Subsurface imaging of silicon nanowires circuits and iron oxide nanoparticles with sub-10 nm spatial resolution

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Abstract: Non-destructive subsurface characterization of nanoscale structures and devices is of significant interest in nanolithography and nanomanufacturing. In those areas, the accurate location of the buried structures and their nanomechanical properties are relevant to optimize the nanofabrication process and the functionality of the system. Here we demonstrate the capabilities of bimodal and trimodal force microscopy to image silicon nanowire devices buried under an ultrathin polymer film. We resolve the morphology and periodicities of silicon nanowire pairs. We report a spatial resolution in the sub-10 nm range for nanostructures buried under a 70 nm thick polymer film. By using numerical simulations we explain the roles of the excited modes in the subsurface imaging process. Independently of the bimodal or trimodal AFM approach, the fundamental mode is the most suitable to track the topography while the higher modes modulate the interaction of the tip with the buried nanostructures and provide the subsurface contrast.

1. Introduction

Non-destructive characterization of buried or internal nanometer-scale structures has a wide range of implications in different fields from cell biology[1,2], toxicity[3] polymer sciences[4,5] or semiconductor device fabrication[6]. A variety of methods ranging from confocal, transmission electron[3] or near-field microscopies have been proposed[7] to image subsurface structures. Ultrasound waves and force microscopy configurations[8] have been proposed to investigate embedded structures [9-17]. Electrostatic and Kelvin probe force microscopy have been applied to reveal the structure of buried polymer nanocomposites [18,19]. Anomalies in the flow-shift paradigm of spin-coated films at the submicrometer scale have been exploited for precise overlay fabrication in thermal scanning probe lithography[20]. However, the spatial resolution of the above methods is in the submicrometer range. The long-range character of the electrostatic interactions in bias-based probe methods or the wavelength of ultrasound waves in acoustic-based force microscopy limits the spatial resolution to the 50-100 nm range.

Recently, Solares and co-workers have reported a multifrequency AFM subsurface imaging method[21]. They developed a trimodal AFM approach[21-23] to image glass nanoparticles buried under a polymer film. This approach involves the simultaneous excitation and detection of three eigenmodes to measure the topography, modulate the sample indentation and map compositional contrast. This multifrequency AFM approach has opened a novel and potentially high spatial resolution method for subsurface characterization. However, many key aspects of this approach have not been addressed. Are three modes needed to achieve subsurface imaging? Could the same effect be achieved with monomodal or bimodal excitation and detection? What properties should have a flexural mode to enable subsurface contrast? Only a single system, glass nanoparticles have been imaged by multifrequency AFM. Additional examples are needed to establish the general character of the method.

Here, we demonstrate the capability of both bimodal and trimodal AFM to image silicon nanowire (SiNWs) arrays and iron oxide nanoparticles (NPs) buried under a spin-coated film. The images show a lateral resolution in the sub-10 nm range. Furthermore, we provide some examples that illustrate that the subsurface imaging process preserves the spatial resolution of tapping mode AFM operation. We have developed a theoretical framework to understand the contrast mechanisms and explain the different roles of the excited modes in the subsurface imaging process. We show that both bimodal and trimodal configurations are suitable for subsurface imaging. The first mode provides the topography mapping. The second mode enhances the contrast of the buried nanostructures and facilitates the interaction with the nanostructures. If a third mode is used, its role is to favor the tip interaction with the buried nanostructures.

2. Experimental methods

2.1. Silicon Substrates

Silicon substrates have been cleaned with 2-propanol, acetone and distilled water (Sigma-Aldrich) by ultrasonic treatment for 5 minutes each. The substrates were then immersed in a H_2O_2 -NH₄OH-H₂O (1 : 1 : 2) mixture and four ultrasound cycles of 10 minutes have been performed.

2.2. Fabrication of silicon nanowires by oxidation SPL

Silicon on insulator substrates with a Si active layer 12 nm thick, p-doped and a nominal resistivity of 9-15 Ω cm (MEMC/SunEdison, US) and a buried oxide layer (BOX) 25 nm thick were used for the fabrication of the devices. The substrates were firstly cleaned with a NH₄OH-H₂O₂–H₂O (1: 1: 4) mixture in three ultrasound cycles of 10 minutes and a last ultrasound cycle of 5 minutes in deionized water . Then Ti/Au marker electrodes were defined by photolithography to localize the structures. An interdigitated array of silicon nanowires were fabricated after definition of SiO₂ masks by oxidation scanning probe lithography and pattern transfer by dry etching processing. The o-SPL was performed by operating the AFM (Dimension V, Bruker, USA) in the amplitude modulated mode with a free amplitude of 5 nm and a set point amplitude/free amplitude ratio of about 0.9. n+-doped silicon cantilevers (NCH-W, NanoWorld) with a force constant of about 40 N/m and a resonant frequency about 300 kHz were used for the fabrication of the patterns. The relative humidity was kept at about 45% in a sealed chamber. Voltage pulses of 22.5-24 V and 1 ms were used. The silicon oxide mask thickness and width are, respectively, in the range of 1.5-2.2 nm and 40-70 nm. After reactive ion etching (NRE-3000, INNOVA Scientific) process, using a gas mixture proportion, chamber pressure, radiofrequency power and etching time of, respectively, $SF_6:O_2(10:5)$ sccm, 59 mTorr, 15 W and 43 s, silicon nanowires which preserve the original thickness of the top Si layer were produced.

2.3. Nanoparticles deposition

In order to deposit the Fe_2O_3 nanoparticles, the silicon surface was functionalized after the cleaning procedure. The substrates were immersed in a solution containing 11 µl 3-aminopropyl-triethoxysilane (APTES) and 50 mL ethanol for 45 minutes. Finally, the substrates were rinsed with ethanol and water, and dried under N₂. Afterwards, the silicon surface was covered with the dimercaptosuccinic acid (DMSA) coated Fe_2O_3 nanoparticles by the drop casting method. A 20 µl drop taken from a 1.4 mg/ml nanoparticles aqueous solution was deposited on the silicon surface for 60 seconds.

2.4. Polydimethylsiloxane (PDMS) substrates

The chips containing the silicon nanowires and the nanoparticles were spin coated by a mixture of PDMS (Sylgard 184, Sigma Aldrich) curing agent: PDMS elastomer base: hexane (Scharlau, Scharlab, S.L.) with a proportion of 1:10:1000 (by weight) at 6000 rpm for 180 s and then cured

on a hot plate at 150 °C for 10 minutes. Under these conditions, the PDMS films have thicknesses between 30 nm- 60 nm.

2.5. AFM Measurements

The experiments have been performed with a Cypher S microscope (Asylum Research, Santa Barbara, USA). We have used PPP-FMAuD cantilevers (Nanosensors) with typical values of $k_1 \approx 2.3 \text{ N/m}$, $f_{01} \approx 66 \text{ kHz}$ and $Q_1 \approx 190 Q_1 \approx 190$. The cantilever has been driven at the three first eigenmodes simultaneously in the trimodal AM scheme[24]. The amplitudes of the modes have been adjusted to achieve compositional contrast, stable imaging and the indentation needed for the subsurface imaging process.

2.6. Multifrequency AFM theory and simulations

The solution of the modified Euler-Bernouilli beam equation for a uniform, continuous and rectangular microcantilever is approximated by a system of three point-mass equations coupled by the force term [25],

$$m \frac{d^2 z_i(t)}{dt_i^2} + \frac{m\omega_{0i}}{Q_i} \frac{dz_i(t)}{dt_i} + k_i z_i(t) = \sum_{j=1}^3 F_{0j} \cos \omega_j t + F_{ts}(z_c + z_1(t) + z_2(t) + z_3(t)) \quad i = 1,2,3$$
(1)

where *m* is the reduced mass of the cantilever and z_i , Q_i , f_i , $\omega_i = 2\pi f_i$, $\omega_{0i} = 2\pi f_{0i}$, k_i , A_{0i} and F_{0i} are respectively the deflection, quality factor, driving frequency, angular driving frequency, resonant frequency, angular resonant frequency, stiffness, free amplitude and driving force amplitude of the *i*-th flexural mode.

 F_{ts} is the tip – sample force and z_c is the cantilever – sample distance. The flexural modes parameters are $f_1 = f_{01} = 66.7$ kHz, $f_2 = f_{02} = 421$ kHz, $f_3 = f_{03} = 1.18$ MHz, $k_1 = 2.3$ N/m, $k_2 = 88.15$ N/m, $k_3 = 800$ N/m, $Q_1 = 173$, $Q_2 = 400$, $Q_3 = 771$. The equations of motion were integrated numerically using a fourth order Runge–Kutta algorithm. The solutions of (1) are given approximately by

$$z_i(z_c,t) = A_i(z_c)\cos(\omega_i t - \phi_i(z_c))$$
⁽²⁾

where A_i , ϕ_i are the amplitude and phase shift of the i-th flexural mode. The tip–sample interactions are described by the Van der Waals interaction for the attractive regime and by the Derjaguin-Muller-Toporov and Kelvin – Voigt model as is described in [26]

$$F_{ts}(d) = \begin{cases} -HR_t / 6d^2 & d \ge a_0 \\ 4/3E_{ff} \sqrt{R_t \delta^3} - HR_t / 6a_0^2 - \eta \sqrt{R_t \delta} (d\delta / dt) & d \le a_0 \end{cases}$$
(3)

where $d \equiv d(t) = z_c + z_1(t) + z_2(t) + z_3(t)$ is the instantaneous tip – sample distance, $a_0 = 0.165$ nm is the intermolecular distance, $\delta(t)$ is the indentation, $\eta = 5$ Pa is the dynamic viscous coefficient R_t is the tip radius,

$$E_{ff} = \left(\frac{1 - v_t^2}{E_t} + \frac{1 - v_s^2}{E_s}\right)^{-1}$$
(4)

is the effective Young modulus and v_x , E_x are the sample (s) and tip (t) Poisson's ratios and Young modulus respectively. We use $v_s = v_t = 0.3$, $E_s = 10$ MPa, $E_t = 170$ GPa. We consider a spherical tip of negligible mass ($R_t = 10$ nm) only for interaction purposes.

3. Results and discussion

3.1. Multifrequency AFM: Bimodal or trimodal excitation/detection schemes

The multifrequency AFM approach for subsurface imaging involves the simultaneous excitation of several cantilever modes, either two (bimodal) or three (trimodal). Bimodal AFM is the most robust multifrequency-based probe approach for topography and quantitative mechanical characterization of surfaces at the nanoscale. The fundamentals of bimodal AFM and its extensions are described in several reviews[24][27,28] and research contributions[29-37].

Subsurface imaging in a trimodal configuration is more complex than in bimodal AFM, for this reason, trimodal AFM operation for subsurface imaging is depicted in Figure 1(a). The first three flexural modes of the microcantilever are excited by a signal that contains three sinusoidal components. The components are tuned, respectively, to the first, second and third cantilever flexural modes. In the optimum operating conditions, the amplitudes of the components are asymmetric, the largest amplitude value corresponds to the 1st mode A_1 , next is A_3 and then A_2 . This scheme enhances the indentation of the polymer layer ($A_3 > A_2$) and improves compositional contrast. In the case of bimodal AFM A_1 is larger than A_2 .

Figure 1b-c illustrate the type of nanostructures investigated to demonstrate the subsurface imaging capabilities. The topography (Fig. 1(d)) and thickness (Fig. 1(e)) of the spin-coated film are shown. For example, the cross-section across the edge of a spin-coated silicon region reveals a thickness of 70 nm (Fig. 1(e)). The thickness values that are directly extracted from the AFM data are smaller (~45 nm). The force applied during the imaging process introduces some deformation on the polymer[38,39]. The true profile (continuous line) is reconstructed by using a numerical simulation code [26]. The topography peak observed at the edge of the coated/uncoated region is due to the scratching process applied to generate coated and uncoated regions on the Si substrate.

3.2. Silicon nanowire and iron oxide nanoparticles buried under a polymer film

Two different nanoscale systems have been chosen to test multifrequency AFM for subsurface imaging, silicon nanowires and nanoparticles. Figure 2(a) shows an optical image of the gold contacts of a silicon nanowire circuit. The square indicates the region that contains the SiNW array. The amplitude modulated (tapping mode) AFM [40] image (Fig. 2(b)) shows a set of interdigitated SiNWs fabricated by oxidation scanning probe lithography (o-SPL)[41,42]. The SiNWs are 2 μ m in length, 40 nm in width (average value at half maximum). The separation between nanowires ranges between 49 and 98 nm. To demonstrate the capability to sub-10 nm particles, we have deposited iron oxide nanoparticles [43,44] on a Si(100) surface (Fig. 2(c)). The nanoparticles have a diameter distribution between 7 and 10 nm with an average diameter centered at 8.2 nm (Fig. 2(d)).

To illustrate that both the amplitude and the phase shift observables in multifrequency AFM are sensitive to subsurface features we plot the topography and phase shift contrast $(\Delta \phi_1)$ images of an array of SiNWs and a random distribution of NPs after they have been buried under an approximately 70 nm layer of polydimethylsiloxane (PDMS). Both observables provide good contrast of the spin-coated SiNWs (Fig. 3(a), (b)) and iron oxide NPs (Fig. 3(c), (d)).

The capabilities of multifrequency AFM to reveal the morphology of nanostructures buried under polymer films are demonstrated by comparing the images obtained, respectively, by tapping mode and trimodal AFM of the same region of the sample. Figure 4a and 4b show, respectively, the trimodal and the tapping mode AFM images of the same SiNW circuit. The SiNWs are only resolved in the trimodal AFM image. Identical results are obtained by performing the comparison with the buried NPs (Fig. 4(c) and 4(d)). The above results are independent of the imaging acquisition sequence (first tapping mode then trimodal or first trimodal then tapping mode).

The cross-sections along the marked lines shown in Fig. 2(b) and Fig. 4 illustrate the genuine character of the subsurface contrast of multifrequency AFM (Fig. 5(a)-(b)). The tapping mode AFM cross-section of an array of SiNWs coated by PDMS does not provide any hint on the presence of the nanostructures. However, the buried nanowires are resolved if the AFM configuration is switched to trimodal AFM. Furthermore, the trimodal AFM profile matches the one obtained by tapping mode AFM before the deposition of the PDMS film. We observe that the subsurface cross-section is slightly sharper than the cross-section of the same nanowires before PDMS deposition. This could be attributed to a change of the tip geometry. It also points out that the subsurface imaging process does not necessarily imply less spatial resolution. Similar result is obtained with the NPs (Fig. 5(b)).

In the above comparison, we have used the same cantilever for tapping and trimodal AFM modes. The free amplitude A_0 was higher in tapping mode AFM ($A_0 = 160$ nm) than in trimodal

AFM (total amplitude ~142 nm). This demonstrates that the operational amplitudes for imaging are not the dominant parameters in the subsurface imaging contrast.

3.3. Simulations of trimodal AFM imaging

We have performed numerical simulations of the cantilever dynamics under the simultaneous excitation of the first three flexural modes. The simulations are intended to clarify the contrast mechanism and to explain the role of the different eigenmodes in the subsurface imaging process. It has been shown that the vertical resolution Δh (the minimum step height variation that can be measured) is inversely proportional to the slope of the amplitude curve (dA/dz_c) [45],

$$\Delta h = \frac{\Delta A}{\left| dA / dz_c \right|} \tag{5}$$

where ΔA is the error in the amplitude A and z_c is the probe height. The maximum possible value of the above slope is 1. Equation 5 shows that the vertical resolution increases by decreasing the slope. At the same time, slopes below 1 will diminish the ability of the AFM to track the surface topography.

Figure 6(a) shows the dependence of the amplitude with respect to the tip-sample separation (probe height) for different monomodal amplitude modulation AFM (AM-AFM) configurations. The slope in the central region decreases with the mode order from 1 (1st mode) to 0.4 (2nd mode) and 0.02 (3rd mode). Definitely a feedback set on A_3 will have an extremely poor microscopy performance. The second mode is still suitable for imaging flat surfaces but unsuitable to track surfaces with features of tens or hundreds of nanometers in height. This result is related to the sensitivity of the cantilever modes which for the phase shift scales with [27] [45],

$$\Delta \phi_i = K \frac{Q_i}{k_i} \tag{6}$$

where K is a factor that depends on the type of interaction force acting on the cantilever and Q_i and k_i are, respectively, the quality factor and force constant of the *i*-th flexural mode of the microcantilever.

In multifrequency AFM the slope dA/dz_c shows a sharp dependence on the mode order. Figure 6b shows the slope corresponding to A_1 for different monomodal, bimodal and trimodal configurations. In all the configurations the slope in the central region is approximately 1. Consequently, A_1 is a suitable observable to track the topography. A different result is obtained in multifrequency AFM when the slopes corresponding to A_2 and A_3 are plotted (Fig. 6(c)). The dA_i/dz_c values (i = 2,3) are close to zero.

We have compared the experimental and theoretical dependence of the amplitude curves on the excited mode for monomodal excitations. Figure 7(a) shows some experimental amplitude curves obtained on a PDMS sample. The curves show that the slope decreases with the excited mode. The first mode has a slope close to 1 while for the second and third modes the slopes are, respectively, 0.5 and 0.05. The simulations shown in Fig. 7(b) confirm the experimental observation by reproducing the trend described above. In addition, the slope values reported for the different modes are close to the experimental values.

A critical aspect in the subsurface imaging process is the tip's ability to interact with the nanostructures buried under the polymer film. We have studied the tip penetration as a function of the AFM configurations. Figure 8(a) shows that in monomodal excitation and detection the tip penetration increases with the mode order. For $z_c = 60$ nm, the penetration is 23.5 nm for the 1st mode, 70 nm for the 2nd mode and 78 nm for the 3rd mode. This indicates that for the same operational conditions, the penetration of the first mode is significantly smaller than that of the 2nd and 3rd modes. This could be explained in terms of the maximum potential energy of the microcantilever. The energy is proportional to the k_i which grows with the resonant frequency of the excited mode (f_{0i}). For a rectangular cantilever [27],

$$k_{i} = k_{1} \left(\frac{f_{0i}}{f_{01}}\right)^{2}$$
(7)

Consequently, for the same set-point amplitude the maximum potential energy stored in the microcantilever grows with the mode order.

A different result is obtained in multifrequency AFM. At $z_c = 60$ nm, the penetration is about 32 nm for the three configurations studied here (Fig. 8(b)). That value is in between the value calculated for the tip penetration of the 1st mode and that of the 2nd and 3rd modes.

The phase shift of the 2nd mode is very sensitive to the compositional properties of the material [25,46]. Figure 9 compares the phase shift contrast $\Delta\phi$ obtained between a region of 10 MPa and another region of 1 GPa for different multifrequency AFM configurations. In bimodal AFM, the configuration with the smallest contrast is obtained in $\Delta\phi_3$ by exciting the 1st and the 3rd modes (Fig. 9(a)). For trimodal AFM, the contrast is maximized by using the phase shift of the 2nd mode. Nonetheless, it should be emphasized that in both cases the phase shift contrast is well above the noise level.

Imaging soft matter with the AFM always involves some sort of sample deformation. This property has enabled the use of force microscopy to study the nanomechanical properties of tissues[2] and cells [47,48]. Here, the deformation is exploited to detect the nanostructrures buried under a thin polymer film. The existence of deformations is independent of the single frequency or the multifrequency character of the probe excitation and detection schemes,

however, bimodal or trimodal AFM optimizes the indentation and the subsurface imaging contrast versus standard (single frequency) AFM methods.

The tip's potential energy depends on the force constant and the amplitude. For a rectangular cantilever the force contact ratio grows with the square of the resonant frequency that would favour the use of higher modes for penetration. However, the sensitivity of higher modes decreases with the mode order [49] (see Fig. 6) so the ability to properly track the topography is reduced. On the other hand, the slope of the first mode hardly depends on the AFM configuration (single mode, bimodal or trimodal). This implies that the ability A_1 to track the sample topography remains unaffected in multifrequency AFM. Then the introduction of an additional excitation will open additional channels and increase the tip indentation (Fig. 8).

The above considerations have been supported by the numerical simulations. The simulations confirm that the feedback to track the sample topography must be connected to the A_1 . The slope A_1 with the probe height is the highest (Fig. 6(a)), which guarantees the optimum conditions for topography imaging. Second, the additional excitation of one or two higher modes enhances the tip penetration. This property enables the tip to interact with the buried nanostructures. It has been established that the material properties contrast depends on the observable. An enhanced phase contrast is found by plotting the phase shift of the 3rd mode. The above factors explain the capabilities of trimodal AFM to perform subsurface imaging. The data also illustrates that the spatial resolution in trimodal AFM is similar to the one obtained with the SiNWs before being coated by the PDMS layer.

Figure 8 illustrates that trimodal excitation is not strictly needed for achieving subsurface imaging. The slope of the A_1 curve in bimodal AFM (130,12, 0) and (130,0,12) is almost identical to the slope of the A_1 curve in a trimodal configuration (130, 2, 10). The compositional contrast is smaller ($\Delta \phi_{\text{bimodal}} \approx 15^\circ$ versus $\Delta \phi_{\text{trimodal}} \approx 40^\circ$). However, in both configurations $\Delta \phi$ is well above the noise level (~0.5°). In practice, this means that both bimodal or trimodal AFM are suitable to perform subsurface imaging.

The similar dependence of the tip's indentation on the probe height (Fig. 8(b)) emphasizes a property of multifrequency AFM operation. It reflects that in multifrequency AFM, the amplitude reduction of the higher modes is not controlled by the Q_i/k_i ratio (Eq. 6) provided that the amplitudes (A_2 , A_3) are smaller than A_1 . The energy transfer from the first mode to the higher excited modes is a factor which compensates the amplitude reduction due to the tip-surface forces that happens in monomodal excitation and detection (Fig. 6(a)).

4. Conclusions

We have studied and demonstrated the capability of bimodal and trimodal AFM to image nanostructures buried under thin polymer films. Silicon nanowires of 12 nm in height and about 40 nm in diameter buried under a 70 nm soft polymer film (1-10 MPa) have been imaged. Iron

oxide nanoparticles with a mean diameter of 8.2 nm have also been detected. By combining theory and numerical simulations we have explained the role of the different excited modes in the subsurface imaging process. The amplitude of first mode is the optimum observable to track the sample topography of the external interface while the higher excited modes contribute to enhance the subsurface contrast or facilitate the sample deformation. In bimodal AFM, the signal of the second mode has a dual role, it provides the tip-subsurface structure interaction and the subsurface contrast. Trimodal AFM assigns specific roles to the three excited modes. The first mode carries the topography contrast, the second mode optimizes the compositional contrast and the third mode modulates the tip penetration. Trimodal AFM operation offers a higher subsurface contrast than bimodal AFM.

The success of multifrequency AFM to image buried nanostructures depends on the softness of the interface. This method will not be suitable to inspect embedded nanostructures in a rigid matrix, say with a Young modulus above 500 MPa. However, there is a variety of applications in nanofabrication and nanolithography where soft resists layers are used. The inspection of those samples before further processing represents an application niche for subsurface imaging by multifrequency AFM. In cell biology and nanotoxicity studies, the cell's uptake of nanoparticles and nanotubes, and its location respect to cell organelles could also be examined by this method.

Acknowledgments

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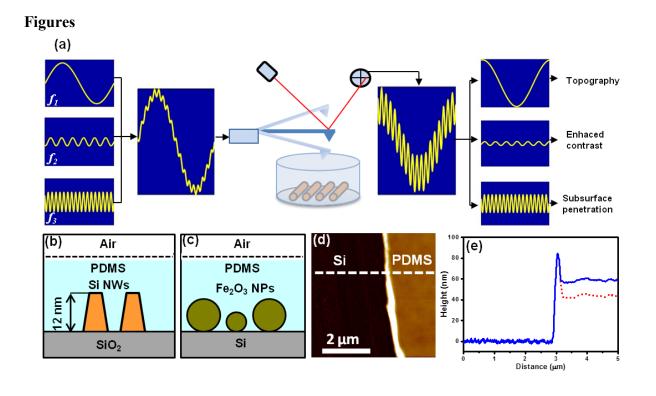


Figure 1. (a) Scheme of trimodal AFM for subsurface imaging. The driving force of the cantilever involves three components tuned at the first, second and third flexural modes of the cantilever beam. The 1st mode tracks the topography of the sample; the 2nd mode provides subsurface contrast and the 3rd mode modulates the subsurface penetration. (b) Scheme of the silicon nanowires buried under a spin-coated PDMS film. (c) Scheme of the iron oxide nanoparticles buried under a spin-coated PDMS film; $A_{sp} = 120$ nm; $A_0 = 200$ nm. (e) Cross-section along the line marked in panel (d). The profile reveals the thickness of the film (continuous line). The dash line represents the unreconstructed height profile. The peak at the edge is an artifact generated during the scratching process to generate a coated/uncoated interface.

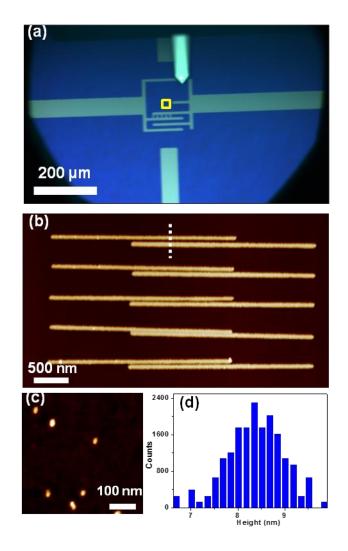


Figure 2. (a) Optical image of the metallic contacts of a silicon nanowire circuit. The marked square indicates the region that contains the SiNWs. The AFM cantilever is also seen in the image. (b) Tapping mode (amplitude modulation) AFM image of an array of SiNWs that have been fabricated by oxidation SPL in the marked region. $A_{sp} = 4.5$ nm; $A_0 = 5$ nm. The image shows the array before the spin-coated process. (c) Tapping mode AFM image of NPs deposited on a Si surface. $A_{sp} = 6.5$ nm; $A_0 = 10$ nm. (d) Height histogram of the NPs obtained from (c).

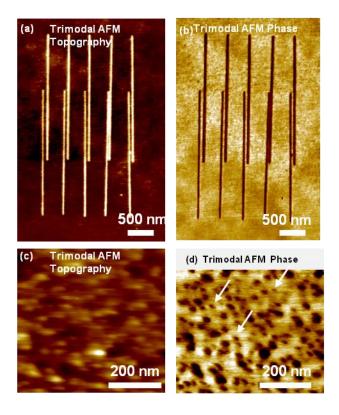


Figure 3. Trimodal AFM images of buried nanostructures. (a) Topography and (b) phase shift images of SiNWs buried under a ~70 nm PDMS film. $A_{sp1} = 113$ nm; $A_0 = (130, 2, 12)$ nm (c) Topography and (d) phase shift contrast of iron oxide NPs buried under a ~65 nm PDMS film. The topography is acquired by keeping $A_{sp1} = 120$ nm; $A_0 = (130, 1.5, 40)$ nm and the phase shift corresponds to ϕ_2 . The arrows indicate the position of NPs with a diameter below 9 nm.

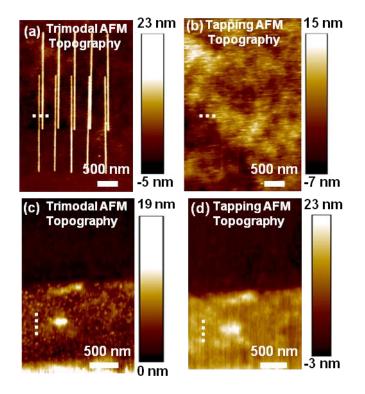


Figure 4. Comparison of tapping and trimodal AFM images. (a) Trimodal AFM image (topography) of an array of SiNWs buried under a 70 nm PDMS film; A_{sp1} =113 nm; A_0 = (130, 1.5, 40) nm (b) Tapping mode AFM image of the same array; A_{sp} =128 nm ; A_0 =160 nm. (c) Trimodal AFM image of a region that has a bare silicon surface and a section of iron oxide NPs coated by a 65 nm PDMS film. Under the PDMS there is a random distribution of NPs. A_{sp1} =100 nm; A_0 = (200, 1.5, 40) nm. (d) Tapping mode AFM image of the region shown in c. ; A_{sp} = 160 nm; A_0 = 200 nm.

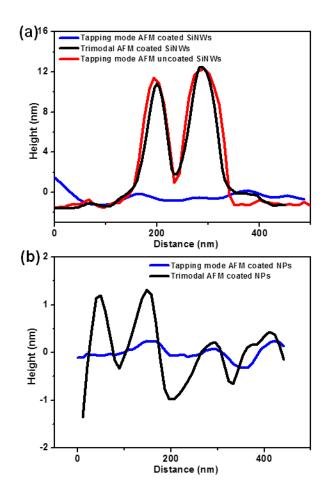


Figure 5. (a) Height cross-sections of the same section of a pair of SiNWs before and after the deposition of a PDMS film. The trimodal AFM profile matches the one obtained by tapping mode AFM before the deposition of PDMS. The tapping mode AFM image after PDMS deposition does not reveal the existence of SiNWs. (b) Height cross-section of NPs buried under PDMS; in black is the trimodal AFM cross-section; in blue is the tapping mode AFM cross-section.

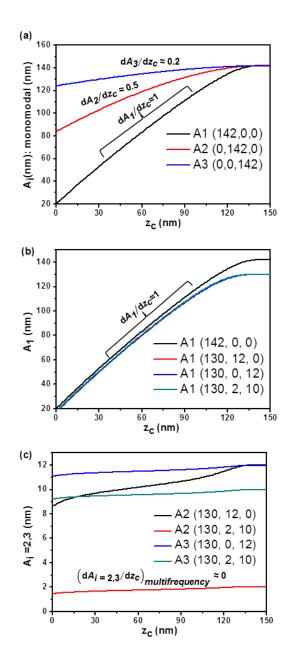


Figure 6. Dependence of the modal amplitude on the tip height distance z_c (amplitude curve) for different dynamic AFM configurations. (a) Amplitude versus the z_c for different monomodal excitations. The value of slope of the amplitude curves (central region) is also shown. (b) Amplitude curve of the first mode for monomodal and multifrequency configurations. (c) Amplitude curves for the second and third mode in multifrequency AFM. The slopes are nearly zero indicating that higher eigenmodes are unsuitable for feedback control in multifrequency AFM. The free oscillation amplitudes (A_{01}, A_{02}, A_{03}) of the excited modes are expresses in nm.

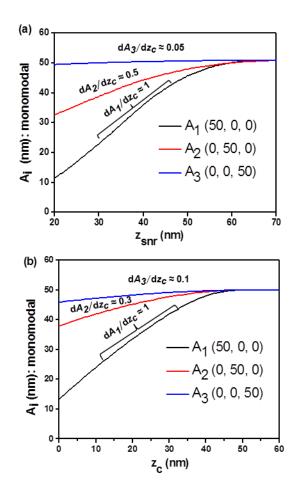


Figure 7. Experimental and numerical simulations amplitude curves. (a) Experimental amplitude curves for different monomodal excitations. (b) Numerical simulation amplitude curve for different monomodal excitations. The free oscillation amplitudes (A_{01}, A_{02}, A_{03}) of the excited modes are expresses in nm.

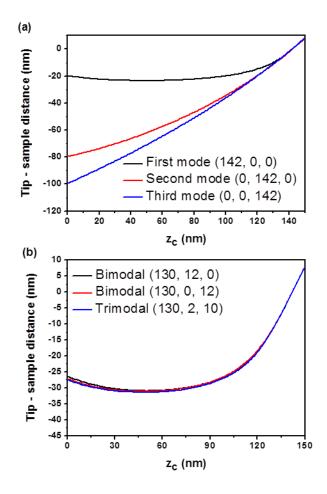


Figure 8. Tip penetration for different AFM configurations. (a) Tapping mode AFM. (b) Bimodal and trimodal AFM configurations. The free oscillation amplitudes of the excited modes (A_{01}, A_{02}, A_{03}) are expressed in nm.

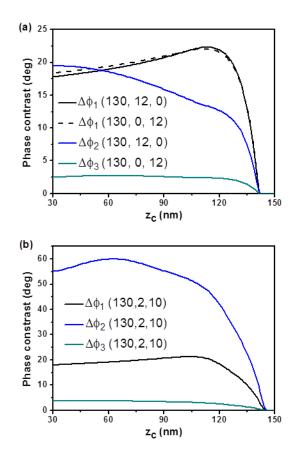


Figure 9. Subsurface phase shift contrast between a soft region (10 MPa) and relatively rigid region (1 GPa). (a) Bimodal AFM. (b) Trimodal AFM. The phase shift contrast is $\Delta \phi_i = \phi_i$ (10 MPa) – ϕ_i (1GPa).

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