RVC OPEN ACCESS REPOSITORY – COPYRIGHT NOTICE

This is the peer reviewed version of the following article:

Swann, J. W., Maunder, C. L., Roberts, E., McLauchlan, G. and Adamantos, S. (2016), Prevalence and risk factors for development of hemorrhagic gastro-intestinal disease in veterinary intensive care units in the United Kingdom. Journal of Veterinary Emergency and Critical Care, 26: 419–427. doi: 10.1111/vec.12434

which has been published in final form at <u>http://dx.doi.org/10.1111/vec.12434</u>.

This article may be used for non-commercial purposes in accordance with Wiley Terms and Conditions for Self-Archiving.

The full details of the published version of the article are as follows:

TITLE: Prevalence and risk factors for development of hemorrhagic gastro-intestinal disease in veterinary intensive care units in the United Kingdom

AUTHORS: Swann, J. W., Maunder, C. L., Roberts, E., McLauchlan, G. and Adamantos, S.

JOURNAL TITLE: Journal of Veterinary Emergency and Critical Care

PUBLISHER: Wiley

PUBLICATION DATE: 7 December 2015 (online)

DOI: 10.1111/vec.12434



1 Prevalence and Risk Factors for Development of Hemorrhagic Gastro-Intestinal Disease in Veterinary 2 Intensive Care Units in the United Kingdom 3 Abstract 4 5 **Objective:** To determine the prevalence of hemorrhagic gastro-intestinal (GI) disease developing in dogs 6 and cats admitted for management of non-GI disease in veterinary intensive care units (ICUs). 7 **Design**: Retrospective study of animals presented between October 2012 and July 2013. 8 *Setting*: Three ICUs located in veterinary teaching hospitals in the United Kingdom. 9 Animals: Dogs (n=272) and cats (n=94) were consecutively enrolled from three ICUs if they were hospitalized in the unit for at least twenty-four hours. Cases were excluded if they had hemorrhagic GI 10 11 disease in the forty-eight hour period before presentation or in the twenty-four hour period after admission. Cases were also excluded if they suffered skull fracture, epistaxis or hemoptysis, if they 12 underwent surgical procedures of the GI or upper respiratory tracts, or if they were presented for 13 14 management of GI disease. 15 Measurements and Main Results: Hemorhagic GI disease was observed in dogs at all three units, but at different rates (Center 1: 10.3%, Center 2: 4.8%, Center 3: 2.2%). Hemorrhagic GI disease was not 16 observed in cats at any of the participating centers. Construction of a multivariable logistic regression 17 model revealed that serum albumin concentration, administration of prophylactic gastro-protectant drugs 18 19 and institution were significantly associated with the development of hemorrhagic GI disease in dogs.

Development of hemorrhagic GI disease and placement of a feeding tube were significantly associated with mortality during the period of hospitalization in dogs. Thirty-seven (13.6%) dogs and 12 (12.8%) cats died or were euthanized while hospitalized, with a higher mortality rate (42.1%) in dogs with hemorrhagic GI disease. Conclusions: Hemorrhagic GI disease does develop in dogs hospitalized for management of non-GI

3

disease, but this phenomenon was not observed in cats. Development of hemorrhagic GI disease 25 26 appeared to have a significant impact on survival in veterinary ICUs. 27 Keywords: Stress-related mucosal disease, stress ulcer prophylaxis, omeprazole, enteral feeding 28 29 30 Abbreviations: GI: gastro-intestinal; GMB: gastric mucosal barrier; ICU: intensive care unit; MODS: multiple organ dysfunction syndrome; NSAID: non-steroidal anti-inflammatory drug; ROC: receiver 31 operator characteristic; SIRS: systemic inflammatory response syndrome; SRMD: stress-related mucosal 32 33 disease 34 35 Introduction 36 37 In human medicine, stress-related mucosal disease (SRMD) refers to the development of erosive lesions of the stomach and intestines in patients admitted to ICUs for management of severe illness¹. The 38 term covers a spectrum of disease, from superficial mucosal injury detectable only by 39 gastroduodenoscopy to severe ulceration that results in clinically important hemorrhage. Overt clinical 40

bleeding due to SRMD was reported to occur in approximately 4% of humans admitted to a group of
ICUs in Canada², and development of this disease significantly increased the risk of death during the
period of hospitalization.

44

24

Impaired perfusion of the gastric mucosal barrier (GMB) is the proximate cause of SRMD, but development of the disease is reflective of systemic changes in hemodynamic status and inflammatory cascade³. Splanchnic hypoperfusion caused by sympathetic stimulation or hypovolemia is likely to be an important factor in the development of SRMD but it may be difficult to detect in patients that appear to have adequate macrohemodynamic markers of systemic perfusion⁴. Reduced splanchnic blood flow also increases the risk of reperfusion injury caused by oxygen free radicals if blood flow is restored after appropriate resuscitation⁵. Local or systemic production of pro-inflammatory cytokines such as tumour necrosis factor alpha and interleukins 1 and 8 causes further alterations in perfusion of the gastric mucosa and disrupts the production of mucus and bicarbonate⁶, which are required for neutralization of gastric acid. If the GMB is sufficiently disrupted by these changes, gastric acid may cause direct damage to the mucosa, and this process can progress to cause substantial ulceration and hemorrhage.

56

57 Several factors have been identified in human patients that increase the risk of development of 58 SRMD², particularly respiratory failure necessitating mechanical ventilation and coagulopathy. 59 Administration of prophylactic gastro-protectant medications reduces the risk of SRMD², but this may be 60 associated with development of other complications, such as aspiration pneumonia, because increased 61 gastric pH permits bacterial colonization of the stomach¹³.

62

63 Hemorrhagic GI disease has not been described specifically in veterinary ICUs, but two studies identified subclinical gastric erosions in dogs that underwent decompressive surgery for intervertebral 64 disc disease, some of which also received glucocorticoids^{7,8}. These lesions did not appear to be 65 responsive to administration of gastro-protectant medications. The pathogenesis of gastric ulceration in 66 Alaskan sled dogs at the Iditarod race may also share some features with that of SRMD in people. 67 Strenuous exercise in these dogs resulted in increased gastric permeability and increased frequency of 68 gastric lesions observed by endoscopy^{9,10}. The authors of this study speculated that these changes could 69 70 occur due to increased circulating glucocorticoid concentrations or diversion of cardiac output to skeletal 71 muscle for exercise. Two further studies described development of gastroduodenal ulceration in critically ill animals in association with various underlying causes, including hepatic disease, pancreatitis, 72 hypoadrenocorticism, and administration of non-steroidal anti-inflammatory drugs (NSAIDs)^{11,12}. 73

74

97

75	The primary aim of this study was to determine the proportion of animals that developed overt
76	hemorrhagic GI disease in veterinary ICU patients. It was hypothesized that this would occur at similar
77	rates to those reported in human ICUs, and that dogs would develop the disease more frequently than cats
78	based on previous evidence suggesting that the GI tract is not the shock organ of cats. Secondary aims
79	were to investigate risk factors for the development of hemorrhagic GI disease, and to determine whether
80	development of these signs was associated with mortality during the period of hospitalization.
81	
82	Materials and Method
83	Study design A retrospective multi-center survey was conducted at three intensive care units
84	(Centers 1, 2 and 3) located in teaching hospitals in the UK. These units accept referral cases from
85	veterinarians in general practice and from other specialist services within the same institutions. Case
86	management in each unit is supervised by board certified clinicians in emergency and critical care,
87	internal medicine, surgery or neurology. Entry and egress of patients from the ICUs and use of any drugs,
88	including gastro-protectant medications, were at the discretion of the attending clinician.
89	
90	All cases presenting consecutively to the ICUs were considered eligible for enrolment during the
91	period of the study if they were hospitalized for at least twenty-four hours. Cases were excluded if they
92	had a history of hemorrhagic GI disease in the forty-eight hours prior to hospitalization or if they
93	developed signs within the first twenty-four hours after admission. Cases were also excluded if they
94	underwent surgical procedures involving the GI or upper respiratory tracts, if they presented with or
95	developed epistaxis or hemoptysis, if they presented for management of GI disease, or if they had
96	sustained one or more skull fractures. Cases were not excluded if they received gastro-protectant drugs,

98 diagnosed with diseases that may cause secondary GI signs, such as hypoadrenocorticism.

NSAIDs, glucocorticoids or anticoagulants prior to admission or during hospitalization, nor if they were

100 **Data collection:** A single entry form was produced for each case enrolled in the study 101 (Supplementary Data 1), and this was completed by a veterinarian after the patient was discharged from 102 the ICU. The veterinarian completing the enrolment form did not necessarily have primary responsibility 103 for the case. The following data were collected from the medical records of each case: signalment, 104 presenting problem and problems identified after initial consultation, concurrent diseases and 105 medications, clinical examination findings, GI signs observed while hospitalized, and results of complete blood cell count, serum biochemistry and coagulation profiles performed on admission. Types of feeding 106 tube placed in individual patients were recorded, as were the types and doses of any gastro-protectant, 107 108 NSAID, glucocorticoid or antithrombotic medications administered in the ICU. The length of 109 hospitalization, the nature of any surgical procedures conducted immediately before or during the period 110 of ICU hospitalization, and mortality or euthanasia while hospitalized were also noted.

111

112 SRMD was defined as hemorrhagic GI disease manifesting as hematemesis, melena or 113 hematochezia or as mucosal erosions and hemorrhage observed during GI endoscopy. Dogs were 114 diagnosed with systemic inflammatory response syndrome (SIRS) if they fulfilled two or more of the 115 following four conditions: rectal temperature <37.2C or >39.2C, heart rate >140 beats per minute, 116 respiratory rate >30 breaths per minute, or total white blood cell count <6x10⁹/l (6,000/µl) or >19x10⁹/l 117 (19,000/µl)¹⁵.

118

119 Statistical analysis: All statistical analyses were conducted using a commercial software 120 program^a. Shapiro-Wilks tests and visual assessments of histograms were used to determine whether 121 variables were parametrically distributed. Parametric and non-parametric variables were compared using 122 Student's *t* tests and Mann-Whitney *U* tests, respectively. Categorical data were compared using Fisher's 123 exact tests or Chi squared tests. Confidence intervals, where stated, are at the 95% level.

124

125 The proportion of veterinary patients that developed SRMD was determined by dividing the 126 number of cases with hematemesis, melena or hematochezia by the total number of included cases 127 collectively and for each ICU, and 95% confidence intervals were calculated.

128

129 Multivariable logistic regression was used to evaluate risk factors for development of SRMD and 130 for mortality during hospitalization. Univariable analyses were first conducted using Mann-Whitney U131 tests, Chi squared tests or Fisher's exact tests, and variables with p values <0.2 were retained. These variables were entered together in the multivariable analysis, and a model was fitted using a forward entry 132 method based on calculation of likelihood ratios. A categorical variable describing whether prophylactic 133 134 gastro-protectant medications were administered was forced into the final model for development of SRMD, and a variable describing whether cases fulfilled the SIRS criteria was forced into the model of 135 mortality as these factors were considered to be of considerable *a priori* importance for each model based 136 on published evidence^{2,16}. Institute was included as a factor in both models to account for possible 137 138 differences between centers. Hosmer-Lemeshow tests were performed to assess the adequacy of model 139 fit, and receiver operator characteristic (ROC) curves were produced using probabilities derived from 140 each logistic regression model to determine the predictive capability of each model.

141

142 Results

Study populations: After removal of duplicate cases and application of exclusion criteria, 272 dogs and 94 cats were included in the study (Figure 1). Of the dogs included, 159 (58.5%) were from Center 1, 21 (7.7%) were from Center 2, and 92 (33.8%) were from Center 3. Of the cats, 67 (71.3%) were from Center 1, 7 (7.4%) were from Center 2, and 20 (21.3%) were from Center 3. Seventy-one dogs were intact males, 37 intact females, 82 neutered males and 82 neutered females, whereas 3 cats were intact males, 4 intact females, 51 neutered males and 36 neutered females.

150 There was no difference in median age between dogs and cats, nor between animals of either151 species at different centers.

152

Prevalence of GI disease: The proportion of dogs that developed SRMD was 10.3% (CI: 6.3-153 154 15.7) at Center 1, 4.8% (CI: 0.85-22.7) at Center 2 and 2.2% (CI: 0.6-7.7) at Center 3, with a combined 155 proportion of 7.0% (CI: 4.5-10.7). The difference in proportions between centers was not significant (Chi square 5.2, p=0.075). SRMD did not occur in any of the cats observed during this study at any center. 156 Among the dogs that received prophylactic gastro-protectant medications, the proportion that developed 157 158 SRMD was 16.4% (CI: 8.9-28.3), compared to 4.2% (CI: 2.2-7.8) in dogs that did not receive 159 prophylaxis, and there was a significant difference in these proportions (Chi square 10.3, p=0.001). Rates of development of SRMD during hospitalization in dogs and cats at each center are shown in Table 2. 160

161

Of the dogs that developed SRMD (n=19), the most common diagnoses were immune-mediated disease (4), neoplasia (2), trauma (2), and *Angiostrongylus vasorum* infestation (2). The remaining dogs were diagnosed with pyelonephritis (1), fasciitis (1), traumatic brain injury (1), hepatic disease (1), intervertebral disc protrusion (1), hypoadrenocorticism (1), sudden acute retinal degeneration syndrome (1), sepsis and disseminated intravascular coagulation (1), and intra-abdominal hemorrhage following ovariohysterectomy (1).

168

Feeding tubes were placed in 27 animals across all centers. Only esophagostomy tubes were placed in cats (n=7), but naso-esophageal (n=12), esophagostomy (n=5) and gastrostomy (n=3) tubes were placed in dogs. The majority of the tubes were placed at Center 1 (n=23), with a smaller number at Center 3 (n=4). Placement of a feeding tube was undertaken at Center 1 after a median period of anorexia of 3 days (IQR: 3-5, range 2-9, n=13). The duration of anorexia could not be determined in 6 dogs, and a

naso-esophageal feeding tube was placed in the remaining 4 dogs as a standard preparation formechanical ventilation.

176

177 Evaluation of risk factors for development of SRMD: Univariable analyses revealed that multiple factors were associated with development of SRMD (Table 3). When these variables were 178 entered into the multivariable analysis, decreased serum albumin concentration, institute and 179 180 administration of prophylactic gastro-protectant medications were retained in the final model. Dogs with SRMD were 4.3 (CI: 1.2-15.5) times more likely to have decreased serum concentrations of albumin, 4.3 181 182 (CI: 1.4-13.7) times more likely to have received prophylactic gastro-protectant medications and 10.0 (CI: 183 1.7-33.3) times less likely to have been hospitalized at Center 3 than those that did not develop SRMD. 184 Performance of the Hosmer-Lemeshow test indicated good model fit (Chi Square 2.2, p=0.826), and the area under the ROC curve constructed using model probabilities was 0.79 (CI: 0.68-0.90)(Supplementary 185 Figure 1A), showing that the model was able to discriminate adequately between cases with and without 186 SRMD. A ROC curve was also generated using individual values for serum albumin concentration 187 188 (n=201), and the area under the curve was smaller using this model (0.68, CI: 0.52-0.84) (Supplementary 189 Figure 1B). Using a cut-off value of 28.0 g/l (2.8 g/dl), the sensitivity and specificity values for 190 prediction of development of SRMD were 0.67 and 0.62, respectively.

191

192 *Hospitalization and survival:* The median durations of hospitalization and mortality rates for 193 dogs and cats at different centers are shown in Table 2. There was no difference in duration of 194 hospitalization between cats and dogs from all centers, but average length of hospitalization was greater at 195 both Centers 2 (p=0.048) and 3 (p<0.001) compared to Center 1. Thirty-seven (13.6%) dogs and 12 196 (12.8%) cats died or were euthanized while hospitalized. Mortality rates were similar between centers 197 and species, but the proportion of dogs with SRMD that did not survive to discharge (8/19 dogs, 42.1%) was significantly greater than for dogs that did not develop SRMD (29/252, 11.5%; Chi square 14.0, p < 0.001).

200

201 Evaluation of risk factors for mortality during hospitalization: Significant associations were detected between several variables and mortality during hospitalization using univariable analysis (Table 202 203 4). When these variables were entered into the multivariable analysis, placement of a feeding tube and 204 development of SRMD were associated with mortality. Dogs that died or were euthanized while hospitalized were 13.3 (CI: 4.0-43.5) times more likely to have had a feeding tube placed and 5.1 (CI: 205 1.6-15.9) times more likely to have developed SRMD than those that were discharged. Fulfilment of the 206 207 SIRS criteria and institute were forced into the final model, but these variables were not significantly 208 associated with mortality. The model fit was adequate (Hosmer-Lemeshow Chi square 5.7, p=0.338), and generation of a ROC curve using model probabilities yielded an area under the curve of 0.77 (CI: 0.68-209 0.86)(Supplementary Figure 2). 210

211

Naso-esophageal feeding tubes were placed routinely in dogs that were mechanically ventilated. When these animals were excluded from the analysis (n=4), the odds ratio for tube placement decreased to 8.5 (CI: 2.4 - 30.3, p=0.001), while that for development of SRMD did not change considerably (OR: 5.1, CI: 1.6 - 15.9, p=0.006). Model fit parameters were similar to those reported above (data not shown).

- 217
- 218 Discussion

The results of this study show that hemorrhagic GI disease, defined here as SRMD, does occur in dogs hospitalized in veterinary ICUs, but was not observed in any of the cats that were included. Dogs with SRMD were more likely to have decreased serum concentrations of albumin on presentation, but this parameter had a low sensitivity and specificity for prediction of development of this disease. Affected

animals were also more likely to have received prophylactic gastro-protectant medications and less likely
to have been hospitalized at Center 3. SRMD and placement of a feeding tube were significantly more
likely to occur in dogs that died or were euthanized while hospitalized.

226

The proportion of dogs that developed SRMD varied considerably between Centers, and dogs at Center 3 were at significantly reduced risk compared to either Center 1 or 2. The cause of this difference is not apparent: SRMD could have developed at similar rates at Centers 2 and 3 as at Center 1 but was not recorded, although the authors consider this scenario to be unlikely as occurrence of all forms of GI disease is considered to be a notable event among nursing staff and attending clinicians at all three centers. The true prevalence of SRMD at Center 2 may also differ from that reported due to the small number of cases observed in this study, as indicated by the wide confidence intervals for this parameter.

234

Measurable hemorrhage due to stress-related GI injury is reported to occur in approximately 4-235 6% of affected humans^{2,16}, which is broadly comparable to the proportion of dogs affected with SRMD 236 237 defined according to this study. In contrast to human intensive care, none of the animals included in this 238 study underwent gastroduodenoscopy, which is a much more sensitive technique for detection of superficial erosions in the acid-secreting sections of the stomach. When this technique was applied in 239 dogs undergoing surgery for management of inter-vertebral disc disease, subclinical lesions were 240 observed in approximately 75% of patients^{7,8}. The animals considered in this study are therefore likely to 241 represent the most severely affected patients in a spectrum of stress-related GI disease, similar to the 242 243 syndrome of clinically important bleeding in humans with SRMD.

244

SRMD was not observed in any of the 94 cats that were included in this study, and this finding is consistent with the hypothesis that the GI tract is not a 'stress organ' in cats. Previous experimental¹⁷ and epidemiologic^{18,19} studies indicate that cats are susceptible to pulmonary injury when suffering from

systemic inflammation or sepsis, and there are no previous reports suggestive of SRMD in this species.
Observation of a greater number of cats in ICUs is likely to be required to determine whether SRMD
occurs at lower prevalence than could be detected during this study.

251

Dogs that developed SRMD were more likely to have decreased serum albumin concentrations at 252 253 presentation. Albumin is an essential product used in maintaining the GMB, and dogs with decreased 254 albumin concentrations are reported to be at increased risk of dehiscence following incisional biopsy of the small intestine²⁰. It is therefore possible that dogs with hypoalbuminemia are at increased risk of 255 SRMD and other GI signs due to their inability to maintain an effective mucosal wall. Alternatively, the 256 albumin concentration could be decreased in patients that have clinically undetectable GI injury resulting 257 258 in increased GI permeability and protein-losing enteropathy, prior to the onset or recognition of hemorrhagic GI disease. Investigation of Alaskan sled dogs indicated that increased gastric permeability 259 was an early event in development of erosive gastric disease in this cohort¹⁰, suggesting that 260 hypoalbuminemia could be an effect rather than a cause of the GI signs observed. Hypoalbuminemia in 261 262 this study may also represent a non-specific marker of illness, as serum albumin is a negative acute phase 263 protein. Decreased serum concentrations of albumin have also been identified as negative independent prognostic factors in two studies of dogs admitted to veterinary ICUs^{21,22}. 264

265

With a cut off value of 28.0 g/l (2.8 g/dl, the lower limit of the reference range in use at Center 1), serum albumin concentration had a poor sensitivity and specificity for prediction of the development of SRMD, limiting the usefulness of this parameter in guiding the use of prophylactic interventions. Further studies will be required to establish whether patients with hypoalbuminemia would benefit from administration of gastro-protectant medications.

271

Gastro-protectant medications were administered to a large proportion of the animals included in this study, which complicated the interpretation of the results obtained, particularly because animals that ultimately developed SRMD were more likely to have received one or more of these products. This variable was included when fitting the logistic regression model due to its *a priori* importance², but it would have been preferable to evaluate groups of treated and untreated dogs separately in a stratified model²³. This approach was not attempted in this study as the number of cases in each subgroup would have been insufficient to evaluate the number of risk factors included.

279

Univariable analysis identified several other factors that were significantly associated with 280 281 development of SRMD, including several that have previously been associated with the analogous disease in humans, such as hepatopathy, nephropathy and thrombocytopenia. Cook and colleagues² reported 282 hepatic and renal failure as significant risk factors for development of clinically important bleeding after 283 284 univariable analysis, but only secondary coagulopathy and respiratory failure necessitating mechanical ventilation were retained in the multivariable model. Failure to identify these variables as risk factors 285 286 for SRMD in this study probably relates to the relatively low prevalence of these problems in this sample, 287 and in veterinary ICU caseloads in the UK.

288

SRMD and placement of a feeding tube were significantly more likely to occur in dogs that died 289 290 or were euthanized compared to those that were discharged. Placement of a feeding tube is considered to be a relatively benign procedure²⁴ and, in this group of patients, was usually performed at the same time 291 292 as imaging or other procedures that necessitated general anesthesia or sedation. Placement of a feeding 293 tube in this model is more likely to be a proxy variable that could represent a prolonged history of 294 anorexia or anticipated anorexia, or a patient likely to require a long period of intensive care following placement. Feeding tubes were also placed in four patients in preparation for mechanical ventilation, 295 which is also likely to be a poor prognostic indicator in veterinary ICU patients²⁵. The authors do not 296

consider that use of a feeding tube *per se* should increase the risk of death as this procedure is usually
well tolerated or is associated with only minor complications²⁴. Procedures are also employed at all three
centers to minimize the risk of refeeding syndrome in dogs with prolonged anorexia.

300

301 Evidence from human medicine suggests that enteral nutrition should be beneficial for patients with stress-related GI disease²⁶, and early re-introduction of enteral feeding may reduce the requirement 302 303 for gastro-protectant medications. A recent pilot study of dogs with pancreatitis further indicated that enteral feeding was well tolerated in critical care patients and was not associated with a greater prevalence 304 of adverse effects compared to administration of parenteral nutrition²⁷, and a study of dogs with septic 305 peritonitis suggested that introduction of early enteral nutrition was associated with shorter duration of 306 hospitalization²⁸. Nevertheless, it remains to be determined in future studies whether re-introduction of 307 308 enteral nutrition would prevent the hemorrhagic GI disease reported in patients in this study.

309

Development of hemorrhagic GI disease in patients that did not present for investigation or 310 management of GI disease is likely to cause increased morbidity, either due to development of anemia, 311 312 production and release of further inflammatory mediators, or increased risk of bacterial translocation across the wall of the stomach or upper small intestine. SRMD may itself act as a proxy variable for 313 severe systemic disease, such as SIRS, sepsis, or multiple organ dysfunction syndrome (MODS). A 314 315 variable describing fulfilment of established SIRS criteria was included in the final model of factors associated with mortality during hospitalization to try to account for this possibility as this factor was 316 shown to be significant in a previous study¹⁵. Despite this, development of SRMD remained an 317 independent predictor of mortality, and the higher risk of mortality among patients that developed SRMD 318 319 is consistent with findings in humans with overt clinical hemorrhage due to GI disease².

320

321 *Limitations:* Limitations of this study include the relatively small number of cases included, especially for investigation of risk factors for development of SRMD and mortality. Animals with SRMD 322 323 were more likely to have been hospitalized at Centers 1 and 2 than Center 3, and, although institution was 324 included as an independent factor in all multivariable analyses, it is possible that unmeasured differences between centers could have acted as confounding or modifying factors. 325 326 327 Although much of the data included in this study was collected prospectively, some information regarding development of GI disease was collected retrospectively from clinical records, reducing the 328 reliability and consistency of these findings. Data were also collected by a number of different 329 330 investigators who may not have been involved in the primary care of the case. 331 *Conclusions:* SRMD was observed in dogs from three different veterinary ICUs but was not 332 observed in cats. Decreased serum albumin concentration was associated with development of SRMD, 333 but, using a clinically relevant cut off value, this variable had a poor sensitivity and specificity for 334 335 prediction of the disease. Development of SRMD and placement of a feeding tube were independently 336 associated with increased mortality while hospitalized, but further studies will be required to determine the effects and potential benefits of prophylactic gastro-protectant therapy in veterinary ICU patients. 337 338 Footnotes 339 ^a IBM Corp. Released 2011. IBM SPSS Statistics for Windows, Version 20.0. Armonk, NY: IBM Corp. 340 341 References 342 343 1. Monnig AA, Prittie JE. A review of stress-related mucosal disease. J Vet Emerg Crit Care (San

Antonio). 2011; 21(5): 484-495.

345	2.	Cook DJ, Fuller HD, Guyatt GH, et al. Risk factors for gastrointestinal bleeding in critically ill
346		patients. Canadian Critical Care Trials Group. N Eng J Med. 1994; 330(6): 377-381.
347	3.	Stollman N, Metz DC. Pathophysiology and prophylaxis of stress ulcer in intensive care unit
348		patients. J Crit Care. 2005; 20(1): 35-45.
349	4.	Maynard N, Bihari D, Beale R, et al. Assessment of splanchnic oxygenation by gastric tonometry
350		in patients with acute circulatory failure. JAMA. 1993; 270(10): 1203-1210.
351	5.	Fennerty MB. Pathophysiology of the upper gastrointestinal tract in the critically ill patient:
352		rationale for the therapeutic benefits of acid suppression. Crit Care Med. 2002; 30(6 Suppl):
353		S351-355.
354	6.	Ali T, Harty RF. Stress-induced ulcer bleeding in critically ill patients. Gastroenterol Clin North
355		Am. 2009; 38(2): 245-265.
356	7.	Dowdle SM, Joubert KE, Lambrechts NE, et al. The prevalence of subclinical gastroduodenal
357		ulceration in Dachshunds with intervertebral disc prolapse. J S Afr Vet Assoc. 2003; 74(3): 77-
358		81.
359	8.	Neiger R, Gaschen F, Jaggy A. Gastric mucosal lesions in dogs with acute intervertebral disc
360		disease: characterization and effects of omeprazole or misoprostol. J Vet Intern Med. 2000;
361		14(1): 33-36.
362	9.	Davis MS, Willard MD, Nelson SL, et al. Prevalence of gastric lesions in racing Alaskan sled
363		dogs. J Vet Intern Med. 2003; 17(3): 311-314.
364	10.	Davis M, Willard M, Williamson K, et al. Temporal relationship between gastrointestinal protein
365		loss, gastric ulceration or erosion, and strenuous exercise in racing Alaskan sled dogs. J Vet
366		Intern Med. 2006; 20(4): 835-839.
367	11.	Hinton LE, McLoughlin MA, Johnson SE, et al. Spontaneous gastroduodenal perforation in 16
368		dogs and seven cats (1982-1999). J Am Anim Hosp Assoc. 2002;38(2):176-187.

369	12.	Stanton ME, Bright RM. Gastroduodenal ulceration in dogs. Retrospective study of 43 cases and
370		literature review. J Vet Intern Med. 1989;3(4):238-244.
371	13.	Johnstone J, Nerenberg K, Loeb M. Meta-analysis: proton pump inhibitor use and the risk of
372		community-acquired pneumonia. Aliment Pharmacol Ther. 2010; 31(11): 1165-1177.
373	14.	Silverstein D, Otto C. Sepsis. In: Greene CE, editor. Infectious Disease of the Dog and Cat. 4th
374		ed. Missouri: Elsevier Saunders; 2012, pp. 359-369.
375	15.	Okano S, Yoshida M, Fukushima U, et al. Usefulness of systemic inflammatory response
376		syndrome criteria as an index for prognosis judgement. Vet Rec. 2002; 150(8): 245-246.
377	16.	Duerksen DR. Stress-related mucosal disease in critically ill patients. Best Pract Res Clin
378		Gastroenterol. 2003;17(3):327-344
379	17.	Schutzer KM, Larsson A, Risberg B, Falk A. Lung protein leakage in feline septic shock. The
380		Am Rev Respir Dis. 1993; 147(6 Pt 1): 1380-1385.
381	18.	Brady CA, Otto CM, Van Winkle TJ, King LG. Severe sepsis in cats: 29 cases (1986-1998). J
382		Am Vet Med Assoc. 2000; 217(4): 531-535.
383	19.	Declue AE, Delgado C, Chang CH, Sharp CR. Clinical and immunologic assessment of sepsis
384		and the systemic inflammatory response syndrome in cats. J Am Vet Med Assoc. 2011; 238(7):
385		890-897.
386	20.	Shales CJ, Warren J, Anderson DM, et al. Complications following full-thickness small intestinal
387		biopsy in 66 dogs: a retrospective study. J Small Anim Pract. 2005; 46(7): 317-321.
388	21.	King LG, Wohl JS, Manning AM, et al. Evaluation of the survival prediction index as a model of
389		risk stratification for clinical research in dogs admitted to intensive care units at four locations.
390		Am J Vet Res 2001;62(6):948-954.
391	22.	Hayes G, Mathews K, Doig G, et al. The acute patient physiologic and laboratory evaluation
392		(APPLE) score: a severity of illness stratification system for hospitalized dogs. J Vet Intern Med
393		2010;24(5):1034-1047.

Page 17 of 25

394	23. Hosmer D, Lemeshow S. Applied Logistic Regression. 2 nd ed. Wiley-Blackwell; 2000.
395	24. Devitt CM, Seim HB, 3rd. Clinical evaluation of tube esophagostomy in small animals. J Am
396	Anim Hosp Assoc. 1997; 33(1): 55-60.
397	25. Hopper K, Haskins SC, Kass PH, et al. Indications, management, and outcome of long-term
398	positive-pressure ventilation in dogs and cats: 148 cases (1990-2001). J Am Vet Med Assoc.
399	2007; 230(1): 64-75.
400	26. Marik PE, Vasu T, Hirani A, Pachinburavan M. Stress ulcer prophylaxis in the new millennium: a
401	systematic review and meta-analysis. Critical Care Medicine. 2010; 38(11): 2222-2228.
402	27. Mansfield CS, James FE, Steiner JM, et al. A pilot study to assess tolerability of early enteral
403	nutrition via esophagostomy tube feeding in dogs with severe acute pancreatitis. J Vet Intern
404	Med. 2011; 25(3): 419-425.
405	28. Liu DT, Brown DC, Silverstein DC. Early nutritional support is associated with decreased length
406	of hospitalization in dogs with septic peritonitis: a retrospective study of 45 cases (2000-2009). J
407	Vet Emerg Crit Care (San Antonio). 2012; 22(4): 453-9.
408	
409	Figure 1: Flow diagram of cases included in study
410	
411	Supplementary Figure 1: Receiver operator characteristic curves generated using (A) probabilities from
412	the multivariable regression model of risk factors for development of SRMD and (B) serum
413	concentrations of albumin
414	
415	Supplementary Figure 2: Receiver operator characteristic curve generated using probabilities derived
416	from multivariable logistic regression model of risk factors for mortality during hospitalization



		Center 1		Center 2		Center 3		Combined	
		Canine	Feline	Canine	Feline	Canine	Feline	Canine	Feline
N		159	67	21	7	92	20	272	94
Time period		October 2012		September to		January 2013 to			
		to July 2013		November		April 2013			
				2012					
Age (years)	Median	5.9	6.9	4.8	5.0	5.0	8.5	5.1	7.2
	Inter-	2.6 -	2.3 -	2.8 -	0.8 -	3.0 -	4.0 -	2.8-9.0	2.9 -
	quartile	9.4	12.3	6.0	10.0	8.0	12.0		12.0
	range								

Table 1: Summary of demographic data obtained from included patients

*Inter-quartile range

		Canine				Feline			
		Cente	Cente	Cente	Combine	Cente	Cente	Cente	Combine
		r 1	r 2	r 3	d	r 1	r 2	r 3	d
N		159	21	92	272	67	7	20	94
SRMD* (%)		16	1	2	19 (7.0)	0	0	0	0
		(10.3	(4.8)	(2.2)					
)							
	Melena or	15	1	2	18 (6.6)				
	hematochezia	(9.4)	(4.8)	(2.2)					
	(%)								
	Hematemesis	1	0	0	1 (0.4)				
	(%)	(0.6)							
	Hemorrhage	0	0	0	0				
	observed on								
	endoscopy								
	(%)								
	Died/euthaniz	7	0	1	8 (42.1)				
	ed while	(43.8		(50.0					
	hospitalized))					
	(% of SRMD								
	cases)								
Duration of	Median	3.0	4.0	2.0	3.0	2.0	4.0	6.0	3.0
hospitalization									
(days)									

Table 2: Summary of clinical and gastro-intestinal disease data obtained from cases

	Inter-quartile	2.0 -	2.0 -	4.0 -	2.0 - 5.0	2.0 -	1.0 -	2.25	2.0 -
	range	4.0	7.5	6.0		4.0	8.0	_	5.25
								8.75	
Died/euthanzi		27	1	9	37 (13.6)	4	1	7	12 (12.8)
ed while		(17.0	(4.8)	(9.8)		(6.0)	(14.3	(35.0	
hospitalized)))	
(%)									
Received GI§		28	9	19	56 (20.6)	1	3	1	5 (5.3)
prophylaxis		(17.6	(42.9	(20.7		(1.5)	(42.9	(5.0)	
(%)))))		
	Subsequently	6	1	2	9 (16.1)	0	0	0	0
	developed	(21.4	(11.1	(10.5					
	SRMD* (% of)))					
	those				3				
	receiving								
	prophylaxis)								

*SRMD: Stress-related mucosal disease, §GI: gastro-intestinal



	Univariable	factors	Multivariable model**				
		Developed	Did not	<i>p</i> value	Odds	95%	р
		SRMD [¶]	develop		ratio	Confidence	value
		(%)	SRMD [¶]			interval	
			(%)				
Median age (years)(i	nterquartile	80(50-	50(26	0.007			
range)		11.3)	- 8 5)	(Mann-			
Talige)		11.3)	- 8.5)				
				Whitney U			
				test)			
Institute	Center 1	16 (5.9)	143	0.059	1.0		
			(52.8)				
	Center 2	1 (0.4)	20		0.3	0.03 - 2.8	0.304
			(7.4)				
	Center 3	2 (0.7)	89		0.1	0.03 - 0.6	0.012
			(32.8)				
Packed cell volume	< 35%	12 (4.4)	72	0.003			
			(26.6)				
	≥ 35%	7 (2.6)	180				
			(66.4)				
Platelet count	< lower	6 (3.0)	30	0.045			
	RL§		(15.0)				
	\geq lower RL	10 (5.0)	154				
			(77.0)				
Serum albumin	< lower RL	11 (5.5)	94	0.110	4.3	1.2 – 15.5	0.026
concentration			(46.8)				

Table 3: Results of univariable and multivariable analysis of risk factors for development of SRMD

	\geq lower RL	4 (2.0)	92				
			(45.8)				
Serum ALT*	\leq 4 x upper	11 (5.7)	168	0.019			
activity	RL		(86.6)				
	> 4 x upper	4 (2.1)	11 (5.7)				
	RL						
Serum creatinine	\leq 2 x upper	15 (6.8)	194	0.176			
concentration	RL		(88.6)				
	> 2 x upper	2 (0.9)	8 (3.7)				
	RL	1					
SIRS ^Π	No	5 (2.7)	85	0.086			
			(45.2)				
	Yes	13 (6.9)	85				
			(45.2)				
Prophylactic	No	10 (3.7)	206	0.002	4.3	1.4 – 13.7	0.013
administration of			(76.0)				
gastro-protectant							
drugs							
	Yes	9 (3.3)	46				
			(17.0)		7		

*ALT: alanine aminotransferase, [§]RL: reference limit, ^{II}SIRS: systemic inflammatory response

syndrome, [¶]SRMD: stress-related mucosal disease. **n=201.

		Univar	iable factors	5	Multivariable model [¶]			
		Died	Survived	<i>p</i> value	Odds	95%	р	
		(%)	(%)		ratio	confidence	value	
						interval		
Median age (years))(interquartile	7.5	5.0 (2.6	0.037				
range)		(4.0 –	- 8.4)	(Mann-				
		10.5)		Whitney				
				U test)				
Institute	Center 1	27	132	0.130	1.0			
		(9.9)	(48.5)					
	Center 2	1	20 (7.4)		1.3	0.2-11.3	0.808	
		(0.4)						
	Center 3	9	83		0.9	0.3-2.8	0.836	
		(3.3)	(30.5)					
Packed cell	< 35%	18	66	0.012				
volume		(6.6)	(24.3)					
	≥ 35%	19	169					
		(7.0)	(62.1)		4	•		
Platelet count	< lower RL*	8	28	0.187				
		(4.0)	(13.9)					
	\geq lower RL	21	144					
		(10.4)	(71.6)					
Serum creatinine	\leq 2 x upper RL	26	184	0.006				
concentration		(11.8)	(83.6)					
	> 2 x upper RL	5	5 (2.3)					

Table 4: Results of univariable and multivariable analysis of risk factors for death while hospitalized

		(2.3)					
SIRS§	No	8	82	0.016	2.0	0.8-5.2	0.164
		(4.2)	(43.4)				
	Yes	22	77				
		(11.6)	(40.7)				
SRMD ^{II}	No	29	223	0.001	5.1	1.6-15.9	0.006
		(10.7)	(82.3)				
	Yes	8	11 (4.1)				
		(3.0)					
Placement of	None	26	226	< 0.001	13.3	4.0-43.5	< 0.001
feeding tube		(9.6)	(83.1)				
	Naso-	7	5 (1.8)				
	esophageal	(2.6)					
	Esophagostomy	2	3 (1.1)				
		(0.7)		5			
	Gastrostomy	2	1 (0.4)				
		(0.7)					
Mechanically	No	34	233	0.019			
ventilated		(12.5)	(85.7)				
	Yes	3	2 (0.7)			•	
		(1.1)					

*RL: reference limit, [§]SIRS: systemic inflammatory response syndrome, ^ΠSRMD: stress-related

mucosal disease. [¶]n=188.