

PARAMETER EVALUATION IN MICHAELIS-MENTEN KINETICS

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ABSTRACT

Parameter estimation reliability in enzyme kinetics depends upon the substrate range concentrations under assay. An inappropriate concentration set may lead to spurious values of K_m and V_{max} in the Michaelis-Menten approach. In this paper, the theoretical arguments for a practical criterium concerning the best work range of substrate concentrations are discussed on a velocity ratio basis (V_1/V_n) as response to the pertinent substrate concentration ratio (S_1/S_n).

INTRODUCTION

The Michaelis-Menten equation (1) has been subjected to a variety of comments on what may be referred to as textbook distortions, such as goodness-of-fit for estimates of error (1) and the comparison of reciprocal plots with microcomputer

softwares (2).

$$V = \frac{V_{max} \cdot S}{K_m + S} \quad (*) \quad (1)$$

In fact, inaccurate graphs of velocity vs concentration are particularly common in biochemical textbooks as has been pointed out recently by NAQUI (3), who alludes to the fact that in plots of V vs S the expression (1) applies

$$\left(\frac{dV}{dS}\right)_{S=0} = V_{max}/K_m \quad (II)$$

As a suggestion we should like to recall a further helpful drawing aid: the slope (III)

$$\left(\frac{dV}{dS}\right)_{S=K_m} = V_{max}/4K_m. \quad (III)$$

At this point $V = V_{max}/2$ as evidenced by the Michaelis-Menten equation (1) for $S = K_m$.

The estimatives of parameters (K_m and V_{max}) by linearization (such as in Lineweaver-Burk representations, FIGURE 1) are at present somewhat obsolete having been superseded by computer techniques (4).

(*)

The following symbols apply:

V: Initial velocity of an enzyme-catalyzed reaction, obeying Michaelis-Menten kinetics; V_{max} : Maximum conventional velocity of such a reaction; K_m : Michaelis-Menten constant; S_i ($i = 1 \dots n$): Substrate concentration, $S_1 > \dots > S_n$; GC: Geometric Center. $GC = (S_1 \times S_n)^{1/2}$

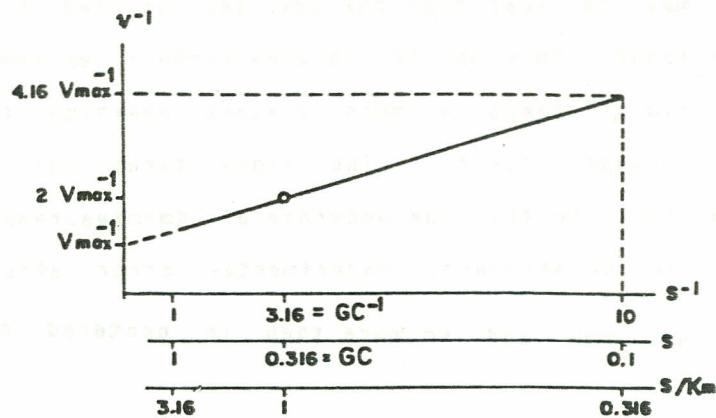


FIGURE 1. Lineweaver-Burk plot. K_m lies within the experimental range $S_n/S_1 = 10$; $K_m = 0.316$.

This, however, does not exempt from searching for the most reliable range of S 's in order to minimize the consequences of experimental error. As a premise, interpolations cause smaller distortions than extrapolations. In other words, a range of experimental S 's encompassing K_m is preferable to those not comprising it.

In enzyme kinetics the array of S 's represents the independent variable and that of V 's the dependent variable.

By geometric arguments it is well established that slopes near unit yield more dependable readings than those deviating considerably from it (5).

In Lineweaver-Burk plots the slope representing Michaelis-Menten kinetics is K_m/V_{max} . The intersection of the straight line with the ordinate registers V_{max}^{-1} . So, $2 V_{max}^{-1}$ of the ordinate yields K_m on the abscissa (FIGURE 1). The value of K_m in this figure may be labelled as "centered" ($S_n < K_m < S_1$) identifying itself with GC .

If the range of experimental S 's does not encompass the hitherto unknown K_m -value, there follows one of two possible alternatives:

a) K_m may be less than the smallest applied S of the experimental range, $K_m < S_n$. In Lineweaver-Burk representations such a K_m^{-1} -value takes a more distant position than the experimental range itself. The slope turns out to be precariously low, so that the accuracy of K_m -measurements are endangered. In consequence, experimental error affects the computation of V_{max} and K_m more than in centered positions (FIGURE 2).

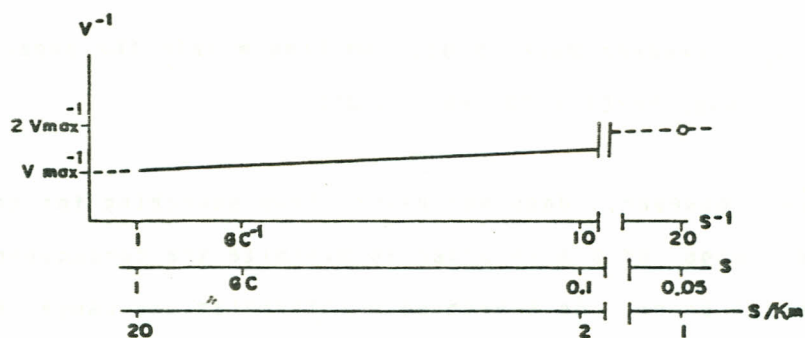


FIGURE 2. Lineweaver-Burk plot. K_m is smaller than S_n , the smallest experimental value. $S_1/S_n = 10$; $K_m = 0.05$

b) K_m may turn out to be greater than the largest applied S , $K_m > S_1$. As a consequence in Lineweaver-Burk plots K_m^{-1} is located closer to the ordinate (FIGURE 3) which leads to unconveniently steep slopes, again imperiling accuracy of ordinate intersection readings (6, 7).

METHODOLOGY

An experimental set of data dealing with the beta-glucosidase of a fungus (*Humicola* sp) hydrolysing p-nitrophenyl beta-glucoside (8) may help make our point. The unabridged set of these observations runs from $S_n = 0.05$ to $S_1 = 2$, expressed in empirical units. FIGURE 4 records the observed initial velocity as a function of the applied substrate concentration by way of

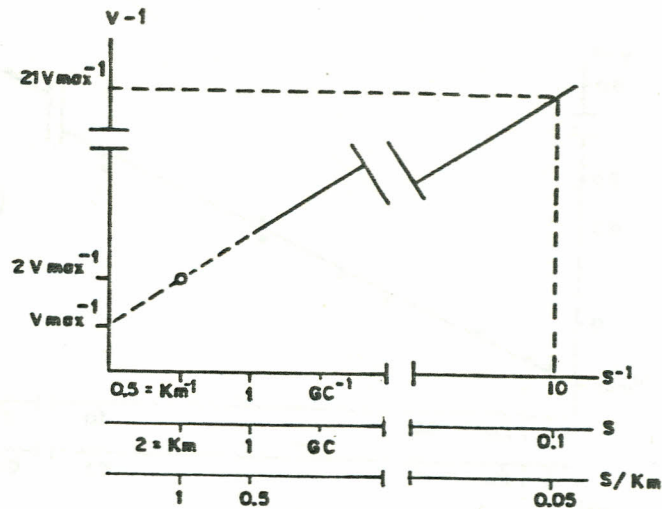


FIGURE 3. Lineweaver-Burk plot. K_m is larger than S_1 , the largest experimental value $S_1/S_n = 10$; $K_m = 2$

the Lineweaver-Burk representation. V_{max} and K_m have been evaluated in the usual way, while the geometric center between S_n^{-1} and S_1^{-1} is located at $GC^{-1} = (S_1 \times S_n)^{-1/2}$.

DISCUSSION

In a range, e. g., from $S_1^{-1} = 1$ to $S_n^{-1} = 10$, the geometric center lies at $S^{-1} = 3.16$ (see also FIGURE 1). The goodness-of-centering can be expressed by the ratio V_1/V_n where the subscripts correspond to those of their pertinent S 's. This ratio depends not only on the centering but also upon the width of the substrate range, but not on the empirical scale. This can be taken as a reliable criterium for the centering of K_m , stretching from unit ($K_m \ll S_n$, FIGURE 2) to S_1/S_n (for $K_m \gg S_1$, FIGURE 3): ten in the present case.

The original equation of Michaelis-Menten (1) in its "reduced" form, where $y_i = V_i/V_{max}$ and $x_i = S_i/K_m$, takes the shape

$$y_i^{-1} = 1 + x_i^{-1} \quad (IV)$$

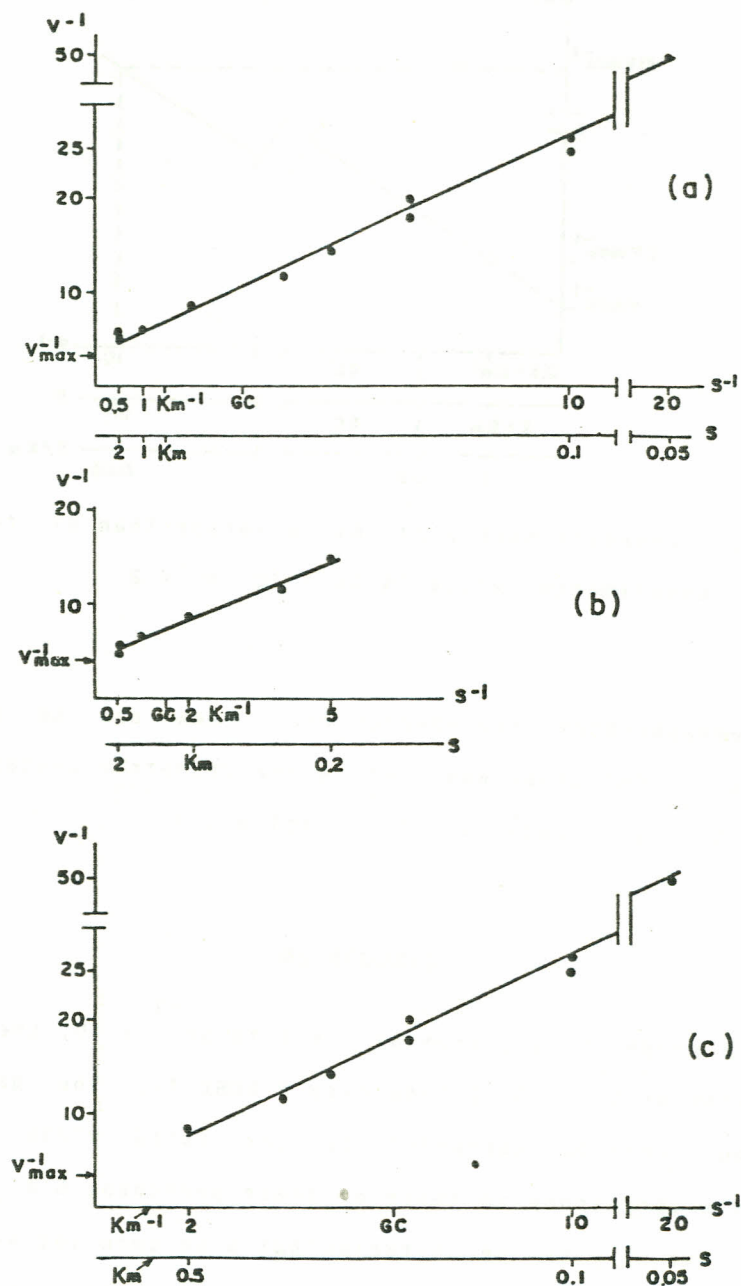


FIGURE 4. Lineweaver-Burk plot of a set of data dealing with the beta-glucosidase of a fungus (*HUMICOLA* sp) hydrolysing p-nitrophenyl beta-glucoside (8). GC: Geometric center. (a) The whole set ranging from $S_1 = 2$ to $S_n = 0.05$; (b) A subset from $S_1 = 2$ to $S_n = 0.2$; (c) A subset from $S_1 = 0.5$ to $S_n = 0.05$. Arbitrary units.

The velocity ratio $R = V_1/V_n$ may be expressed by

$$R = (1 + x_n^{-1}) / (1 + x_1^{-1}), \quad (V)$$

since it has the same meaning as y_1/y_n , the pertinent velocity ratio (FIGURE 5).

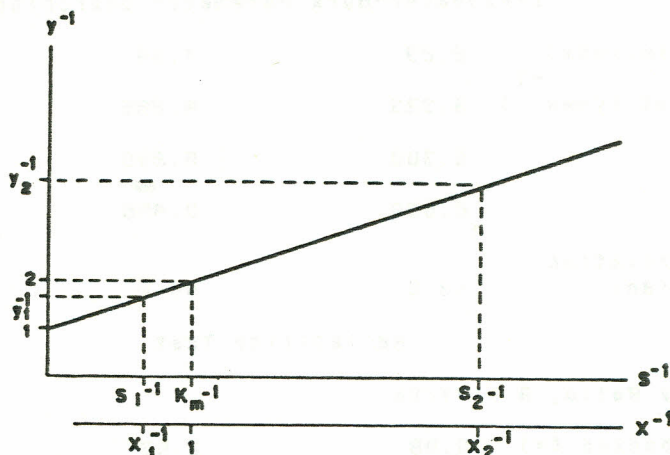


FIGURE 5. Michaelis-Menten equation in its reduced form, expressed through the Lineweaver-Burk plot. $x^{-1} = K_m/S$; $y^{-1} = V_{max}/V$.

For goodness-of-fit judgements arbitrary tolerance limits at 0.5 GC and 2 GC can be established, being equidistant from the GC in the geometric sense by 100%.

The Lineweaver-Burk data treatment to the above quoted Humicola set yields $V_{max} = 0.300$ and $K_m = 0.689$ (TABLE 1).

The correlation coefficient ($r^2 = 99.8\%$) discloses a highly reliable set: On the other hand the K_m value 0.689 lies far apart from the geometric center (GC = 0.316) of the whole set: its position lies above the proposed tolerance limits 0.158 (= 0.316 x 0.5) and 0.632 (= 0.316 x 2). So the assayed range of S's turns out to be rather low for K_m evaluation notwithstanding the fact that K_m still lies within the applied S's. To emphasize the

TABLE I - Parameter analysis and reliability test of a set of data dealing with the beta-glucosidase of a fungus (*HUMICOLA* sp) hydrolysing p-nitrophenyl beta-glucoside (8).

	Range of S's (S _n to S ₁)		
	Whole Set	Bisected Sets	
		0.05 to 2	0.2 to 2
Lineweaver-Burk Parameter Searching			
Slope (Km/Vmax)	2.29	1.94	2.33
Intercept (Vmax ⁻¹)	3.333	4.255	2.786
Vmax	0.300	0.235	0.359
Km	0.889	0.456	0.838
r % - Correlation coefficient	99.6	98.8	99.7
Reliability Test			
Velocity Ratio, R = V ₁ /V _n			
Expected (*)	10.98	2.67	6.63
Observed (**)	9.40	2.89	5.91
R-Tolerance limits (***)			
Lower	3.86	2.23	2.23
Upper	10.37	4.49	4.49
Km ⁻¹	1.451	2.193	1.183
GC ⁻¹	3.16	1.58	6.32
(Km.GC) ⁻¹	4.590	3.467	7.547
u = ln [1/(Km.GC)] ⁻¹	1.524	1.243	2.021
y' at u	1.702	1.439	2.420

(*) From the fitted Lineweaver-Burk straight line.

(**) From experimental data.

(***) Replacing Km either by 0.5 x GC (lower limit) or by 2 x GC (upper limit) on equation (V).

importance of the proper choice of S's, we bisected the whole range into one portion of low and another one of high S's, each fragment comprising a ratio of S₁/S_n = 10. Details of fractioning and results can be followed in TABLE I as well as in FIGURES 4 and 6.

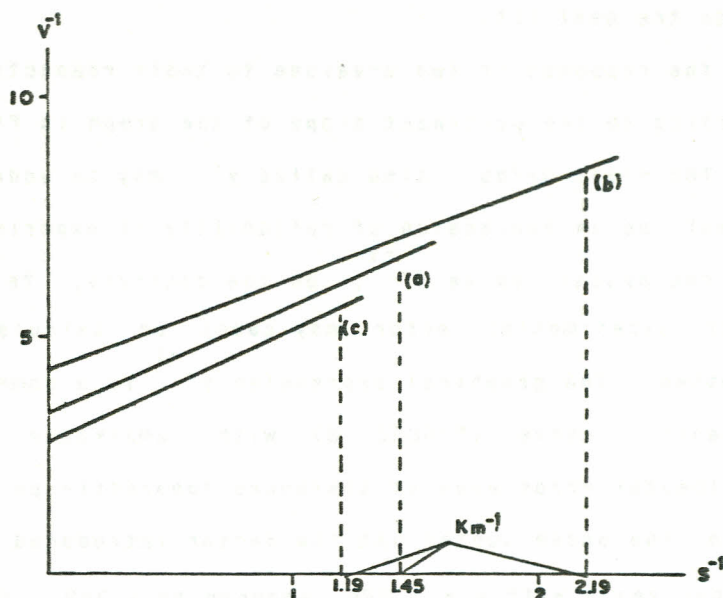


FIGURE 6. Set of data used in FIGURE 4, demonstrating the interference of range-width and position on K_m and V_{max} estimations. (a) The whole set ranging from $S_1 = 2$ $S_n = 0.05$; (b) A subset from $S_1 = 2$ to $S_n = 0.2$; (c) A subset from $S_1 = 0.5$ to $S_n = 0.05$.

In search for the best position of a range of S 's it seems advisable to introduce a logarithmic scale (5, 9) whereby unified S_1/S_n ratios acquire equal lengths. So, e. g., a range chosen as $S_1/S_n = e$, i. e.,

$$\ln S_1 - \ln S_n = 1, \quad (VI)$$

has only one definite length, wherever the range may lie.

Introducing $x = e^u$, the equation (IV) assumes the form

$$y^{-1} = 1 + e^{-u} \quad (VII)$$

A plot of y vs u produces the graph in FIGURE 7. In this figure one may verify the assumption that a range in the neighbourhood of $u = 0$ exhibits the highest slope. As a consequence the experimental error has the smallest influence on

the results in this region. Thus, a range in such a position yields the best fit.

The response of two u -values to their respective V 's varies according to the pertinent slope of the graph in FIGURE 7.

The slope dy/du , also called y' , may be understood in this context as an expression of reliability of experimental values. Its reciprocal value y'^{-1} , on the contrary, tells about the effect experimental error may cause on calculated parameter estimates. The graphical expression y'^{-1} vs u comes out to be an exponential curve (FIGURE 8) with remarkable influences of experimental error even at distances apparently as small as $u = \pm 2$. In the above quoted set the factor introduced by a somewhat improper range with $u = 1.524$ amounts to 1.702. The correct K_m -value of the considered *Humicola* experiment thus may be found through the expression $\ln K_m \pm \ln(y'^{-1})$. In consequence, taking into account the whole set, labelled by $K_m = 0.689$ and $y'^{-1} = 1.702$ (TABLE 1), the most plausible value for K_m lies between 1.173 (= 0.689×1.702) and 0.405 (= $0.689/1.702$). This span for the site of K_m refers to what may be called a "systematic" error. It contributes to the error obtained through inevitable

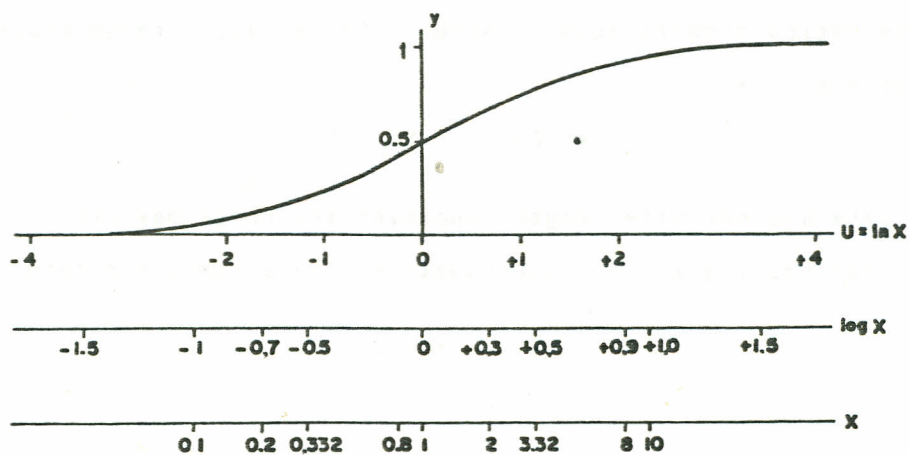


FIGURE 7. Reduced Michaelis-Menten equation $y = \frac{1}{1 + e^{-u}}$.
Substrate concentrations x are recorded as e^u .

measurement shortcomings expressed by its standard deviation. TABLE 1 also makes clear that the range from 0.2 to 2 offers a more dependable reading for K_m than the improperly centered though broader range from 0.05 to 2. An analogous result can be drawn of course for V_{max} -evaluations.

In conclusion we suggest the following procedure for a reasonably dependable estimate of K_m and V_{max} : In a preliminary trial, one may choose two S 's with the ratio $S_1/S_n = 10$. If the observed V 's turn out to result in a velocity ratio $R < 2.2$, measurements with lower S 's should be tried. On the other hand, if $R > 4.5$, higher S 's should be tested. These two arbitrary R -limits are then a practical prior criterium of reliability in enzyme kinetics studies.

Such recommendations should not lead one to underestimate the otherwise useful information on low and high S 's. So, cooperativity, positive as well as negative, shows a peculiar profile at low S/K_m , while substrate-excess inhibition itself is more strongly characterized the higher S/K_m is.

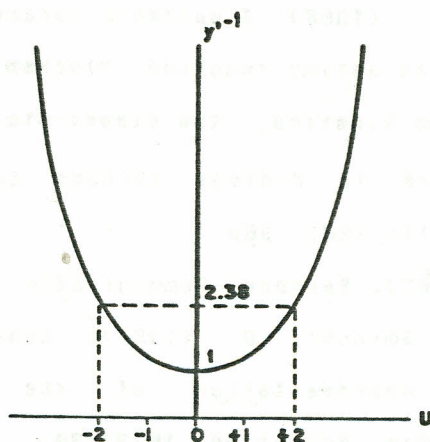


FIGURE 8. Reduced and derived Michaelis-Menten equation. Effect of experimental errors $y' = e^{-u} + 2 + e^{-u}$ as function of u . The y' -values were still normalized by dividing them by 4, yielding $y' = 1$ when $u = 0$. See text for further details.

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