK through signal transducer and activator of transcription 3^(74,75) or by post-transcriptional regulation,⁽⁷⁶⁾ whereupon it contributes to the transformation and survival of ALCL cells.⁽⁷⁷⁾

The pathogenesis of multiple myeloma remains unclear and, at present, this plasma cell disorder is incurable. A recent report shows that C/EBP β is overexpressed in myeloma cells

and is involved in regulating several transcription factors, including IRF4, XBP1, and BLIMP1, all of which are critical for the proliferation and survival of myeloma cells.⁽⁷⁸⁾ Inhibiting C/EBP β translation in myeloma cells using immunomodulatory derivatives of thalidomide has been proposed as a novel therapeutic strategy for multiple myeloma.⁽⁷⁹⁾

Conclusions

The expression and/or function of C/EBP β are upregulated in the hematopoietic system in response to various kinds of cell-extrinsic stress, including infections and cancer. This upregulation increases the supply of myeloid cells. Dysregulation of C/EBP β is observed in several hematological malignancies, resulting in the maintenance or progression of disease. Although the roles of C/EBP β in hematopoiesis have not been fully elucidated, it appears to play a key role in non-steady-state hematopoiesis, including hematological malignancies, and hematopoiesis in host with cancers in addition to hematopoietic responses against infections (Fig. 5). Even though direct targeting of this transcription

factor might be technically difficult, identifying the upstream and downstream networks involving C/EBP β will lead to a better understanding of the pathogenesis and pathophysiology of diseases mediated by non-steady-state hematopoiesis.

Acknowledgments

This work was supported by a Grant-in-Aid for Scientific Research from the Japanese Ministry of Education, Culture, Sports, Science and Technology (MEXT) (to H.H., A.Y., and T.M.), a grant from the Project for Development of Innovative Research on Cancer Therapeutics from MEXT (to H.H. and T.M.), a Grant-in-Aid from the Ministry of Health, Labour and Welfare in Japan (to T.M.), the National Cancer Center Research and Development Fund (to T.M.), the Takeda Science Foundation (to H.H.), a research grant from the Princess Takamatsu Cancer Research Fund (to T.M.), and the Kobayashi Foundation for Cancer Research (to T.M.).

Disclosure Statement

The authors have no conflict of interest.

References

- Ramji DP, Foka P. CCAAT/enhancer-binding proteins: structure, function and regulation. *Biochem J* 2002; **365**: 561–75.
- Nerlov C. The C/EBP family of transcription factors: a paradigm for interaction between gene expression and proliferation control. *Trends Cell Biol* 2007; **17**: 318–24.
- Kowenz-Leutz E, Leutz A. A C/EBP beta isoform recruits the SWI/SNF complex to activate myeloid genes. *Mol Cell* 1999; **4**: 735–43.
- Ruffell D, Mourkioti F, Gambardella A et al. A CREB-C/EBPbeta cascade induces M2 macrophage-specific gene expression and promotes muscle injury repair. *Proc Natl Acad Sci USA* 2009; **106**: 17475–80.
- Hirai H, Kamio N, Huang G et al. Cyclic AMP responsive element binding proteins are involved in ‘emergency’ granulopoiesis through the upregulation of CCAAT/enhancer binding protein beta. *PLoS One* 2013; **8**: e54862.
- Descombes P, Schibler U. A liver-enriched transcriptional activator protein, LAP, and a transcriptional inhibitory protein, LIP, are translated from the same mRNA. *Cell* 1991; **67**: 569–79.
- Trautwein C, Caelles C, van der Geer P, Hunter T, Karin M, Chojkier M. Transactivation by NF-IL6/LAP is enhanced by phosphorylation of its activation domain. *Nature* 1993; **364**: 544–7.
- Bradley MN, Zhou L, Smale ST. C/EBPbeta regulation in lipopolysaccharide-stimulated macrophages. *Mol Cell Biol* 2003; **23**: 4841–58.
- Descombes P, Chojkier M, Lichtsteiner S, Falvey E, Schibler U. LAP, a novel member of the C/EBP gene family, encodes a liver-enriched transcriptional activator protein. *Genes Dev* 1990; **4**: 1541–51.
- An MR, Hsieh CC, Reisner PD et al. Evidence for posttranscriptional regulation of C/EBPalpha and C/EBPbeta isoform expression during the lipopolysaccharide-mediated acute-phase response. *Mol Cell Biol* 1996; **16**: 2295–306.
- Stoilova B, Kowenz-Leutz E, Scheller M, Leutz A. Lymphoid to myeloid cell trans-differentiation is determined by C/EBPbeta structure and post-translational modifications. *PLoS One* 2013; **8**: e65169.
- Natsuka S, Akira S, Nishio Y et al. Macrophage differentiation-specific expression of NF-IL6, a transcription factor for interleukin-6. *Blood* 1992; **79**: 460–6.
- Tanaka T, Akira S, Yoshida K et al. Targeted disruption of the NF-IL6 gene discloses its essential role in bacteria killing and tumor cytotoxicity by macrophages. *Cell* 1995; **80**: 353–61.
- Screpanti I, Romani L, Musiani P et al. Lymphoproliferative disorder and imbalanced T-helper response in C/EBP beta-deficient mice. *EMBO J* 1995; **14**: 1932–41.
- Huber R, Pietsch D, Panterodt T, Brand K. Regulation of C/EBPbeta and resulting functions in cells of the monocytic lineage. *Cell Signal* 2012; **24**: 1287–96.
- Cain DW, O’Koren EG, Kan MJ et al. Identification of a tissue-specific, C/EBPbeta-dependent pathway of differentiation for murine peritoneal macrophages. *J Immunol* 2013; **191**: 4665–75.
- Hirai H, Zhang P, Dayaram T et al. C/EBPbeta is required for ‘emergency’ granulopoiesis. *Nat Immunol* 2006; **7**: 732–9.
- Satake S, Hirai H, Hayashi Y et al. C/EBPbeta is involved in the amplification of early granulocyte precursors during candidemia-induced ‘emergency’ granulopoiesis. *J Immunol* 2012; **189**: 4546–55.
- Hayashi Y, Hirai H, Kamio N et al. C/EBPbeta promotes BCR-ABL-mediated myeloid expansion and leukemic stem cell exhaustion. *Leukemia* 2013; **27**: 619–28.
- Manz MG, Boettcher S. Emergency granulopoiesis. *Nat Rev Immunol* 2014; **14**: 302–14.
- Solinas G, Germano G, Mantovani A, Allavena P. Tumor-associated macrophages (TAM) as major players of the cancer-related inflammation. *J Leukoc Biol* 2009; **86**: 1065–73.
- Gabrilovich DI, Ostrand-Rosenberg S, Bronte V. Coordinated regulation of myeloid cells by tumours. *Nat Rev Immunol* 2012; **12**: 253–68.
- Cortez-Retamozo V, Eitzrodt M, Newton A et al. Origins of tumor-associated macrophages and neutrophils. *Proc Natl Acad Sci USA* 2012; **109**: 2491–6.
- Fujii W, Ashihara E, Hirai H et al. Myeloid-derived suppressor cells play crucial roles in the regulation of mouse collagen-induced arthritis. *J Immunol* 2013; **191**: 1073–81.
- Nauseef WM, Borregaard N. Neutrophils at work. *Nat Immunol* 2014; **15**: 602–11.
- Li Y, Esain V, Teng L et al. Inflammatory signaling regulates embryonic hematopoietic stem and progenitor cell production. *Genes Dev* 2014; **28**: 2597–612.
- He Q, Zhang C, Wang L et al. Inflammatory signaling regulates hematopoietic stem and progenitor cell emergence in vertebrates. *Blood* 2015; **125**: 1098–106.
- Chambers SM, Shaw CA, Gatzka C, Fisk CJ, Donehower LA, Goodell MA. Aging hematopoietic stem cells decline in function and exhibit epigenetic dysregulation. *PLoS Biol* 2007; **5**: e201.
- Akashi K, Traver D, Miyamoto T, Weissman IL. A clonogenic common myeloid progenitor that gives rise to all myeloid lineages. *Nature* 2000; **404**: 193–7.
- Zhang DE, Zhang P, Wang ND, Hetherington CJ, Darlington GJ, Tenen DG. Absence of granulocyte colony-stimulating factor signaling and neutrophil development in CCAAT enhancer binding protein alpha-deficient mice. *Proc Natl Acad Sci USA* 1997; **94**: 569–74.
- Zhang P, Iwasaki-Arai J, Iwasaki H et al. Enhancement of hematopoietic stem cell repopulating capacity and self-renewal in the absence of the transcription factor C/EBP alpha. *Immunity* 2004; **21**: 853–63.
- Radomska HS, Huettner CS, Zhang P, Cheng T, Scadden DT, Tenen DG. CCAAT enhancer binding protein alpha is a regulatory switch sufficient for induction of granulocytic development from bipotential myeloid progenitors. *Mol Cell Biol* 1998; **18**: 4301–14.
- Akagi T, Saitoh T, O’Kelly J, Akira S, Gombart AF, Koeffler HP. Impaired response to GM-CSF and G-CSF, and enhanced apoptosis in C/EBPbeta-deficient hematopoietic cells. *Blood* 2008; **111**: 2999–3004.

- 34 Zhang H, Nguyen-Jackson H, Panopoulos AD, Li HS, Murray PJ, Watowich SS. STAT3 controls myeloid progenitor growth during emergency granulopoiesis. *Blood* 2010; **116**: 2462–71.
- 35 Hall CJ, Flores MV, Oehlers SH *et al*. Infection-responsive expansion of the hematopoietic stem and progenitor cell compartment in zebrafish is dependent upon inducible nitric oxide. *Cell Stem Cell* 2012; **10**: 198–209.
- 36 Jones LC, Lin ML, Chen SS *et al*. Expression of C/EBPbeta from the C/ebpalpha gene locus is sufficient for normal hematopoiesis in vivo. *Blood* 2002; **99**: 2032–6.
- 37 Porse BT, Pedersen TA, Xu XF *et al*. E2F repression by C/EBP alpha is required for adipogenesis and granulopoiesis in vivo. *Cell* 2001; **107**: 247–58.
- 38 Wang HM, Iakova P, Wilde M *et al*. C/EBP alpha arrests cell proliferation through direct inhibition of cdk2 and cdk4. *Mol Cell* 2001; **8**: 817–28.
- 39 Johansen LM, Iwama A, Lodie TA *et al*. c-Myc is a critical target for c/EBPalpha in granulopoiesis. *Mol Cell Biol* 2001; **21**: 3789–806.
- 40 Sebastian T, Johnson PF. Stop and go: anti-proliferative and mitogenic functions of the transcription factor C/EBPbeta. *Cell Cycle* 2006; **5**: 953–7.
- 41 Donadieu J, Fenneteau O, Beaupain B, Mahlaoui N, Chantelot CB. Congenital neutropenia: diagnosis, molecular bases and patient management. *Orphanet J Rare Dis* 2011; **6**: 26.
- 42 Klein C. Genetic defects in severe congenital neutropenia: emerging insights into life and death of human neutrophil granulocytes. *Annu Rev Immunol* 2011; **29**: 399–413.
- 43 Hiramoto T, Ebihara Y, Mizoguchi Y *et al*. Wnt3a stimulates maturation of impaired neutrophils developed from severe congenital neutropenia patient-derived pluripotent stem cells. *Proc Natl Acad Sci USA* 2013; **110**: 3023–8.
- 44 Morishima T, Watanabe K, Niwa A *et al*. Genetic correction of HAX1 in induced pluripotent stem cells from a patient with severe congenital neutropenia improves defective granulopoiesis. *Haematologica* 2014; **99**: 19–27.
- 45 Zeidler C, Germeshausen M, Klein C, Welte K. Clinical implications of ELA2-, HAX1-, and G-CSF-receptor (CSF3R) mutations in severe congenital neutropenia. *Br J Haematol* 2009; **144**: 459–67.
- 46 Skokowa J, Cario G, Uenal M *et al*. LEF-1 is crucial for neutrophil granulocytopenia and its expression is severely reduced in congenital neutropenia. *Nat Med* 2006; **12**: 1191–7.
- 47 Skokowa J, Klimiankou M, Klimenkova O *et al*. Interactions among HCLS1, HAX1 and LEF-1 proteins are essential for G-CSF-triggered granulopoiesis. *Nat Med* 2012; **18**: 1550–9.
- 48 Skokowa J, Lan D, Thakur BK *et al*. NAMPT is essential for the G-CSF-induced myeloid differentiation via a NAD(+)-sirtuin-1-dependent pathway. *Nat Med* 2009; **15**: 151–8.
- 49 Solinas G, Marchesi F, Garlanda C, Mantovani A, Allavena P. Inflammation-mediated promotion of invasion and metastasis. *Cancer Metastasis Rev* 2010; **29**: 243–8.
- 50 Taketo MM. Roles of stromal microenvironment in colon cancer progression. *J Biochem* 2012; **151**: 477–81.
- 51 Jensen HK, Donskov F, Marcussen N, Nordmark M, Lundbeck F, von der Maase H. Presence of intratumoral neutrophils is an independent prognostic factor in localized renal cell carcinoma. *J Clin Oncol* 2009; **27**: 4709–17.
- 52 Steidl C, Lee T, Shah SP *et al*. Tumor-associated macrophages and survival in classic Hodgkin's lymphoma. *N Engl J Med* 2010; **362**: 875–85.
- 53 Zhang H, Maric I, DiPrima MJ *et al*. Fibrocytes represent a novel MDSC subset circulating in patients with metastatic cancer. *Blood* 2013; **122**: 1105–13.
- 54 van Deventer HW, Palmieri DA, Wu QP, McCook EC, Serody JS. Circulating fibrocytes prepare the lung for cancer metastasis by recruiting Ly-6C⁺ monocytes via CCL2. *J Immunol* 2013; **190**: 4861–7.
- 55 Hirai H, Fujishita T, Kurimoto K *et al*. CCR1-mediated accumulation of myeloid cells in the liver microenvironment promoting mouse colon cancer metastasis. *Clin Exp Metastasis* 2014; **31**: 977–89.
- 56 Gabrilovich DI, Nagaraj S. Myeloid-derived suppressor cells as regulators of the immune system. *Nat Rev Immunol* 2009; **9**: 162–74.
- 57 Marigo I, Bosio E, Solito S *et al*. Tumor-induced tolerance and immune suppression depend on the C/EBPbeta transcription factor. *Immunity* 2010; **32**: 790–802.
- 58 Lechner MG, Megiel C, Russell SM *et al*. Functional characterization of human Cd33⁺ and Cd11b⁺ myeloid-derived suppressor cell subsets induced from peripheral blood mononuclear cells co-cultured with a diverse set of human tumor cell lines. *J Transl Med* 2011; **9**: 90.
- 59 Sawyers CL. Chronic myeloid leukemia. *N Engl J Med* 1999; **340**: 1330–40.
- 60 Minami Y, Stuart SA, Ikawa T *et al*. BCR-ABL-transformed GMP as myeloid leukemic stem cells. *Proc Natl Acad Sci USA* 2008; **105**: 17967–72.
- 61 Guerzoni C, Bardini M, Mariani SA *et al*. Inducible activation of CEBPB, a gene negatively regulated by BCR/ABL, inhibits proliferation and promotes differentiation of BCR/ABL-expressing cells. *Blood* 2006; **107**: 4080–9.
- 62 Nerlov C. C/EBP alpha mutations in acute myeloid leukaemias. *Nat Rev Cancer* 2004; **4**: 394–400.
- 63 Koschmieder S, Halmos B, Levantini E, Tenen DG. Dysregulation of the C/EBPalpha differentiation pathway in human cancer. *J Clin Oncol* 2009; **27**: 619–28.
- 64 Friedman AD. C/EBPalpha in normal and malignant myelopoiesis. *Int J Hematol* 2015; **111**: 33041.
- 65 Vegesna V, Takeuchi S, Hofmann WK *et al*. C/EBP-beta, C/EBP-delta, PU.1, AML1 genes: mutational analysis in 381 samples of hematopoietic and solid malignancies. *Leuk Res* 2002; **26**: 451–7.
- 66 Watanabe-Okochi N, Yoshimi A, Sato T *et al*. The shortest isoform of C/EBPbeta, liver inhibitory protein (LIP), collaborates with Evl1 to induce AML in a mouse BMT model. *Blood* 2013; **121**: 4142–55.
- 67 Haas SC, Huber R, Gutsch R *et al*. ITD- and FL-induced FLT3 signal transduction leads to increased C/EBPbeta-LIP expression and LIP/LAP ratio by different signalling modules. *Br J Haematol* 2010; **148**: 777–90.
- 68 Guibal FC, Alberich-Jorda M, Hirai H *et al*. Identification of a myeloid committed progenitor as the cancer-initiating cell in acute promyelocytic leukemia. *Blood* 2009; **114**: 5415–25.
- 69 Li J, Zhu H, Hu J *et al*. Progress in the treatment of acute promyelocytic leukemia: optimization and obstruction. *Int J Hematol* 2014; **100**: 38–50.
- 70 Duprez E, Wagner K, Koch H, Tenen DG. C/EBPbeta: a major PML-RARA-responsive gene in retinoic acid-induced differentiation of APL cells. *EMBO J* 2003; **22**: 5806–16.
- 71 Chen X, Liu W, Ambrosino C *et al*. Impaired generation of bone marrow B lymphocytes in mice deficient in C/EBPbeta. *Blood* 1997; **90**: 156–64.
- 72 Yoshioka S, Miura Y, Yao H *et al*. CCAAT/enhancer-binding protein beta expressed by bone marrow mesenchymal stromal cells regulates early B-cell lymphopoiesis. *Stem Cells* 2014; **32**: 730–40.
- 73 Akasaka T, Balasas T, Russell LJ *et al*. Five members of the CEBP transcription factor family are targeted by recurrent IGH translocations in B-cell precursor acute lymphoblastic leukemia (BCP-ALL). *Blood* 2007; **109**: 3451–61.
- 74 Piva R, Pellegrino E, Mattioli M *et al*. Functional validation of the anaplastic lymphoma kinase signature identifies CEBPB and BCL2A1 as critical target genes. *J Clin Invest* 2006; **116**: 3171–82.
- 75 Anastasov N, Bonzheim I, Rudelius M *et al*. C/EBPbeta expression in ALK-positive anaplastic large cell lymphomas is required for cell proliferation and is induced by the STAT3 signaling pathway. *Haematologica* 2010; **95**: 760–7.
- 76 Bergalet J, Fawal M, Lopez C *et al*. HuR-mediated control of C/EBPbeta mRNA stability and translation in ALK-positive anaplastic large cell lymphomas. *Mol Cancer Res* 2011; **9**: 485–96.
- 77 Bonzheim I, Irmeler M, Klier-Richter M *et al*. Identification of C/EBPbeta target genes in ALK⁺ anaplastic large cell lymphoma (ALCL) by gene expression profiling and chromatin immunoprecipitation. *PLoS One* 2013; **8**: e64544.
- 78 Pal R, Janz M, Galson DL *et al*. C/EBPbeta regulates transcription factors critical for proliferation and survival of multiple myeloma cells. *Blood* 2009; **114**: 3890–8.
- 79 Li S, Pal R, Monaghan SA *et al*. IMiD immunomodulatory compounds block C/EBPbeta translation through eIF4E down-regulation resulting in inhibition of MM. *Blood* 2011; **117**: 5157–65.