

Hymenopteran Parasitoids Associated with Frugivorous Larvae in a Brazilian Caatinga-Cerrado Ecotone

Author(s): A. R. De Souza, G. N. Lopes-Mielezrski, E. N. Lopes, R. B. Querino,

C.D.A. Corsato, T. A. Giustolin, and R. A. Zucchi

Source: Environmental Entomology, 41(2):233-237. 2012.

Published By: Entomological Society of America

DOI: http://dx.doi.org/10.1603/EN11121

URL: http://www.bioone.org/doi/full/10.1603/EN11121

BioOne (www.bioone.org) is a nonprofit, online aggregation of core research in the biological, ecological, and environmental sciences. BioOne provides a sustainable online platform for over 170 journals and books published by nonprofit societies, associations, museums, institutions, and presses.

Your use of this PDF, the BioOne Web site, and all posted and associated content indicates your acceptance of BioOne's Terms of Use, available at www.bioone.org/page/terms_of_use.

Usage of BioOne content is strictly limited to personal, educational, and non-commercial use. Commercial inquiries or rights and permissions requests should be directed to the individual publisher as copyright holder.

BioOne sees sustainable scholarly publishing as an inherently collaborative enterprise connecting authors, nonprofit publishers, academic institutions, research libraries, and research funders in the common goal of maximizing access to critical research.

Hymenopteran Parasitoids Associated With Frugivorous Larvae in a Brazilian Caatinga-Cerrado Ecotone

A. R. DE SOUZA, G. N. LOPES-MIELEZRSKI, E. N. LOPES, R. B. QUERINO, C.D.A. CORSATO, T. A. GIUSTOLIN, AND R. A. ZUCCHI

Environ. Entomol. 41(2): 233-237 (2012); DOI: http://dx.doi.org/10.1603/EN11121

ABSTRACT The purpose of this study was to investigate native species of parasitoids of frugivorous larvae and their associations with host plants in commercial guava orchards and in typical native dry forests of a caatinga-cerrado ecotone in the State of Minas Gerais, Brazil. Nine species of parasitoids were associated with larvae of Anastrepha (Tephritidae) and Neosilba (Lonchaeidae) in fruit of Psidium guajava L. (Myrtaceae), Ziziphus joazeiro Mart. (Rhamnaceae), Spondias tuberosa Arruda (Anacardiaceae), Spondias dulcis Forst. (Anacardiaceae), Syzygium cumini (L.) Skeels (Myrtaceae), and Randia armata (Sw.) DC. (Rubiaceae). Doryctobracon areolatus was the most abundant species, obtained from puparia of Anastrepha zenildae, An. sororcula, An. fraterculus, An. obliqua, and An. turpiniae. This is the first report of Asobara obliqua in Brazil and of As. anastrephae and Tropideucoila weldi in dry forests of Minas Gerais State. The number of species of parasitoids was higher in areas with greater diversity of cultivated species and lower pesticide use. The forest fragments adjacent to the orchards served as shelter for parasitoids of frugivorous larvae.

KEY WORDS Doryctobracon, Asobara, Aganaspis, Anastrepha, Neosilba

Studies on tritrophic interactions involving frugivorous larvae, their parasitoids, and host fruits have been conducted basically on commercial crops, aiming to maximize parasitoid use to control fruit flies of economic concern (Aluja et al. 2003). Little is known about these flies and their associations in native areas or in areas with anthropic impact. Also, little attention has been attributed to the natural history and behavior of fruit flies in nature, therefore underestimating the complexity of fruit fly biology and ecology (Aluja 1999). Inventories of unaltered native environments are essential means of retrieving information to further current understanding on the relationships among parasitoids, frugivorous larvae and their host fruits.

The vegetation adjacent to agricultural areas has been studied to reveal the importance of these areas to supply food and alternative habitats for natural enemies. It is estimated that natural enemies increase in number, thus enhancing biological control, when native vegetation is present at the edges of a crop area or is associated with a plantation (Altieri et al. 2003). Studies have shown that the fragmentation of native habitats affects natural enemies more than it does phytophagous insects (Kruess and Tscharntke 2000).

The potential use of fruit fly parasitoids for biological control has gained relevance within Integrated Pest Management programs, in which a crucial stage is to know the native fauna of natural enemies. Objectives of this study are to investigate native species of parasitoids and their associations with frugivorous larvae and respective host fruits, in commercial guava orchards and in an undisturbed environmental, and to determine if the undisturbed environmental is a shelter for these parasitoids in a caatinga-cerrado ecotone.

Materials and Methods

Fruit was sampled in guava orchards and fragments of dry forest vegetation in areas near the cities of Jaíba, Janaúba, and Matias Cardoso of the State of Minas Gerais, Brazil (Table 1). The mean temperature in the

In the northern State of Minas Gerais, Brazil, there is an ecosystem known as dry forest, whose native vegetation is part of the caatinga domain, with enclaves of cerrado. This ecotone shows the typical vegetation of a dry forest (Drummond et al. 2005), also known as Deciduous Seasonal Forest. Dry forests typically are tropical, occurring in patches within a larger expanse of cerrado or caatinga. Cerrado is one of the world's biodiversity hotspots. In the last 35 yr, >50% of its $\approx \! 2$ million km² has been transformed into pasture and agricultural lands planted in cash crops. Cerrado has the richest flora among the world's savannas and high levels of endemism. Species richness of insects is equally high in cerrado (Klink and Machado 2005). Deforestation rates have very high in the caatinga too.

¹ Departamento de Entomologia e Acarologia, Escola Superior de Agricultura Luiz de Queiroz - ESALQ/USP, Av. Pádua Dias, 11, 13418-900, Piracicaba, SP, Brazil.

² Corresponding author: Empresa Brasileira de Pesquisa Agropecuária - Embrapa Meio-Norte Av. Duque de Caxias, 5650, CEP 64006–220, Teresina, PI, Brazil (email: ranyse@cpamn.embrapa.br).

³ Universidade Estadual de Montes Claros - Unimontes, Rua Reinaldo Viana, 2630, 39440-000, Janaúba, MG, Brazil.

Table 1. Location, geographical coordinates, and characteristics of the areas sampled (guava or chards adjacent to fragments of native forest)

Cities	Areas	Geographical coordinates	Management of guava orchards	Fruit diversity in nearby	Agricultural systems of guava orchards	Fruit diversity in the forest fragment
Jaíba	Area 1	15° 6′1.9″ S; 43° 58′56.5″ W	Management min., with cultural practices and no chemical control	Citrus spp., Cocos nucifera, Cucurbita spp., Manihot esculenta, Musa spp., Spondias mombi	Smallholder farming	Ziziphus joazeiro, Syzygium cumini, Spondias dulcis
Jaíba	Area 2	15° 08′26.4″ S; 44° 02′04.7″ W	Without management and without cultural practices	Musa spp.	Smallholder farming	Ziziphus joazeiro, Randia armata
Janaúba	Area 3	15° 49′52.8″ S; 43° 16′20.1″ W	Management min., with cultural practices and no chemical control	Citrus spp., Cocos nucifera, Musa spp.	Experimental orchard	Ziziphus joazeiro, Spondias tuberosa
Matias Cardoso	Area 4	15° 1′56.3″ S; 43° 50′20.8″ W	Intensive management, with cultural practices and chemical control	Annona sp., Mangifera indica	Commercial Orchard	Spondias tuberosa

region ranges from 21 to 25°C, with total annual rainfall of 700–1,200 mm (rainy period in October-April and dry period in May–September) and relative humidity ranging from 60 to 70% (Antunes 1994).

Collections from guava, Psidium guajava L. (Myrtaceae), were made in areas of smallholder farmers, private company and an experimental area. Other host fruit species, including juá, Ziziphus joazeiro Mart. (Rhamnaceae); Brazil plum or umbu, Spondias tuberosa Arruda (Anacardiaceae); tahitian apple or cajá, Spondias dulcis Forst. (Anacardiaceae); jambolan, Syzygium cumini (L.) Skeels (Myrtaceae); and indigoberry, Randia armata (Sw.) DC. (Rubiaceae) were collected in fragments of native forest (Table 1). Guava orchards were adjacent to fragments of native forest. The orchards were cultivated with the variety 'Paluma', with different types of management (in terms of culture treatment and use of chemical pesticides) and different levels of plant diversity in the surrounding areas (Table 1).

Fruit collection and processing procedures followed methods commonly used (e.g., Aluja et al. 2003). Ripe or ripening fruits were collected randomly from trees canopies and as recently fallen fruit on the ground (if the fruit was undamaged). Fruit was collected every 2 wk, from March 2005 to March 2006. The sample sizes were dependent of fruit availability. Fruit was kept in plastic trays containing a layer of vermiculite and covered with voil. Puparia were individually placed in glass flasks containing vermiculite, until the flies or the parasitoids emerged. The specimens were fixed in 70% ethanol. Association between a fly species and a parasitoid species was considered when a single species of fruit fly and of parasitoid emerged in the same flask.

Voucher specimens were deposited at the Departamento de Entomologia e Acarologia da Escola Superior de Agricultura Luiz de Queiroz (ESALQ/USP, in Piracicaba, São Paulo, Brazil), at Universidade Estadual de Montes Claros (UNIMONTES, in Janaúba, Minas Gerais, Brazil), and at the Departamento de Ecologia e Biologia Evolutiva, Universidade Federal de São Carlos (UFSCar, in São Carlos, São Paulo, Brazil).

Results and Discussion

Fruit was collected from six host plants: *P. guajava; Z. joazeiro; S. tuberosa; S. dulcis; S. cumini* and *R. armata.* In total, 544 parasitoids emerged (475 braconids, 68 figitids, and 1 pteromalid) from frugivorous larvae in three species of fruit (*P. guajava, S. tuberosa,* and *Z. joazeiro*). Four *Anastrepha* species and three *Neosilba* species (*N. pendula, N. zadolicha,* and *Neosilba* sp.) were reared (Table 2).

Infestation levels (pupae per gram of fruit) were variable. The highest level of infestation of *Anastrepha* and Lonchaeidae was observed in *Z. joazeiro* in area 3. The lowest level of infestation of *Anastrepha* was in *P. guajava* in area 4, and the lowest level of infestation of Lonchaeidae was in *S. tuberosa* in area 3 (Table 3).

Nine parasitoids species, all native of Neotropical region, were recovered from frugivorous larvae. These were Doryctobracon areolatus (Szépligeti, 1911); D. brasiliensis (Szépligeti, 1911); Utetes anastrephae (Viereck 1913); Opius bellus Gahan, 1930 (Braconidae, Opiinae); Asobara anastrephae (Muesebeck 1958); and Asobara obliqua (Papp 1969) (Braconidae, Alysiinae); Aganaspis pelleranoi (Brèthes, 1924); Tropideucoila weldi Lima, 1940 (Figitidae, Eucoilinae); and Pachycrepoideus vindemmiae (Rondani, 1875) (Pteromalidae) (Table 2).

Six parasitoid species were associated with tephritids and lonchaeids (Table 4). Aganaspis pelleranoi and D. areolatus parasitized the highest number of fruit fly species, whereas As. obliqua, O. bellus, and P. vindemmiae were obtained from just one unique fruit fly host.

Doryctobracon areolatus was the most abundant parasitoid and associated with *An. obliqua*, *An. sororcula*, and *An. zenildae* (Table 4). It was collected in both ecosystems (guava orchards and dry forest fragments), confirming the abundance of this parasitoid in the northern of Minas Gerais as observed by Alvarenga et al. (2009), and its potential relevance for biological control programs. This parasitoid stands out in various regions of Brazil for its abundance and frequency (Zucchi 2008).

The other braconids were restricted to a smaller number of fruit hosts. *Doryctobracon brasiliensis* was found only on guava. *Utetes anastrephae* parasitized

Table 2. Parasitoid and their association with fruit flies in orehards of guava and fragments of dry forest in Minas Gerais, Brazil, from March 2005 to March 2006

$egin{array}{c} L \ A \ \end{array}$ Figitidae A	Doryctobracon areolatus Doryctobracon brasiliensis Utetes anastrephae Asobara anastrephae	1 28 1 102 12 34 6 8 2 2 1	0.18 5.15 0.18 18.75 2.21 6.25 1.10 1.47 0.37	An. zenildae An. zenildae An. Sororcula An. zenildae; An. sororcula An. zenildae; An. fraterculus An. zenildae; An. sororcula; An. fraterculus An. zenildae; An. obliqua; An. fraterculus An. zenildae; An. obliqua; An. turpiniae An. zenildae; An. sororcula	Psidium guajava Ziziphus joazeip Psidium guajava Psidium guajava Psidium guajava Psidium guajava Psidium guajava Psidium guajava
L A Figitidae A	Doryctobracon brasiliensis Utetes anastrephae	28 1 102 12 34 6 8 2 2	5.15 0.18 18.75 2.21 6.25 1.10 1.47 0.37 0.37	An. zenildae An. Sororcula An. zenildae; An. sororcula An. zenildae; An. fraterculus An. zenildae; An. sororcula; An. fraterculus An. zenildae; An. obliqua; An. fraterculus An. zenildae; An. sororcula	Ziziphus joazeiro Psidium guajava Psidium guajava Psidium guajava Psidium guajava Psidium guajava Psidium guajava
Figitidae A	Utetes anastrephae	1 102 12 34 6 8 2 2	0.18 18.75 2.21 6.25 1.10 1.47 0.37 0.37	An. Sororcula An. zenildae; An. sororcula An. zenildae; An. fraterculus An. zenildae; An. sororcula; An. fraterculus An. zenildae; An. obliqua; An. fraterculus An. zenildae; An. obliqua; An. turpiniae An. zenildae; An. sororcula	Psidium guajava Psidium guajava Psidium guajava Psidium guajava Psidium guajava Psidium guajava
Figitidae A	Utetes anastrephae	102 12 34 6 8 2 2	18.75 2.21 6.25 1.10 1.47 0.37 0.37	An. zenildae; An. sororcula An. zenildae; An. fraterculus An. zenildae; An. sororcula; An. fraterculus An. zenildae; An. obliqua; An. fraterculus An. zenildae; An. obliqua; An. turpiniae An. zenildae; An. sororcula	Psidium guajava Psidium guajava Psidium guajava Psidium guajava Psidium guajava
Figitidae A	Utetes anastrephae	12 34 6 8 2 2	2.21 6.25 1.10 1.47 0.37 0.37	An. zenildae; An. fraterculus An. zenildae; An. sororcula; An. fraterculus An. zenildae; An. obliqua; An. fraterculus An. zenildae; An. obliqua; An. turpiniae An. zenildae; An. sororcula	Psidium guajava Psidium guajava Psidium guajava Psidium guajava
Figitidae A	Utetes anastrephae	34 6 8 2 2	6.25 1.10 1.47 0.37 0.37	An. zenildae; An. sororcula; An. fraterculus An. zenildae; An. obliqua; An. fraterculus An. zenildae; An. obliqua; An. turpiniae An. zenildae; An. sororcula	Psidium guajava Psidium guajava Psidium guajava
Figitidae A	Utetes anastrephae	6 8 2 2 1	1.10 1.47 0.37 0.37	An. zenildae; An. obliqua; An. fraterculus An. zenildae; An. obliqua; An. turpiniae An. zenildae; An. sororcula	Psidium guajava Psidium guajava
Figitidae A	Utetes anastrephae	8 2 2 1	1.47 0.37 0.37	An. zenildae; An. obliqua; An. turpiniae An. zenildae; An. sororcula	Psidium guajava
Figitidae A	Utetes anastrephae	2 2 1	0.37 0.37	An. zenildae; An. sororcula	
Figitidae A	Utetes anastrephae	2 1	0.37		Psidium guajava
Figitidae A		1			
Figitidae A				An. zenildae; An. fraterculus	Psidium guajava
Figitidae A		3	0.18	An. zenildae; An. sororcula; An. fraterculus	Psidium guajava
Figitidae A			0.55	An. zenildae; An. obliqua; An. fraterculus	Psidium guajava
Figitidae A	Asobara anastrephae	1	0.18	An. zenildae	Ziziphus joazeiro
т	,	1	0.18	An. zenildae	Psidium guajava
т		4	0.74	An. zenildae	Ziziphus joazeiro
т		1	0.18	An. sororcula	Psidium guajava
т		15	2.76	An. zenildae; An. sororcula	Psidium guajava
	Aganaspis pelleranoi	13	2.39	An. zenildae	Psidium guajava
		1	0.18	An. zenildae; An. sororcula	Ziziphus joazeiro
		46	8.46	An. zenildae; An. sororcula	Psidium guajava
		1	0.18	An. zenildae; An. fraterculus	Psidium guajava
		1	0.18	An. zenildae; An. fraterculus; An.sororcula	Psidium guajava
		2	0.37	An. zenildae; An. obliqua; An. fraterculus	Psidium guajava
	Tropideucoila weldi	1	0.18	An. fraterculus; An. zenildae; An. sororcula	Psidium guajava
	.,			,,,,,,	8
Braconidae L	Doryctobracon areolatus	177	32.54	An. zenildae	Ziziphus joazeiro
	3	6	1.10	An. zenildae	Psidium guajava
		i	0.18	An. sororcula	Psidium guajava
		1	0.18	An. zenildae; An. sororcula	Ziziphus joazeiro
T.	Utetes anastrephae	26	4.78	An. zenildae	Ziziphus joazeiro
_		1	0.18	An. zenildae: An. sororcula	Ziziphus joazeiro
A	Asobara anastrephae	4	0.74	An. zenildae	Ziziphus joazeiro
	2505ara arrasir opriac	i	0.18	An. sororcula	Psidium guajava
		10	1.84	An. zenildae	Psidium guajava
Figitidae A	Aganaspis pelleranoi	1	0.18	An. zenildae	Psidium guajava
1.8	agantaspis petieranoi	î	0.18	N. pendula; Neosilba sp.	Psidium guajava
Area 3-experimental orchard		1	0.10	11. penetuta, 11costibu sp.	1 мани диадаса
	Doryctobracon areolatus	15	2.76	An. zenildae	Ziziphus joazeiro
	y ar contaction	5	0.92	An. obliqua	Spondias tuberosa
I	Utetes anastrephae	3	0.55	An. zenildae	Ziziphus joazeiro
	Asobara obliqua	1	0.18	N. pendula; Neosilba sp.	Ziziphus joazeiro
	Opius bellus	2	0.13	Anastrepha sp.	Ziziphus joazeiro
	Aganaspis pelleranoi	1	0.18	N. zadolicha; N. pendula; Neosilba sp.	Ziziphus joazeiro
Area 4–commercial orchard Pteromalidae	зданаэріз ренегиног	1	0.18	N. zadoucha; N. penatua; Neosuba sp. Neosilba sp.	Psidium guajava

only larvae in juá, which has adequate characteristics (small size; thin epicarp, and mesocarp) for the short ovipositor of this parasitoid (e.g., Sivinski et al. 1997, Carvalho 2005). Among the Alysiinae, we highlight the first record of *As. obliqua* in *Neosilba* sp. on juá in Brazil. *Asobara anastrephae* parasitizes *An. obliqua* on

mango in the State of Tocantins (Bomfim et al. 2007), *An. striata* on guava and cajá and *An. antunesi* on cajá in the State of Amapá (Silva et al. 2007), and it was recorded in Bahia (Bomfim et al. 2010). This is the first record of *As. anastrephae* in the state of Minas Gerais.

Table 3. Degree of infestation of Anastrepha and Lonchaeidae on fruit in the orchards and in the dry forest in Minas Gerais, Brazil

Host fruit	Total no. of pupae of Anastrepha	Number of pupae of Anastrepha per g of fruit	Total no. of pupae of Lonchaeidae	Number of pupae of Lonchaeidae/g of fruit
Area 1-smallholder farming				
Psidium guajava	4,680	2.128	163	0.309
Ziziphus joazeiro	202	0.158	55	0.034
Area 2-smallholder farming				
Psidium guajava	122	0.039	54	0.017
Ziziphus joazeiro	1,903	0.172	428	0.055
Area 3-experimental orchard				
Ziziphus joazeiro	212	3.068	317	3.282
Spondias tuberosa	298	0.107	0	0
Area 4-commercial orchard				
Psidium guajava	0	0	3	0.002

Table 4. Tritrophic relation between host fruits, frugivorous larvae, and their parasitoid in orchards of guava and in the fragments of dry forest in Minas Gerais, Brazil

Host plants	Host flies	Parasitoid species
Psidium guajava	Anastrepha zenildae	Aganaspis pelleranoi
Psidium guajava	Anastrepha sororcula	Asobara anastrephae
Psidium guajava	Anastrepha zenildae	Asobara anastrephae
Psidium guajava	Anastrepha sororcula	Doryctobracon areolatus
Psidium guajava	Anastrepha zenildae	Doryctobracon areolatus
Psidium guajava	Neosilba sp.	Pachycrepoideus vindemmiae
Spondias tuberosa	Anastrepha obliqua	Doryctobracon areolatus
Žiziphus joazeiro	Anastrepha zenildae	Asobara anastrephae
Ziziphus joazeiro	Anastrepha zenildae	Doryctobracon areolatus
Ziziphus joazeiro	Anastrepha sp.a	Opius bellus
Ziziphus joazeiro	Anastrepha zenildae	Utetes anastrephae

a Male.

236

Aganaspis pelleranoi was the unique figitid associated exclusively with fruit fly and lance fly larvae. This parasitoid is common and widely distributed in Brazil (Guimarães et al. 2004). The figitid *T. weldi* parasitized *An. fraterculus*, *An. zenildae*, and *An. sororcula*, as well as drosophilids in guava. This species previously had been associated only with *N. pendula* (Lonchaeidae) (Guimarães et al. 2003). *Pachycrepoideus vindemmiae* was associated with *Neosilba* sp. (first report in a lonchaeid species) in guava (Table 4).

Fruit fly research and monitoring programs have been implemented in the irrigated perimeters of Jaíba and Gorutuba, in the northern of Minas Gerais state, since 1994 (Canal-Daza 1997, Alvarenga et al. 2009). Besides parasitoid species discussed herein, *D. fluminensis* (Costa Lima 1938) and *U. anastrephae*; *O. bellus* and *Diachasmimorpha longicaudata* (Ashmead) (introduced species) also were recorded in this region.

We observed differences in species composition of parasitoids between guava orchard and dry forest. Asobara obliqua, O. bellus, and U. anastrephae were collected exclusively in forest areas, whereas D. brasiliensis, P. vindemmiae, and T. weldi were found only in orchards (Table 5). However, Ag. pelleranoi, As. anastrephae and D. areolatus were collected in both environments. The latter species was the most abundant in both, but with higher numbers in forest sites.

Higher number of species of parasitoids was found in areas with greater diversity of crops and lower pesticide use, namely area 1 (six species), area 3 (five species) and, area 2 (four species). In areas 1 and 2, the orchards were subject to minimal management, consisting basically of routine pruning. These two areas

Table 5. Composition of parasitoids in the orchards and in the dry forest and in Minas Gerais, Brazil

Orchard	Forest		
Aganaspis pelleranoi	Aganaspis pelleranoi		
Asobara anastrephae	Asobara anastrephae		
Doryctobracon areolatus	Asobara obliqua		
Doryctobracon brasiliensis	Doryctobracon areolatus		
Pachycrepoideus vindemmiae	Opius bellus		
Tropideucoila weldi	Utetes anastrephae		

also presented higher number of fruit trees in neighboring areas. The characteristics of these areas propitiated a constant succession of fruit flies in the orchards, which was reflected in the higher number of species of parasitoids. Guava production in these areas is extensive and many fruit are left on the ground. Area 3 was different from the other two because its orchard was not for commercial purposes and contained a smaller number of guava trees. Area 4 was subject to intensive, systematic management, with few fruit on the ground, and featuring intensive use of chemical pesticides applied every 15 d to the guava orchard and surrounding orchards. Consequently, just one species of parasitoid was collected in area 4 (Table 2).

The natural parasitism of fruit flies is affected mainly by the host fruit, host flies, and location (Canal-Daza and Zucchi 2000). It would be interesting to emphasize that in northern Minas Gerais, the parasitoid species that occur in unperturbed environments (dry forests) are practically the same of those found in commercial orchards in other Brazilian regions.

The neighboring vegetation fragments with greater availability of native fruit also contributed to higher number of species. This can be exemplified by areas 1, 2, and 3. In these areas, the dry forest fragments were less affected and had more food resources available to the fruit flies. In area 3, there was higher number of native fruit fly host plants, and consequently in this area the variety of parasitoid species was high. The fragment in area 4 was deeply degraded, with low availability of native fruits. This was reflected as lower incidence of parasitoids in this area. Therefore, our data reveal that the forest fragments adjacent to the orchards served as shelter for fruit fly parasitoids in a caatinga-cerrado ecotone. Moreover, we report for first time As. obliqua in Brazil and As. anastrephae and T. weldi in dry forests of Minas Gerais State.

Acknowledgments

We acknowledge the Minas Gerais Research Support Foundation (FAPEMIG) for granting financial support; Angélica M.M. Penteado Dias (UFSCar) for identification of Asobara obliqu; Jorge A. Guimarães (EMBRAPA) for help with Figitidae; and Pedro Strikis (UNICAMP) for identifying the specimens of Lonchaeidae.

References Cited

Altieri, M. A., E. N. Silva, and C. I. Nicholls. 2003. O papel da biodiversidade no manejo de pragas. Holos Editora. Ribeirão Preto, São Paulo, Brazil.

Aluja, M. 1999. Fruit fly (Diptera: Tephritidae) research in Latin America: myths, realities and dreams. An. Soc. Entomol. Bras. 28: 565–594.

Aluja, M., J. Rull, J. Sivinski, A. L. Norrbom, R. A. Wharton, R. Macias-Ordónez, F. Dias-Fleischer, and M. López. 2003. Fruit flies of the genus Anastrepha (Diptera: Tephritidae) and associated native parasitoids (Hymenoptera) in the Tropical Rainforest Biosphere Reserve of Montes Azules, Chiapas, Mexico. Environ. Entomol. 32: 1377–1385.

Alvarenga, C. D., C. A. Matrangolo, G. N. Lopes, M. A. Silva, E. N. Lopes, D. A. Alves, A. S. Nascimento, and R. A.

- Zucchi. 2009. Moscas-das-frutas (Diptera: Tephritidae) e seus parasitóides em plantas hospedeiras de três municípios do norte do Estado de Minas Gerais. Arq. Inst. Biol. 76: 195–204.
- Antunes, F. Z. 1994. Caracterização climática. Inf. Agrop. 17: 18–19.
- Bomfim, D. A., M. A. Uchôa-Fernandes, and M.A.L. Bragança. 2007. Hosts and parasitoids of fruit flies (Diptera: Tephritoidea) in the State of Tocantins, Brazil. Neotrop. Entomol. 36: 984–986.
- Bomfim, Z. V., R. S. Carvalho, and C.A.L. Carvalho. 2010. Relações interespecíficas entre parasitóides nativos de moscas-das-frutas e o braconídeo exótico *Diachasmimor*pha longicaudata em frutos de 'umbú-cajá'. Cienc. Rural 40: 77–82.
- Canal-Daza, N. A. 1997. Levantamento, flutuação populacional e análise faunística das espécies de moscas-dasfrutas (Dip., Tephritidae) em quatro municípios do Norte de Minas Gerais. Ph.D. Dissertation, Escola Superior de Agricultura Luiz de Queiroz/Universidade de São Paulo, Piracicaba, Brazil.
- Canal-Daza, N. A. and R. A. Zucchi. 2000. Parasitóides Braconidae, pp. 119–126. In A. Malavasi and R. A. Zucchi (eds.), Moscas-das-frutas de Importância Econômica no Brasil: Conhecimento Básico e Aplicado. Holos Editora, Ribeirão São Paulo, Brazil.
- Carvalho, R. S. 2005. Avaliação das liberações inoculativas do parasitóide exótico *Diachasmimorpha longicaudata* (Ashmead) (Hymenoptera: Braconidae) em pomar diversificado em Conceição do Almeida, BA. Neotrop. Entomol. 34: 799–805.
- Drummond, G. M., C. S. Martins, A.B.M. Machado, F. A. Sebaio, and Y. Antonini. 2005. Biodiversidade em Minas

- Gerais: um atlas para sua conservação. Fundação Biodiversitas, Belo Horizonte, Minas Gerais.
- Guimarães, J. A., F. E. Gallardo, N. B. Diazand, and R. A. Zucchi. 2003. Eucoilinae species (Hymenoptera: Cynipoidea: Figitidae) parasitoids of fruit-infesting dipterous larvae in Brazil: identity, geographical distribution and host associations. Zootaxa 278: 1–23.
- Guimarães, J. A., M. F. Souza Filho, A. Raga, and R. A. Zucchi. 2004. Levantamento e interações tritróficas de figitídeos (Hymenoptera: Eucoilinae) parasitóides de larvas frugívoras (Diptera) no Brasil. Arq. Inst. Biol. 71: 51–56.
- Klink, C. A., and R. B. Machado. 2005. Conservation of the Brazilian Cerrado. Conserv. Biol. 19: 707–713.
- Kruess, A., and T. Tscharntke. 2000. Species richness and parasitism in a fragmented landscape: experiments and field studies with insects on *Vicia sepium*. Oecologia 122: 129–137
- Silva, R. A., D. B. Nascimento, E. G. Deus, G. D. Souza, and L.P.S. Oliveira. 2007. Hospedeiros e parasitóides de Anastrepha spp. (Diptera: Tephritidae) em Itaubal do Piririm, Estado do Amapá, Brasil. Cienc. Rural 37: 557– 560.
- Sivinski, J., M. Aluja, and M. Lopes. 1997. Spatial and temporal distributions of parasitoids of Mexican Anastrepha species (Diptera: Tephritidae) within the canopies of fruit trees. Ann. Entomol. Soc. Am. 90: 604–618.
- Zucchi, R. A. 2008. Fruit flies in Brazil Anastrepha species and their hosts plants. (http://www.lea.esalq.usp.br/anastrepha/).

Received 13 May 2011; accepted 11 January 2012.