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Acaricide resistance of Rhipicephalus (Boophilus) microplus in State of Mato Grosso do Sul, Brazil

Resistência do Rhipicephalus (Boophilus) microplus aos acaricidas no Estado de Mato Grosso do Sul, Brasil

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Abstract

This study was conducted to obtain an epidemiological view of acaricide resistance in populations of Rhipicephalus (Boophilus) microplus in the State of Mato Grosso do Sul. Twenty-four tick samples were collected from municipalities in the State where farmers had reported concerns about resistance to or failure of tick control. These ticks were subjected to in vitro resistance detection assays using the adult immersion test (AIT). The efficacy of alpha-cypermethrin, cypermethrin and amitraz treatments on samples collected throughout the State was generally poor. AIT showed efficacy ≥ 90% from the use of DDVP + chlorfenvinphos) (20 out of 21 municipalities), dichlorvos + cypermethrin (10 out of 16 municipalities) and cypermethrin + citronella + chlorpyrifos + piperonyl butoxide (20 out of 21 municipalities). PCR assays were used to detect the presence of pyrethroid resistance-associated sodium channel gene mutation. Larvae from three different populations that had previously been diagnosed as pyrethroid-resistant, through AIT, were evaluated. The PCR assays showed that the pyrethroid resistance-associated gene mutation was absent from these three populations. This study confirms that the emergence of resistance is a constant challenge for the livestock industry, and that development of resistance continues to be a major driver for new antiparasitic drugs to be developed.

Keywords: Tick, control, bovine, acaricide, PCR.

Resumo

Este estudo foi realizado com o objetivo de se obter uma visão epidemiológica da resistência aos acaricidas nas populações de Rhipicephalus (Boophilus) microplus no Estado de Mato Grosso do Sul. Vinte e quatro amostras de carrapatos foram coletadas em cidades do estado onde os fazendeiros relataram preocupação com resistência ou falha no controle dos carrapatos. Estes carrapatos foram submetidos a testes de detecção de resistência in vitro usando o teste de imersão de adultos (TIA). A eficácia do tratamento com alfa-cipermetrina, cipermetrina e amitraz foi geralmente pobre nas amostras coletadas em todo o Estado. Obteve-se eficácia de ≥90% no TIA com o DDVP + chlorfenvinphos (20 das 21 cidades), Diclorvós + Cipermetrina (10 das 16 cidades), e Cipermetrina + Citronela + Clorpirifós + Butóxido de piperonila (20 das 21 cidades). O ensaio da PCR foi utilizado para detectar a presença de mutação do gene do canal de sódio associado à resistência a piretróide. Avaliaram-se larvas de três diferentes populações onde anteriormente foi diagnosticada à resistência por TIA. Os ensaios da PCR mostraram que a mutação no gene associado à resistência ao piretróide estava ausente nessas três populações. Esse estudo confirma que o surgimento da resistência é um constante desafio para a pecuária e o desenvolvimento de resistência continua a ser um importante estímulo para o desenvolvimento de novas drogas antiparasitárias.

Palavras-chave: Carrapato, controle, bovinos, acaricida, PCR.

Introduction

Over 170 million cattle are managed by the Brazilian livestock industry (ANUALPEC, 2009). However, the industry's productivity is undermined by the tick species *Rhipicephalus* (*Boophilus*) *microplus* (Canestrini, 1887) (Acari: Ixodidae). In addition to the direct effects of *R.* (*B.*) *microplus* infestation on milk, meat and leather production, this tick species also serves as a vector for the infectious agents that cause bovine babesiosis and anaplasmosis. The annual economic loss suffered by the Brazilian livestock industry attributable to *R.* (*B.*) *microplus* parasitism is estimated to be over US\$ 2 billion (GRISI et al., 2002).

One common practice for controlling *R*. (*B*.) *microplus* is to use commercial acaricides and, historically, this proved to be an effective strategy for mitigating the tick's economic impact on the livestock industry. However, the evolution of acaricide resistance in populations of *R*. (*B*.) *microplus* is a cause of grave concern among cattle producers, government agencies and technical personnel.

Information concerning the regional distribution of *R. (B.) microplus* and the prevalence of acaricide resistance among Brazilian tick populations is limited. Consequently, selection and use of acaricides is based on criteria other than effectiveness, and applications are often inadequate, which may result in costly and ineffective treatment. Compounding this situation in Brazil, horn flies (*Haematobia irritans*) can be controlled using products with acaricidal activity (pyrethroids and organophosphates) at dosages that are insufficient to ensure tick control. This indirectly contributes towards increasing the likelihood that resistant tick populations will develop (BARROS et al., 2007).

The earliest records of acaricide resistance in Brazilian populations of *R.* (*B.*) *microplus* relate to arsenic use and date from 1953 (FREIRE, 1953). Resistance has been confirmed for organochlorines, organophosphates, synthetic pyrethroids (ARANTES et al., 1995), amidines and macrocyclic lactones (MARTINS; FURLONG, 2001; KLAFKE et al., 2006). There have not yet been any reports on the evolution of fluazuron or spinosad resistance in field populations of *R.* (*B.*) *microplus* (ANDREOTTI, 2010).

Between 1975 and 1984, Mexico conducted a national campaign against *R.* (*B.*) *microplus*. The campaign failed, and this was largely due to development of resistance to organophosphates. Since then, widespread and increasing resistance to organophosphate and pyrethroid acaricides in Mexico has become a major concern for the United States, since the organophosphate coumaphos is the only approved acaricide for use against cattle fever ticks in the United States (MILLER et al., 2005).

An evaluation on the susceptibility of *R*. (*B*.) *microplus* to acaricides in Mato Grosso do Sul within the last decade showed that pyrethroid effectiveness was generally less than 70%, although mixtures of DDVP-chlorfenvinphos and cypermethrin-chlorpyrifospiperonyl-citronella showed 97.68% and 100% effectiveness, respectively (KOLLER et al., 2009). There have not been any reports of mechanistic studies within these pyrethroid-resistant populations. Information on the molecular basis of resistance is important for tick control program managers, because molecular and biochemical diagnostic tests developed from knowledge of the mechanisms of resistance can be used to monitor acaricide

susceptibility and to measure the success of interventions involving rotation of acaricides with different modes of action.

A polymerase chain reaction-based assay to detect the presence of pyrethroid resistance-associated sodium channel gene mutation is a useful tool for investigating resistance mechanisms in cattle ticks. This assay detects a nucleotide difference that causes Phe to Ile amino acid substitution in the S6 transmembrane segment of domain III of the para-like sodium channel, and it clearly distinguishes heterozygotes from homozygotes (GUERRERO et al., 2001). Studies have shown that this mutation correlates with flumethrin, deltamethrin and cypermethrin resistance in Mexican tick populations (JAMROZ et al., 2000; ROSARIO-CRUZ et al., 2005).

The aim of the present study was to obtain an emerging view of the epidemiology of acaricide resistance among populations of *R.* (*B.*) *microplus* in the State of Mato Grosso do Sul.

Material and Methods

1. Tick samples

Adult females of *R.* (*B.*) *microplus* were collected from cattle in the State of Mato Grosso do Sul, which is located between the latitudes 17° 34' and 23° 56' S, and the longitudes 51° 40' and 57° 39' W. This area is characterized by an annual mean temperature of 26 °C (range: 17-35 °C), annual mean relative humidity of 80% and annual rainfall of 1,500 mm. Collection of these specimens formed part of a monitoring project in which farmers accessed the internet to send information and ticks to the laboratory at Embrapa Beef Cattle in Campo Grande, Mato Grosso do Sul.

In 2009, *R.* (*B.*) *microplus* ticks were collected from ranches in Mato Grosso do Sul where farmers had expressed concerns about resistance to or failure of tick control management and these ticks were subjected to *in vitro* resistance detection assays. Twenty-four samples were collected from the following 21 municipalities in Mato Grosso do Sul: Anastácio, Bataguassu, Caarapó, Chapadão do Sul (three farms), Corguinho, Dourados, Iguatemi, Inocência, Itaquiraí, Naviraí, Nova Alvorada do Sul, Paraiso, Paranaíba, Ponta Porã, Ribas do Rio Pardo, Rio Negro, Rochedo, São Gabriel do Oeste, Sidrolândia (two farms), Sonora and Terenos. The location of each municipality within the State is shown in Figure 1.

2. Adult immersion test (AIT)

The AIT was used to test fully engorged females, of size greater than 4.5 mm (DRUMMOND et al., 1973), under standard laboratory conditions of 28 °C and 80% relative humidity. Acaricidal products containing alpha-cypermethrin (0.015%), cypermethrin (0.015%) and amitraz (0.025%) as the single active ingredient were tested separately using the AIT. The following acaricidal products with active ingredient combinations were also tested: dichlorvos (0.1125%) + cypermethrin (0.0125%); chlorpyrifos (0.04%) + cypermethrin (0.0165); DDVP (0.15%) + deltamethrin (0.0025%); DDVP (0.15%) + chlorfenvinphos (0.05%); ethion (0.06%) + cypermethrin (0.008%); cypermethrin (0.015%) + citronella (0.001%) + chlorpyrifos (0.025%) + piperonyl butoxide

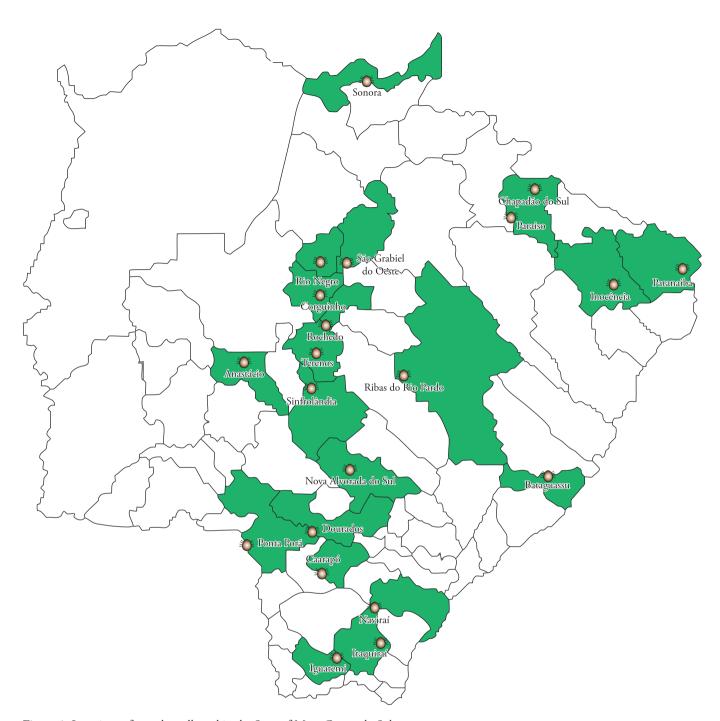


Figure 1. Locations of samples collected in the State of Mato Grosso do Sul.

(0.015%); and dichlorvos (0.15%) + chlorpyrifos (0.05%); DDVP (0.15%) + cypermethrin (0.1125%).

These products were commercially available and used in accordance with the manufacturer's recommendations and following the commercial dosages of active ingredient used by farmers. Each test was performed in duplicate with 20 females per group, with an average tick weight of 250 mg. The ticks were immersed for 5 minutes in the solution containing the acaricide diluted in water, after which they were dried with absorbent paper and transferred to Petri dishes. The ticks were fastened to

A colony of the Mozzo strain that originated from the region of the same name in Uruguay was established in our laboratory and has been maintained under standard laboratory conditions in the absence of acaricide pressure for multiple generations. This strain was used as a pyrethroid-sensitive reference strain.

3. DNA purification

A subset of females was allowed to oviposit, and the resulting larvae were used to purify genomic DNA. The larvae were frozen in plastic vials placed in dry ice or liquid nitrogen and were kept frozen, in storage, until the time of analysis. Individual larvae were then transferred onto a petri plate on dry ice and placed in individual 1.5 mL microcentrifuge tubes, which were also kept on dry ice.

Twenty microliters of sample buffer (100 mM Tris, pH 8.3; 500 mM KCl) was added to each tube. A disposable pellet pestle for 1.5 mL centrifuge tubes (Kontes, Vineland, NJ) was used to crush and grind the larva against the tube wall for 20 seconds, until close visual inspection ensured that the larva was broken into several fragments. The tube was transferred back to dry ice until a set of larvae had been prepared. The tubes were briefly microcentrifuged to ensure that their content of liquid and crushed larva were at the bottom of the tube, and then the tubes were placed in a boiling water bath for 3 minutes. After cooling, 1 μL was used for PCR as described by Guerrero et al. (2001).

Larval genomic DNA isolation and genotyping by means of PASA was performed as described by Guerrero et al. (2001) in thin-walled 0.5 mL microcentrifuge tubes using 20 μ L reactions optimized for primer annealing temperature and MgCl₂ concentration. The final optimized reaction conditions consisted of 1 μ L of genomic DNA homogenate from a single tick larva, 20 pmol of each primer, 10 mM Tris (hydroxymethyl) aminomethane hydrochloride pH = 8.3, 50 mM KCl, 0.05 mM of each dNTP, 1.75 mM MgCl₂ and 0.1 μ L of 1:1 vol/vol mix of AmpliTaq DNA polymerase (5 U. μ L⁻¹ stock) and TaqStart antibody (1.1 μ g, μ L⁻¹ stock).

Amplification was carried out using an MJ DNA engine programmed for 96 °C for 2 minutes followed by 37 cycles, each consisting of denaturation at 94 °C for 1 minute, annealing at 60 °C for 1 minute and extension at 72 °C for 1 minute. The program also included a final extension step at 72 °C for 7 minutes. The following primers were used: 5′-TTATCTTCGGCTCCTTCT-3′ (wild type-specific sense), 5′-TTATCTTCGGCTCCTTCA-3′ (mutant-specific sense) and 5′-TTGTTCATTGAAATTGTCGA-3′ (non-specific antisense). At least twenty individual larvae from each tick family were tested to determine kDr-genotype frequencies. Electrophoresis was carried out on 2.5% agarose gels, followed by ethidium bromide staining and viewing under UV illumination.

Results

1. AIT bioassay

Commercial products that are used in Mato Grosso do Sul were selected for use in this investigation and their use was based on the manufacturer's recommendations. Samples from 21 municipalities

that represent all the microregions of Mato Grosso do Sul were tested in this study.

Only alpha-cypermethrin, cypermethrin and amitraz were commercially available as products with a single active ingredient, for use in the AIT evaluations. Table 1 shows that at the manufacturer's recommended application doses, the efficacy of the alphacypermethrin, cypermethrin and amitraz treatments was generally poor, on samples collected throughout the State. Table 2 shows the results from eight products that consisted of more than one active ingredient. The results indicated that acceptable tick control would be achieved on most farms across the State with three of the combination products. The AIT showed efficacy ≥90% from the use of DDVP + chlorfenvinphos (20 out of 21 farms), dichlorvos + cypermethrin (10 out of 16 farms) and cypermethrin + citronella + chlorpyrifos + piperonyl butoxide (20 out of 21 farms). Two mixtures showed <90% efficacy in samples from every municipality: ethion + cypermethrin and DDVP + cypermethrin. The combinations of dichlorvos + chlorpyrifos, DDVP + deltamethrin and chlorpyrifos + cypermethrin showed mixed results depending on the specific city sample.

Ticks from the farm near Sonora were the most susceptible, since the efficacy was ≥90% from all the products tested, including those containing a single active ingredient. The Rio Negro sample was susceptible to 7 of the 11 active ingredient products tested. Conversely, the sample from Dourados was only susceptible to the combination of four active ingredients consisting of cypermethrin + citronella + chlorpyrifos + piperonyl butoxide. The Dourados region has a large number of dairy cattle operations, and the preferred breed in the Brazilian dairy production industry is based on *Bos taurus* and crossbred bovines, which require more intensive use of acaricides to control ticks.

2. kDr - genotype frequency

The presence of a sodium channel gene mutation in *R.* (*B.*) *microplus* from the municipalities of Paraiso, Rio Negro and Rochedo was assessed by testing 20 individual larvae from each sample. These samples were all pyrethroid resistant, as determined by the AIT assay. Only the wild-type pyrethroid-susceptible alleles were detected in our tests (data not shown). This assay only detects the single nucleotide difference that results in phenylalanine to isoleucine amino acid substitution in the S6 transmembrane segment of domain III. Although this mutation correlates with pyrethroid target-site resistance in Mexican ticks, its absence in these three samples from Mato Grosso do Sul does not preclude the presence of other sodium channel gene mutations or elevated metabolic enzyme activity that can cause pyrethroid resistance in *R.* (*B.*) *microplus*.

Discussion

The poor results from amitraz are of interest because this is one of the products most used in the State of Rio Grande do Sul, yet it only provided effective control in six of the 42 farms. However, in a previous report, amitraz in combination with chlorpyrifos was the most efficient product (CAMILLO et al., 2009). In surveys

Table 1. Efficacy of acaricides in products containing a single active ingredient against *R.* (*B.*)*microplus* on farms in municipalities in the State of Mato Grosso do Sul.

Municipalities	lities Active Ingredient*							
	Alpha- Cypermethrin		Amitraz					
	cypermethrin	1						
Anastácio	63	89	52					
Bataguassu	14	28	39					
Caarapó	36	_**	-					
Chapadão do Sul	54	70-75	46-70					
Corguinho	3	100	1					
Dourados	30	52	26					
Iguatemi	2	71	62					
Inocência	-	-	95					
Itaquiraí	11	85	0					
Naviraí	76	96	99					
Nova Alvorada Sul	-	-	-					
Paraíso	3	-	-					
Paranaíba	2	72	56					
Ponta Porá	66	88	75					
Ribas do Rio Pardo	29	51	61					
Rio Negro	47	100	100					
Rochedo	0	-	79					
São Gabriel	0	66	0					
Sidrolândia	3-60	-	32- 96					
Sonora	100	100	-					
Terenos	10	-	84					

*Products containing one acaricide were tested at the application doses recommended by the manufacturer: alpha-cypermethrin (0.015%), cypermethrin (0.015%) and amitraz (0.025%). Efficacy is reported as the proportion (%) of the estimated reproduction (ER) of the adult females in a treatment group. **Not tested.

in other Brazilian States, amitraz alone also performed poorly (CAMPOS JÚNIOR et al., 2005; PEREIRA, 2006).

The ineffectiveness of alpha-cypermethrin and cypermethrin as single active-ingredient products in Mato Grosso do Sul was clear. This reflects patterns seen in earlier resistance surveys. In Rio Grande do Sul, cypermethrin alone was effective in only 65% of the tests (CAMILLO et al., 2009) and in the State of Bahia, deltamethrin was effective in only 65% of the tests (CAMPOS JÚNIOR et al., 2005). Fernandes (2001) reported that in the State of Goiás, ticks were resistant to commercial concentrations of deltamethrin and cypermethrin. Pereira (2006) reported that the efficiency of permethrin in the State of São Paulo was only 28%, and Mendes et al. (2007) reported resistance to cypermethrin at levels of 75 and 41.7%. Finally, Silva et al. (2005) reported that the resistance to cypermethrin in Paraiba was 70.5%.

The Brazilian Santa Luiza strain of the southern cattle tick (*R*. (*B*.) *microplus*) is also resistant to permethrin. Studies by Li et al. (2008) suggested that permethrin resistance was inherited as an incomplete recessive trait and that one major gene was responsible for permethrin resistance. Additionally, selection of F3 larvae with either permethrin or amitraz led to significantly increased resistance to both permethrin and amitraz, thus indicating a close linkage between the genes responsible for permethrin and amitraz resistance.

These results demonstrate that most drugs used for controlling *R*. (*B*.)*microplus* in Mato Grosso do Sul had low efficacy on adult ticks, as measured by *in vitro* tests, with similar results reported in other States. Clearly, selection for resistance to acaricides has occurred throughout the State. However, from the research presented here, it is not known whether the resistance levels detected among adult ticks *in vitro* amount to actual field control failure at the larval, nymph and partially engorged adult stages of these ticks. Adequate control may still be possible with these products, even with indications of resistance among engorged adult females, if the concentrations used are sufficient to kill other life stages that are present in the host animal, which may be more susceptible to this same concentration of acaricide (DAVEY et al. 2004).

For field efficacy measurements, there is no substitute for on-animal efficacy studies using a formulated acaricide with a known commercial concentration, strictly following the label recommendations. It is likely that acaricides that are effective on adult females would also be effective on earlier life stages. However, further research using the abovementioned techniques needs to be conducted on suspected resistant populations in order to confirm the resistance and study the mechanistic basis of this resistance, before strong conclusions can be drawn regarding the state of acaricide resistance in this area of Brazil. Another possibility to consider when addressing questions of acaricide resistance and suspected product failure is product quality. We did not chemically analyze the products used in our study, and active ingredient degradation could not conclusively be ruled out as a factor in the AIT results.

Currently, the level of resistance in a tick population is determined by means of bioassay techniques. The bioassay procedures are valuable for their portability and low cost. However, these tests do not reveal direct information about the genetics of the samples and cannot detect resistance in its early stages of development with much precision. It often takes several weeks to obtain results. An assay that could determine the resistance status of individual Brazilian ticks within a day would be helpful: hence our interest in the rapid diagnostic capability of the sodium channel PCR assay.

Susceptibility to synthetic pyrethroids was evaluated in Mexican R. (B.) microplus tick populations (ROSARIO-CRUZ et al., 2009). Knockdown resistance (kdr) PCR allele-specific assays were conducted in tick populations that were resistant to cypermethrin, flumethrin and deltamethrin. A significant correlation (p < 0.01) was found between presence of the sodium channel mutation, and resistance to synthetic pyrethroids as measured by means of PCR and the larval packet test (LPT) respectively. Just over half the populations (16/28) were cross-resistant to flumethrin, deltamethrin and cypermethrin, 21.4% of the samples (6/28) were susceptible to all of the three pyrethroids, 10.7% (3/28) were resistant to flumethrin, 3.4% (1/28) were resistant to deltamethrin only and 7.1% (2/28) were resistant to both flumethrin and deltamethrin. The presence of the kdr mutation correlated with resistance to synthetic pyrethroids as a class. Target-site insensitivity was the major mechanism of resistance to synthetic pyrethroids in Mexican R. (B.) microplus field strains, involving the presence of a sodium channel mutation. However, esterase-based mechanisms,

Table 2. Efficacy of acaricides in combination products against R. (B.) microplus on farms in municipalities in the State of Mato Grosso do Sul.

Municipalities	Acaricide combinations*								
	DLV CPT	CPF CPT	DDVP DMT	DDVP CFF	ETH CPT	CPT, CPF CTR, PBO	DLV CPF	DDVP CPT	
Anastácio	96	95	25	100	77	100	100	54	
Bataguassu	86	93	14	99	-	97	90	-	
Caarapó	_**	48	-	100	-	100	-	-	
Chapadão do Sul	91-99	94	4-58	100	64-89	82-100	43-94	50-80	
Corguinho	-	-	-	100	-	100	39	-	
Dourados	83	54	22	34	34	98	67	16	
Iguatemi	100	0	88	100	20	100	71	19	
Inocência	-	-	99	100	-	100	99	-	
Itaquiraí	100	94	13	100	62	100	100	15	
Naviraí	97	100	83	100	89	100	100	60	
Nova Alvorada Sul	-	-	-	90	12	100	12	-	
Paraíso	-	-	-	100	28	100	-	-	
Paranaíba	51	3	22	100	3	100	45	41	
Ponta Porá	100	-	-	100	88	100	100	75	
Ribas Rio Pardo	100	-	12	100	-	100	100	66	
Rio Negro	100	73	100	100	44	100	100	77	
Rochedo	63	78	0	100	-	100	-	-	
São Gabriel	88	0	43	100	24	100	56	16	
Sidrolândia	100	80	100	100	28	100	40-99	-	
Sonora	100	-	100	100	-	100	100	-	
Terenos	46	-	-	100	-	100	-		

Abbreviations: DLV: dichlorvos; CPT: cypermethrin; CPF: chlorpyrifos; CFF: chlorfenvinphos; DMT: deltamethrin; CTR: citronella; PBO: piperonyl butoxide; DDVP: DDVP; ETH: ethion. *Combination products were tested at the application dose recommended by the manufacturer: dichlorvos (0.1125%) + cypermethrin (0.0125%); chlorpyrifos (0.04%) + cypermethrin (0.0165); DDVP (0.15%) + deltamethrin (0.0025%); DDVP (0.15%) + chlorfenvinphos (0.05%); ethion (0.06%) + cypermethrin (0.008%); cypermethrin (0.015%) + citronella (0.001%) + chlorpyrifos (0.025%) + piperonyl butoxide (0.015%); dichlorvos (0.15%) + chlorpyrifos (0.05%); DDVP (0.15%) + cypermethrin (0.1125%). Efficacy is reported as the proportion (%) of the estimated reproduction (ER) of the adult females in a treatment group. **Not tested.

other mutations or combinations of mechanisms can also occur (JAMROZ et al., 2000; ROSARIO-CRUZ et al., 2009).

We did not detect the presence of this mutation in the three suspected pyrethroid-resistant samples tested in this study. We also tested seven other samples from other locations in the state and did not find the sodium channel gene mutation (data not shown). Potentially, there is a different resistance mechanism involved, such as esterase detoxification, as detected in Mexican ticks by Jamroz et al. (2000). Alternatively, there may be a different sodium channel gene mutation in these ticks. Jonsson et al. (2010) recently reported on a new sodium channel gene mutation that confers pyrethroid resistance in Australian ticks.

Our results provide further evidence supporting the notion that resistance to the products available in Brazil is developing. There is concern regarding the future availability of efficient products for tick control. Emergence of resistance is a constant challenge and resistance is always a major driver of new antiparasitic drug development.

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