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Pesticides in the Ebro River basin: Occurrence and risk assessment \star

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ABSTRACT

In this study, 50 pesticides were analyzed in the Ebro River basin in 2010 and 2011 to assess their impact in water, sediment and biota. A special emphasis was placed on the potential effects of both, individual pesticides and their mixtures, in three trophic levels (algae, daphnia and fish) using Risk Quotients (RQs) and Toxic Units (TUs) for water and sediments. Chlorpyrifos, diazinon and carbendazim were the most frequent in water (95, 95 and 70% of the samples, respectively). Imazalil (409.73 ng/L) and diuron (150 ng/L) were at the highest concentrations. Sediment and biota were less contaminated. Chlorpyrifos, diazinon and diclofenthion were the most frequent in sediments (82, 45 and 21% of the samples, respectively). The only pesticide detected in biota was chlorpyrifos (up to 840.2 ng g^{-1}). Ecotoxicological risk assessment through RQs showed that organophosphorus and azol presented high risk for algae; organophosphorus, benzimidazoles, carbamates, juvenile hormone mimic and other pesticides for daphnia, and organophosphorus, azol and juvenile hormone mimics for fish. The sum TU_{site} for water and sediments showed values < 1 for the three bioassays. In both matrices, daphnia and fish were more sensitive to the mixture of pesticide residues present.

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1. Introduction

Pesticides are a widespread group of chemical substances used to improve agricultural production. However, these substances could be persistent in water, accumulative in sediment or bioaccumulative in biota, depending on their solubility and Log K_{ow}. They are hazardous for living organisms, human health or environment, even at low concentrations (Campo et al., 2013; Claver et al., 2006; Damásio et al., 2011; Giordano et al., 2009; Masiá et al., 2015a). Furthermore, physical, chemical and biological processes degrade pesticides into one or more transformation products that could be more toxic or persistent than the parent one. There is a need of data on the real occurrence of pesticide residues in environmental matrices (De Gerónimo et al., 2014; KöckSchulmeyer et al., 2014; Palma et al., 2014a; Bruzzoniti et al., 2014; Martínez-Domínguez et al., 2015; Masiá et al., 2014, 2015b; Wei et al., 2015).

The potential ecotoxicological risks associated with pesticide residue contamination are addressed through toxic units (TUs) and/ or risk quotients (RQs) (EC, 2003; Ginebreda et al., 2014; Kökc et al., 2010). Their application in most studies is restricted to water samples (Ginebreda et al., 2014; Kuzmanović et al., 2016). However, pesticide residues can also be adsorbed into sediments (Masiá et al., 2015b). WFD (EC, 2000) and environmental quality standards (EQS) (EC, 2008; EU, 2013) unquestionably support to include sediments in the risk assessment. A variety of methods were proposed but only scarcely applied to evaluate the potential toxicity of sediments (e.g., toxic equivalent factor approach, TUs summation, hazard index) (Schwarzenbach and Westall, 1981; Booij et al., 2015; de Castro-Catalá et al., 2016).

Another problem caused by pesticides contamination is the simultaneous occurrence of several of them and the need to establish the real impact of these mixtures on biota (Cedergreen,

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2014; Roig et al., 2015), which can be predicted by independent action (IA) or concentration addition (CA). The former assumes that the components have different mechanisms of action —ignoring synergies/antagonisms and effect summation and therefore underestimating the effect— and the latter that have a similar one —overestimating the effects. (Cedergreen, 2014; Ginebreda et al., 2014; Kuzmanović et al., 2016). CA is often the recommended first step on a tiered process because presents the worst case scenario (even that synergies are not considered) (de Castro-Catalá et al., 2016).

Mediterranean area is one of the most affected by climatic fluctuations that alter hydrological conditions and originate the great wavering on concentrations of the cocktail of pesticide residues present in water (Batalla et al., 2004). Ebro River is the second largest river of the Iberian Peninsula and the first one that flows into the Mediterranean area of Spain. Previous studies performed in the Ebro River linking occurrence of pollutants, concentrations and toxicity, but most of them have focused on a single chemical family or select one environmental matrix (water, soils, sediments or biota) (Claver et al., 2006; Damásio et al., 2010; Köck-Schulmeyer et al., 2013; Köck et al., 2010; Navarro et al., 2010; Silva et al., 2011).

The objective of this study was to establish pesticide's occurrence, spatial distribution and transport and to evaluate the ecotoxicological risk in three trophic levels (Algae, daphnia and fish), using RQs for each pesticide and sumTUs for each sampling site. The partial objectives of this study were to (i) monitor the concentration of 50 pesticides and transformation products in the surface waters, sediments and biota of the Ebro River basin in two consecutive campaigns (2010–2011) (ii) compare the concentration of the pesticides found in the present study with those detected since 2001 and with the EQS values of the pesticides included in the Directive 2013/39/EU (EU, 2013), and (iii) perform an environmental risk assessment not only for water concentrations but also sediments based on the RQs and TUs methods.

2. Experimental design

2.1. Physical setting and sampling

The Ebro River is located at the northeast of Spain and drains an area of approximately 85,000 km². It has 928 km in length and receives waters from several tributaries, which altogether represent 12,000 km of waterway network, ending into Mediterranean Sea and forms a delta of more than 300 km² (Lacorte et al., 2006; Navarro et al., 2010; Roig et al., 2015). The basin is characterized by a Mediterranean valley, which forms a triangular morphological unit, surrounded by mountains. Mean annual precipitation and temperature vary with altitude, ranging respectively from 1800 mm to 8 °C in the Pyrenees to 320 mm and 18 °C in the Ebro valley. Traditionally, the Ebro River basin is agricultural land, but lately industry has been a growing sector. In 2008, one third of the total surface of the basin was agricultural and it is still the most irrigated area in Spain (906.000 ha) (Herrero-Hernández et al., 2013), the most important crops are herbaceous plants (all over the basin), grapes for wine production (La Rioja), fruit trees (Lleida) and rice (Ebro Delta) (Silva et al., 2011). The Spanish statistics estimated that ca. 14,000 T of pesticides were used in 2010 and ca. 13,500 T in 2011. The monitoring in this study comprised two sampling campaigns, 2010 and 2011, including 24 sampling stations for water and sediments covering the whole River Basin (see Fig. S-1 and S-2) and finally five for biota sampling in 2010. These sites are representative of the whole basin (geo - references are in Table S-2).

Samples were taken in October in both years. Grab water samples (2 L) were collected in clean amber glass bottles, from the

middle of the river width. Each bottle was thoroughly rinsed with MilliQ water at the laboratory and with the river water at the sampling point before collection. Sediment samples (approx. 250 g) were taken in the same point as the water ones using a Van Veen grab sampler (0.5 L capacity). They were transferred and wrapped into an aluminum foil (previously washed with methanol and dried in oven at 100 °C) that was put inside an aluminum box. Fish samples were only collected in 2010 at five selected sites of the River course: EBR2, EBR3, EBR4, EBR5 and OCA using electro-fishing because the complexity of the basin, the difficulties to perform electrofishing and the small sample sizes obtained.

All samples were transported in hermetic boxes refrigerated with ice upon arrival at the laboratory. There, the water samples were kept at 4°°C and pre-treated and processed in a period not exceeding 5 days. Before the analysis, water samples were vacuum filtered through 1 μ m glass fiber filters followed by 0.45 μ m nylon membrane filters (VWR, Barcelona, Spain). Sediment and fish samples were frozen, lyophilized (Hetosicc CD4, Birkerod, Denmark), pulverized, thoroughly mixed, passed through a 2 mm Ø sieve and kept at -20 °C until the analysis that was performed within 3 months.

2.2. Extraction procedures and instrumental analysis: water, sediment and fish samples

For this study, 42 pesticides including some of their transformation products were determined in the 2010 campaign. Carbendazim, thiabendazole, terbumeton, terbumeton deethyl, terbuthylazine, terbuthylazine deethyl, terbuthylazine-2-hydroxy and tebuconazole were added in the next year. These pesticides belong to different chemical families, with a variety of uses as well as different physicochemical characteristics and toxicity (see Table S-1).

The water extraction was carried out according to Masiá et al. (2013b). Very briefly, water samples (200 mL) were extracted using an Oasis HLB solid-phase extraction (SPE) cartridge (200 mg sorbent/6 mL cartridge, Waters, Milford, MA, USA). The cartridge was dried under vacuum for 10 min and the analytes eluted with 10 mL of dichloromethane—methanol (50:50, v/v). The extract was evaporated to dryness and reconstituted with 1 mL of methanol. The fish and sediment samples were extracted using the QuEChERS method as described by Masiá et al. (2015b). Lyophilized sediment (1 g) or fish (2 g) were extracted with 8 ml of H₂O MilliQ, 15 ml of acetonitrile, 6 g of MgSO₄ and 1.5 g of NaCl. Then, 2 mL of the resulting supernatant were cleaned-up by dispersive SPE with 0.3 g of MgSO₄, 0.1 g of PSA, 0.1 g of C₁₈ and 0.015 g of GCB. All samples were analyzed in triplicate. The results presented are the average of the three values.

The chromatographic instrument was an HP1200 series LC – automatic injector, degasser, quaternary pump and column oven – combined with an Agilent 6410 triple quadrupole (QQQ) mass spectrometer, equipped with an electrospray ionization interface (Agilent Technologies, Waldbronn, Germany). Data were processed using a MassHunter Workstation Software for qualitative and quantitative analysis (A GL Sciences, Tokyo, Japan). The detailed conditions are in the Supplementary material Tables S-3 and S-4).

2.3. Quality assurance and quality control

The analytical methods validation was detailed in the SM Table S-5. The method's limits of detection (MLDs) and quantification (MLQs) ranged from 0.01 to 2 ng L^{-1} for water, from 0.03 to 1.67 ng g^{-1} for sediment and from 0.08 to 3.75 ng g^{-1} for biota. Recovery tests were carried out in quintuplicate in order to evaluate the precision of the method. In water samples, recoveries varied

from 48% to 70% and precision was below 20% for all pesticides. In sediment and biota samples, recoveries were higher than 40% and precision ${\leq}22\%$.

Pesticide concentrations were assessed though a comprehensive quality control scheme that included: laboratory and field blanks, matrix spikes and triplicate samples. Blank contamination is the most common problem observed in the determination of pesticides at trace levels. Thus, precautions were taken to prevent contamination from personnel, organic solvents, equipment and glassware. Blank assays were performed employing MilliQ water samples, to check for laboratory background levels of the studied compounds.

2.4. Risk assessment

The Toxic Units (TUS) and Risk Quotient (RQ) were calculated according to the European guidelines for each pesticide (EC, 2003) in at least three representative taxons (algae, *Daphnia magna*, and fish) of three trophic levels in the ecosystem. Acute 48 h EC50 for *D. magna*, 72 h EC50 for algae and 96 h LC50 for fish, as well as Chronic 96 h NOEC data for algae and 21 days NOEC for fish and *D. magna* of each chemical was collected from the website http://sitem.herts.ac.uk/aeru/ppdb/en/atoz.htm. In this database the EC50 for *D. magna* is refereed to immobilization, for algae (un-known species) to growth inhibition and for fish (*Oncorhynchus mykiss* mostly) to survival. Values of any compound not available in this site were calculated using the ECOSARTM v. 1.11 (ECOlogical Structure Activity Relationship), in which the lowest toxicity prediction for each taxon was chosen to set in the worst-case scenario.

The toxic unit (TU_i) (Sprague, 1971) is used for the ecotoxicological risk assessment of measured concentrations of compounds (C_i). The TU of each compound was based on acute toxicity values. The following equation was applied for water and sediment samples.

$$TU_i$$
 (algae, daphnia, fish) = $\frac{C_i}{EC50_i}$

where TUi is the toxic unit of a compound i; C_i measured concentration (ng L⁻¹) in the water samples; EC50_i (ng L⁻¹) is the effective concentration of 50% of individuals when exposed to the substance concerned.

Site specific toxic stress (TUsite) was calculated by summing all the individual TUi of each detected compound at all of the 24 studied sites.

$$Sum TU_{site} = \sum_{i=1}^{n} TU_{i}$$

Sediment-associated pesticide concentrations were converted to pore-water concentrations according to the equilibriumpartitioning approach to comply with the sediment benchmark toxicity tests that are based on dissolved phase pesticides in pore water. Pore water concentrations from sediments were calculated according to Di Toro et al. (1991) as:

$$C_{pw} = \frac{C_s}{K_d}$$

where K_d is the partitioning coefficient, C_S is the sediment concentration and C_{PW} the pore water concentration of the pesticide. K_d was calculated as:

where K_{OC} is the dimensionless organic carbon–water partitioning coefficient for the pesticide and f_{OC} is the fraction of total organic carbon measured in the sediment samples. The K_{OC} was calculated as:

$$logK_{OC} = a \times logK_{OW} + b$$

where K_{OW} is the octanol—water partitioning coefficient. The constants a and b were set to 0.72 and 0.49, respectively (Schwarzenbach and Westall, 1981). TUs > 1 indicates environmental concern.

RQ was calculated using the following equation:

RQ = EC/PNEC

where, EC is the mean or maximum concentration of pesticides detected in the water samples and PNEC is the predicted no-effect concentration. PNEC can be calculated for acute or chronic toxicity, dividing the lowest short-term EC50 or long-term NOEC respectively by an assessment factor (AF), in this case 1000. The AF is an arbitrary factor to consider the inherent uncertainty in the obtained laboratory toxicity data. If RQ > 1, harmful effects could be expected due to the presence of the pollutant in water. On the contrary, if RQ < 0.1, the environmental risk is low. The intermediate situation in which the RQ is between 0.1 and 1 involves medium risk.

3. Results and discussion

Pollutants were more frequent in water than in sediment and biota (more apolar matrices). The low frequency can be explained because of the 50 target pesticides, only 21 had values Log K_{ow} > 3 (high), 6 between 2.5 and 3 (moderate) and 17 had values < 2.5 (low). Tables 1–3 show the minimum, maximum, mean and frequency of detection of the studied pesticides in the water, sediment and biota samples, respectively.

3.1. Residues of pesticides in water samples

The frequency was higher in 2010 than 2011. Organophosphorus, juvenile hormone mimics, azols, triazines, ureas and other pesticides were detected in both campaigns (See Table 1). In 2010, pyriproxyphen, chlorpyrifos, diazinon, buprofezin and hexythiazox were the most frequents (>90% of the samples) followed by imazalil and prochloraz (70% of the samples). In 2011, carbendazim was the most frequent (70% of the samples), whereas, diazinon, terbuthylazine and terbutryn frequency was >45% of the samples. Chlorpyrifos (95% of the samples in 2010) was already reported as the most commonly detected pesticide in the Ebro River (Claver et al., 2006; Navarro et al., 2010) even though is not usually persistent in water systems. Diazinon had a high frequency in 2010 (95% of the samples) but a medium-low one in 2011 (45%). This compound is stable in water, moderately soluble and slightly volatile (Table S-1). In 2011, carbendazim (not analyzed in 2010) was present in 70% of the sampling points. This fungicide has a low water solubility, can be persistent in water under certain conditions and is moderately persistent in soil. Herbicides terbuthylazine and terbutryn not analyzed in 2010 were detected in 50% of the samples in 2011. On the legal or illegal use of pesticides, of 50 target compounds analyzed, 14 —withdrawn by the European Union— were detected in both campaigns including carbendazim, metolachlor, azinphos methyl, chlorfenvinphos, diazinon, fenitrothion, fenthion, omethoate, parathion-methyl, atrazine, propazine, simazine, terbumeton and terbutryn (See Table S-1).

The pollution profile in both campaigns was marked by azoles, organophosphorus and triazines (detailed concentration at each

 $K_d = K_{oc} \times \, f_{oc}$

Table 1

Minimum, maximum and mean concentrations and frequency of detection of the studied pesticides in water samples.

Pollutants	2010				2011			
	Concentra	tion (ng L^{-1})			Concentration (ng L ⁻¹)			
	Min	Max	Mean	Freq (%) ^a	Min	Max	Mean	Freq (%) ^a
Azol								
Imazalil	4.91	409.76	61.01	17 (70)	1.28	121.70	7.50	8 (33)
Prochloraz	2.24	34.47	15.59	17 (70)	2.14	2.14	0.09	1 (4)
Benzimidazole				()				- (-)
Carbendazim	n.a	n.a	n.a	n.a	0.04	11.63	2.78	17 (70)
Thiabendazole	n.a	n.a	n.a	n.a	0.43	48.77	3.58	5 (20)
Carbamates								- ()
3-Hydroxycarbofuran	8.47	8.47	0.35	1 (4)	0.20	0.20	0.01	1 (4)
Methiocarb	n.d	n.d	n.d	n.d	1.24	2.52	0.30	4 (16)
Chloroacetanilide								
Metoalachlor	n.d	n.d	n.d	n.d	1.10	4.86	0.55	7 (29)
Juvenile Hormone Mimics	1110	ind	ma			100	0.00	, (20)
Pyriproxyphen	0.89	37.74	24.38	23 (95)	4.76	4.76	0.20	1 (4)
Neonicotinoid	0.05	57.71	21.50	23 (33)	1.70	1.70	0.20	1 (1)
Imidacloprid	1.84	2.77	1.06	11 (45)	1.64	14.96	1.66	9 (37)
Organophosphorus	110 1	2	1100	11(10)	110 1	1 1100	1100	0(07)
Azinphos Methyl	n.d	n.d	n.d	n.d	2.31	2.31	0.10	1 (4)
Chlorfenvinphos	2.54	41.24	17.97	18 (75)	1.57	1.57	0.07	1 (4)
Chlorpyrifos	2.64	16.40	5.97	23 (95)	1.01	2.86	0.32	5 (20)
Diazinon	0.12	13.58	5.65	23 (95)	0.53	20.39	1.35	11 (45)
Diclofenthion	13.62	22.73	12.86	18 (75)	n.d	n.d	n.d	n.d
Dimethoate	2.33	3.19	0.47	4 (16)	61.56	61.56	2.57	
Fenitrothion	2.55	2.64	0.47		36.49	36.49	1.52	1 (4)
	2.64	2.64	0.11	1 (4)				1 (4)
Fenoxon	2.64			1 (4)	n.d	n.d	n.d	n.d
Fenoxon Sulfone		2.64	0.11	1 (4)	n.d	n.d	n.d	n.d
Fenoxon Sulfoxide	2.64	20.84	4.43	9 (37)	n.d	n.d	n.d	n.d
Fenthion	2.64	2.64	0.11	1 (4)	0.33	0.33	0.01	1 (4)
Fenthion Sulfone	2.64	2.64	0.11	1 (4)	n.d	n.d	n.d	n.d
Fenthion Sulfoxide	2.64	2.64	0.11	1 (4)	n.d	n.d	n.d	n.d
Malathion	n.d	n.d	n.d	n.d	7.93	7.93	0.33	1 (4)
Omethoate	n.d	n.d	n.d	n.d	3.47	3.47	0.14	1 (4)
Parathion-Ethyl	14.01	14.45	1.19	2 (8)	n.d	n.d	n.d	n.d
Parathion-Methyl	n.d	n.d	n.d	n.d	2.00	2.00	0.08	1 (4)
Tolclophos-Methyl	8.30	16.07	3.50	7 (29)	0.50	0.50	0.02	1 (4)
Other Pesticides								
Buprofezin	2.32	8.25	5.82	22 (91)	n.d	n.d	n.d	n.d
Hexythiazox	1.90	10.57	7.41	22 (91)	1.21	1.21	0.05	1 (4)
Triazines								
Atrazine	8.13	12.22	1.99	5 (20)	n.d	n.d	n.d	n.d
Deisopropylatrazine	4.35	13.15	1.30	4 (16)	6.96	19.16	2.72	6 (25)
Deethylatrazine	6.57	58.82	7.67	7 (29)	4.99	4.99	0.21	1 (4)
Propazine	3.26	3.26	0.14	1 (4)	n.d	n.d	n.d	n.d
Simazine	30.71	47.95	3.28	2 (8)	n.d	n.d	n.d	n.d
Terbumeton	n.a	n.a	n.a	n.a	5.22	5.22	0.22	1 (4)
Terbumeton-Deethyl	n.a	n.a	n.a	n.a	0.42	9.72	0.89	8 (33)
Terbuthylazine	n.a	n.a	n.a	n.a	0.11	10.10	2.21	12 (50)
Terbuthylazine Deethyl	n.a	n.a	n.a	n.a	0.77	2.41	0.29	4 (16)
Terbuthylazine-2 Hydroxy	n.a	n.a	n.a	n.a	0.23	11.59	1.41	6 (25)
Terbutryn	14.85	14.85	0.65	1 (4)	0.92	30.54	7.66	12 (50)
Triazole	1.00			- (-)				(00)
Tebuconazole	n.a	n.a	n.a	n.a	1.66	15.38	2.36	8 (33)
Ureas	11.0	11,0	11.0	11.0	1.00	13.30	2,30	0(33)
Diuron	2.64	150.96	6.40	2 (8)	7.52	24.47	1.95	3 (12)
Isoproturon	2.58	25.48	1.60	2 (8) 4 (16)	2.41	2.41	0.10	1 (4)
isopioturon	2.30	23.40	1.00	- (10)	2.41	2,71	0.10	1 (4)

 $\mathbf{n.d} = \text{Not detected.}$ $\mathbf{n.a} = \text{Not analyzed.}$

^a Number of findings (percentage of positive samples).

point is shown in Fig S3A), Samples of 2010 were more contaminated than those of 2011. The annual pesticide loads from the Ebro River to the Mediterranean Sea were estimated to be 4359 kg in 2010 and 1606 kg in 2011. These estimations correspond to the October—November period, which is characterized by lower pesticide discharge compared to spring time. These annual pesticide loads released to the sea could affect the Ebro Delta, biota and marine ecosystems. There are several estimations in different Mediterranean rivers of the pesticide loads that arrives yearly to the Sea: Jucar River 539 kg and Turía River 156 kg (Ccanccapa et al., 2016; Mai et al., 2013; Soubaneh et al., 2015). Mediterranean Sea receives already 2301 kg of pesticides yearly just from these three rivers. Tables 4 and 5 outline concentration of pesticides in water samples of the Ebro River and of other Mediterranean Rivers from 2001 to present. Regarding pollutants found in the Ebro River organophosphorus, carbamates, triazine, azol and ureas were always the most detected compounds. The concentrations were within the range from 3 to 12,597 ng L⁻¹. The main pesticides found were atrazine, molinate, propanil, diazinon, diuron, malathion, terbuthylazine, imidacloprid, tebuconalezole and dimethoate in

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Table 2

Minimum, maximum and mean concentrations and frequency of detection of the studied pesticides in sediment samples.

Pollutants	2010			2011				
	Concentration (ng g ⁻¹ dw)			Concentration (ng g ⁻¹ dw)				
	Min	Max	Mean	Freq (%) ^a	Min	Max	Mean	Freq (%) ^a
Azol								
Imazalil	7.35	7.35	0.33	1 (4)	4.20	4.20	0.18	1 (4)
Prochloraz	4.60	4.60	0.21	1 (4)	n.d	n.d	n.d	n.d
Chlorpyrifos	0.18	9.59	1.06	10 (45)	0.88	36.17	7.66	19 (82)
Diazinon	0.28	8.85	0.63	10 (45)	0.62	3.30	0.20	3 (13)
Diclofenthion	n.d	n.d	n.d	n.d	1.44	28.82	1.73	5 (21)
Ethion	n.d	n.d	n.d	n.d	5.10	5.10	0.22	1 (4)
Malathion	1.84	1.84	0.08	1 (4)	n.d	n.d	n.d	n.d
Other Pesticid	les							
Hexythiazox	n.d	n.d	n.d	n.d	0.50	0.50	0.02	1 (4)
Triazines								
Terbutryn	3.97	21.61	1.16	2 (9)	0.10	0.10	0.00	1 (4)

^a Number of findings (percentage of positive samples). $\mathbf{n.d} = \mathbf{Not}$ detected. $\mathbf{n.a} = \mathbf{Not}$ analyzed.

agreement with this study. Although the profile of contamination is variable, since 2005 the pesticide residue concentration increased from 4680 ng L^{-1} to 12,597 ng L^{-1} in 2011.

The spatial distribution (See Fig. 1A) of pesticides along the Ebro River and its tributaries could be related to the land use (Belenguer et al., 2014; Ccanccapa et al., 2016; Vryzas et al., 2009). Pesticide concentrations were moderate to low in most of the river course. The most polluted sites are Zadorra (ZAD) in the head and Segre (SEG) as well as the Ebro Delta in the mouth. Station ZAD -located in Alava (Basque Country)— is part of the Natura 2000 Network but surrounding by cereals, sugar beets and potatoes crops and influenced by the Crispijana wastewater treatment plant. In this point, diuron exceed 100 ng L⁻¹, limit established for individual concentrations in drinking water according to EU legislation (EC, 1998). The sampling point of the Segre River (SEG) had the highest concentrations of all tributaries. In 2010, this point exceed 500 ng L⁻¹, limit established for group pesticides in drinking water, and imazalil exceed 100 ng L^{-1} , individual limit established (EC, 1998) and in 2011 the total concentration was 233.33 ng L^{-1} . These high concentrations are only punctual. Fruit trees, corn, wheat and barley crops are characteristics of this area. The high concentrations of fungicide imazalil in both campaigns could be related to the postharvest treatments of apples and pears. The Ebro Delta receives a high load of pesticides because of the intensive agricultural activities that are carried out upstream and in the Delta itself (rice cultivation) (Kuster et al., 2008). The spatial distribution showed clearly the increasing concentration gradient for both campaigns in

Table 3

Minimum, maximum and mean concentrations and frequency of detection of the studied pesticides in biota samples.

Pollutants	2010					
	Concentration ng g^{-1} dw					
	Min	Max	Mean	Freq %		
ORGANOPHOSPHORUS						
CHLORPYRIFOS	n.d	n.d	n.d	n.d		
Barbus (Barbus guiraonis)	n.d	n.d	n.d	n.d		
Barbus (Barbus guiraonis): Adult	n.d	n.d	n.d	n.d		
Barbus (Barbus guiraonis): Young	n.d	n.d	n.d	n.d		
Carp (Cyprinus carpus)	840.25	840.25	420.13	1 (20)		
Carp (Cyprinus carpus): Adult	n.d	n.d	n.d	n.d		
European catfish (Silurus glanis): Adult	168.62	168.62	84.31	1 (20)		

n.d = Not detected.

the points sampled EBR-7, EBR-8 and EBR-9 (see Fig. 1 -A). In 2010, the concentrations go up from 2.32 ng L^{-1} to 109.24 ng L^{-1} and in 2011 from 1.11 ng L^{-1} to 30.54 ng L^{-1} (Kuster et al., 2008; Ochoa et al., 2012).

The co-occurrence of different pesticides in the water samples are shown in Fig. S-4A. In 2010, 38% of the samples contained less than 5 pesticides and 22% of the samples contained more than 16 pesticides. This means that even though concentrations were low, and there was one point (SEG) that exceed the European threshold for drinking water, the number of pesticides in each sample was high. In 2011, 42% of the samples present less than 5 pesticides but 22% of the samples present among 6 to 16 pesticides. In 2011 the co-occurrence was lower than in 2010.

The differences between both sampling campaigns could be related to the river flow (see Table S-6). Considering all the flow measurements in the last ten years in each point where there are data available and normalizing them to 100, the water flow in the first campaign ranged from 0.03 m³ s⁻¹ (MAT) to 213.40 m³ s⁻¹ (EBR-7), these values represent percentiles 18% and 50% that could be considered medium-high. On the contrary, in 2011 the flows ranged from 0.01 m³ s⁻¹ (MAT) to 155.43 m³ s⁻¹ (EBR-7), percentiles 5% and 20%, respectively. These values are below of 50% percentile and could be considered low. Apparently, the higher flow, the greater frequency and co-occurrence of pesticides, and consequently in 2010 the frequency and co-occurrence was higher than 2011 (Table 1 and Fig. S-4A). Regarding the low flow, there are reports that point out that lower flows are related with higher concentrations (Masiá et al., 2015a). However, this work shows low concentrations also at low flows. The concentration could vary taking into account the physico-chemical properties of pesticides but also other environmental conditions as precipitation or temperatures (see Table S-1) (Ccanccapa et al., 2016).

3.2. Residues of pesticides in sediment samples

Pesticides detected in sediment samples in both campaigns are outlined in Table 2. Out of the 42 pesticides analyzed in 2010 and 50 pesticides in 2011, 6 and 7 respectively, were detected at the concentrations over the MLODs. In 2010, 14% of the analytes —imazalil, prochloraz, chlorpyrifos, diazinon, malathion and terbutryn— were found. The concentrations detected ranged from 1.84 to 21.61 ng g⁻¹ of dry weight (d.w). In 2011, pesticides detected were imazalil, chlorpyrifos, diazinon, diclofenthion, ethion, hexythiazox and terbutryn, and their concentrations ranged from 0.10 to 36.17 ng g⁻¹ of d.w.

Regarding the frequency, diazinon and chlorpyrifos were the most prevalent compounds in 2010, which appeared in 45% of the samples. In 2011, chlorpyrifos (82%) and diclofenthion (21%) were the most frequently detected compounds. These pesticides had high octanol/water partition coefficient (log k_{ow}) (see Table S-1), consequently, are hydrophobic, low water soluble and tend to accumulate in sediment. However, other factors influence pesticides accumulation such as the application moment and the time elapses before the next major storm event. Chlorpyrifos was detected at high frequency in both campaigns and there are other reports that also remark their presence in the Mediterranean area (Ccanccapa et al., 2016; Masiá et al., 2015a, 2013a).

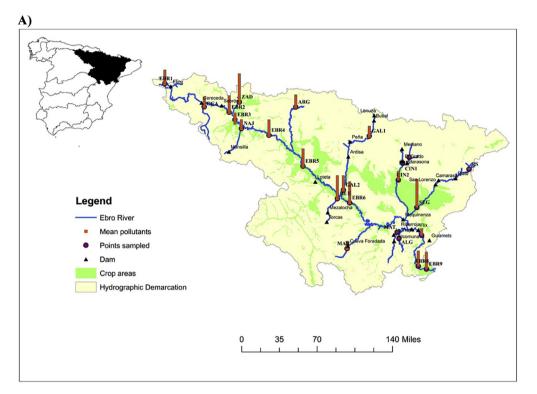
The spatial distribution of pesticides in sediment is shown in Fig. 1B and the contribution of each family of pesticides is detailed in Fig. S-4B. In 2010, the most ubiquitous pesticides were organophosphorus (38.99 ng L⁻¹), triazine (25.57 ng L⁻¹) and azol (11.94 ng L⁻¹). However, in 2011 only organophosphorus (225.62 ng L⁻¹) were found in all points sampled. Regarding the highest concentrations, in 2010 were for terbutryn (21.61 ng L⁻¹) and chlorpyrifos (9.59 ng L⁻¹) in points sampled ZAD and EBR-9

Table 4Historical data of the pesticides concentrations in the Ebro Basin.

Year	Location	Family	Pesticide	Concentrati	on (ng L^{-1})	Ref.	
				Max	Mean		
2001-2004	Ebro River	Urea	Diuron	_	105	(Claver et al., 2006)	
		Carbamates	Molinate	_	751		
		Triazine	Atrazine	-	451		
		Chloroacetanilide	Metolachlor	-	200		
		Organophosphates	Chlorpyrifos	-	312		
2004-2006	Ebro River	Triazine	Atrazine	825	62	(Navarro et al., 2010)	
		Organophosphates	Dimethoate	259	115		
		Chloroacetanilide	Alachlor	272	32		
		Carbamate	Molinate	344	107		
		Anilide	Propanil	156	34		
2005	Ebro Delta	Triazine	Triazines	935	697	(Damásio et al., 2010)	
		Anilide	Propanil	4680	1757		
		Carbamate	Molinate	485	318		
2007-2009	Ebro Basin	Chloroacetanilide	Alachlor	3	3	(Köck-Schulmeyer et al., 2013)	
		Anilide	Propanil	36	9		
		Organophosphates	Diazinon	684	133		
		Urea	Diuron	452	93		
		Triazine	Terbuthylazine	71	21		
2008	Ebro Delta	Organophosphates	Malathion	5825	1072	(Köck et al., 2010)	
		Urea	Diuron	408	72		
		Carbamates	Molinate	3590	526		
		Triazine	Terbuthylazine	1550	250		
2011	Ebro River	Triazine	Terbuthylazine	12,597	_	(Herrero-Hernández et al., 2013)	
		Urea	Diuron	8551	_		
		Neonicotinoid	Imidacloprid	656	_		
		Chloroacetanilide	Acetochlor	314	_		
		Triazole	Tebuconazole	3236	_		
		Organophosphates	Dimethoate	7549	_		

Table 5Historical data of pesticides concentration in the Mediterranean area.

Year	Location	Family	Pesticides	Concentration (ng L ⁻¹)		Ref.
				Max	Mean	
2010	Jucar River	Triazine	Atrazine-desethyl	11	_	(Belenguer et al., 2014)
	-	Organophosphorus	Chlorfenvinphos	93	_	
		Azol	Imazalil	172	_	
		Other Pesticides	Hexythiazox	21	_	
		Juvenile Hormone Mimics	Pyriproxyfen	100	_	
2010-2011	Guadalquivir River	Azole	Carbendazim	11	1	(Masiá et al., 2013a)
	•	Juvenile hormone mimics	Imidacloprid	19	2	
		Organophosphorus	Diazinon	457	19	
2010-2011	Llobregat River	Triazine	Terbuthylazine-2-hydroxy	50	13	(Masiá et al., 2015a)
	-	Organophosphorus	Malathion	320	58	
		Benzimidazole	Carbendazim	697	273	
		Carbamates	Carbofuran	7	3	
		Azol	Prochloraz	10	10	
		Other Pesticides	Hexythiazox	24	13	
		Neonicotinoid	Imidacloprid	67	25	
		Urea	Diuron	160	109	
		Chloroacetanilide	Metolachlor	13	11	
		Juvenile Hormone Mimics	Pyriproxyphen	2	2	
2012-2013	Turia River	Anilide	Propanil	46	2	(Ccanccapa et al., 2016)
		Azol	Imazalil	750	43	
		Benzimidazole	Carbendazim	382	23	
		Carbamates	Carbofuran	6845	283	
		Chloroacetanilide	Metolachlor	58	12	
		Juvenile Hormone Mimics	Pyriproxyfen	3	0	
		Neonicotinoid	Imidacloprid	207	23	
		Organophosphorus	Ethion	350	13	
		Other Pesticides	Buprofezin	25	12	
		Thiocarbamates	Molinate	14	1	
		Triazine	Terbutylazine Deethyl	59	10	
		Triazole	Tebuconazole	21	3	
		Urea	Isoproturon	13	2	





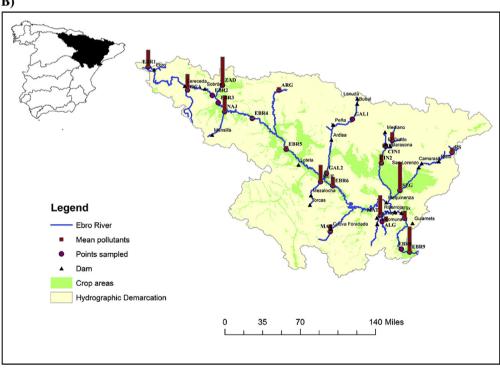


Fig. 1. Spatial distribution of pesticides in Ebro basin. A) 2010–2011 water samples and B) 2010–2011 sediment samples.

respectively. In 2011, chlorpyrifos (36.17 $\,$ ng L⁻¹ in EBR-1) and diclofenthion (28.82 ng L⁻¹ in OCA) had the highest concentrations.

The co-occurrence of pesticides in sediments can be seen in the Fig. S-4B. In both campaigns, 86% of the sediment samples did not present pesticides. In 2010, 9% and 2011, 12% had at least 5 pesticides. Only 5% and 2% samples, consecutively, presented up to 10 pesticides.

3.3. Residues of pesticides in biota samples

Fish samples were taken at five points (EBR-2, EBR-3, EBR-4, EBR-5 and OCA) in one campaign (2010). The collected fish species include, barbus (*Barbus guiraonis*), carp (*Cyprinus carpus*) and european catfish (*Silurus glanis*) (see Table 3). Chlorpyrifos ($K_{ow} = 4$) was the only pesticide detected in two fish species (Carp and European catfish). The concentrations were high, carp presented 840.25 ng g⁻¹ dw and European catfish 168.62 ng g⁻¹ dw. These data indicated possible bioaccumulation of these pesticides in fish. There are studies carried out in Mediterranean Rivers that pointed out chlorpyrifos bioaccumulation's in different fish species (Belenguer et al., 2014; Masiá et al., 2015a). Chlorpyrifos is considered as highly toxic to aquatic organisms.

4. Toxic units and risk quotient for water and sediments

The Sum TU_{site} could help to estimate the toxic effects of the mixture of pollutants per monitoring area by summing single compound TU for each sampling point as well as to study toxicity due to the contaminant present in sediments. However, the obtained Sum TU_{site} for water (Table 6) and sediment (Table 7) were <1 in all sites, evidencing that there is no acute risk associated with pollution either in water or sediments. Among the studied sites EBR-6 (0.26), ARG (0.24), ZAD (0.21), SEG (0.12), HUE (0.21), EBR-5 (0.21) and EBR-2 (0.23) showed the highest Sum TU_{site} values always for daphnia and water (See Fig. 2A). These sites reflected a dispersed pollution along the basin and a corresponding loss of ecological quality. The values do not reach the unit but are indicative of the sensitivity of D. magna to the mixture of pesticide residues in comparison with the other trophic levels. In 2011 the values were very low. These results clearly pointed out that there are not acute effects due to the mixtures of contaminants. However, complex chronic effects and interactions can not be discharged.

Subsequently, to evaluate the impact of the pesticides on the Ebro River basin ecosystems, the risk quotient (RQ) method was used employing, whenever is possible, the NOEC values obtained from chronic toxicity tests for producing the corresponding PNECs. Table 8 (Detailed Table S-7) shows the results obtained for the pesticides exhibiting low to high risk at either average or extreme condition, as calculated from their corresponding mean and

 Table 6

 Toxic units for the different sites and trophic levels for water samples.

	Algae		Daphnia		Fish	
	2010	2011	2010	2011	2010	2011
MAR	E-	E-	0.190	E-	0.028	E-
ALG	E-	E-	0.073	0.001	0.004	E-
ARG	0.002	0.006	0.240	0.001	0.020	E-
CIN1	E	E-	0.053	E-	0.003	E-
CIN2	0.001	E-	0.182	E-	0.017	E-
EBR1	0.001	E-	0.139	E-	0.019	E-
EBR2	0.001	E-	0.232	0.051	0.017	0.003
EBR3	0.001	E-	0.172	E-	0.017	E-
EBR4	0.001	E-	0.221	E-	0.019	E-
EBR5	0.001	0.003	0.210	0.011	0.019	0.001
EBR6	0.001	E-	0.263	0.001	0.020	E-
EBR7	E-	0.004	E-	0.013	E-	0.001
EBR8	0.001	0.004	0.204	E-	0.017	E-
EBR9	0.001	0.004	0.167	E-	0.023	E-
ESE	E-	E-	0.041	E-	0.003	E-
GAL1	0.001	E-	0.126	0.001	0.015	E-
GAL2	0.002	E-	0.153	0.001	0.015	E-
HUE	0.001	0.004	0.215	0.036	0.021	0.001
MAT	0.001	E-	0.029	0.000	0.002	E-
NAJ	E-	0.001	0.149	0.001	0.019	E-
OCA	E-	E-	0.102	E-	0.024	E-
RS	E-	E-	0.077	E-	0.003	E-
SEG	0.001	0.002	0.122	0.016	0.019	0.001
ZAD	0.064	0.012	0.217	0.004	0.019	E-

E- More than four decimals.

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Toxic units for the different sites and trophic levels for sediment samples.

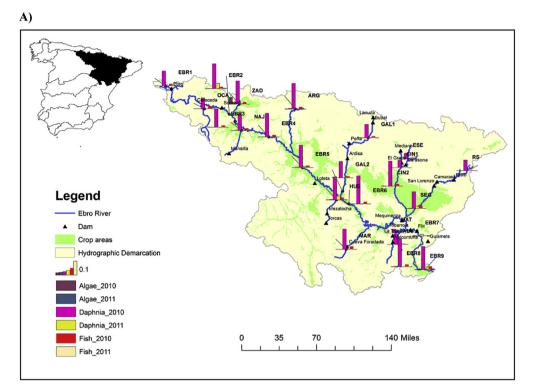
					•	
	Algae	Algae			Fish	
	2010	2011	2010	2011	2010	2011
MAR	E-	E-	0.003	E-	E-	E-
ALG	E-	E-	0.004	E-	E-	E-
ARG	0.001	0.001	0.008	E-	E-	E-
CIN1	E-	n.d	0.001	n.d	E-	n.d
CIN2	E-	E-	0.006	E-	E-	E-
EBR1	E-	E-	0.003	E-	E-	E-
EBR2	E-	E-	0.012	0.002	E-	E-
EBR3	E-	E-	0.004	E-	E-	E-
EBR4	E-	E-	0.022	n.d	0.001	n.d
EBR5	n.a	E-	n.a	E-	n.d	E-
EBR6	E-	E-	0.005	E-	E-	E-
EBR7	E-	E-	E-	E-	E-	E-
EBR8	n.a	n.a	n.a	n.a	n.a	n.a
EBR9	E-	0.001	0.004	E-	E-	E-
ESE	E-	E-	0.006	n.d	E-	n.d
GAL1	E-	E-	0.003	E-	E-	E-
GAL2	0.002	E-	0.009	E-	E-	E-
HUE	E-	0.001	0.004	0.002	E-	E-
MAT	0.001	E-	0.003	E-	E-	E-
NAJ	E-	E-	0.002	E-	E-	E-
OCA	E-	E-	0.002	E-	E-	E-
RS	E-	n.d	0.006	n.d	E-	n.d
SEG	0.001	E-	0.008	E-	E-	E-
ZAD	0.003	0.001	0.002	E-	E-	E-
E						

E- More than four decimals.

n d• Not detected

n.a: Not analyzed.

maximum concentrations (Masiá et al., 2015a; Palma et al., 2014b; Sánchez-Bayo et al., 2002). Hexythiazox and prochloraz were present in some samples at levels that involved a risk, mean and maximum concentrations (RQ values > 1) for algae. Carbendazim, chlorfenvinphos, chlorpyrifos, diazinon, dichlofenthion, fenitrothion, hexythiazox, imazalil, malathion, methiocarb, and pyriproxyfen showed also as a hazard for daphnia at mean and maximum concentrations. Finally, Chlorpyrifos, dichlofenthion, imazalil, and pyriproxyfen presented RQ > 1 for fish at both, mean and maximum concentrations. Chronic toxicity test showed the high risk caused by pesticides in three trophic levels (algae, daphnia and fish): this could cause changes in fish and invertebrate communities and the decrease of the most sensitive species or increase of the more resistant ones, with a consequent loss of biodiversity. On the other hand, out of the 6 pesticides found with values above RQ > 1 for algae, all those are herbicides and fungicides. These compounds affect photosynthesis in microalgae and its reduction in aquatic ecosystems (Booij et al., 2015). For daphnia, 16 pollutants (RQ > 1) —mostly insecticides and fungicides— could produce seriously effect in this trophic level. Finally, for fish, 8 pesticides exceed RQ > 1. Mixtures of organophosphate, azoles and carbamates pesticides were commonly found in water samples. These pesticides inhibit the activity of acetylcholinesterase and have potential to interfere with behaviors that may be essential for the survival of species. There are reports of the Carps exposed to mixtures containing some of the organophosphorus, azoles and carbamates showed concentration additive or synergistic neurotoxicity (Cedergreen, 2014; Wang et al., 2015). This implies that single-chemical assessments systematically underestimate actual risks to aquatic species in watersheds where insecticides mixtures occur. RQ and TU are important indexes to estimate the risk in different trophic levels and for the protection of aquatic ecosystems.



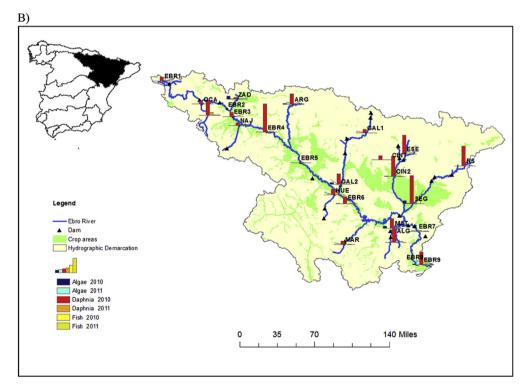


Fig. 2. Sum TU_{site} in sampling site for algae, daphnia and fish 2010–2011 A) Water samples and B) Sediment samples.

5. Conclusions

The survey carried out in 2010 and 2011 in the Ebro River and its tributaries regarding determination, distribution and ecotoxicological effects of 50 pesticides showed a dispersed pattern of concentration and risk on the different trophic levels (algae, daphnia and fish) along the basin. Water samples were the most frequently contaminated in both campaigns and in lesser extent sediment and biota samples. The most ubiquitous pesticides were azoles, organophosphorus and triazines in both years. The annual loads of pesticides for the Ebro basin were estimated in 4359 kg in 2010 and 1606 kg in 2011. This estimation was made in October and

 Table 8

 RO for algae, daphnia and fish.

Pollutants	PNEC Ng L-1	2010		2011	
		RQ-Mean	RQ-Max	RQ-Mean	RQ-Max
Chronic 96/72 h N	OEC in Algae				
Alachlor					
Atrazine	100	<0.1	0.1		
Chlorpyrifos	43	0.1	0.4	<0.1	<0.1
Dichlofenthion	204	0.1	0.1		
Diuron	93	0.1	1.6	<0.1	0.3
Fenitrothion	100	<0.1	<0.1	<0.1	0.4
Hexythiazox	7	1.1	1.5	<0.1	0.2
Imazalil	92	0.7	4.5	0.1	1.3
Isoproturon	52	<0.1	0.5	<0.1	<0.1
Metolachlor	1	4.6	2.4	0.9	8.2
Prochloraz	10	1.6	3.4	<0.1	0.2
Propazine	40	< 0.1	0.1	0.1	0.1
Pyriproxyfen	213	0.1	0.2	<0.1	<0.1
Tebuconazole	100 28	-0.1	0.5	<0.1 0.3	0.2
Terbutryn		<0.1			1.1
Chronic 96/72 h N Azinphos Methyl	0.4	Invertebrate	s (Dupinnu	0.2	5.8
Buprofezin	80	0.1	0.1	0.2	5.8
Carbendazim	1.5	0.1	0.1	1.9	7.8
Chlorfenvinphos	0.1	179.7	412.4	0.7	15.7
Chlorpyrifos	4.6	1.3	3.6	0.1	0.6
Diazinon	0.56	10.1	24.2	2.4	36.4
Dimethoate	40	<0.1	0.1	0.1	1.5
Diuron	96	0.1	1.6	<0.1	0.3
Fenitrothion	0.09	1.3	30.3	17.5	419.4
Hexythiazox	6.1	1.2	1.7	<0.1	0.2
Imazalil	15	4.1	27.3	0.5	8.1
Isoproturon	120	<0.1	0.2	<0.1	<0.1
Malathion	0.06			5.5	132.1
Methiocarb	0.1			3	25.2
Prochloraz	18	0.9	1.9	<0.1	0.1
Pyriproxyfen	0.02	1625.6	2515.7	13.2	317
Tebuconazole	10			0.2	1.5
Terbutryn	205	<0.1	0.1	<0.1	0.1
Thiabendazole	42			0.1	1.2
Chronic 21 days N					
Azinphos Methyl	0.17			0.6	13.6
Buprofezin	52	0.1	0.2		
Carbendazim	3.2			0.9	3.6
Chlorfenvinphos	30	0.6	1.4	<0.1	0.1
Chlorpyrifos	0.14	42.6	117.2	2.3	20.4
Dichlofenthion	4	3.2	5.7	0.1	
Dimethoate	400	<0.1	<0.1	<0.1	0.2
Diuron	410	< 0.1	0.4	<0.1	0.1
Fenitrothion Fenoxon Sulfone	88	<0.1	<0.1	<0.1	0.4
Hexythiazox	23 40	<0.1 0.2	0.1 0.3	<0.1	< 0.1
	40 43		0.5 9.5	<0.1 0.2	2.8
Imazalil Malathion	43 91	1.4	9.5	<0.2	0.1
Methiocarb	50			<0.1	0.1
Prochloraz	30 49	0.3	0.7	<0.1 <0.1	<0.1
Pyriproxyfen	49	0.3 5.7	8.8	<0.1 <0.1	<0.1 1.1
Simazine	700	<0.1	0.1	~0.1	1.1
Tebuconazole	12	~ 0.1	0.1	0.2	1.3
Terbutryn	104	<0.1	0.1	0.2	0.3
Thiabendazole	12		0.1	0.3	4.1

November; a period characterized by lower pesticide discharge, and in 24 points sampled, demonstrating a high impact in the delta and marine ecosystems. The ecotoxicological assessment point out that exist a chronic toxicity (RQ index) caused by pesticides (organophosphorus, azol, carbamates and juvenile hormone mimics) in three trophic levels (algae, daphnia and fish), specially in *Daphnia magna*. The Toxic unit for water and sediments, calculated to assess the effects of the cocktail of pesticide residues and know the specific sites impacted, showed the daphnia as the most sensitive in 2010 along the basin. According to the TUs, there are not acute effects due to pesticide concentrations either in water or

sediments. However, several pesticides showed a RQ > 1 indicating that pesticide risk to the aquatic communities needs further study. A long-term chronic study on assessment of these mixtures is highly required.

Acknowledgment

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Appendix A. Supplementary data

Supplementary data related to this article can be found at http://dx.doi.org/10.1016/j.envpol.2015.12.059.

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