

Functional response of *Telenomus remus* Nixon (Hymenoptera, Scelionidae) to *Spodoptera frugiperda* (J. E. Smith) (Lepidoptera, Noctuidae) eggs: effect of female age

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ABSTRACT. Functional response of *Telenomus remus* Nixon (Hymenoptera, Scelionidae) to *Spodoptera frugiperda* (J. E. Smith) (Lepidoptera, Noctuidae) eggs: effect of female age. Functional response of 24-h and 48-h-old *Telenomus remus* adults was studied on *Spodoptera frugiperda* eggs. The study was carried out in climatic chamber regulated at $25 \pm 1^\circ\text{C}$, $70 \pm 10\%$ RH and 12:12h (L:D). Females of *T. remus* were honey fed and individualized in glass vials along with 25, 50, 75, 100, 150, 200, 250 or 300 eggs of *S. frugiperda* for 24 h. Complete randomized design with ten replications was adopted. The parameters evaluated to construct the functional response curve were daily average parasitism, searching rate and oviposition time. It was observed that the higher the egg density, the higher the parasitism for 24-h and 48-h-old females although there was a tendency of parasitism stabilization at 150-egg density. The results showed a type II functional response curve for both 24-h and 48-h-old female.

KEYWORDS. Biological control; fall armyworm; oviposition time; parasitoid; search rate.

RESUMO. Resposta funcional de *Telenomus remus* Nixon (Hymenoptera, Scelionidae) sobre ovos de *Spodoptera frugiperda* (J. E. Smith) (Lepidoptera, Noctuidae): efeito da idade da fêmea. A resposta funcional de fêmeas de *Telenomus remus* com 24 h e 48 h de idade foi observada em ovos de *Spodoptera frugiperda*. O experimento foi conduzido em câmara climatizada regulada a $25 \pm 1^\circ\text{C}$, $70 \pm 10\%$ UR e fotofase de 12h. As fêmeas de *T. remus* foram alimentadas com mel e mantidas individualmente por 24h em tubos de vidro que continham 25, 50, 75, 100, 150, 200, 250 ou 300 ovos de *S. frugiperda*. Foi adotado o delineamento experimental inteiramente casualizado com 10 repetições. Os parâmetros avaliados para a construção da curva de resposta funcional foram o parasitismo médio diário, a taxa de busca e o tempo de oviposição. Foi observado que com o aumento da densidade de ovos também cresce o parasitismo em fêmeas com 24 h e 48 h de idade e que há uma tendência de estabilização do parasitismo na densidade de 150 ovos/fêmea. Os resultados mostraram uma curva de resposta funcional do tipo II para fêmeas de *T. remus* com 24 h e 48 h de idade.

PALAVRAS-CHAVE. Controle biológico; lagarta-do-cartucho; parasitóide; tempo de oviposição; tempo de busca.

Telenomus remus Nixon is an exclusive parasitoid of lepidopterous pests and several studies have been conducted worldwide to evaluate the efficiency of this natural enemy on *Spodoptera frugiperda* (J. E. Smith) eggs (Joshi *et al.* 1976; Gupta & Pawar 1985; Cruz & Figueiredo 1994; Morales *et al.* 2000). The parasitism of *T. remus* on *S. frugiperda*, *S. latifascia* (Walker), *S. exigua* (Hübner) and *S. eridania* (Cramer) under laboratory conditions varied from 80 to 100%, according to Wojcik *et al.* (1976). In Venezuela (Hernández *et al.* 1989) and other Central American countries (González & Zocco 1996; Cave & Acosta 1999; Cave 2000), studies have shown that *T. remus* can cause up to 90% of parasitism in *S. frugiperda* eggs, releasing 5,000 to 8,000 parasitoids/ha. Also in Venezuela, Ferrer (2001) reports the use of *T. remus* combined with *Trichogramma pretiosum* Riley (Hymenoptera, Trichogrammatidae) to control fall armyworm infestations in 1,600-ha maize field. This program caused 34% reduction in insecticide use.

In Brazil, the insect was first introduced in the mid 1980's aiming at evaluating its efficacy to control fall armyworm in

a classical biological control program. Despite that, very few studies have been conducted to evaluate the performance of *T. remus* under local conditions. However, as Cave & Gaylor (1989) highlighted, it is very important to study *Telenomus* species because they are parasitoids with great biological control potential.

Studies on the interaction of this natural enemy and its host, as the functional response, for instance, can be helpful in integrated pest management programs, once this facilitates to determine predation/parasitism dynamics of a natural enemy. Response to prey density can affect survivorship and fecundity and, therefore, the intrinsic rate of increase as stated by Hassel (1978). As this parasitoid has been mass reared on fall armyworm eggs for many years, the evaluation of functional response is necessary prior to any release.

The study carried out by Morales *et al.* (2001) in Venezuela evaluated only the functional response of 24-hour-old females and adopted polynomial regression to express it which is not appropriate to model the relationship between predation/parasitism rate and prey/host densities.

All functional response curves proposed by Holling (1959) stabilize at a certain predation/parasitism rate level, which is related to the maximum number of preys/hosts attacked per time unit. However, polynomial regression does not allow the determination of such a level. Thus, although it is easy to calculate, it is not helpful.

Several factors can affect the development of natural enemies. Among those factors, the most important is the density and the behavioral characteristics of the predator/parasitoid and prey/host (Holling 1961). When such factors are evaluated through mathematic models as suggested by Holling (1959), three different types of functional responses can be obtained, represented by discrete curves (Van Alphen & Jervis 1996).

The type II curve is more often related to systems involving arthropods, including predator and parasitoid insects (Cave & Gaylor 1989; Garcia 1991), although type III curve is often observed for egg parasitoids (Wang & Ferro 1998; Reay-Jones *et al.* 2006). The main characteristic of the type II functional response is a gradual increase of consumed preys, due to their greater availability, up to a density in which there is stabilization in the consumption rate (Hassel 1978; Hassel *et al.* 1997). Type II curve is a hyperbola that differentiates from the other types by presenting a gradual increase in the number of the eaten preys (Van Alphen & Jervis 1996) or parasitized hosts.

Below is the disk equation as suggested by Holling:

$$Na = (a * Tt * N) / (1 + a * Th * N) \quad (1)$$

In which: Na = number of eaten preys/parasitized hosts, a = predator/parasitoid searching rate, Tt = total exposure time, N = prey/host density, Th = handling/oviposition time.

The ability of predator females to search and attack hosts is a determinant factor to define its population size in a given area, while changes in foraging behavior can be responsible for success or failure of biological control programs (Lewis *et al.* 1990). Therefore, since functional responses can represent key factors in selecting natural enemies for a biological control program (Lester & Harmsen 2002), the present study aimed at demonstrating, by using the curves obtained from the Holling equation, how the *T. remus* egg parasitoid responds to the increase in the density of the eggs of *S. frugiperda*. Moreover, it was intended to demonstrate the effect of parasitoids' age in its functional response and, therefore, the feasibility of releasing older females in biological control programs.

MATERIAL & METHODS

Daily Average Parasitism. The experiment was conducted with the following *S. frugiperda* egg densities: 25, 50, 75, 100, 150, 200, 250 and 300 eggs. In order to obtain such densities eggs were counted multiplying the number of eggs in the superior layer by the number of deposition layers of each egg mass. The number of eggs on the edges was added to the total (Beserra & Parra 2005).

Ten parasitoid females were used for each egg density. The females were placed in glass tubes (8 cm height x 2 cm diameter) individually, and oviposition was allowed for 24 hours. The procedure was performed with 24 and 48 hour-old

T. remus females without previous egg laying experience. All females were fed with small drops of honey placed on the internal walls of the tube.

During the experiment, the tubes containing females and *S. frugiperda* eggs were kept in a controlled environment chamber at 25±1°C, RH 70±10% and 12 h of photophase. Newly hatched larvae were counted and withdrawn from the tubes with a brush so they would not be able to feed on parasitized eggs. Such observations were performed in the first four days after oviposition at 12-hour intervals. Parasitoids that emerged from the remaining eggs were counted after death, and sexed under stereoscopic microscope (Zeiss Stemi V6, Germany).

Oviposition time and searching rate. To determine oviposition time, 20 *T. remus* females from each of the treatments (24 and 48 hour-old individuals) were placed in Petri dishes (1 x 8 cm diameter) individually and exposed to approximately 100 *S. frugiperda* eggs. The time females were in contact with an egg was measured and all parasitoid activities were videotaped for time checking. During observation, the females were kept in a room with controlled temperature and humidity (25±1°C, RH 70±10%).

The searching rate was calculated as described by Cave & Gaylor (1989) and Morales *et al.* (2001), in which the number of parasitized eggs is divided by the total number of eggs exposed to the parasitoid.

Statistical Analysis. The experiment was arranged in completely randomized design and data were submitted to ANOVA. Means were compared by the Tukey test (P≤0.05). The Holling equation (1) was used to build the functional response curves. However, a model which incorporates depletion (host depletion in our case) was also used to analyze the functions according to Juliano (2001). This analysis was performed using PROC CATMOD (SAS Institute 2004).

RESULTS

Oviposition Time, Daily Mean Parasitism and Searching Rate. *T. remus* females did not show significant difference on oviposition time regardless age (df= 2; F= 12.07; P 0.05). Twenty-four-hour-old females spent 40.6 sec/egg (36.0 – 42.0 s/egg), whereas 48-h-old females spent 41.3 sec./egg (34.0 – 45.0 s/egg), on average.

The mean number of eggs parasitized by *T. remus* females increased with higher egg host densities, and stabilized at and beyond the density of 150 eggs (Table I). It can be observed that the age was not a major factor influencing the searching rates as the egg density. *T. remus* showed a decrease in the searching rates beyond the density of 150 eggs (Table I).

Sex Ratio. There was an increase tendency towards females in the progeny of *T. remus* as the age of female parasitoids increased (Table II), indicating that female age affects the sex ratio in the progeny. However, there is no influence of host density on the proportion of males and females in the progeny.

Functional Response. The results for 24-h and 48-h-old *T. remus* females allowed the adjustment of a Type II and Type

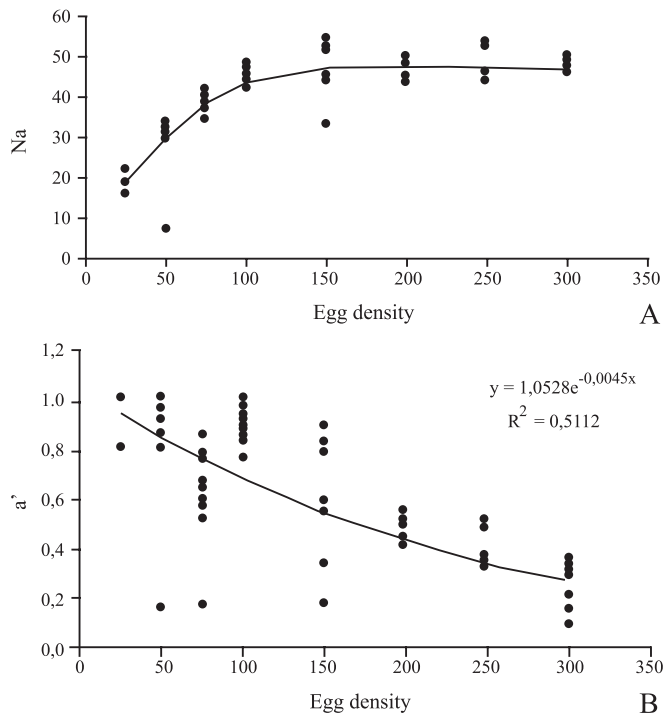


Fig. 1. Functional response of 24-h *T. remus* females on *S. frugiperda* eggs. Na = number of parasitized eggs/day [A]. Proportion of parasitized eggs/available eggs (a') and egg density [B].

III functional responses, respectively, as suggested by Holling (1959). Despite the type, the stabilization of parasitism rate became evident at densities of 150 eggs and higher (Figs. 1 and 2).

DISCUSSION

The average oviposition time from several species of *Telenomus*, including *T. remus* is about 43 s (Cave *et al.* 1987), which is very close to the time found in the present study (40,6–41,8 s). Not many experiments have studied *T. remus* oviposition time, but Schwartz & Gerling (1974) conducted a study to evaluate the oviposition time of 24-hour-old females of that species using *Spodoptera litoralis* (Boisduval) (Lepidoptera, Noctuidae) eggs as host. These authors reported a mean time of 37 s/egg, which is close to the results observed even though a different host was used. Host recognition and handling is the key for successful parasitism, especially in host-specific parasitoids (Cave *et al.* 1987). The manner females handle host eggs and the time they take are directly related to the number of parasitized eggs. They represent the most important factors in functional response (Tillmam 1996).

The increased parasitism with higher egg host densities was also observed by Morales *et al.* (2001) studying *T. remus* 24-hour-old females, although these authors reported lower parasitism and searching rates at lower densities.

The age was not a major factor influencing the searching rates as the egg density was. This was explained by Rogers (1972), who noticed that the higher the host availability, the

lower the searching rate of parasitoids, especially those which massively attack eggs, as *T. remus*.

According to Holling (1961), the host exposition time to the parasitoid, the oviposition time and the searching rate influence the functional response and these are essential parameters in behavior analysis. Therefore, these parameters should be obtained to enhance the information on a natural enemy's behavior and increase its efficacy (Fernandez-Arhex & Corley 2003), although functional response should not be taken into account alone to determine a potential species to be used in biological control programs (Lester & Harmsen 2002; Fernandez-Arhex & Corley 2003).

Insects belonging to the family Scelionidae usually attack hosts that deposit eggs masses and the progeny of these species presents a predominance of females (Gauld & Bolton 1988). Besides, the sex ratio depends on the searching capacity of the females and its energy reserves (Rogers 1972).

Morales *et al.* (2000) demonstrated that 24-hour-old *T. remus* females exposed to *S. frugiperda* eggs produce 1 male for each 5.4 females and this ratio changes with parasitoid age. That is because females with up to 48 h of age produce 1 male for each 7.2 females, with an increase in the number of males starting from the third day (72 h of age). The same has been demonstrated by Schwartz & Gerling (1974) studying biological aspects of *T. remus* on *S. litura*, who verified that 48-hour old *T. remus* females produce more females.

It would be therefore advisable that *T. remus* is kept in laboratory for at least 48 h after hatching before releasing parasitoids, so a higher number of females would be produced in the next generation. However, it must be considered that

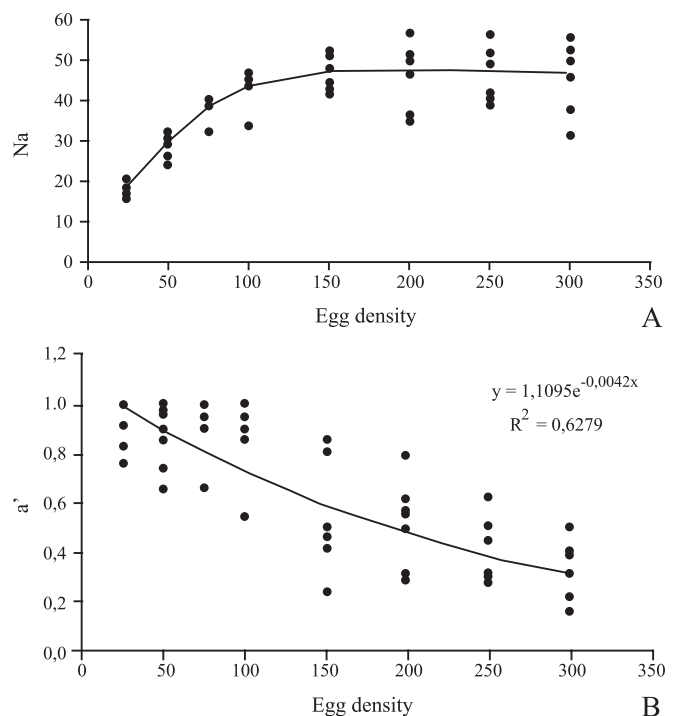


Fig. 2. A. Functional response of 48-h *T. remus* females on *S. frugiperda* eggs. Na = number of parasitized eggs/day [A]. Proportion of parasitized eggs/available eggs (a') and egg density [B].

Table I. Daily mean parasitism and searching rate (\pm SE) of *T. remus* females at different *S. frugiperda* egg densities.

Egg density	Parasitism (eggs/day)		Search rate (parasitized eggs/eggs)	
	24h-old females	48h-old females	24h-old females	48h-old females
25	24.0 \pm 0.67 a	23.2 \pm 1.20 a	0.96 \pm 0.03 a	0.93 \pm 0.05 a
50	43.5 \pm 4.10 a	43.7 \pm 2.50 a	0.87 \pm 0.08 a	0.87 \pm 0.05 a
75	62.6 \pm 2.79 a	68.9 \pm 3.36 a	0.84 \pm 0.04 a	0.92 \pm 0.04 a
100	90.1 \pm 2.45 a	86.7 \pm 6.82 a	0.90 \pm 0.02 a	0.87 \pm 0.07 a
150	97.3 \pm 9.03 a	101.5 \pm 8.43 a	0.68 \pm 0.06 a	0.68 \pm 0.06 a
200	96.6 \pm 2.84 a	103.7 \pm 13.40 a	0.52 \pm 0.10 a	0.52 \pm 0.10 a
250	96.7 \pm 5.95 a	103.0 \pm 14.20 a	0.41 \pm 0.06 a	0.41 \pm 0.06 a
300	95.9 \pm 1.94 a	100.0 \pm 15.50 a	0.33 \pm 0.05 a	0.33 \pm 0.05 a

Means followed by the same letter (row) are not significantly different for the same characteristic according to Tukey test ($P \leq 0.05$).

older adults would be released, and studies should be carried out to evaluate the influence of *T. remus* age on its field efficiency as a biological control agent.

Morales *et al.* (2001) also demonstrated that *T. remus* has a parasitism stabilization tendency at densities of 150 host eggs and higher. However, this stabilization was not so evident and authors have not demonstrated the Type II functional response through Holling's equation or any other more accurate method, but using polynomial regression analysis. The method used by the authors does not consider oviposition times and sex ratio, and uses absolute parasitism numbers. Therefore it does not characterize the functional response of the parasitoid, but solely the daily mean parasitism (Fan & Pettitt 1994; Tillman 1996).

The tendency shown in the present study supports the observations of Holling (1959, 1961), who suggested the use of functional response studies as tools for better understanding predator-prey dynamics and not as parasitism capacity indices. The stabilization at the 150-egg density is confirmed in spite of the changes in *S. frugiperda* egg densities and time for oviposition, because according to Holling (1961) parasitoids are less attracted for oviposition under high host densities and this may reflect in functional response curves.

From the analysis of our results it can be concluded that 24-h and 48-h-old *T. remus* females have effective parasitism at low host densities and are able to continue oviposition when there is high *S. frugiperda* egg density. According to Huffaker & Messenger (1976), these properties are ideal for an efficient natural enemy in biological control programs. Also, parasitoids are more commonly associated to type II functional response than to type III. However, we noticed that the age of an insect can interfere in such aspect. Therefore, this affects the rate of parasitism under low range of host densities; although this might not be of practical importance for biological control (Mills & Laca 2004).

The study of basic aspects of parasitoid-host interaction, as the functional response, searching rates and handling time has major importance, since it can contribute to studies on the impact of *T. remus* on *S. frugiperda* population dynamics. Moreover, in biological control programs that may use

Table II. Sex ratio (\pm SE) of *T. remus* progeny at different *S. frugiperda* egg densities.

Egg density	24h-old females	48h-old females
25	0.56 \pm 0.06Ab	0.71 \pm 0.11Aa
50	0.69 \pm 0.09Ab	0.86 \pm 0.02Aa
75	0.61 \pm 0.08Ab	0.73 \pm 0.09Aa
100	0.61 \pm 0.07Ab	0.78 \pm 0.11Aa
150	0.51 \pm 0.05Ab	0.83 \pm 0.12Aa
200	0.64 \pm 0.07Ab	0.83 \pm 0.03Aa
250	0.62 \pm 0.06Ab	0.76 \pm 0.11Aa
300	0.58 \pm 0.06Ab	0.85 \pm 0.14Aa

Means followed by the same capital letter (columns) or small letter (row) are not significantly different according to Tukey test ($P \leq 0.05$).

this natural enemy in the future, our data will be useful in calculating at which pest density the parasitoid should be released and which is the behavior of *T. remus* under host population changes.

Acknowledgement. The authors thank D. Gerling (Tel Aviv University, Israel) for helpful comments on the manuscript. Assitanship to T.R. Carneiro was provided by Coordenação de Aperfeiçoamento de Pessoal de Nível Superior (CAPES).

REFERENCES

- Beserra, E. B. & J. R. P. Parra. 2005. Impact of the number of *Spodoptera frugiperda* egg layers on parasitism by *Trichogramma atopovirilia*. **Scientia Agricola** 62: 190–193.
- Cave, R. D. 2000. Biology, ecology and use in pest management of *Telenomus remus*. **Biocontrol News and Information** 21: 21–26.
- Cave, R. D. & M. J. Gaylor. 1989. Functional Response of *Telenomus reynoldsi* (Hym: Scelionidae) at five constant temperatures and in an artificial plant arena. **Entomophaga** 34: 3–10.
- Cave, R. D. & N. M. Acosta. 1999. *Telenomus remus* Nixon: un parasitoide en el control biológico del gusano cogollero, *Spodoptera frugiperda* (Smith). **Ceiba** 40: 215–227.
- Cave, R. D.; M. J. Gaylor & J. T. Bradley. 1987. Host handling and recognition by *Telenomus reynoldsi* (Hym.: Scelionidae), an egg parasitoid of *Geocoris* spp. (Heteroptera: Lygaeidae). **Annals of Entomological Society of America** 80: 217–223.
- Cruz, I. & M. L. C. Figueiredo. 1994. Estudos preliminares do parasitóide *Telenomus* sp. Nixon sobre ovos de *Spodoptera frugiperda*. **Relatório Técnico Anual do Centro Nacional de Pesquisa de Milho e Sorgo 1992-1993, Sete Lagoas** 6: 104–105.
- Fan, Y. & F. L. Pettitt. 1994. Parameter Estimation of the Functional Response. **Environmental Entomology** 23: 785–794.
- Fernandez-Arhex, V. & J. C. Corley. 2003. The functional response of parasitoids and its implications for biological control. **Biocontrol Science and Technology** 13: 403–413.
- Ferrer, F. 2001. Biological control of agricultural insect pests in Venezuela; advances, achievements, and future perspectives. **Biocontrol News and Information** 22: 67–74.
- Gauld, I. & B. Bolton. 1988. **The Hymenoptera**. Oxford University Press, 332 p.
- Garcia, M. A. 1991. Ecologia nutricional de parasitóides e predadores terrestres, p. 289–305. In: J. R. P. Parra & A. R. Panizzi (eds.). **Ecologia nutricional de insetos e suas implicações no manejo de pragas**. Brasília, Manole, 359 p.
- González, C. E. & J. L. Zocco. 1996. Control integrado de *Spodoptera frugiperda* (Smith) utilizando *Telenomus remus* (Nixon) em *Zea mays* L. **Revista de Investigación Agrícola-DANAC** 1: 201–219.
- Gupta, M. & A. D. Pawar. 1985. Multiplication of *Telenomus remus* Nixon on *Spodoptera litura* (Fabricius) reared on artificial diet. **Journal of Advanced Zoology** 6: 13–17.

- Hassel M. P. 1978. **The dynamic of arthropod predator-prey systems.** Princeton, Princeton University Press, 237 p.
- Hassel, M. P.; J. H. Lawton & J. R. Beddington. 1997. Sigmoid functional responses by invertebrate predator and parasitoids. **Journal of Animal Ecology** **46**: 249–262.
- Hernández, D.; F. Ferrer & B. Linares. 1989. Introducción de *Telenomus remus* Nixon (Hym.: Scelionidae) para controlar *Spodoptera frugiperda* (Lep.: Noctuidae) en Yaritagua, Venezuela. **Agronomia Tropical** **39**: 199–205.
- Holling C. S. 1959. Some characteristics of simple types of predation and parasitism. **Canadian Entomologist** **91**: 385–398.
- Holling C. S. 1961. Principles of insect predation. **Annual Review of Entomology** **6**: 163–182.
- Huffaker, C. B. & P. S. Messenger. 1976. **Theory and practice of biological control.** New York, Academic Press, 788 p.
- Joshi, B. G.; G. Ramaprasad; S. Sitaramaiah & C. V. V. Sathyanarayana. 1976. Some observations on *Telenomus remus* Nixon, an egg parasitoid of the tobacco caterpillar, *Spodoptera litura* (F.). **Tobacco Research** **2**: 17–20.
- Juliano, S. A. 2001. Nonlinear curve fitting: predation and functional response curves, p.159–182. In: Scheiner, S. M. & J. Gurevitch (eds.). **Design and analysis of ecological experiments**, 2nd ed., New York, Chapman and Hall, p. 373.
- Lester, P. J. & R. Harmsen. 2002. Functional and numerical responses do not always indicate the most effective predator for biological control: an analysis of two predators in a two-prey system. **Journal of Applied Ecology** **39**: 455–468.
- Lewis, W. J.; L. E. M. Vet; J. H. Tumlinson; J. C. Van Lenteren & D. R. Papaj. 1990. Variations in parasitoid foraging: essential element of a sound biological control theory. **Environmental Entomology** **19**: 1183–1193.
- Mills, N. J. & I. Lacan. 2004. Ratio dependence in the functional response of insect parasitoids: evidence from *Trichogramma minutum* foraging for eggs in small host patches. **Ecological Entomology** **29**: 208–216.
- Morales, J.; J. S. Gallardo; C. Vásquez & Y. Ríos. 2000. Patrón de emergencia, longevidad, parasitismo y proporción sexual de *Telenomus remus* (Hymenoptera: Scelionidae) con relación al cogollero del maíz. **Bioagro** **12**: 47–54.
- Morales, J.; J. S. Gallardo; C. Vásquez & Y. Ríos. 2001. Respuesta funcional de *Telenomus remus* (Hymenoptera: Scelionidae) a los huevos de *Spodoptera frugiperda* (Lepidoptera: Noctuidae). **Bioagro** **13**: 49–55.
- Reay-Jones, F. P. F.; J. Rochat; R. Goebel & E. Tabone. 2006. Functional response of *Trichogramma chilonis* to *Galleria mellonella* and *Chilo sacchariphagus* eggs. **Entomologia Experimentalis et Applicata** **118**: 229–236.
- Rogers, D. 1972. Random Search and Insect Populations Models. **Journal of Animal Ecology** **41**: 369–383.
- SAS Institute. 2004. **SAS/STAT user's guide.** Release 9.1 ed. SAS Institute, Cary, NC.
- Schwartz, A. & D. Gerling. 1974. Adult biology of *Telenomus remus* (Hymenoptera: Scelionidae) under laboratory conditions. **Entomophaga** **19**: 482–492.
- Tillman, P. G. 1996. Functional response of *Microplitis croceipes* and *Cardiochiles nigriceps* (Hymenoptera: Braconidae) to variation in density of Tobacco Budworm (Lepidoptera: Noctuidae). **Biological Control** **25**: 254–528.
- Van Alphen, J. J. M. & M. Jervis 1996. Foraging behavior. In: Jervis M. & N. Kidd (eds.). **Insect natural enemies.** New York, Chapman and Hall, 491 p.
- Wang, B. & D. N. Ferro 1998. Functional response of *Trichogramma ostrinia* (Hymenoptera: Trichogrammatidae) to *Ostrinia nubilalis* (Lepidoptera: Pyralidae) under laboratory and field conditions. **Environmental Entomology** **27**: 752–758.
- Wojcik, B.; W. H. Whitcomb & D. H. Habeck. 1976. Host range testing of *Telenomus remus* (Hymenoptera: Scelionidae). **Florida Entomologist** **59**: 195–198.