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Annual variations of carbonaceous PM_{2.5} in Malaysia: influence by Indonesian peatland fires

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Abstract. In this study, we quantified carbonaceous PM_{2.5} in Malaysia through annual observations of PM2.5, focusing on organic compounds derived from biomass burning. We determined organic carbon (OC), elemental carbon and concentrations of solvent-extractable organic compounds (biomarkers derived from biomass burning sources and *n*-alkanes). We observed seasonal variations in the concentrations of pyrolyzed OC (OP), levoglucosan (LG), mannosan (MN), galactosan, syringaldehyde, vanillic acid (VA) and cholesterol. The average concentrations of OP, LG, MN, galactosan, VA and cholesterol were higher during the southwestern monsoon season (June-September) than during the northeastern monsoon season (December-March), and these differences were statistically significant. Conversely, the syringaldehyde concentration during the southwestern monsoon season was lower. The PM2.5 OP / OC4 mass ratio allowed distinguishing the seven samples, which have been affected by the Indonesian peatland fires (IPFs). In addition, we observed significant differences in the concentrations between the Indonesian peatland fire (IPF) and other samples of many chemical species. Thus, the chemical characteristics of PM_{2.5} in Malaysia appeared to be significantly influenced by IPFs during the southwestern monsoon season. Furthermore, we evaluated two indicators, the vanillic acid / syringic acid (VA / SA) and LG / MN mass ratios, which have been suggested as indicators of IPFs. The LG / MN mass ratio ranged from 14 to 22 in the IPF samples and from 11 to 31 in the other samples. Thus, the respective variation ranges partially

overlapped. Consequently, this ratio did not satisfactorily reflect the effects of IPFs in Malaysia. In contrast, the VA / SA mass ratio may serve as a good indicator, since it significantly differed between the IPF and other samples. However, the OP / OC4 mass ratio provided more remarkable differences than the VA / SA mass ratio, offering an even better indicator. Finally, we extracted biomass burning emissions' sources such as IPF, softwood/hardwood burning and meat cooking through varimax-rotated principal component analysis.

1 Introduction

Peatland is a terrestrial wetland ecosystem where organic matter production exceeds its decomposition, resulting in net accumulation (Page et al., 2006). Indonesia has the third largest peatland area and the largest tropical peatland area in the world (270 000 km²; Joosten, 2010). Peatland fires occur predominantly in the Sumatra and Kalimantan islands, Indonesia (Fujii et al., 2014; Page et al., 2002), during the dry season (June–September) mostly due to illegal human activities (Harrison et al., 2009). Because peatland fires are usually underground fires, they are extremely difficult to extinguish. The resulting haze comprises gases and particulates that are emitted because of biomass burning. It extends beyond Indonesia to the neighbouring countries including Malaysia and Singapore (Betha et al., 2014; Engling et al., 2014; Fujii et al., 2015; He et al., 2010; See et al., 2006, 2007), limiting

visibility and causing health problems to the local population (Emmanuel, 2000; Othman et al., 2014; Pavagadhi, et al., 2013; Sahani et al., 2014). Therefore, Indonesian peatland fires (IPFs) have been recognised as an international problem (Yong and Peh, 2014; Varkkey, 2014).

The main constituent of particulates derived from biomass burning is PM2.5 defined as particles having aerodynamic diameters below 2.5 µm, which has been associated with serious health problems (Federal Register, 2006; Schlesinger, 2007). These particulates are primarily composed of organic carbon (OC), which constitutes 50-60% of the total particle mass (Reid et al., 2005). At present, there are only four papers concerning the PM2.5 chemical speciation resulting from IPFs; these papers are based on surface-recorded source-dominated data (Betha et al., 2013; Fujii et al., 2014, 2015a; See et al., 2007). Organic matter is the main component of PM2.5 from IPFs as well as from biomass burning in general (Fujii et al., 2014; See et al., 2007). The primary organic compounds such as cellulose and lignin pyrolysis products have been quantified and potential Indonesian peatland fire (IPF) indicators at the receptor site have been suggested by Fujii et al. (2015a). Additional compounds have been discussed by Betha et al. (2013) (metals) and See et al. (2007) (water-soluble ions, metals and polycyclic aromatic hydrocarbons).

Several studies exist on the chemical characteristics of haze ambient particulates, which have been potentially affected by IPFs in Malaysia and Singapore (e.g. Abas et al., 2004a, b; Betha et al., 2014; Engling et al., 2014; Fang et al., 1999; Fujii et al., 2015b; He et al., 2010; Keywood et al., 2003; Narukawa et al., 1999; Okuda et al., 2002; See et al., 2006; Yang et al., 2013). In most cases, the field observation periods were short. Even when long-term observations have been obtained, however, only typical chemical species such as ions and metals have been analysed. Nevertheless, organic compounds significantly contribute to the IPF aerosols (Fujii et al., 2014). In Malaysia especially, there are no available quantitative data regarding variations of several organic compound concentrations based on long-term observations of $PM_{2.5}$.

The three major sources of air pollution in Malaysia are mobile, stationary and open burning sources including the burning of solid wastes and forest fires (Afroz, et al., 2003). The annual burned biomass in Malaysia has been estimated to be 23 Tg on average (Streets et al., 2003). Therefore, it is necessary to distinguish the effects of IPFs from those of other sources, particularly local biomass burning. Fujii et al. (2015b) reported the total suspended particulate matter (TSP) concentrations in the different carbon fractions (OC1, OC2, OC3, OC4 and pyrolysed OC (OP)) defined by the IM-PROVE_A protocol (Chow et al., 2007) in Malaysia during the haze periods affected by IPFs. They proposed the OP / OC4 mass ratio as a useful indicator of transboundary haze pollution from IPFs at receptor sites even in light haze; the ratio during the haze periods were higher (>4) than during the non-haze periods (<2).

In the present study, the carbonaceous $PM_{2.5}$ components are quantitatively characterised using annual $PM_{2.5}$ observations in Malaysia, with special regard to the organic compounds resulting from biomass burning. Furthermore, the OP / OC4 mass ratio is used as an indicator to investigate the effects of IPFs on carbonaceous $PM_{2.5}$ species in this area. In addition, other indicators that potentially record the effects of IPFs are investigated. Finally, possible carbonaceous $PM_{2.5}$ sources are suggested using varimax-rotated principal component analysis (PCA).

2 Experimental method

2.1 Sampling site and period

The sampling site is the Malaysian Meteorological Department (MMD) located in Petaling Jaya (PJ), Selangor, Malaysia ($\sim 100 \text{ m}$ above sea level; $3^{\circ}06'09'' \text{ N}$, $101^{\circ}38'41'' \text{ E}$). Eighty-one PM_{2.5} samples were collected on the roof of the MMD's main building (eight stories) from August 2011 to July 2012. A detailed description of the sampling site has been provided by Jamhari et al. (2014). In brief, PJ is located in an industrial area (Department of Environment, 2014) $\sim 10 \text{ km}$ from Kuala Lumpur. It is predominantly residential and industrial with high-density road traffic.

2.2 Sample collection and analysis

PM_{2.5} samples were continuously collected with a Tisch high-volume air sampler (model TE-3070V-2.5-BL) on a quartz-fibre filter for 24 h at a flow rate of $1.13 \text{ m}^3 \text{ min}^{-1}$. Before sampling, the quartz-fibre filters were heated to 500 °C for 3 h. After sampling, OC, elemental carbon (EC) and solvent-extractable organic compounds (SEOC; biomarkers derived from biomass burning sources and *n*-alkanes) were measured.

The carbonaceous content was quantified using a DRI model 2001 OC / EC carbon analyser, which employs the thermal optical-reflectance method following the IM-PROVE_A protocol. As shown in our former report (Fujii et al., 2014), the IMPROVE_A temperature protocol defines temperature plateaus for thermally derived carbon fractions as follows: 140 °C for OC1, 280 °C for OC2, 480 °C for OC3 and 580 °C for OC4 in helium (He) carrier gas; 580 °C for EC1, 740 °C for EC2 and 840 °C for EC3 in a mixture of 98 % He and 2 % oxygen (O₂) carrier gas. OC and EC are calculated from the eight carbon fractions as follows:

$$OC = OC1 + OC2 + OC3 + OC4 + OP,$$
 (1)

$$EC = EC1 + EC2 + EC3 - OP,$$
 (2)

where OP is defined as the carbon content measured after the introduction of O_2 until reflectance returns to its initial value

at the start of analysis. Blank corrections were performed on the OC and EC data by subtracting the blank filter value from the loaded filter values.

SEOC obtained from the quartz-fibre filters were quantified by gas chromatography/mass spectrometry (GC/MS). Biomarker organic compound speciation was accomplished following the procedures reported previously (Fujii et al., 2015a, b). To quantify *n*-alkanes, aliquots from the quartzfibre filter were spiked with internal standards of eicosane d_{42} and triacontane- d_{62} before extraction. Each spiked filter was extracted by ultrasonic agitation for 2×20 min periods using 8 mL hexane (Kanto Chemical, purity > 96.0%). The combined extracts were filtered through a polytetrafluoroethylene syringe filter (pore size 0.45 µm), dried completely under a gentle stream of nitrogen gas and re-dissolved to 0.1 mL in hexane. Before the GC/MS analysis, $\sim 1.05 \,\mu g$ of tetracosane- d_{50} dissolved in 50 µL of hexane was added as a second internal standard. The n-alkanes values were reported in carbon numbers, ranging from 22 to 33 (C_{22} – C_{33}). The extract samples were analysed on a Shimadzu GC/MS system (GCMS-QP2010-Plus, Shimadzu) equipped with a 30 m HP-5MS column (0.25 µ m film thickness, 0.25 mm ID). The carrier gas was helium (purity >99.9%) at a pressure of 73.0 kPa (37.2 cm s⁻¹ at 100 °C). The GC oven temperature program was as follows: isothermal at 100 °C for 5 min, 100–300 °C at 10 °C min⁻¹ and then 300 °C for 20 min. The injection port and transfer line were maintained at 300 °C. The data for quantitative analysis were acquired in the electron impact mode (70 eV). The mass spectrometer was operated under the selected ion-monitoring scanning mode, and the monitored ions for the quantification of *n*-alkanes were 85 m/z. The monitored ions corresponding to the internal standards were 66 m/z. The recovery ratios for known amounts of n-alkane standards (1 µg addition) on the quartz-fibre filters ranged from 73 to 110% (mean \pm standard deviation: 94 \pm 6.3 %). Blank corrections were performed on the biomarker and *n*-alkane data by subtracting the blank filter value from the loaded filter values.

2.3 Source apportionment method

Varimax-rotated PCA was used to identify the possible carbonaceous PM_{2.5} sources at PJ. The following two data sets were considered: (i) PJ_A data, which includes 25 variables (all quantified compounds) and 81 samples (all samples), and (ii) PJ_S data, which includes 25 variables and 65 samples (excluded are the samples acquired in September 2011 and June 2012, which are influenced by IPFs as shown in Sect. 3). PCA results with these data sets are expected to show IPF effects on other sources. It has been suggested that the minimum number of samples (*n*) for factor analysis should satisfy the following condition (Henry et al., 1984; Karar and Gupta, 2007):

$$n > 30 + \frac{V+3}{2},\tag{3}$$

where V represents the number of variables. Both data sets satisfy this condition.

Varimax-rotated PCA followed the procedure proposed by Karar and Gupta (2007) and was accomplished with the R-software (http://www.R-project.org). The eigenvalues correspond to the number of factors, which was selected to ensure that the cumulative variance contribution rate is greater than 80 %.

3 Results and discussion

3.1 Air quality and monthly hotspot data

Figure 1 presents the daily variability of the Malaysian Air Pollutant Index (MAPI) and visibility during the sampling periods. The MAPI data were obtained from the Department of Environment Ministry of Natural Resources and Environment website (http://apims.doe.gov.my/apims/hourly2.php, last access: 10 March 2015). Hourly visibility data (07:00-17:00 local time) provided by the MMD were used to produce the daily variation in visibility after removing the hourly data corresponding to periods of rainfall. The MAPI values of 0-50, 51-100, 101-200, 201-300 and > 300 correspond to good, moderate, unhealthy, very unhealthy and hazardous air quality conditions (Department of Environment, 2014; Fujii et al., 2015b). Good MAPI levels dominate the sampling periods except in August 2011, September 2011 and June 2012. However, moderate air quality is observed in August 2011, September 2011 and June 2012. The two MAPI values for 15 and 16 June 2012 indicate unhealthy air quality conditions. The average visibility during these two sampling periods (Fig. 1) was below 2.7 km, corresponding to extremely low visibility compared with other intervals.

Figure 2 presents the monthly hotspot counts on Sumatra Island detected by the NOAA-18 satellite (Indofire, http://www.indofire.org/indofire/hotspot, last access: 17 July 2013). During the southwestern monsoon season in September 2011 and June 2012, hotspots exceeded 3000 on several occasions. The hotspot counts in September 2011 and June 2012 mainly derived from the South Sumatra (60 % of the hotspot counts) and the Riau (42 %) provinces respectively. The sampling site is predominantly downwind of the Sumatra Island during the southwestern monsoon season. Thus, some samples have probably been affected by IPFs. The 3-day backward air trajectories for the sampling periods (Fig. S1 in the Supplement) support this conclusion.



Figure 1. Daily variability of the MAPI and visibility during the sampling periods.



Figure 2. Monthly hotspot counts in the Sumatra Island.

3.2 PM_{2.5} chemical characteristics and seasonal variations

3.2.1 OC and EC

The annual average concentrations of OC and EC are 7.0 ± 5.4 and $3.1 \pm 1.1 \,\mu g \, C \, m^{-3}$ respectively. The OC and EC concentrations' statistical results for each monsoon season appear in Table 1. The average OC concentration during the southwestern monsoon season (June-September) is higher than that during other seasons. In particular, an extremely high OC concentration (>25 μ g m⁻³) is observed on 12 September 2011 and on 15 and 16 June 2012. There is no statistically significant difference in the EC concentration between the southwestern and northeastern (December-March) monsoon seasons according to the two-sided Wilcoxon rank sum test (p value: p > 0.05) with R software. In Bangi (\sim 30 km southeast of the sampling site), the OC concentration was $11 \pm 3.2 \,\mu\text{g}\,\text{C}\,\text{m}^{-3}$ in September 2013 (Fujii et al., 2015c), in good agreement with the present results for the southwestern monsoon season. The OC / EC mass ratios during the southwestern monsoon, post-monsoon (October-November), northeastern monsoon and pre-monsoon (AprilMay) season range among 1.2-6.5, 1.4-2.4, 0.99-3.0 and 1.2-2.3 respectively. A high OC / EC mass ratio value (>4) is found only for some samples collected in September 2011 and June 2012. These values have probably been affected by biomass burning, because aerosols emitted from biomass burning usually present higher OC / EC mass ratios (Cong et al., 2015).

The daily variations of the OC fractions' mass concentrations during the sampling periods are presented in Fig. 3. The annual average concentrations of OC1, OC2, OC3, OC4 and OP are 0.51 ± 0.80 , 1.9 ± 1.1 , 2.3 ± 1.4 , 1.2 ± 0.36 and $1.1 \pm 2.2 \,\mu g \, m^{-3}$ respectively. Statistically significant differences among the OP concentrations during the southwestern and northeastern monsoon seasons are observed according to the two-sided Wilcoxon rank sum test (p < 0.001). In particular, high OP concentrations are clearly observed in September 2011 and June 2012, in addition to the higher OC / EC mass ratios described above. Fujii et al. (2015b) supported that the enhanced OP concentrations in TSP, which are observed in Malaysia during the haze periods, are affected by the IPFs. The enhanced OP concentrations in PM_{2.5} during the southwestern monsoon season, which are observed in the present study, are also probably affected by IPFs from Sumatra Island. The increased number of hotspots recorded (Fig. 2) and backward air trajectories (Fig. S1) further support this conclusion.

3.2.2 Biomarkers

Ten biomarkers are identified in this study that have been suggested as indicators of biomass burning processes such as wood burning and meat cooking. The annual average concentrations of levoglucosan (LG), mannosan (MN), galactosan, *p*-hydroxybenzoic acid, vanillic acid (VA) and syringic acid (SA) are 86 ± 95 , 4.8 ± 5.7 , 1.2 ± 1.6 , 1.1 ± 1.3 , 0.19 ± 0.28 and 0.25 ± 0.28 ng m⁻³ respectively; notably, they exhibit great variability. The annual average concentrations of vanillin, syringaldehyde, dehydroabietic acid

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Table 1. Statistical results of chemical species concentrations. Av is the average; SD is the standard deviation.

	Southwester (June-Sep	rn monsoon ptember)	Post-monsoon (October–November)		Northeasterr (December	Northeastern monsoon (December–March)		Pre-monsoon (April–May)	
Compounds	$Av \pm SD$	Range	$Av \pm SD$	Range	$Av \pm SD$	Range	$Av \pm SD$	Range	
OC and EC ($\mu gm^{-3})$									
OC	10 ± 7.8	3.6-36	5.6 ± 2.4	2.5-11	5.2 ± 1.4	2.7-8.2	4.2 ± 1.4	2.8-7.3	
EC	3.0 ± 0.95	1.0-5.6	3.2 ± 1.3	1.1–5.9	3.4 ± 1.1	1.6-6.1	2.6 ± 1.2	1.4-4.5	
Biomarkers (ng m $^{-3}$)									
Levoglucosan	160 ± 130	32-490	64 ± 39	19–130	40 ± 14	17-64	49 ± 21	23-86	
Mannosan	8.4 ± 8.2	1.5-30	3.4 ± 2.6	0.95-9.1	2.6 ± 1.2	0.84-5.3	2.5 ± 1.2	1.2-5.3	
Galactosan	2.3 ± 2.3	0.38-8.3	0.86 ± 0.72	0.29 - 2.8	0.60 ± 0.35	0.13-1.3	0.62 ± 0.34	0.33-1.5	
p-Hydroxybenzoic acid	1.9 ± 1.9	0.18 - 7.5	0.79 ± 0.67	0.036-2.2	0.64 ± 0.30	0.20 - 1.2	0.50 ± 0.25	0.24 - 1.0	
Vanillin	1.6 ± 1.1	0.54-5.5	1.2 ± 0.66	0.45 - 2.2	1.0 ± 0.38	0.21 - 1.7	0.96 ± 0.42	0.30 - 1.7	
Syringaldehyde	0.29 ± 0.22	0.085 - 1.0	0.59 ± 0.22	0.26 - 1.2	0.77 ± 0.54	0.074 - 2.2	0.36 ± 0.22	0.093-0.77	
Vanillic acid	0.39 ± 0.39	0.074 - 1.9	0.11 ± 0.070	0.031-0.22	0.073 ± 0.057	0.013-0.26	0.066 ± 0.027	0.034-0.12	
Syringic acid	0.35 ± 0.41	0.075-2.4	0.26 ± 0.21	0.058-0.59	0.17 ± 0.13	0.029-0.64	0.16 ± 0.084	0.049-0.28	
Dehydroabietic acid	1.7 ± 1.1	0.10-5.4	1.1 ± 0.69	0.31-2.4	1.1 ± 1.1	0.14-4.6	0.67 ± 0.24	0.16-0.98	
Cholesterol	1.8 ± 0.82	0.50-3.7	1.2 ± 0.51	0.57 - 2.0	0.98 ± 0.51	0.026-2.0	1.3 ± 0.56	0.51-2.0	
<i>n</i> -Alkanes (ng m ^{-3})									
Docosane	3.2 ± 0.82	1.8-5.0	2.9 ± 0.61	2.0-4.0	3.0 ± 0.53	1.9-4.2	4.0 ± 4.8	2.1–19	
Tricosane	3.6 ± 1.2	2.0-7.2	3.2 ± 0.91	2.0-4.8	3.2 ± 0.65	1.8-4.4	5.0 ± 7.6	2.1-29	
Tetracosane	5.8 ± 3.2	2.5-19	5.7 ± 1.7	3.3-8.7	6.1 ± 2.3	2.9-15	6.3 ± 8.5	2.7-33	
Pentacosane	8.9 ± 6.7	3.5-34	5.7 ± 2.3	3.1-11	6.0 ± 1.6	3.7-9.2	5.8 ± 5.5	3.2-23	
Hexacosane	13 ± 9.8	4.3-49	8.6 ± 3.7	3.6-18	9.7 ± 2.8	5.0-16	7.1 ± 5.3	3.5-23	
Heptacosane	16 ± 14	4.7-64	7.2 ± 2.6	3.6-12	8.2 ± 2.4	3.7-14	5.8 ± 3.4	3.3-16	
Octacosane	12 ± 12	2.6-54	4.3 ± 1.8	1.7-7.9	5.9 ± 3.0	2.3-17	3.6 ± 1.7	2.3-8.2	
Nonacosane	13 ± 13	3 0-55	49 + 21	15-87	63 + 22	3 3-13	45 ± 14	2.6-7.8	
Triacontane	79 ± 78	2.0-36	38 ± 2.0	16-90	52 ± 2.2	2.0-16	33+17	17-83	
Hentriacontane	14 ± 14	2.8-59	48 ± 1.0	18-84	5.2 ± 2.7 5.7 ± 2.0	3 3-11	43+12	29-69	
Dotriacontane	67 ± 55	1.6-27	34+0.72	2 4-4 5	46 ± 13	28-78	31 ± 0.88	18-44	
Tritriacontane	6.7 ± 3.3 6.8 ± 7.1	1.0 27	2.5 ± 0.97	1 1-4 2	2.8 ± 0.92	1 2-5 0	2.1 ± 0.00	1.5-3.8	
munucontaile	0.0 ± 7.1	1.2 33	2.5 ± 0.57	1.1 1.2	2.0 ± 0.92	1.2 5.0	2.1 ± 0.72	1.5 5.6	
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Figure 3. Daily variation of the OC fractions' mass concentrations during the sampling periods.

and cholesterol are 1.2 ± 0.80 , 0.51 ± 0.42 , 1.3 ± 1.0 and 1.3 ± 0.72 ng m⁻³ respectively. The biomarker statistical results for each monsoon season are listed in Table 1.

LG is a specific indicator for cellulose burning emissions and is generally formed during cellulose pyrolysis at temperatures above 300 °C (Fujii et al., 2015b; Lin et al., 2010; Shafizadeh, 1984; Simoneit et al., 1999). The MN and galactosan are derived from hemicellulose pyrolysis products; they can also be used as tracers of biomass burning besides LG (e.g. Engling et al., 2014; Fujii et al., 2014, 2015b; Zhu et al., 2015). Statistically significant differences are observed among the concentrations of LG, MN and galactosan obtained during the southwestern and northeastern monsoon seasons on the basis of the two-sided Wilcoxon rank sum test (p < 0.001); high concentrations of these compounds are mostly observed during the southwestern monsoon season (especially September 2011 and June 2012; Fig. S2). In Singapore, Engling et al. (2014) suggested that the enhanced concentrations of these compounds during the haze periods were due to the IPFs during the southwestern monsoon season. Thus, the presently observed enhanced concentrations of these compounds may also be attributed to the IPFs.

In a previous report, $PM_{2.5}$ lignin unit-originating compounds in samples collected at the IPF source were quantified (Fujii et al., 2015a). Lignin is an aromatic polymer consisting of phenylpropane units linked through many ether and C-C linkages. Its aromatic structure varies depending on the species; softwood lignins exclusively contain guaiacyl(G) types and hardwood ligning include both G and syringyl (S) types, whereas herbaceous plants include G, S and *p*-hydroxyphenyl (*H*) types (Fujii et al., 2015a, b). The composition of these aromatic nuclei within the lignin pyrolysis products resulting from biomass burning may be useful in identifying the biomass type (Fujii et al., 2015a; Simoneit et al., 1993). In the present study, vanillin and VA (compounds derived from G units), syringaldehyde and SA (compounds derived from S units) as well as and p-hydroxybenzoic acid (compounds derived from H units or the secondary decomposition of G and S units) (Fujii et al., 2015b) have been quantified. There are significant differences between the concentrations of syringaldehyde and VA derived from lignin pyrolysis during the southwestern and northeastern monsoon seasons on the basis of the two-sided Wilcoxon rank sum test (p < 0.001), corresponding to seasonal variations. The average VA concentration during the southwestern monsoon season is 5.3 times greater than that during the northeastern monsoon season. In contrast, the average concentration of syringaldehyde during the northeastern monsoon season is 2.6 times greater than that during the southwestern monsoon season. This may be due to the transboundary pollution by prevailing winds from the Chinese region including Thailand and Vietnam during the northeastern monsoon season (Fig. S1; Khan et al., 2015).

Dehydroabietic acid and cholesterol are quantified as indicators of softwood burning and meat cooking respectively (Fujii et al., 2015b; Lin et al., 2010). The two-sided Wilcoxon rank sum test indicates that the difference between the cholesterol concentration during the southwestern and northeastern monsoon seasons is statistically significant (p < 0.001). The dehydroabietic acid and cholesterol concentrations recorded in the interval between June and July 2014 in Bangi, which is located ~ 30 km southeast of the sampling site, range between 2.6–8.7 and 1.5–5.7 ng m⁻³ respectively (Fujii et al., 2015b). The PJ industrial area's concentrations of these compounds are lower than those in the Bangi suburban area owing to the decreased impact of softwood burning and meat cooking in PJ.

3.2.3 N-alkanes

The total annual average concentrations of *n*-alkanes is 79 ± 63 ng m⁻³. The total *n*-alkanes concentrations during the southwestern monsoon, post-monsoon, northeastern monsoon and pre-monsoon seasons are 110 ± 93 , 57 ± 20 , 67 ± 18 and 55 ± 41 ng m⁻³ respectively. The highest concentration is observed during the southwestern monsoon season. Figure 4 illustrates the molecular distribution of *n*-alkanes during the southwestern and northeastern monsoon seasons. There are no significant differences among the concentrations of C₂₂-C₂₆, C₂₉, C₃₀ and C₃₂ in the two seasons (*p* > 0.05). High concentrations of > C₂₄ are mainly observed



Figure 4. Box-and-whisker plots of molecular distributions of n-alkanes during the (**a**) southwestern and (**b**) northeastern monsoon seasons. The horizontal lines in the box represent the 25th, 50th and 75th percentiles. The whiskers represent the 10th and 90th percentiles.

in September 2011 and June 2012 when many hotspots are detected in the Sumatra Island (Fig. 2). Fujii et al. (2015a) suggested that IPFs increase the C_{27} , C_{28} and C_{29} concentrations in $PM_{2.5}$ at the receptor site relative to other sources such as vehicle and biomass burning. Thus, the enhanced *n*-alkanes concentrations in $PM_{2.5}$ during the southwestern monsoon season may be mainly attributed to IPFs.

The carbon number maximum (C_{max}) in *n*-alkanes during the southwestern and northeastern monsoon seasons is C_{27} (in 83 % of the samples) and C_{26} (89 %) respectively (Fig. 5). Reported C_{max} values range from 27 to 33, characteristic of biogenic sources (higher plant wax), whereas lower C_{max} values may indicate major petrogenic input (Abas et al., 2004a; Gogou et al., 1996; He et al., 2010). The C_{max} during the southwestern monsoon season (C_{27}) suggests primarily biogenic sources and is in perfect agreement with the measured value for the IPF source (Fujii et al., 2015b).

The carbon preference index (CPI) has been widely used to roughly estimate the effects of anthropogenic or biogenic sources (e.g. Bray and Evans, 1961; Chen et al., 2014; He et al., 2010; Yamamoto et al., 2013). The CPI values are calculated by the following equation based on the suggestion by Bray and Evans (1961).



Figure 5. Number fraction of C_{max} in the $PM_{2.5}$ samples for each monsoon season.

$$CPI = 0.5 \times \left(\frac{C_{25} + C_{27} + C_{29} + C_{31}}{C_{26} + C_{28} + C_{30} + C_{32}} + \frac{C_{25} + C_{27} + C_{29} + C_{31}}{C_{24} + C_{26} + C_{28} + C_{30}} \right)$$
(4)

The CPI values are generally high (CPI > 5) when there is no serious input from fossil fuel hydrocarbons (CPI = 1) (Yamamoto et al., 2013, and references therein). The CPI values during the southwestern and northeastern monsoon seasons are 1.3 ± 0.12 and 1.0 ± 0.14 respectively; these values are close to one for both seasons, indicating an anthropogenic *n*alkane source. Thus, the CPI value is not susceptible to IPF influence, since the CPI value at the IPF source is 1.6 ± 0.13 (Fujii et al., 2015a), which is not high. Consequently, the CPI cannot be used to identify IPF sources at a receptor site.

3.3 Indonesian peatland fire effect

The hotspot data and backward air trajectories suggest that IPFs strongly modify many chemical species concentrations mostly during the southwestern monsoon season. However, IPFs do not always occur during the southwestern monsoon season. Therefore, significant differences in some chemical species concentrations among samples affected by IPF and other sources should be observed. To distinguish IPF samples from other samples obtained during the southwestern monsoon season, the OP / OC4 mass ratio is used, which is a useful indicator for IPF (Fujii et al., 2015b). The ratio value is >4 for seven samples (11–13 September 2011 and 14–17 June 2012); these samples are regarded as the IPF samples. The OP / OC4 mass ratio for the IPF and other samples is 7.4 ± 3.4 and 0.44 ± 0.49 respectively, exhibiting significant differences among them according to the two-



Figure 6. *P* values to determine significance in the two-sided Wilcoxon rank sum test between the IPF and other samples.

sided Wilcoxon rank sum test (p < 0.001). Figure 6 shows the *p* values used to determine the statistical significance in a hypothesis test of the differences between the IPF and other samples for all the quantified species. Significant differences (p < 0.001) are recorded for many chemical species. Thus, the chemical characteristics of PM_{2.5} in Malaysia are significantly influenced by IPFs.

Furthermore, the VA / SA and LG / MN mass ratios in the IPF source are investigated as potential indicators, as suggested in previous studies (Fujii et al., 2014, 2015a). The VA / SA mass ratio for IPF and other samples is 1.7 ± 0.36 and 0.59 ± 0.27 respectively, providing a good indicator (p < 0.001). Although the VA / SA mass ratio at the IPF source is 1.1 ± 0.16 (Fujii et al., 2015a), the ratios for IPF samples are higher. Opsahl and Benner (1998) reported photochemical reactivity of VA and SA in the Mississippi River water. They demonstrated that the early degradation of SA in the water is mostly due to its higher photochemical reactivity compared with VA. Even though there are no reports of such degradations in air, SA is considered to be less stable than VA in air as well as in water, which leads to an increased VA / SA ratio after long-range transportation. The LG / MN mass ratio for the IPF and other samples ranges from 14 to 22 and 11 to 31 respectively (Fig. S3). Therefore, the LG / MN mass ratio is inappropriate to extract the effects of IPF in Malaysia, because its value's ranges in the IPF and other samples partially overlap.

The daily variability of the C_{27} and LG concentration as well as the VA / SA and OP / OC4 mass ratios are presented in Fig. 7; similar trends are observed in all cases. However, the concentrations of LG, MN and galactosan (Fig. S2) increase abruptly on 10 August 2011, although this sample is not categorised as an IPF sample. We hypothesised that this increase results from local biomass burning, since LG emissions are produced by several different biomass burning sources (Oros and Simoneit, 2001a, b; Oros et al., 2006). Therefore, LG levels are not directly indicative of the IPF



Figure 7. Daily variability of the C₂₇ and LG concentration as well as the VA / SA and OP / OC4 mass ratios during the sampling periods.

contribution in Malaysia; instead, C_{27} may be a useful indicator (Fig. 7). Although the VA / SA mass ratio can be used as an IPF indicator, as we mentioned before, the OP / OC4 mass ratio highlights the differences between the IPF and other samples better than the VA / SA mass ratio (Fig. 7).

3.4 Carbonaceous PM_{2.5} contributions

The possible sources of carbonaceous PM2.5 are investigated through varimax-rotated PCA of the PJ A and PJ S data sets. Over 80% of the cumulative variance in the PJ A and PJ_S data sets is explained by three and five factors respectively (Table 2). For the PJ_A data (Table 2a), the total variance explained by the three factors is 80%. Factor A1, which explains 60 % of the variance, is heavily loaded (loading factor: >0.65) with OC, LG, MN, galactosan, phydroxybenzoic acid, VA and C₂₅–C₃₃, which direct towards an IPF source. Factor A2, which corresponds to 12% of the variance, is heavily loaded with C22-C24, suggesting a petrogenic source (Abas et al., 2004a; Gogou et al., 1996; He et al., 2010). Factor A3, which explains 8.0% of the variance in the data set, is heavily loaded with SA and dehydroabietic acid, indicating mixed (softwood and hardwood) biomass burning sources. For the PJ S data set (Table 2b), the total variance explained by five factors is 82%. Factor S1 explains 43% of the data's variance and is heavily loaded with C_{27} - C_{33} , which suggests tire wear emission (Rogge et al., 1993). Factor S2 explains 19% of the variance and is heavily loaded with LG, MN, galactosan, VA and SA, which correspond to a biomass burning source. Factor S3, which explains 11%

of the variance, is heavily loaded with $C_{22}-C_{26}$, which indicate a petrogenic source, similar to factor A2. Although heavy loading with only syringaldehyde is found in factor S4 (5.0% of the variance), its source could not be identified. Finally, factor S5 explains 4.5% of the variance and is heavily loaded with EC and cholesterol, which are produced when cooking meat.

Differences of the factor loadings between PJ_A and PJ_S data are observed. For the PCA result of the PJ_A data set, the factors such as tire wear (factor S1) and cooking (factor S5) as shown in Table 2b are not extracted due to the strong influence of the IPFs. Although a petrogenic source is identified from both results, C_{25} and C_{26} are not heavily loaded for the PJ_A data set. This is also considered to be due to the strong influence of the IPFs.

Wahid et al. (2013) reported varimax-rotated PCA results on the distribution of inorganic ions within fine-mode aerosols (< 1.5 µm) at Kuala Lumpur, which is close to the present study's sampling site (\sim 10 km). They extracted three principal components from this analysis: (1) motor vehicles, (2) soil and the earth's crust and (3) sea spray. Jamhari et al. (2014) applied varimax-rotated PCA on polycyclic aromatic hydrocarbon data in PM₁₀ at Kuala Lumpur. They extracted two factors, which were attributed to (1) natural gas emission and coal combustion and (2) vehicles and gasoline emissions. In the present study, only biomass burning could be identified as a factor through comparison with these previous analyses. Factors such as soil, sea spray and coal combustion could not be identified, because the key inor-

Table 2. (a) Factor loadings from varimax-rotated	PCA	of PJ_A
data. A1-A3 indicate factors. (b) Factor loadings	from	varimax-
rotated PCA of PJ_S data. S1–S5 indicate factors.		

(a)	A1	A2	A3		
OC	0.97	0.10	0.16		
EC	0.29	0.37	0.51		
Levoglucosan	0.81	-0.05	0.17		
Mannosan	0.89	0.00	0.11		
Galactosan	0.90	0.02	0.08		
p-Hydroxybenzoic acid	0.94	0.04	0.22		
Vanillin	0.61	0.15	0.25		
Syringaldehyde	-0.17	0.12	0.40		
Vanillic acid	0.65	-0.10	0.55		
Syringic acid	0.28	-0.11	0.81		
Dehydroabietic acid	0.15	-0.01	0.86		
Cholesterol	0.36	0.14	0.39		
C ₂₂	0.03	0.95	0.05		
C ₂₃	0.07	0.95	0.05		
C ₂₄	0.30	0.92	0.06		
C ₂₅	0.81	0.54	0.14		
C ₂₆	0.86	0.43	0.13		
C ₂₇	0.95	0.23	0.13		
C ₂₈	0.96	0.18	0.07		
C ₂₉	0.97	0.13	0.12		
C ₃₀	0.92	0.25	0.05		
C ₃₁	0.97	0.10	0.13		
C ₃₂	0.93	0.15	0.11		
C ₃₃	0.97	0.10	0.13		
% variance	60	12	8.0		
% cumulative	60	72	80		
(b)	S 1	S2	S 3	S4	S5
(b) OC	S1 0.47	S2 0.47	S3 0.10	S4	85 0.57
(b) OC EC	S1 0.47 0.39	S2 0.47 0.20	\$3 0.10 0.25	S4 0.08 0.26	S5 0.57 0.65
(b) OC EC Levoglucosan	S1 0.47 0.39 0.09	S2 0.47 0.20 0.71	S3 0.10 0.25 -0.03	S4 0.08 0.26 -0.52	S5 0.57 0.65 0.19
(b) OC EC Levoglucosan Mannosan	S1 0.47 0.39 0.09 0.19	S2 0.47 0.20 0.71 0.84	S3 0.10 0.25 -0.03 0.02	S4 0.08 0.26 -0.52 -0.26	S5 0.57 0.65 0.19 0.28
(b) OC EC Levoglucosan Mannosan Galactosan	S1 0.47 0.39 0.09 0.19 0.17	S2 0.47 0.20 0.71 0.84 0.83	S3 0.10 0.25 -0.03 0.02 0.06	S4 0.08 0.26 -0.52 -0.26 -0.09	S5 0.57 0.65 0.19 0.28 0.41
(b) OC EC Levoglucosan Mannosan Galactosan <i>p</i> -Hydroxybenzoic acid	S1 0.47 0.39 0.09 0.19 0.17 0.26	S2 0.47 0.20 0.71 0.84 0.83 0.62	S3 0.10 0.25 -0.03 0.02 0.06 0.08	S4 0.08 0.26 -0.52 -0.26 -0.09 0.23	S5 0.57 0.65 0.19 0.28 0.41 0.42
(b) OC EC Levoglucosan Mannosan Galactosan <i>p</i> -Hydroxybenzoic acid Vanillin	S1 0.47 0.39 0.09 0.19 0.17 0.26 0.22	S2 0.47 0.20 0.71 0.84 0.83 0.62 0.32	S3 0.10 0.25 -0.03 0.02 0.06 0.08 0.07	S4 0.08 0.26 -0.52 -0.26 -0.09 0.23 0.05	S5 0.57 0.65 0.19 0.28 0.41 0.42 0.61 0.61
(b) OC EC Levoglucosan Mannosan Galactosan <i>p</i> -Hydroxybenzoic acid Vanillin Syringaldehyde	S1 0.47 0.39 0.09 0.19 0.17 0.26 0.22 0.24	S2 0.47 0.20 0.71 0.84 0.83 0.62 0.32 0.13	S3 0.10 0.25 -0.03 0.02 0.06 0.08 0.07 0.01	S4 0.08 0.26 -0.52 -0.26 -0.09 0.23 0.05 0.74	S5 0.57 0.65 0.19 0.28 0.41 0.42 0.61 0.07
(b) OC EC Levoglucosan Mannosan Galactosan <i>p</i> -Hydroxybenzoic acid Vanillin Syringaldehyde Vanillic acid	S1 0.47 0.39 0.09 0.19 0.17 0.26 0.22 0.24 -0.12	S2 0.47 0.20 0.71 0.84 0.83 0.62 0.32 0.13 0.81	S3 0.10 0.25 -0.03 0.02 0.06 0.08 0.07 0.01 -0.04	$\begin{array}{c} S4\\ 0.08\\ 0.26\\ -0.52\\ -0.26\\ -0.09\\ 0.23\\ 0.05\\ 0.74\\ 0.22\\ \end{array}$	85 0.57 0.65 0.19 0.28 0.41 0.42 0.61 0.07
(b) OC EC Levoglucosan Mannosan Galactosan <i>p</i> -Hydroxybenzoic acid Vanillin Syringaldehyde Vanillic acid Syringic acid	S1 0.47 0.39 0.09 0.19 0.17 0.26 0.22 0.24 -0.12 0.02	S2 0.47 0.20 0.71 0.84 0.83 0.62 0.32 0.13 0.81 0.81	S3 0.10 0.25 -0.03 0.02 0.06 0.08 0.07 0.01 -0.04 0.00	$\begin{array}{c} 84\\ 0.08\\ 0.26\\ -0.52\\ -0.26\\ -0.09\\ 0.23\\ 0.05\\ 0.74\\ 0.22\\ 0.37\\ \end{array}$	85 0.57 0.65 0.19 0.28 0.41 0.42 0.61 0.07 -0.01 0.26
(b) OC EC Levoglucosan Mannosan Galactosan <i>p</i> -Hydroxybenzoic acid Vanillin Syringaldehyde Vanillic acid Syringic acid Dehydroabietic acid	S1 0.47 0.39 0.09 0.19 0.17 0.26 0.22 0.24 -0.12 0.02 0.18	S2 0.47 0.20 0.71 0.84 0.83 0.62 0.32 0.13 0.81 0.81 0.44	S3 0.10 0.25 -0.03 0.02 0.06 0.08 0.07 0.01 -0.04 0.00 0.04	$\begin{array}{c} 84\\ 0.08\\ 0.26\\ -0.52\\ -0.26\\ -0.09\\ 0.23\\ 0.05\\ 0.74\\ 0.22\\ 0.37\\ 0.12\\ \end{array}$	$\begin{array}{c} 85\\ \hline 0.57\\ 0.65\\ 0.19\\ 0.28\\ 0.41\\ 0.42\\ 0.61\\ 0.07\\ -0.01\\ 0.26\\ 0.60\\ \end{array}$
(b) OC EC Levoglucosan Mannosan Galactosan <i>p</i> -Hydroxybenzoic acid Vanillin Syringaldehyde Vanillic acid Syringic acid Dehydroabietic acid Cholesterol	S1 0.47 0.39 0.19 0.17 0.26 0.22 0.24 -0.12 0.02 0.18 0.01	S2 0.47 0.20 0.71 0.84 0.83 0.62 0.32 0.13 0.81 0.81 0.44 0.17	$\begin{array}{c} \text{S3} \\ 0.10 \\ 0.25 \\ -0.03 \\ 0.02 \\ 0.06 \\ 0.08 \\ 0.07 \\ 0.01 \\ -0.04 \\ 0.00 \\ 0.04 \\ 0.15 \end{array}$	$\begin{array}{c} 84\\ 0.08\\ 0.26\\ -0.52\\ -0.26\\ -0.09\\ 0.23\\ 0.05\\ 0.74\\ 0.22\\ 0.37\\ 0.12\\ -0.21\\ \end{array}$	S5 0.57 0.65 0.19 0.28 0.41 0.42 0.61 0.07 -0.01 0.26 0.60 0.77
(b) OC EC Levoglucosan Mannosan Galactosan <i>p</i> -Hydroxybenzoic acid Vanillin Syringaldehyde Vanillic acid Syringic acid Dehydroabietic acid Cholesterol C ₂₂	\$1 0.47 0.39 0.19 0.17 0.26 0.22 0.24 -0.12 0.02 0.18 0.01 0.05	$\begin{array}{c} \text{S2} \\ 0.47 \\ 0.20 \\ 0.71 \\ 0.84 \\ 0.83 \\ 0.62 \\ 0.32 \\ 0.13 \\ 0.81 \\ 0.81 \\ 0.44 \\ 0.17 \\ -0.02 \end{array}$	$\begin{array}{c} \text{S3} \\ 0.10 \\ 0.25 \\ -0.03 \\ 0.02 \\ 0.06 \\ 0.08 \\ 0.07 \\ 0.01 \\ -0.04 \\ 0.00 \\ 0.04 \\ 0.15 \\ 0.97 \end{array}$	$\begin{array}{c} 84\\ 0.08\\ 0.26\\ -0.52\\ -0.26\\ 0.09\\ 0.23\\ 0.05\\ 0.74\\ 0.22\\ 0.37\\ 0.12\\ -0.21\\ -0.04 \end{array}$	\$5 0.57 0.65 0.19 0.28 0.41 0.42 0.61 0.07 -0.01 0.26 0.60 0.77 0.05
(b) OC EC Levoglucosan Mannosan Galactosan <i>p</i> -Hydroxybenzoic acid Vanillin Syringaldehyde Vanillic acid Syringic acid Dehydroabietic acid Cholesterol C ₂₂ C ₂₃	\$1 0.47 0.39 0.09 0.17 0.26 0.22 0.24 -0.12 0.02 0.18 0.01 0.05 0.05	$\begin{array}{c} S2\\ 0.47\\ 0.20\\ 0.71\\ 0.84\\ 0.83\\ 0.62\\ 0.32\\ 0.13\\ 0.81\\ 0.81\\ 0.44\\ 0.17\\ -0.02\\ 0.00\\ \end{array}$	S3 0.10 0.25 -0.03 0.06 0.08 0.07 0.01 -0.04 0.00 0.04 0.15 0.97	$\begin{array}{c} 84\\ 0.08\\ 0.26\\ -0.52\\ -0.26\\ 0.09\\ 0.23\\ 0.05\\ 0.74\\ 0.22\\ 0.37\\ 0.12\\ -0.21\\ -0.04\\ -0.04\\ \end{array}$	\$5 0.57 0.65 0.19 0.28 0.41 0.42 0.61 0.07 -0.01 0.26 0.60 0.77 0.05 0.04
(b) OC EC Levoglucosan Mannosan Galactosan <i>p</i> -Hydroxybenzoic acid Vanillin Syringaldehyde Vanillic acid Syringic acid Dehydroabietic acid Cholesterol C ₂₂ C ₂₃ C ₂₄	S1 0.47 0.39 0.09 0.17 0.26 0.22 0.24 -0.12 0.02 0.18 0.01 0.05 0.05 0.28	$\begin{array}{c} \text{S2} \\ 0.47 \\ 0.20 \\ 0.71 \\ 0.84 \\ 0.83 \\ 0.62 \\ 0.32 \\ 0.13 \\ 0.81 \\ 0.81 \\ 0.44 \\ 0.17 \\ -0.02 \\ 0.00 \\ -0.03 \end{array}$	S3 0.10 0.25 -0.03 0.02 0.06 0.08 0.07 0.01 -0.04 0.00 0.04 0.15 0.97 0.94	$\begin{array}{c} 84\\ 0.08\\ -0.52\\ -0.26\\ -0.09\\ 0.23\\ 0.05\\ 0.74\\ 0.22\\ 0.37\\ 0.12\\ -0.21\\ -0.04\\ -0.04\\ 0.04\\ \end{array}$	S5 0.57 0.65 0.19 0.28 0.41 0.42 0.61 0.07 -0.01 0.26 0.60 0.77 0.05 0.04 -0.01
(b) OC EC Levoglucosan Mannosan Galactosan <i>p</i> -Hydroxybenzoic acid Vanillin Syringaldehyde Vanillic acid Syringic acid Dehydroabietic acid Cholesterol C ₂₂ C ₂₃ C ₂₄ C ₂₅	S1 0.47 0.39 0.09 0.17 0.26 0.22 0.24 -0.12 0.02 0.18 0.01 0.05 0.05 0.28 0.33	$\begin{array}{c} S2\\ 0.47\\ 0.20\\ 0.71\\ 0.84\\ 0.83\\ 0.62\\ 0.32\\ 0.13\\ 0.81\\ 0.81\\ 0.44\\ 0.17\\ -0.02\\ 0.00\\ -0.03\\ 0.10\\ \end{array}$	$\begin{array}{c} S3\\ 0.10\\ 0.25\\ -0.03\\ 0.02\\ 0.06\\ 0.08\\ 0.07\\ 0.01\\ -0.04\\ 0.00\\ 0.04\\ 0.15\\ 0.97\\ 0.97\\ 0.94\\ 0.85\end{array}$	$\begin{array}{c} 84\\ 0.08\\ -0.52\\ -0.26\\ 0.23\\ 0.05\\ 0.74\\ 0.22\\ 0.37\\ 0.12\\ -0.21\\ -0.04\\ -0.04\\ 0.04\\ 0.05\\ \end{array}$	S5 0.57 0.65 0.19 0.28 0.41 0.42 0.61 0.07 -0.01 0.26 0.60 0.77 0.05 0.04 -0.01 0.35
(b) OC EC Levoglucosan Mannosan Galactosan p-Hydroxybenzoic acid Vanillin Syringaldehyde Vanillic acid Syringic acid Dehydroabietic acid Cholesterol C ₂₂ C ₂₃ C ₂₄ C ₂₅ C ₂₆	S1 0.47 0.39 0.09 0.17 0.26 0.22 0.24 -0.12 0.02 0.18 0.01 0.05 0.05 0.28 0.33 0.61	$\begin{array}{c} \text{S2} \\ 0.47 \\ 0.20 \\ 0.71 \\ 0.84 \\ 0.83 \\ 0.62 \\ 0.32 \\ 0.13 \\ 0.81 \\ 0.81 \\ 0.44 \\ 0.17 \\ -0.02 \\ 0.00 \\ -0.03 \\ 0.10 \\ 0.05 \\ \end{array}$	$\begin{array}{c} S3\\ 0.10\\ 0.25\\ -0.03\\ 0.02\\ 0.06\\ 0.08\\ 0.07\\ 0.01\\ -0.04\\ 0.00\\ 0.04\\ 0.15\\ 0.97\\ 0.97\\ 0.97\\ 0.94\\ 0.85\\ 0.68\\ \end{array}$	$\begin{array}{c} 84\\ 0.08\\ -0.52\\ -0.26\\ -0.09\\ 0.23\\ 0.05\\ 0.74\\ 0.22\\ 0.37\\ 0.12\\ -0.21\\ -0.04\\ -0.04\\ 0.04\\ 0.05\\ 0.14\\ \end{array}$	S5 0.57 0.65 0.19 0.28 0.41 0.42 0.61 0.07 -0.01 0.26 0.60 0.77 0.05 0.04 -0.01 0.35 0.24
(b) OC EC Levoglucosan Mannosan Galactosan p-Hydroxybenzoic acid Vanillin Syringaldehyde Vanillic acid Syringic acid Dehydroabietic acid Cholesterol C ₂₂ C ₂₃ C ₂₄ C ₂₅ C ₂₆ C ₂₇	S1 0.47 0.39 0.09 0.17 0.26 0.22 0.24 -0.12 0.02 0.18 0.01 0.05 0.05 0.28 0.33 0.61 0.67	S2 0.47 0.20 0.71 0.84 0.83 0.62 0.32 0.13 0.81 0.81 0.81 0.44 0.17 -0.02 0.00 -0.03 0.10 0.05 0.08	$\begin{array}{c} S3\\ 0.10\\ 0.25\\ -0.03\\ 0.02\\ 0.06\\ 0.08\\ 0.07\\ 0.01\\ -0.04\\ 0.00\\ 0.04\\ 0.15\\ 0.97\\ 0.97\\ 0.94\\ 0.85\\ 0.68\\ 0.53\\ \end{array}$	$\begin{array}{c} 84\\ 0.08\\ -0.52\\ -0.26\\ 0.23\\ 0.05\\ 0.74\\ 0.22\\ 0.37\\ 0.12\\ -0.21\\ -0.04\\ -0.04\\ 0.05\\ 0.14\\ 0.05\\ 0.14\\ 0.10\\ \end{array}$	S5 0.57 0.65 0.19 0.28 0.41 0.42 0.61 0.07 -0.01 0.26 0.60 0.77 0.05 0.04 -0.01 0.35 0.24 0.35
(b) OC EC Levoglucosan Mannosan Galactosan p-Hydroxybenzoic acid Vanillin Syringaldehyde Vanillic acid Syringic acid Dehydroabietic acid Cholesterol C ₂₂ C ₂₃ C ₂₄ C ₂₅ C ₂₆ C ₂₇ C ₂₈	S1 0.47 0.39 0.09 0.17 0.26 0.22 0.24 -0.12 0.02 0.18 0.01 0.05 0.05 0.28 0.33 0.61 0.67 0.86	$\begin{array}{c} \text{S2} \\ 0.47 \\ 0.20 \\ 0.71 \\ 0.84 \\ 0.83 \\ 0.62 \\ 0.32 \\ 0.13 \\ 0.81 \\ 0.81 \\ 0.44 \\ 0.17 \\ -0.02 \\ 0.00 \\ -0.03 \\ 0.10 \\ 0.05 \\ 0.08 \\ 0.06 \\ \end{array}$	$\begin{array}{c} S3\\ \hline 0.10\\ 0.25\\ -0.03\\ 0.02\\ 0.06\\ 0.08\\ 0.07\\ 0.01\\ -0.04\\ 0.00\\ 0.04\\ 0.15\\ 0.97\\ 0.97\\ 0.97\\ 0.94\\ 0.85\\ 0.68\\ 0.53\\ 0.27\\ \end{array}$	$\begin{array}{c} 84\\ 0.08\\ -0.52\\ -0.26\\ -0.09\\ 0.23\\ 0.05\\ 0.74\\ 0.22\\ 0.37\\ 0.12\\ -0.21\\ -0.04\\ -0.04\\ 0.05\\ 0.14\\ 0.05\\ 0.14\\ 0.10\\ -0.01\\ \end{array}$	S5 0.57 0.65 0.19 0.28 0.41 0.42 0.61 0.07 -0.01 0.26 0.60 0.77 0.05 0.04 -0.01 0.35 0.24 0.35 0.01
(b) OC EC Levoglucosan Mannosan Galactosan p-Hydroxybenzoic acid Vanillin Syringaldehyde Vanillic acid Syringic acid Dehydroabietic acid Cholesterol C ₂₂ C ₂₃ C ₂₄ C ₂₅ C ₂₆ C ₂₇ C ₂₈ C ₂₉	S1 0.47 0.39 0.09 0.17 0.26 0.22 0.24 -0.12 0.02 0.18 0.01 0.05 0.05 0.28 0.33 0.61 0.67 0.86 0.89	$\begin{array}{c} S2\\ 0.47\\ 0.20\\ 0.71\\ 0.84\\ 0.83\\ 0.62\\ 0.32\\ 0.13\\ 0.81\\ 0.81\\ 0.44\\ 0.17\\ -0.02\\ 0.00\\ -0.03\\ 0.10\\ 0.05\\ 0.08\\ 0.06\\ 0.14\\ \end{array}$	S3 0.10 0.25 -0.03 0.02 0.06 0.08 0.07 0.01 -0.04 0.00 0.04 0.15 0.97 0.97 0.94 0.85 0.68 0.53 0.27 0.18	$\begin{array}{c} 84\\ 0.08\\ 0.26\\ -0.52\\ -0.26\\ 0.09\\ 0.23\\ 0.05\\ 0.74\\ 0.22\\ 0.37\\ 0.12\\ -0.21\\ -0.04\\ -0.04\\ 0.05\\ 0.14\\ 0.05\\ 0.14\\ 0.10\\ -0.01\\ 0.08\\ \end{array}$	S5 0.57 0.65 0.19 0.28 0.41 0.42 0.61 0.07 -0.01 0.26 0.60 0.77 0.05 0.04 -0.01 0.35 0.24 0.35 0.01 0.29
(b) OC EC Levoglucosan Mannosan Galactosan <i>p</i> -Hydroxybenzoic acid Vanillin Syringaldehyde Vanillic acid Syringic acid Dehydroabietic acid Cholesterol C ₂₂ C ₂₃ C ₂₄ C ₂₅ C ₂₆ C ₂₇ C ₂₈ C ₂₉ C ₃₀	S1 0.47 0.39 0.09 0.17 0.26 0.22 0.24 -0.12 0.02 0.18 0.01 0.05 0.28 0.33 0.61 0.67 0.86 0.89 0.84 0.84 0.84 0.89 0.84 0.95 0.95 0.95 0.95 0.95 0.95 0.95 0.95 0.95 0.95 0.95 0.95 0.95 0.95 0.95 0.95 0.95 0.95 0.95 0.95 0.95 0.95 0.95 0.95 0.95 0.95 0.95 0.95 0.95 0.95 0.95 0.95 0.95 0.95 0.95 0.95 0.95 0.95 0.95 0.95 0.95 0.95 0.95 0.95 0.95 0.95 0.95 0.95 0.95 0.95 0.95 0.95 0.95 0.95 0.95 0.95 0.95 0.95 0.95 0.95 0.95 0.95 0.95 0.95 0.95 0.95 0.95 0.95 0.95 0.95 0.95 0.95 0.95 0.95 0.95 0.95 0.95 0.95 0.95 0.95 0.95 0.95 0.95 0.95 0.95 0.95 0.95 0.95 0.95 0.95 0.95 0.95 0.95 0.95 0.95 0.95 0.95 0.95 0.95 0.95 0.95 0.95 0.95 0.95 0.95 0.95 0.95 0.95 0.95 0.95 0.95 0.95 0.95 0.95 0.95 0.95 0.95 0.95 0.95 0.95 0.95 0.95 0.95 0.95 0.95 0.95 0.95 0.95 0.95 0.95 0.95 0.95 0.95 0.95 0.95 0.95 0.95 0.95 0.95 0.95 0.95 0.95 0.95 0.95 0.95 0.95 0.95 0.95 0.95 0.95 0.95 0.95 0.95 0.95 0.95 0.95 0.95 0.95 0.95 0.95 0.95 0.95 0.95 0.95 0.95 0.95 0.95 0.95 0.95 0.95 0.95 0.95 0.95 0.95 0.95 0.95 0.95 0.95 0.95 0.95 0.95 0.95 0.95 0.95 0.95 0.95 0.95 0.95 0.95 0.95 0.95 0.95 0.95 0.95 0.95 0.95 0.95 0.95 0.95 0.95 0.95 0.95 0.95 0.95 0.95 0.95 0.95 0.95 0.95 0.95 0.95 0.95 0.95 0.95 0.95 0.95 0.95 0.95 0.95 0.95 0.95 0.95 0.95 0.95 0.95 0.95 0.95 0.95 0.95 0.95 0.95 0.95 0.95 0.95 0.95 0.95 0.95 0.95 0.95 0.95 0.95 0.95 0.95 0.95 0.95 0.95 0.95 0.95 0.95 0.95 0.95 0.95 0.95 0.95 0.95 0.95 0.95 0.95 0.95 0.95 0.95 0.95 0.95 0.95 0.95 0.95 0.95 0.95 0.95 0.95 0.95 0.95 0.95 0.95 0.95 0.95 0.95 0.95 0.95 0.95 0.95 0.95 0.95 0.95 0.95 0.95 0.95 0.95 0.95 0.95 0.95 0.95 0.95 0.95 0.95 0.95 0.95 0.95 0.95 0.95 0.95 0.95 0.95 0.95 0.95 0.95 0.95 0.95 0.95 0.95 0.95 0.95 0.95 0.95 0.95 0.95 0.95 0	$\begin{array}{c} S2\\ 0.47\\ 0.20\\ 0.71\\ 0.84\\ 0.83\\ 0.62\\ 0.32\\ 0.13\\ 0.81\\ 0.81\\ 0.81\\ 0.44\\ 0.17\\ -0.02\\ 0.00\\ -0.03\\ 0.10\\ 0.05\\ 0.08\\ 0.06\\ 0.14\\ 0.03\\ \end{array}$	S3 0.10 0.25 -0.03 0.02 0.06 0.08 0.07 0.01 -0.04 0.00 0.04 0.15 0.97 0.94 0.85 0.68 0.53 0.27 0.18 0.33	$\begin{array}{c} 84\\ 0.08\\ 0.26\\ -0.52\\ -0.26\\ 0.09\\ 0.23\\ 0.05\\ 0.74\\ 0.22\\ 0.37\\ 0.12\\ -0.21\\ -0.04\\ -0.04\\ 0.05\\ 0.14\\ 0.05\\ 0.14\\ 0.10\\ -0.01\\ 0.08\\ 0.04\\ 0.04\\ 0.04\\ 0.04\\ 0.04\\ 0.05\\ 0.14\\ 0.00\\ 0.04\\ 0.04\\ 0.04\\ 0.04\\ 0.04\\ 0.04\\ 0.04\\ 0.04\\ 0.04\\ 0.04\\ 0.04\\ 0.04\\ 0.04\\ 0.04\\ 0.04\\ 0.04\\ 0.04\\ 0.04\\ 0.04\\ 0.04\\ 0.04\\ 0.04\\ 0.04\\ 0.04\\ 0.04\\ 0.04\\ 0.04\\ 0.04\\ 0.04\\ 0.04\\ 0.04\\ 0.04\\ 0.04\\ 0.04\\ 0.04\\ 0.04\\ 0.04\\ 0.04\\ 0.04\\ 0.04\\ 0.04\\ 0.04\\ 0.04\\ 0.04\\ 0.04\\ 0.04\\ 0.04\\ 0.04\\ 0.04\\ 0.04\\ 0.04\\ 0.04\\ 0.04\\ 0.04\\ 0.04\\ 0.04\\ 0.04\\ 0.04\\ 0.04\\ 0.04\\ 0.04\\ 0.04\\ 0.04\\ 0.04\\ 0.04\\ 0.04\\ 0.04\\ 0.04\\ 0.04\\ 0.04\\ 0.04\\ 0.04\\ 0.04\\ 0.04\\ 0.04\\ 0.04\\ 0.04\\ 0.04\\ 0.04\\ 0.04\\ 0.04\\ 0.04\\ 0.04\\ 0.04\\ 0.04\\ 0.04\\ 0.04\\ 0.04\\ 0.04\\ 0.04\\ 0.04\\ 0.04\\ 0.04\\ 0.04\\ 0.04\\ 0.04\\ 0.04\\ 0.04\\ 0.04\\ 0.04\\ 0.04\\ 0.04\\ 0.04\\ 0.04\\ 0.04\\ 0.04\\ 0.04\\ 0.04\\ 0.04\\ 0.04\\ 0.04\\ 0.04\\ 0.04\\ 0.04\\ 0.04\\ 0.04\\ 0.04\\ 0.04\\ 0.04\\ 0.04\\ 0.04\\ 0.04\\ 0.04\\ 0.04\\ 0.04\\ 0.04\\ 0.04\\ 0.04\\ 0.04\\ 0.04\\ 0.04\\ 0.04\\ 0.04\\ 0.04\\ 0.04\\ 0.04\\ 0.04\\ 0.04\\ 0.04\\ 0.04\\ 0.04\\ 0.04\\ 0.04\\ 0.04\\ 0.04\\ 0.04\\ 0.04\\ 0.04\\ 0.04\\ 0.04\\ 0.04\\ 0.04\\ 0.04\\ 0.04\\ 0.04\\ 0.04\\ 0.04\\ 0.04\\ 0.04\\ 0.04\\ 0.04\\ 0.04\\ 0.04\\ 0.04\\ 0.04\\ 0.04\\ 0.04\\ 0.04\\ 0.04\\ 0.04\\ 0.04\\ 0.04\\ 0.04\\ 0.04\\ 0.04\\ 0.04\\ 0.04\\ 0.04\\ 0.04\\ 0.04\\ 0.04\\ 0.04\\ 0.04\\ 0.04\\ 0.04\\ 0.04\\ 0.04\\ 0.04\\ 0.04\\ 0.04\\ 0.04\\ 0.04\\ 0.04\\ 0.04\\ 0.04\\ 0.04\\ 0.04\\ 0.04\\ 0.04\\ 0.04\\ 0.04\\ 0.04\\ 0.04\\ 0.04\\ 0.04\\ 0.04\\ 0.04\\ 0.04\\ 0.04\\ 0.04\\ 0.04\\ 0.04\\ 0.04\\ 0.04\\ 0.04\\ 0.04\\ 0.04\\ 0.04\\ 0.04\\ 0.04\\ 0.04\\ 0.04\\ 0.04\\ 0.04\\ 0.04\\ 0.04\\ 0.04\\ 0.04\\ 0.04\\ 0.04\\ 0.04\\ 0.04\\ 0.04\\ 0.04\\ 0.04\\ 0.04\\ 0.04\\ 0.04\\ 0.04\\ 0.04\\ 0.04\\ 0.04\\ 0.04\\ 0.04\\ 0.04\\ 0.04\\ 0.04\\ 0.04\\ 0.04\\ 0.04\\ 0.04\\ 0.04\\ 0.04\\ 0.04\\ 0.04\\ 0.04\\ 0.04\\ 0.04\\ 0.04\\ 0.04\\ 0.04\\ 0.04\\ 0.04\\ 0.04\\ 0.04\\ 0.04\\ 0.04\\ 0.04\\ 0.04\\ 0.04\\ 0.04\\ 0.04\\ 0.04\\ 0.04\\ 0.04\\ 0.04\\ 0.04\\ 0.04\\ 0.04\\ 0.04\\ 0.04\\ 0.04\\ 0.04\\ 0.04\\ 0.04\\ 0.04\\ 0.04\\ 0.04\\ 0.04\\ 0.04\\ 0.04\\ 0.04\\ 0.04\\ 0.04\\ 0.04\\ 0.04\\ 0.04\\ 0.04\\ 0.04\\ 0.04\\ 0.04\\ 0.04\\ 0.04\\ 0.04\\ 0.04\\ 0.04\\ 0.04\\ 0.04\\ 0.04\\ 0$	S5 0.57 0.65 0.19 0.28 0.41 0.42 0.61 0.07 -0.01 0.26 0.60 0.77 0.05 0.04 -0.01 0.35 0.24 0.35 0.01 0.29 -0.12
(b) OC EC Levoglucosan Mannosan Galactosan <i>p</i> -Hydroxybenzoic acid Vanillin Syringaldehyde Vanillic acid Syringic acid Dehydroabietic acid Cholesterol C ₂₂ C ₂₃ C ₂₄ C ₂₅ C ₂₆ C ₂₇ C ₂₈ C ₂₉ C ₃₀ C ₃₁	S1 0.47 0.39 0.09 0.17 0.26 0.22 0.24 -0.12 0.02 0.18 0.01 0.05 0.28 0.33 0.61 0.67 0.86 0.89 0.84 0.77	$\begin{array}{c} \text{S2} \\ 0.47 \\ 0.20 \\ 0.71 \\ 0.84 \\ 0.83 \\ 0.62 \\ 0.32 \\ 0.13 \\ 0.81 \\ 0.81 \\ 0.44 \\ 0.17 \\ -0.02 \\ 0.00 \\ -0.03 \\ 0.10 \\ 0.05 \\ 0.08 \\ 0.06 \\ 0.14 \\ 0.03 \\ 0.24 \\ \end{array}$	S3 0.10 0.25 -0.03 0.02 0.06 0.08 0.07 0.01 -0.04 0.00 0.04 0.15 0.97 0.94 0.85 0.68 0.53 0.27 0.18 0.33 0.07	$\begin{array}{c} 84\\ 0.08\\ 0.26\\ -0.52\\ -0.26\\ 0.09\\ 0.23\\ 0.05\\ 0.74\\ 0.22\\ 0.37\\ 0.12\\ -0.21\\ -0.04\\ 0.04\\ 0.05\\ 0.14\\ 0.05\\ 0.14\\ 0.00\\ 0.08\\ 0.04\\ 0.10\\ \hline \end{array}$	S5 0.57 0.65 0.19 0.28 0.41 0.42 0.61 0.07 -0.01 0.26 0.60 0.77 0.05 0.04 -0.01 0.35 0.04 -0.01 0.35 0.24 0.35 0.01 0.29 -0.12 0.47
(b) OC EC Levoglucosan Mannosan Galactosan p-Hydroxybenzoic acid Vanillin Syringaldehyde Vanillic acid Syringic acid Dehydroabietic acid Cholesterol C ₂₂ C ₂₃ C ₂₄ C ₂₅ C ₂₆ C ₂₇ C ₂₈ C ₂₉ C ₃₀ C ₃₁ C ₃₂	S1 0.47 0.39 0.19 0.17 0.26 0.22 0.24 -0.12 0.02 0.18 0.01 0.05 0.28 0.33 0.61 0.67 0.86 0.89 0.84 0.77 0.86 0.89 0.84 0.77 0.84 0.77 0.84 0.77 0.85 0.85 0.85 0.85 0.85 0.85 0.85 0.85 0.85 0.85 0.85 0.85 0.85 0.85 0.85 0.85 0.85 0.85 0.85 0.85 0.85 0.85 0.85 0.85 0.85 0.85 0.85 0.85 0.85 0.85 0.85 0.85 0.85 0.85 0.85 0.85 0.85 0.85 0.85 0.85 0.85 0.85 0.85 0.85 0.85 0.85 0.85 0.85 0.85 0.85 0.85 0.85 0.85 0.85 0.85 0.85 0.85 0.85 0.85 0.85 0.85 0.85 0.85 0.85 0.85 0.85 0.85 0.85 0.85 0.85 0.85 0.85 0.85 0.85 0.85 0.85 0.85 0.85 0.85 0.85 0.85 0.85 0.85 0.85 0.85 0.85 0.85 0.85 0.85 0.85 0.85 0.85 0.85 0.85 0.85 0.85 0.85 0.85 0.85 0.85 0.85 0.85 0.85 0.85 0.85 0.85 0.85 0.85 0.85 0.85 0.85 0.85 0.85 0.85 0.85 0.85 0.85 0.85 0.85 0.85 0.85 0.85 0.85 0.85 0.85 0.85 0.85 0.85 0.85 0.85 0.85 0.85 0.85 0.85 0.85 0.85 0.85 0.85 0.85 0.85 0.85 0.85 0.85 0.85 0.85 0.85 0.85 0.85 0.85 0.85 0.85 0.85 0.85 0.85 0.85 0.85 0.85 0.85 0.85 0.85 0.85 0.85 0.85 0.85 0.85 0.85 0.85 0.85 0.85 0.85 0.85 0.85 0.85 0.85 0.85 0.85 0.85 0.85 0.85 0.85 0.85 0.85 0.85 0.85 0.85 0.85 0.85 0.85 0.85 0.85 0.85 0.85 0.85 0.85 0.85 0.85 0.85 0.85 0.85 0.85 0.85 0.85 0.85 0.85 0.85 0.85 0.85 0.85 0.85 0.85 0.85 0.85 0.85 0.85 0.85 0.85 0.85 0.85 0.85 0.85 0.85 0.85 0.85 0.85 0.85 0.85 0.85 0.85 0.85 0.85 0.85 0.85 0.85 0.85 0.85 0.85 0.85 0.85 0.85 0.85 0.85 0.85 0.85 0.85 0.85 0.85 0.85 0.85 0.85 0.85 0.85 0.85 0.85 0.85 0.85 0.85 0.85 0.85 0.85 0.85 0.85 0.85 0.85 0.85 0.85 0.85 0.85 0.85 0.85 0.85 0.85 0.85 0.85 0.85 0.85 0.85 0.85 0.85 0.85 0.85 0.85 0.85 0.85 0.85 0.85 0.85 0.85 0.85 0.85 0.85 0.85 0.85 0.85 0.85 0.85 0.85 0.85 0.85 0.85 0.85 0.85 0.85 0.85 0.85 0.85 0.85 0.85 0.85 0.85 0.85 0.85 0.85 0	$\begin{array}{c} S2\\ 0.47\\ 0.20\\ 0.71\\ 0.84\\ 0.83\\ 0.62\\ 0.32\\ 0.13\\ 0.81\\ 0.81\\ 0.44\\ 0.17\\ -0.02\\ 0.00\\ -0.03\\ 0.10\\ 0.05\\ 0.08\\ 0.06\\ 0.14\\ 0.03\\ 0.24\\ -0.04\\ \end{array}$	S3 0.10 0.25 -0.03 0.02 0.06 0.08 0.07 0.01 -0.04 0.00 0.04 0.15 0.97 0.94 0.85 0.68 0.53 0.27 0.18 0.33 0.07 0.02	$\begin{array}{c} 84\\ 0.08\\ 0.26\\ -0.52\\ -0.26\\ 0.09\\ 0.23\\ 0.05\\ 0.74\\ 0.22\\ 0.37\\ 0.12\\ -0.21\\ -0.04\\ -0.04\\ 0.05\\ 0.14\\ 0.00\\ 0.10\\ 0.08\\ 0.04\\ 0.10\\ 0.10\\ 0.10\\ 0.10\\ 0.10\\ 0.10\\ 0.10\\ 0.10\\ 0.10\\ 0.10\\ 0.10\\ 0.10\\ 0.10\\ 0.10\\ 0.10\\ 0.10\\ 0.10\\ 0.10\\ 0.10\\ 0.10\\ 0.10\\ 0.10\\ 0.10\\ 0.10\\ 0.10\\ 0.10\\ 0.10\\ 0.10\\ 0.10\\ 0.10\\ 0.10\\ 0.10\\ 0.10\\ 0.10\\ 0.10\\ 0.10\\ 0.10\\ 0.10\\ 0.10\\ 0.10\\ 0.10\\ 0.10\\ 0.10\\ 0.10\\ 0.10\\ 0.10\\ 0.10\\ 0.10\\ 0.10\\ 0.10\\ 0.10\\ 0.10\\ 0.10\\ 0.10\\ 0.10\\ 0.10\\ 0.10\\ 0.10\\ 0.10\\ 0.10\\ 0.10\\ 0.10\\ 0.10\\ 0.10\\ 0.10\\ 0.10\\ 0.10\\ 0.10\\ 0.10\\ 0.10\\ 0.10\\ 0.10\\ 0.10\\ 0.10\\ 0.10\\ 0.10\\ 0.10\\ 0.10\\ 0.10\\ 0.10\\ 0.10\\ 0.10\\ 0.10\\ 0.10\\ 0.10\\ 0.10\\ 0.10\\ 0.10\\ 0.10\\ 0.10\\ 0.10\\ 0.10\\ 0.10\\ 0.10\\ 0.10\\ 0.10\\ 0.10\\ 0.10\\ 0.10\\ 0.10\\ 0.10\\ 0.10\\ 0.10\\ 0.10\\ 0.10\\ 0.10\\ 0.10\\ 0.10\\ 0.10\\ 0.10\\ 0.10\\ 0.10\\ 0.10\\ 0.10\\ 0.10\\ 0.10\\ 0.10\\ 0.10\\ 0.10\\ 0.10\\ 0.10\\ 0.10\\ 0.10\\ 0.10\\ 0.10\\ 0.10\\ 0.10\\ 0.10\\ 0.10\\ 0.10\\ 0.10\\ 0.10\\ 0.10\\ 0.10\\ 0.10\\ 0.10\\ 0.10\\ 0.10\\ 0.10\\ 0.10\\ 0.10\\ 0.10\\ 0.10\\ 0.10\\ 0.10\\ 0.10\\ 0.10\\ 0.10\\ 0.10\\ 0.10\\ 0.10\\ 0.10\\ 0.10\\ 0.10\\ 0.10\\ 0.10\\ 0.10\\ 0.10\\ 0.10\\ 0.10\\ 0.10\\ 0.10\\ 0.10\\ 0.10\\ 0.10\\ 0.10\\ 0.10\\ 0.10\\ 0.10\\ 0.10\\ 0.10\\ 0.10\\ 0.10\\ 0.10\\ 0.10\\ 0.10\\ 0.10\\ 0.10\\ 0.10\\ 0.10\\ 0.10\\ 0.10\\ 0.10\\ 0.10\\ 0.10\\ 0.10\\ 0.10\\ 0.10\\ 0.10\\ 0.10\\ 0.10\\ 0.10\\ 0.10\\ 0.10\\ 0.10\\ 0.10\\ 0.10\\ 0.10\\ 0.10\\ 0.10\\ 0.10\\ 0.10\\ 0.10\\ 0.10\\ 0.10\\ 0.10\\ 0.10\\ 0.10\\ 0.10\\ 0.10\\ 0.10\\ 0.10\\ 0.10\\ 0.10\\ 0.10\\ 0.10\\ 0.10\\ 0.10\\ 0.10\\ 0.10\\ 0.10\\ 0.10\\ 0.10\\ 0.10\\ 0.10\\ 0.10\\ 0.10\\ 0.10\\ 0.10\\ 0.10\\ 0.10\\ 0.10\\ 0.10\\ 0.10\\ 0.10\\ 0.10\\ 0.10\\ 0.10\\ 0.10\\ 0.10\\ 0.10\\ 0.10\\ 0.10\\ 0.10\\ 0.10\\ 0.10\\ 0.10\\ 0.10\\ 0.10\\ 0.10\\ 0.10\\ 0.10\\ 0.10\\ 0.10\\ 0.10\\ 0.10\\ 0.10\\ 0.10\\ 0.10\\ 0.10\\ 0.10\\ 0.10\\ 0.10\\ 0.10\\ 0.10\\ 0.10\\ 0.10\\ 0.10\\ 0.10\\ 0.10\\ 0.10\\ 0.10\\ 0.10\\ 0.10\\ 0.10\\ 0.10\\ 0.10\\ 0.10\\ 0.10\\ 0.10\\ 0.10\\ 0.10\\ 0.10\\ 0.10\\ 0.10\\ 0.10\\ 0.10\\ 0.10\\ 0.10\\ 0.10\\ 0.10\\ 0.10\\ 0.10\\ 0.10\\ 0.10\\ 0.10\\ 0.10\\ 0.10\\ 0.10\\ 0.10\\ 0.10\\ 0.10\\ 0.10\\ 0.10\\ 0.10\\ 0.10\\ 0.10\\ 0.10\\ 0.10\\ 0.10\\ 0.10\\ 0.10\\ 0.10\\ 0.10\\ 0.10\\ 0.10\\ 0.10\\ 0.10\\ 0.$	$\begin{array}{c} 85\\ \hline 0.57\\ 0.65\\ 0.19\\ 0.28\\ 0.41\\ 0.42\\ 0.61\\ 0.07\\ -0.01\\ 0.26\\ 0.60\\ 0.77\\ 0.05\\ 0.04\\ -0.01\\ 0.35\\ 0.04\\ 0.035\\ 0.24\\ 0.35\\ 0.01\\ 0.29\\ -0.12\\ 0.47\\ 0.16\\ \end{array}$
(b) OC EC Levoglucosan Mannosan Galactosan <i>p</i> -Hydroxybenzoic acid Vanillin Syringaldehyde Vanillic acid Syringic acid Dehydroabietic acid Cholesterol C ₂₂ C ₂₃ C ₂₄ C ₂₅ C ₂₆ C ₂₇ C ₂₈ C ₂₉ C ₃₀ C ₃₁ C ₃₂ C ₃₃	S1 0.47 0.39 0.09 0.17 0.26 0.22 0.24 -0.12 0.02 0.18 0.01 0.05 0.28 0.33 0.61 0.67 0.86 0.89 0.84 0.77 0.88 0.72	$\begin{array}{c} S2\\ 0.47\\ 0.20\\ 0.71\\ 0.84\\ 0.83\\ 0.62\\ 0.32\\ 0.13\\ 0.81\\ 0.81\\ 0.81\\ 0.44\\ 0.17\\ -0.02\\ 0.00\\ -0.03\\ 0.10\\ 0.05\\ 0.08\\ 0.06\\ 0.14\\ 0.03\\ 0.24\\ -0.04\\ 0.28\\ \end{array}$	$\begin{array}{c} \text{S3} \\ 0.10 \\ 0.25 \\ -0.03 \\ 0.02 \\ 0.06 \\ 0.08 \\ 0.07 \\ 0.01 \\ -0.04 \\ 0.00 \\ 0.04 \\ 0.15 \\ 0.97 \\ 0.97 \\ 0.97 \\ 0.97 \\ 0.97 \\ 0.94 \\ 0.85 \\ 0.68 \\ 0.53 \\ 0.27 \\ 0.18 \\ 0.33 \\ 0.07 \\ 0.02 \\ -0.03 \end{array}$	$\begin{array}{c} 84\\ 0.08\\ 0.26\\ -0.52\\ -0.26\\ 0.09\\ 0.23\\ 0.05\\ 0.74\\ 0.22\\ 0.37\\ 0.12\\ -0.21\\ -0.04\\ 0.05\\ 0.14\\ 0.05\\ 0.14\\ 0.00\\ 0.01\\ 0.08\\ 0.04\\ 0.10\\ 0.10\\ 0.10\\ 0.14\\ \end{array}$	$\begin{array}{c} 85\\ \hline 0.57\\ 0.65\\ 0.19\\ 0.28\\ 0.41\\ 0.42\\ 0.61\\ 0.07\\ -0.01\\ 0.26\\ 0.60\\ 0.77\\ 0.05\\ 0.04\\ -0.01\\ 0.35\\ 0.04\\ 0.35\\ 0.01\\ 0.29\\ -0.12\\ 0.47\\ 0.16\\ 0.49\\ \end{array}$
(b) OC EC $Levoglucosan$ $Mannosan$ $Galactosan$ $p-Hydroxybenzoic acid$ Vanillin Syringaldehyde Vanillic acid Syringic acid Dehydroabietic acid Cholesterol C ₂₂ C ₂₃ C ₂₄ C ₂₅ C ₂₆ C ₂₇ C ₂₈ C ₂₉ C ₃₀ C ₃₁ C ₃₂ C ₃₃ % variance	S1 0.47 0.39 0.09 0.17 0.26 0.22 0.24 -0.12 0.02 0.18 0.01 0.05 0.28 0.33 0.61 0.67 0.86 0.89 0.84 0.77 0.88 0.72 43	$\begin{array}{c} S2\\ 0.47\\ 0.20\\ 0.71\\ 0.84\\ 0.83\\ 0.62\\ 0.32\\ 0.13\\ 0.81\\ 0.81\\ 0.44\\ 0.17\\ -0.02\\ 0.00\\ -0.03\\ 0.10\\ 0.05\\ 0.08\\ 0.06\\ 0.14\\ 0.03\\ 0.24\\ -0.04\\ 0.28\\ \hline\end{array}$	S3 0.10 0.25 -0.03 0.02 0.06 0.08 0.07 0.01 -0.04 0.00 0.04 0.15 0.97 0.97 0.94 0.85 0.68 0.53 0.27 0.18 0.33 0.07 -0.03 11	$\begin{array}{c} 84\\ 0.08\\ 0.26\\ -0.52\\ -0.26\\ 0.09\\ 0.23\\ 0.05\\ 0.74\\ 0.22\\ 0.37\\ 0.12\\ -0.21\\ -0.04\\ -0.04\\ 0.05\\ 0.14\\ 0.05\\ 0.14\\ 0.00\\ -0.01\\ 0.08\\ 0.04\\ 0.10\\ 0.10\\ 0.10\\ 0.10\\ 0.14\\ \hline 5.0\\ \end{array}$	$\begin{array}{c} \text{S5} \\ \hline 0.57 \\ 0.65 \\ 0.19 \\ 0.28 \\ 0.41 \\ 0.42 \\ 0.61 \\ 0.07 \\ -0.01 \\ 0.26 \\ 0.60 \\ 0.77 \\ 0.05 \\ 0.04 \\ -0.01 \\ 0.35 \\ 0.04 \\ -0.01 \\ 0.35 \\ 0.24 \\ 0.35 \\ 0.01 \\ 0.29 \\ -0.12 \\ 0.47 \\ 0.16 \\ 0.49 \\ \hline \end{array}$

ganic compounds produced from these sources were not determined.

4 Conclusions

Annual $PM_{2.5}$ observations in Malaysia have been conducted to quantitatively characterise carbonaceous $PM_{2.5}$, especially focusing on organic compounds derived from biomass burning for the first time. The main conclusions are summarised as follows:

Concentrations of OP, LG, MN, galactosan, syringaldehyde, VA and cholesterol exhibit seasonal variability. The average concentrations of OP, LG, MN, galactosan, VA and cholesterol during the southwestern monsoon season are higher than those during the northeastern monsoon season, and the differences are statistically significant. In contrast, the syringaldehyde concentration during the southwestern monsoon season is lower.

Seven IPF samples are distinguished on the basis of the PM_{2.5} OP / OC4 mass ratio. In addition, significant differences are observed for the concentrations of many chemical species between the IPF and other samples. Thus, the $PM_{2.5}$ chemical characteristics in Malaysia are clearly influenced by IPFs during the southwestern monsoon season. Furthermore, two previously suggested indicators of IPF sources have been evaluated: the VA / SA and LG / MN mass ratios. The LG / MN mass ratio ranges from 14 to 22 in the IPF samples and from 11 to 31 in the other samples. The two ratio distributions partially overlap. Thus, the LG / MN mass ratio is not considered appropriate for extracting the effects of IPFs in Malaysia. In contrast, significant differences among the VA / SA mass ratios in the IPF and other samples suggest that it may serve as a good indicator. However, the OP / OC4 mass ratio differentiates the IPF samples better than the VA/SA mass ratio. Consequently, the OP/OC4 mass ratio is proposed as a better indicator than the VA / SA mass ratio. Finally, varimax-rotated PCA enabled to discriminate biomass burning components such as IPFs, softwood/hardwood burning and meat cooking.

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