

1 **JET MILLING EFFECT ON WHEAT FLOUR CHARACTERISTICS AND**
2 **STARCH HYDROLYSIS**

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11 **Highlights:**

- 12 • The effect of jet milling settings on wheat flour characteristics was evaluated.
- 13 • Large aggregates were reduced in size separating starch granules and proteins.
- 14 • Jet milled flours showed lower viscosity and faster enzymatic starch hydrolysis.
- 15 • Controlling jet milling settings allow obtaining flours with diverse functionality.

16

17 **Abstract**

18 The interest for producing wheat flour with health promoting effect and improved
19 functionality has led to investigate new milling techniques that can provide finer flours.

20 In this study, jet milling treatment was used to understand the effect of ultrafine size
21 reduction onto microstructure and physicochemical properties of wheat flour. Three
22 different conditions of jet milling, regarding air pressure (4 or 8 bars) feed rate and
23 recirculation, were applied to obtain wheat flours with different particle size (control,
24 F1, F2 and F3 with d50 127.45 μm , 62.30 μm , 22.94 μm and 11.4 μm , respectively).

25 Large aggregates were gradually reduced in size, depending on the intensity of the
26 process, and starch granules were separated from the protein matrix. Damaged starch
27 increased while moisture content decreased because of milling intensity. Notable
28 changes were observed in starch hydrolysis kinetics, which shifted to higher values with
29 milling. Viscosity of all micronized samples was reduced and gelatinization
30 temperatures (T_o , T_p , T_c) for F2 and F3 flours increased. Controlling jet milling
31 conditions allow obtaining flours with different functionality, with greater changes at
32 higher treatment severity that induces large particle reduction.

33

34 **Keywords:** jet milling; wheat flour; starch hydrolysis; pasting properties; thermal
35 parameters.

36 **Introduction**

37 Nowadays, alternative milling procedures and micronizing technologies are tested in
38 order to produce flours with enhanced functional properties, which are suitable for
39 making new edible products or for improving the properties of the current ones. Milling
40 technologies focused on producing finer flours with improved properties are getting
41 increased attention (de la Hera et al. 2013; Protonotariou et al. 2014; Sakhare et al.
42 2015). Jet milling is a new technological development that aims at the production of
43 super fine flours by accelerating the particles in a high-velocity air stream, the size
44 reduction being the result of inter-particle collisions or impacts against solid surface
45 (Létang et al. 2002; Protonotariou et al. 2014, 2015). The particles impact at high
46 velocities produces superfine powders and reduces the size of all aggregates (Létang et
47 al. 2002). It is a fluid energy impact-milling technique, commonly used to produce
48 particle sizes lower than 40 μm (Chamayou and Dodds 2007), which are greatly
49 appreciated in the chemical, pharmaceutical and mineral industry (Midoux et al. 1999).
50 In food applications, smaller particle size results in faster starch digestion (de la Hera et
51 al. 2014). Small particles have high surface-to-volume ratio increasing the access of
52 enzymes to the interior of the particle taking advantage of the absence of intact cell
53 walls (Heaton et al. 1988). An increased surface area of food materials could increase
54 the rate of water absorption of materials, improving solubility of dry products, and
55 increase site accessibility for chemical reactions (e.g., oxidation, digestion, flavor
56 release, catalyst, and enzyme activity) (Augustin and Sanguansri 2009). Jet milling
57 combined with air classification has been successfully used to separate starch from
58 protein in order to produce starch-rich fine flours (Graveland and Henderson 1991).
59 Furthermore, differential scanning calorimetry showed lower gelatinization enthalpy
60 values for the doughs (flour:water, 60:40) of fine flours than their coarse flour

61 counterparts (6.76-7.09 and 9.92-10.12 mJ/mg respectively) (Vouris et al. 2013).
62 Overall, particle size of wheat flour seems to have an impact on dough mechanical and
63 starch gelatinization properties. Therefore, there is a consensus that particle size
64 reduction promotes changes in the majority of physicochemical properties due to the
65 increase of a particle's surface area (Tóth et al. 2006), although it must be assessed if
66 there is a critical point that leads to an increase of damaged starch (Protonotariou et al.
67 2014). The higher the specific surface area per weight unit, the higher the rate of
68 hydration and water absorption is (Manley et al. 2011). Generally, starch granules
69 become physically injured with milling's shearing and scrapping, i.e., starch damage
70 occurs (Oladunmoye et al. 2010), which could also increase water holding capacity.
71 Moreover the production of ultrafine powders from cereals flours may present benefits
72 to human health. Sanguansri and Augustin (2006) suggested that jet milling may be
73 useful for modifying or improving functionality and availability of bioactive
74 compounds.

75 Therefore, the objective of the present study was to determine the impact of jet milling
76 conditions on the wheat flour characteristics compared to conventional wheat flour.
77 Specifically, this study evaluate the enzymatic starch hydrolysis, chemical composition,
78 thermal and pasting features of different mill fractions of wheat flour obtained from jet
79 milling varied in the severity of the process.

80 **2. Materials and Methods**

81 2.1. Flour

82 Commercial soft wheat flour (T70) donated by the Company Loulis Mills S.A was
83 pulverized in a jet mill (Model 0101S Jet-O-Mizer Milling, Fluid Energy Processing
84 and Equipment Company, Telford, Pennsylvania, USA) using three different conditions
85 regarding air pressure, feed rate, vibration rate of the feeder and feedback (Table 1).

86 2.2. Particle size distribution

87 Particle size distributions were determined by laser technology with a Malvern
88 Mastersizer 2000 diffraction laser particle sizer (Malvern Instruments, Worcestershire,
89 UK), equipped with a Scirocco dry powder unit (Malvern Instruments, Worcestershire,
90 UK). Median diameter (d₅₀) is the value of the particle size, which divides the
91 population exactly into two equal halves i.e. there is 50% of the distribution above this
92 value and 50% below. The particles were assumed to have a refractive index of 1.53.

93 2.3. Scanning Electronic Microscopy (SEM)

94 Wheat flours were mounted on metal stubs with double-sided stick tape and sputter-
95 coated with a 100–200 Å thick layer of gold and palladium by ion sputter (Bio-Rad SC-
96 500, Aname, Madrid, Spain). Analysis of the specimens was performed at 10 kV
97 accelerating voltage with a SEM (S-4800, Hitachi, Ibaraki, Japan) equipped with a field
98 emission gun, a backscattered detector of RX Bruker, transmission detector, the
99 QUANTAX 400 programmed for microanalysis and the five motorized axes Scanning
100 electron microscope with a spotlight of field emission (FEG) and a resolution of 1.4nm
101 at 1KV. The microstructure analysis was carried out in the Central Service for
102 Experimental Research of the Universidad de Valencia.

103 2.4. Flours composition

104 Moisture and protein content were determined in all the samples. Moisture content was
105 determined by ICC Standard Method (ICC 2011) and protein content was determined
106 according to AACC method (AACCI 2012) with a Foss 2300 Kjeltex Analyzer Unit
107 (Foss, Hillerød, Denmark). Starch damage (iodine absorption) was measured with a
108 SDmatic (Chopin, Villeneuve-la-Garenne, France) according to AACC (AACCI 2012).
109 These measurements were carried out in Loulis Mills S.A company. All determinations
110 were carried out in triplicate.

111 2.5. Pasting properties

112 The pasting properties were determined with rapid visco analyser (RVA) (model 4-SA,
113 Perten, Instruments, Hägersten, Sweden) by following the AACC Method (AACCI
114 2012), with minor modifications. Distilled water (25 mL) was added to 3.5 g of flour
115 placed into the aluminum RVA canister. RVA settings during assessment were: heating
116 from 50 to 95 °C in 282 s, holding at 95 °C for 150 s and then cooling to 50 °C. Each
117 cycle was initiated by a 10 s at 960 rpm paddle speed for getting an even suspension
118 followed by 160 rpm paddle speed for the rest of the assay. Viscosity was recorded
119 during a heating–cooling cycle using Thermocline software for Windows (Newport
120 Scientific Pvt. Limited, Warriewood, Australia). Peak onset, peak viscosity, holding
121 through, breakdown, final viscosity and setback (difference between final viscosity and
122 peak viscosity) were evaluated.

123 2.6. Thermal parameters

124 Thermal behavior from wheat flour samples were determined using a differential
125 scanning calorimeter (DSC) from Perkin–Elmer (DSC 7, Perkin–Elmer Instruments,
126 Norwalk, CT), equipped with a thermal analysis data station (Pyris software, Perkin–
127 Elmer Instruments, Norwalk, CT). For the study, flour samples were accurately weighed
128 into aluminum DSC pans, and de-ionized water was added by micropipette to achieve a
129 water-sample ratio of 3:1 (9 mg : 3 mg). The sample pans were sealed and equilibrated
130 at room temperature for one hour before analysis. Nitrogen was used to purge analyses
131 cells. Instruments were calibrated with indium, using an empty pan as reference.
132 Thermal analysis consisted on heating from 30 to 120 °C at a rate of 5 °C/min. The onset
133 temperature T_o , peak temperature T_p , and conclusion temperature T_c were determined
134 from the heating DSC curves. Gelatinization enthalpy (ΔH) was evaluated based on the

135 area of the main endothermic peak, and peak height index (PHI) was calculated as PHI
136 = $\Delta H / (T_p - T_o)$. All DSC experiments were run three times.

137 2.7. Starch hydrolysis kinetics

138 Starch hydrolysis was measured following the method described by Gularte and Rosell
139 (2011) with minor modifications. Briefly, for free sugars removal, flour samples (0.1 g)
140 suspended in 2 mL of 80% ethanol was kept in a shaking water bath at 85 °C for 5 min,
141 and then centrifuged for 10 min at 1000×g. Supernatant was separated to measured free
142 sugar (FS) released.

143 The remaining pellet was incubated with porcine pancreatic α -amylase (0.24 U/mg
144 sample) (Type VI-B, ≥ 10 units/mg solid, Sigma Chemical, St. Louis, USA) in 4 mL of
145 0.1 M sodium maleate buffer (pH 6.9) in a shaking water bath at 37 °C. Aliquots of 200
146 μ L were withdrawn during the incubation period (0.25–16 h) and mixed with 200 μ L of
147 ethanol (96%, w/w) to stop the enzymatic reaction and the sample was centrifuged at
148 10,000 \times g for 5 min at 4 °C. The precipitate was washed twice with 50% ethanol (200
149 μ L) and the supernatants were pooled together and kept at 4 °C for further glucose
150 enzymatic release. Supernatant (100 μ L) was diluted with 850 μ L of 0.1 M sodium
151 acetate buffer (pH 4.5) and incubated with 50 μ L amyloglucosidase (AMG 1100 BG,
152 1100 AGU/g, Novozyme A/S, Bagsvaerd, Denmark) at 50 °C for 30 min in a shaking
153 water bath. For resistant starch (RS) determination after 16h of hydrolysis the sediment
154 was solubilized with 2 mL of 2 M KOH using a Polytron ultraturrax homogenizer IKA-
155 T18 (IKA works, Wilmington, NC, USA) during 1 min at speed 3. The homogenate was
156 diluted with 8 mL 1.2 M sodium acetate (pH 3.8) and incubated with 100 μ L
157 amyloglucosidase (33 U/mL) at 50 °C for 30 min in a shaking water bath. After
158 centrifuging at 2,000 \times g for 10 min, supernatant was kept for glucose determination.
159 Digestible starch (DS) was determined in the supernatant after 16 h of incubation. The

160 glucose content was measured using a glucose oxidase–peroxidase (GOPOD) kit
161 (Megazyme, Dublin, Ireland). The absorbance was measured using an Epoch microplate
162 reader (Biotek Instruments, Winooski, USA) at 510 nm. Starch was calculated as
163 glucose (mg)× 0.9. Replicates (n = 4) were carried out for each determination.

164 Experimental data were fitted to a first-order equation (Goni et al. 1997):

$$165 \quad C_t = C_\infty (1 - e^{-kt}) \quad (1)$$

166 Where C_t is the concentration of product at time t , C_∞ is the concentration at the end
167 point, and k is the pseudo-first order rate constant. The plot of $\ln [(C_\infty - C_t)/ C_\infty] = -kt$
168 against t was used to estimate the slope that corresponded to $-k$.

169 2.8 Statistical analysis

170 Experimental data were statistically analyzed using Statgraphics V.7.1 program
171 (Bitstream, Cambridge, MN) to determine significant differences among them. ANOVA
172 test was applied in order to compare the mean values of studied properties at 95% level
173 of confidence. A correlation analysis was also carried out to determine possible
174 relationships among parameters.

175 3. Results and Discussion

176 3.1. Microstructure and particle size of samples

177 Jet milling process resulted in a significant reduction of median particle size that
178 depended on the process conditions (Table 1). As the intensity of milling conditions
179 increased (feedback or increase of pressure), the size of particles decreased gradually.
180 Milling at 4 bar pressure (F1) decreased the median diameter (d50) from 127.45 μm to
181 62.3 μm , while at 8 bar pressure, particle size decreased to 22.94 (F2) or 11.44 μm (F3)
182 depending on the feeding rate 4.08 kg/h or 1.93 kg/h, respectively. A relationship
183 between pressure applied and the particle size of the samples obtained by jet mill was
184 found ($r=-0.9915$, $P<0.01$).

185 To find out any changes in terms of flour structure with intense milling SEM pictures
186 are presented (Fig.1). According to SEM micrographs, a gradual flour components'
187 disaggregation was observed according to the severity of jet mill treatment. Control
188 flour (Fig.1.A) showed large aggregates ($\geq 200 \mu\text{m}$ length) of protein matrix embedding
189 starch granules. These aggregates displayed smaller size and they were largely
190 fragmented and more separated from the protein matrix as the process became more
191 intense (Fig. 1, B-D). F1 sample showed larger particles than F2 and F3, whereas slight
192 differences between F2 and F3 microstructure were detected. Some starch granules (10-
193 35 μm) appeared deformed, separated from matrix, with rounder shape, disengaged
194 from the protein that was eroded and appeared in smaller aggregates or completely shed
195 having a polygonal shape, as a consequence of milling. The starch granules released
196 from the protein matrix agrees with previous studies confirming that this technology
197 might be an efficient process for the separation of starch and proteins (Sanguansri et al.
198 2006).

199 3.2. Chemical composition

200 As a consequence of jet milling, physicochemical properties of wheat flour changed
201 (Table 2). Moisture content of jet milled flours was reduced gradually, depending on the
202 severity of the process and it was significantly correlated with d50 ($r=0.9925$, $P<0.01$).
203 Treated flours at 8 bars pressure, presented the higher loss of moisture (51% in F3
204 sample) indicating that pressure affected significantly the moisture content ($r=-0.9803$,
205 $P<0.05$). Moisture content decreased as the particle size diminished, because higher
206 surface area was available to interact. Moreover, jet milling reduces moisture content of
207 flours due to their exposure to dry air of high flow rate as has been already observed
208 (Protonotariou et al. 2015).

209 Concerning damaged starch, it increased owing to milling (Table 2). A positive
210 significant correlation was found between damaged starch and air pressure ($r=0.9967$,
211 $P<0.01$). However, there was no significant difference between the samples F2 and F3.
212 Feedback did not increase further the amount of damaged starch, as was observed in F3,
213 indicating that the intensity of the process induced an increase in damaged starch up to
214 certain limit. Thus, it was confirmed that one of the advantages of jet mill is the reduced
215 damaged starch promoted in comparison to other milling processes. Hossen et al.
216 (2011a) reported that jet milled white rice flour with d50 45 μm had less starch damage
217 than rice flour processed by a hammer mill, with d50 53 μm , in spite of similar mean
218 size.

219 Protein amount ranged from 9.69 to 10.28%. Differences in the protein content were
220 statistically significant for F1 and F2 (Table 2), but, there was no general trend
221 considering treatment conditions. It has been reported that finer fractions had lower ash
222 and higher dry gluten than coarser fractions when wheat flour was fractioned by sieving
223 (Sakhare et al. 2014). Nevertheless, Protonotariou et al. (2015) do not observed any
224 trend when studying the impact of jet milling intensity on the protein and ash content of
225 whole wheat flour.

226 3.3. Pasting properties

227 Figure 2 illustrates RVA pasting curves for control flour and jet milled flours. Viscosity
228 of all micronized samples decreased because of milling and changes in the pasting
229 curves were readily evident during heating and cooling stages. The diverse damaged
230 starch content, changes in the particle size of the flour or differences in starch
231 accessibility might explain the different pasting performance. Barrera et al. (2013)
232 mentioned that damaged starch granules facilitated hydration and swelling, increasing
233 the viscosity of unheated starches. In the present study, in spite of the damaged starch

234 content of the samples was significant different, pasting plots did not reveal differences
235 on the shoulder exhibited during heat, which has been related to the amount of damaged
236 starch (Figure 2). In addition, it has been reported that particle size distribution affects
237 pasting properties of rice flour (Hossen et al. 2011a), but it seems that pasting properties
238 become independent on the particle size with fine flours ($<132\ \mu\text{m}$) (Martínez et al.
239 2014). Nevertheless, in the present study particle size was much lower than $132\ \mu\text{m}$ and
240 viscosity plots revealed great impact of particle size on the pasting properties of wheat
241 flour.

242 Calculated pasting parameters from RVA curves can be seen in Table 3. Peak viscosity,
243 which had a negative correlation with feedback ($r=-0.9596$, $P<0.05$), was significantly
244 reduced (control>F1>F2>F3), likewise breakdown viscosity and total setback. It has
245 been reported that peak viscosity was affected linearly by percentage of damaged starch
246 (Hasjim et al. 2013; Hossen et al. 2011b), which agrees with results of the present study,
247 although a non-significant negative correlation was found between peak viscosity and
248 damaged starch content ($r=-0.9132$, $P>0.05$). Hossen et al. (2011a) reported that peak
249 viscosity was almost constant for dry jet milled rice flour with $d_{50}>50\ \mu\text{m}$ but
250 decreased gradually at lower mean size, and dramatically at $d_{50}<10\ \mu\text{m}$. Therefore,
251 reduced peak viscosities of the processed flours indicated that smaller particles are more
252 resistant to swelling or required longer periods and RVA measurements are affected at
253 those levels of particle size. Hossen et al. (2011b) suggested that after pulverization,
254 peak and final viscosities of all flours (rice, wheat, corn maize, potato, sweet potato,
255 cassava) decreased. Final viscosity and setback were progressively reduced according to
256 the intensity of the jet milling treatment. Final viscosity and holding strength differed
257 significantly for samples F2 and F3. This reduction might be attributed to either the
258 breakage of amylose chains with lower ability to retrograde during cooling as has been

259 reported for extrusion (Martínez et al. 2014), or differences in the amylose chains
260 leakage during gelatinization associated to particle size that consequently affected
261 amylose retrogradation. In fact, larger flour particles have greater physical barrier for
262 both heat transfer and water diffusion (Hasjim et al. 2013).

263 *3.4. Thermal parameters*

264 Thermal properties of jet milled samples were investigated to assess the possible impact
265 of this treatment at molecular level (Table 4). In the range of temperature tested (30 to
266 110 °C), flours exhibited one endothermic peak corresponding to amylopectin
267 gelatinization. Specific differences among the treated samples were observed, which
268 grouped the samples in control and F1, and on the other hand F2 and F3. The change in
269 the size of the granules influenced the gelatinization temperatures (T_o , T_p , T_c) and
270 samples F2 and F3 (the lowest particle size), showed significantly higher values
271 comparing to control and F1 samples. Therefore, gelatinization temperatures were
272 progressively shifted to higher values when flours were treated at high milling intensity
273 (8 bar and/or feedback), but the temperature range was not affected significantly.
274 Martínez et al. (2014) stated that gelatinization temperatures were dependent on the
275 particle size, but the present study shows that when applying jet milling no direct
276 correlation was detected between particle size and T_o , T_p , T_c . Gelatinization enthalpy
277 (ΔH) differed significantly only between F2 and F3 but no trend was observed with the
278 intensity of the treatment. Emami et al. (2010) observed that micronization in barley
279 slightly increased T_o , T_p , T_c and reduced ΔH . Moreover, Münzing (1991) referred that
280 gelatinization peak temperature (T_p) increased slightly by milling because of damaged
281 starch. In the present study damaged starch correlated positively with T_o , T_p , T_c , but no
282 significantly.

283 *3.5. Hydrolysis of starch*

284 The potential impact of the jet milling on the integrity of the starch granules was
285 assessed by evaluating the starch susceptibility to enzymatic hydrolysis. The digestion
286 curves of the processed samples were slightly higher than those of the control (Figure
287 3). Jet milled samples presented faster starch hydrolysis compared to control. The finer
288 flours the wider surface area of granules. High surface area of flour particles increases
289 the water diffusion and enzyme accessibility according to de la Hera et al. (2013). The
290 disaggregation of wheat flour constituents favored the alfa amylase accessibility,
291 increasing the starch susceptibility to be hydrolyzed. Detached starch granules from
292 protein matrix could also lead to rapid hydrolysis of starch. F3 sample showed the
293 higher values of hydrolyzed starch (Figure 3) and this can be ascribed to the intensity of
294 the process, since F3 was re-milled. The kinetics parameters, confirmed that jet milled
295 samples showed augmented rate of hydrolysis with significant differences on the
296 hydrolysis constant (k) (Table 5). C_{∞} was positively correlated with d50 ($r=0.9738$,
297 $P<0.05$) and moisture ($r=0.9927$, $P<0.01$) and negatively with damaged starch ($r=-$
298 0.9671 , $P<0.05$). Results suggested that by decreasing particle size (d50), which
299 simultaneously increases damaged starch, lower hydrolysis plateau would be reached.
300 Presumably smaller particles favor the rapid accessibility to the amorphous part of the
301 starch granules, reaching earlier the more resistant crystalline structure of the granules,
302 which would explain the lower plateaus. Starch granules were faster and in lesser extent
303 hydrolyzed at sample F3. Surface components of starch granule, such as proteins, can
304 create a surface membrane that acts as a physical barrier to digestion, proteins layers
305 should be significantly degraded before starch digestion takes place (Svihus et al. 2005).
306 Resistant starch was also measured to determine the potential impact of the jet mill on
307 the structural level of starch (Table 5). RS increased in jet milled samples but
308 differences were not significant. During jet milling, flours were not exposed to thermal

309 stress that may affect the RS amount, which explained the absence of differences. The
310 amount of free sugars was low, but as the intensity of the process increased the amount
311 of FS also augmented. F3 differed significantly from the other samples and presented
312 the higher percentage of free sugars, indicating a high correlation between FS and
313 feedback ($r=0.9856$, $P<0.05$).

314 **Conclusions**

315 Jet milling is an alternative method to produce ultra-fine flour. Milling conditions
316 determine the final particle size and the thereafter flour properties. In the present study,
317 particle size was reduced up to ten times (mean diameter 11.44 μm in F3). Decrease in
318 particle size led to starch granules detached from protein matrix and a significant
319 breakage of aggregates took place. Damaged starch increased but not to a dramatic
320 extent. As long as the treatment was mild (F1), similarities to the control samples were
321 shown in terms of pasting properties. When the process became more intense, small
322 particles presented a retardation to gelatinize and pastes were less viscous either in
323 gelatinization or in the gel forming process. Moreover, starch hydrolysis increased in
324 terms of particle size reduction as higher surface area led to higher starch susceptibility.
325 Therefore, a treatment of 8 bar pressure without feedback (F2) could be used to achieve
326 high particle size reduction. More intense milling treatment, as in F3 with feedback,
327 would not lead additional changes in flour functionality, likely due to reaching the limit
328 of particle sizes. Thermal parameters, hydration and pasting properties of flours are
329 crucial for the developing food products. Therefore, the incorporation of jet milled
330 flours in food process would be of great interest.

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415

416 **Tables**

417 **Table1.** Samples codes describing the jet mill settings for the samples treatment.

Flour code	Air pressure (bar)	Feed Rate (kg/h)	Vibration Rate of Feeder (%)	Feedback	Particle size d50 (µm)
Control	-	-	-	-	127.45
F1	4	2.71	100	No	62.30
F2	8	4.08	100	No	22.94
F3	8	1.93	100	Yes	11.44

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419

420 **Table 2.** Chemical composition of wheat flour and jet milled wheat flours F1, F2 and
 421 F3, expressed as percentage of dry basis (d.b.).

Name	Moisture (%)		Damaged starch (%, db)		Protein content (%, db)	
Control	15.56 ± 1.17	c	2.37 ± 0.33	a	9.80± 0.03	a
F1	10.17 ± 0.17	b	4.61 ± 0.30	b	10.28± 0.02	c
F2	7.83 ± 0.34	a	6.26 ± 0.08	c	10.05± 0.09	b
F3	7.62 ± 0.22	a	6.51 ± 0.13	c	9.69± 0.02	a

422 Values followed by different letters in each column indicate significant differences ($P \leq 0.05$).

423

424

425 **Table 3.** RVA pasting properties of wheat flour and jet milled wheat flours F1, F2 and F3.

Sample	Pasting onset (°C)		Peak viscosity (cP)			Holding strength(cP)			Breakdown (cP)			Final viscosity(cP)		Total Setback (cP)			
Control	64.5	± 1.17	3412	± 16.97	d	1923	± 33.23	c	1490	± 16.26	c	3599	± 43.84	c	1677	± 10.61	d
F1	65.8	± 0.64	3121	± 16.97	c	1901	± 26.87	c	1220	± 43.84	b	3545	± 17.68	c	1644	± 9.19	c
F2	63.2	± 3.04	2846	± 6.36	b	1653	± 20.51	b	1193	± 14.14	b	3187	± 24.75	b	1534	± 4.24	b
F3	65.3	± 1.31	2407	± 25.46	a	1520	± 5.66	a	887	± 19.80	a	2973	± 9.90	a	1453	± 4.24	a

426 Values followed by different letters in each column indicate significant differences ($P \leq 0.05$).

427

Table 4. Thermal parameters of wheat flour (control) and jet milled wheat flours F1, F2, F3 determined by DSC.

Sample	To (°C)	TP (°C)	Tc(°C)	Tp-To(°C)	ΔH (J/g)	PHI= ΔH/To-Tp
Control	57.14 ± 0.79 a	61.34 ± 0.75 a	66.12 ± 0.92 a	4.20 ± 0.39	3.02 ± 0.44 ab	0.72 ± 0.05 ab
F1	56.79 ± 0.25 a	61.21 ± 0.17 a	67.24 ± 0.38 a	4.42 ± 0.24	2.77 ± 0.49 ab	0.62 ± 0.08 ab
F2	59.25 ± 0.73 b	63.87 ± 0.65 b	70.20 ± 1.05 b	4.62 ± 0.26	3.34 ± 0.35 a	0.76 ± 0.10 b
F3	60.36 ± 0.04 b	64.63 ± 0.34 b	69.20 ± 0.67 b	4.27 ± 0.13	2.43 ± 0.37 b	0.57 ± 0.10 a

Values followed by different letters in each column indicate significant differences ($P \leq 0.05$).

To, gelatinization onset; Tp, peak temperature; Tc, conclusion temperature, Tp-To, gelatinization range, ΔH, enthalpy and PHI, peak high index

Table 5. Kinetic parameters of the starch hydrolysis of wheat flour samples (control) and jet milled flours (F1, F2, F3).

Sample	Free sugars (mg/100mg, db)			Resistant Starch (mg/100 mg, db)			Digestible starch (mg/100 mg, db)			C_{∞}	k
Control	0.54	±	0.01 a	6.85	±	3.42	58.94	±	5.86	321.51 ± 24.74 a	0.0002 ± 0.0000 a
F1	0.82	±	0.00 b	9.58	±	2.88	58.78	±	2.14	85.79 ± 1.40 b	0.0014 ± 0.0000 a
F2	0.94	±	0.00 b	9.75	±	2.88	57.85	±	0.39	33.97 ± 5.66 c	0.0046 ± 0.0011 b
F3	1.25	±	0.01 c	11.64	±	0.79	57.84	±	4.43	21.59 ± 1.41 c	0.0135 ± 0.0004 c

Values followed by different letters in each column indicate significant differences ($P \leq 0.05$)

Figures Captions

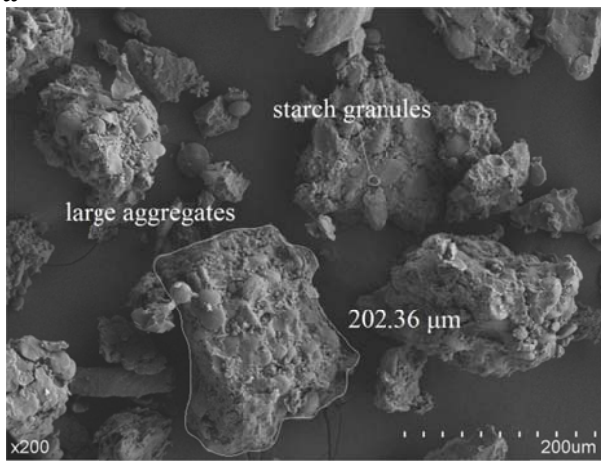
Fig. 1. Scanning electron micrograph of a) control sample of wheat flour and jet milled samples grinded under different milling conditions; b) F1, c) F2 and d) F3. Magnification 200x.

Fig 2. RVA profiles of control sample of wheat flour (▲) and jet milled flours, F1 (●), F2 (◆) and F3 (■) with (—) Temperature.

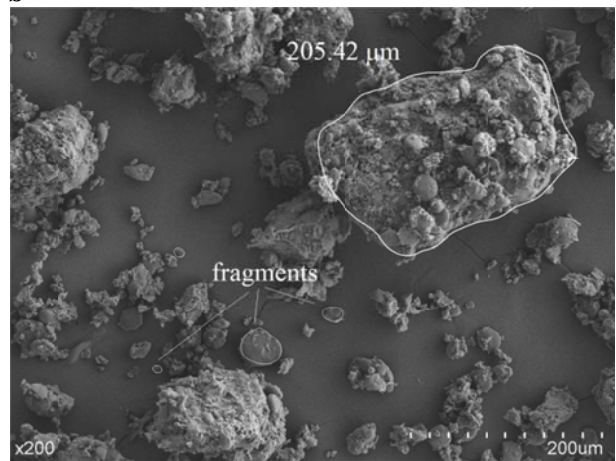
Fig 3. Effect of different jet milling conditions in the enzymatic starch hydrolysis kinetics of wheat flour; control (▲), F1 (●), F2 (◆) and F3 (■).

Fig.1.

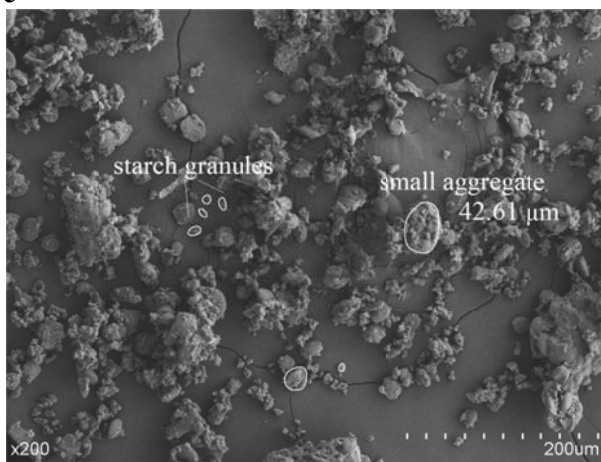
a



b



c



d

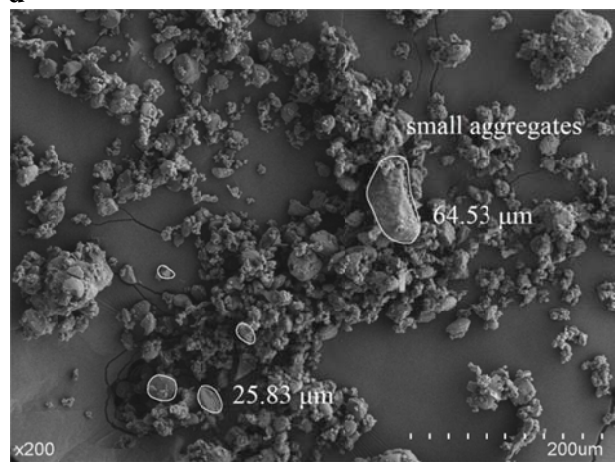


Fig.2

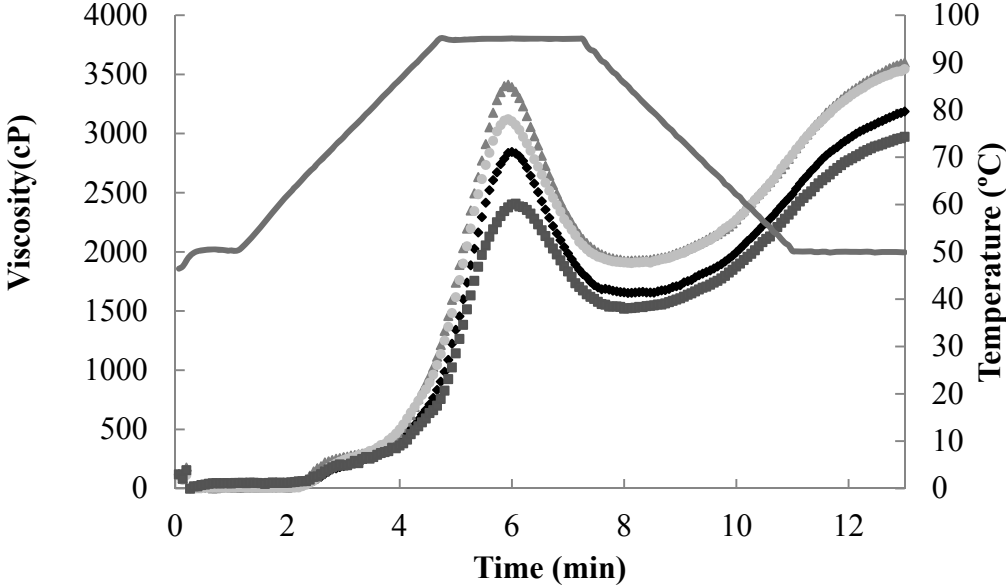


Fig 3.

