

1 **3D Printed Porous Media Columns with Fine Control of Column**
2 **Packing Geometry**

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15
16 **Abstract**

17
18 In this paper we demonstrate, for the first time, the use of 3D printing (also known as
19 additive manufacturing or rapid prototyping) to create porous media with precisely
20 defined packing geometries, directly from computer aided design (CAD) models. We
21 used CAD to design perfectly ordered beds with octahedral beads (115 μm apothem)
22 packed in a simple cubic configuration and monoliths with hexagonal channels (150
23 μm apothem) in parallel and herringbone arrangements. The models were then printed
24 by UV curing of acrylonitrile-butadiene-styrene powder layers. Each porous bed was
25 printed at 1.0, 1.5 and 2.0 mL volumes, within a complete column, including internal
26 flow distributors and threaded 10-32 flow connectors. Close replication of CAD
27 models was achieved. The resultant individual octahedral beads were highly uniform
28 in size, with apothems of $113.6 \pm 1.9 \mu\text{m}$, while the monolith hexagonal cross-section
29 channels had apothems of $148.2 \pm 2.0 \mu\text{m}$. Residence time distribution measurements
30 show that the beds largely behaved as expected from their design void volumes.
31 Radial and fractal flow distributor designs were also tested. The former displayed
32 poor flow distribution in parallel and herringbone pore columns, while the fractal
33 distributors provided uniform flow distribution over the entire cross section. The

34 results show that 3D printing is a feasible method for producing precisely controlled
35 porous media. We expect our approach to revolutionize not only fundamental studies
36 of flow in porous media but methods of chromatography column production.

37

38 **Keywords:** Porous media; 3D printing; Additive manufacturing; Packed bed; Packing
39 geometry; Residence time distribution

40

41

42 **Introduction**

43

44 Porous media are important for fluid-solid contacting in many unit operations,
45 including adsorption, chromatography, catalysis and filtration. Media particles are
46 typically packed into a column, allowing fluid to flow through the interstitial voids,
47 thus bringing the fluid into close contact with the solid phase. Key to the effectiveness
48 of packed columns are the flow-related properties of mass transfer, fluid distribution,
49 back pressure and fluid dispersion, which in turn depend upon packing geometry.

50

51 Packing geometry is determined primarily by particle shape, size and size distribution
52 and the packing method used. While there have been many theoretical studies on
53 optimal packing configurations and their effects on packing density, along with
54 computational studies on theoretical plate height and flow dispersion (e.g. [1]),
55 packed beds have, for practical reasons, invariably been randomly packed to date.
56 Thus, there has been no way before now to translate optimal ordered packing
57 arrangements into practice.

58

59 There have been many studies on flow through randomly packed beds, notably the
60 seminal works of Darcy [2], Kozeny [3], Carman [4] and Ergun [5]. These and other
61 authors have contributed much to our understanding of pressure drop and fluid
62 dispersion as functions of flow rate, particle shape, size and size distribution, largely
63 based upon empirical characterization. Experimental replication of models with
64 specific random or ordered packing geometries has been challenging. For random
65 geometries, no two randomly packed beds are exactly alike so we rely upon
66 generalized correlations and efficiency factors where, to quote Khirevich et al. [6]:
67 “column packing and consolidation are largely treated phenomenologically and

68 considered an art rather than a science”. On the other hand, it has been impracticable
69 to precisely reproduce ordered packing at the micron scale, mainly because there has
70 been no practical way to precisely locate individual particles within a bed. Even if
71 precise placement of the particles were feasible, e.g. through manual placement of
72 each bead, the column walls would almost certainly frustrate attempts to maintain
73 order.

74
75 Some authors (e.g. [7-10]) have characterized existing randomly packed beds through
76 tomography, thus reproducing, *a posteriori*, the geometry of their experimental
77 columns for computational analysis. However, they have had no control over the
78 initial packing of the experimental beds at the individual particle level so it has been a
79 case of accepting, rather than *a priori* designing, the fine structural detail.
80 Furthermore, because individual particles may change their positions with time, the
81 characterization of packing geometries is valid only as a snapshot in time.

82
83 Efforts to optimize the performance of packed beds for chromatography have focused
84 on the manufacture of bed particles (resin) and, because of ease of manufacture and
85 guaranteed bed permeability, these have been predominantly spherical [11]. Many
86 methods have been developed for producing spherical beads in bulk but they typically
87 result in wide particle size distributions, which are minimized in final media products
88 by fractionation, leading to increased costs, inefficient production and ultimately
89 variations in packing geometry through size variation in all but the most expensive of
90 media.

91
92 In this paper, we introduce an entirely new approach to packed column manufacture
93 that solves many of the above problems, using 3D printing (also known as “additive
94 manufacturing” or “rapid prototyping”) to produce packed beds that precisely
95 replicate computer aided design (CAD) models. 3D printing is a generic term for
96 techniques by which solid objects are created from digital models. The first working
97 3D printer was patented by Hull with a priority date of 1984 [12]. Since then, a
98 variety of 3D printing systems have been developed, including fused deposition,
99 selective heat or laser sintering, photopolymerization and thin-film lamination.
100 Several recent reviews of the development and advances in 3D printing are available
101 [13-17].

102

103 We use the term "packed" above advisedly because our technique produces what
104 might better be described as monoliths, although, as described below, they are distinct
105 from monoliths in their current sense in chromatography, which effectively exchange
106 random particle packing for random pore geometries [18, 19]. In contrast, with our
107 approach, we have created and tested exact physical replicas of ordered packed bed
108 CAD models, comprising ordered arrays of uniform particles. Our approach opens up
109 the possibility, for the first time, to precisely locate and orient every individual
110 particle within a porous bed. Here, we also demonstrate the production of monoliths
111 with precise internal pore geometries and, moreover, show that we can print not only
112 the porous bed but the entire column, complete with internal flow distributors,
113 packing, and external fluid connectors, therefore creating single-piece
114 chromatography columns.

115

116 **Materials and Methods**

117 Stereolithography (STL) files for the column models were created on Solidworks
118 2012 (Dassault Systèmes, Paris, France) and printed on a 3DS Projet HD 3500 printer
119 (3D Systems, Rock Hill, SC, USA). The printed components were made from non-
120 porous urethane acrylate oligomers (acrylonitrile butadiene styrene, ABS). A
121 proprietary paraffin wax was also used by the 3D printer during printing to support
122 overhanging features. The wax was removed from the internal structures of the
123 columns by alternating warm water (70°C) and 100% cyclohexane washes for up to 3
124 h.

125

126 The CAD models included the "packed" porous core and the ancillary column
127 elements, namely column walls, fluid distributors and collectors and end fittings for
128 easy connection to the experimental chromatography system. This enabled our
129 "packed" columns to be printed as an all-in-one parts, with no further assembly
130 required before use.

131

~~132~~

134 The porous beds were created with three geometries: beads in a simple cubic
135 arrangement (SC, Fig. 1a), a monolith containing parallel channels (PC, Fig. 1b) and
136 another containing herringbone shaped channels (HC, Fig. 1c).

137 The nominal resolution of the 3D printer was 28 μm , but the limiting dimensions of
138 the lattice elements that could be reliably printed at the desired resolution were about
139 one order of magnitude larger. Polyhedrons and polygons were used to design the
140 “packing” elements rather than spheres and circles, to minimize the file size of the
141 STL models while maintaining a regular shape in the lattice elements. For this reason,
142 octahedral beads (115 μm apothem) were used in the SC arrangement, while channels
143 with hexagonal cross-sections (150 μm apothem) were used for both the PC and HC
144 configurations. The HC geometry was designed with a tortuosity of 1.15, where the
145 tortuosity is defined as the ratio between the total length of the channels and the
146 column height.

147
148 In a standard SC configuration, only the outer diameters of the beads would contact,
149 creating a relatively weak structure prone to movement of the individual beads. The
150 octahedral beads were therefore designed to overlap at the edges (Fig. 1a), ensuring
151 the manufacture of physically robust prototypes with particle positions that do not
152 change with time. An overlap factor, defined as the ratio between the distance
153 between the centers of two adjacent polygons and the external bead face-to-face
154 diameter, was applied. Initial experiments indicated that an overlap value of 1.4
155 would yield a physically robust monolithic structure.

156
157 Columns with total bed volumes of 1, 1.5 and 2 ml were produced for each packed
158 bed geometry studied. The internal diameter and wall thickness of the cylindrical
159 columns were 16 and 2 mm, respectively. Connection with the chromatography
160 system was facilitated by including a 10-32 standard coned, female, fast protein liquid
161 chromatography (FPLC) finger-tight fitting at each end of the columns. All columns
162 contained a flow distributor at the porous bed entrance and an identical flow collector
163 at the outlet. Two geometric designs for the flow distributor and collector were used, a
164 standard radial distributor comprising a set of concentric and radial channels, and a
165 fractal flow distributor with square cross-section and 1024 nodes as proposed by
166 Tondeur and Luo [20]. The corresponding printed columns, therefore, were of circular
167 and square cross-sections, accordingly to the flow distributor considered. Figure 2
168 presents solid models of the flow channels within each distributor (collector) design,
169 which were then subtracted from the solid ends of the columns in the CAD model to

170 produce the flow channels. The characteristics of the 3D printed columns are
171 summarized in Table 1.

172

173 Residence time distribution (RTD) tests were carried out using an ÄKTA explorer
174 10TM FPLC system equipped with an auto-sampler (GE Healthcare, Uppsala,
175 Sweden). The columns were first equilibrated with pure water for 15 column volumes
176 (CV), followed by injection of 30 µl of 2 M NaCl. RTD experiments were carried out
177 at a flow rate of 10 ml/min, which corresponds to superficial velocities of 298 and 295
178 cm/h for the circular and square cross-section columns, respectively. The conductivity
179 peak in the column effluent was recorded and analyzed using the moment method.

180 The injected volume was 6% or less of the void volumes of the columns tested, hence
181 the contribution to the first moment arising from the injection loop can be neglected.

182 Under this assumption, the experimental residence time, t_r^{exp} , can be calculated as:

$$183 \quad t_r^{\text{exp}} = \frac{M_1}{M_0} = \frac{\int_0^{\infty} c(t)t dt}{\int_0^{\infty} c(t) dt} \quad (1)$$

184 where M_i is the i^{th} absolute moment, c is the concentration of the tracer, and t is time.
185 E curves, i.e. normalized elution profiles having unitary area, were calculated from
186 the conductivity signal and the 0th moment:

$$187 \quad E(\theta) = \frac{c(\theta)}{M_0} \quad (2)$$

188 where θ is the dimensionless time defined in terms of the theoretical residence time,
189 t_r^{theo} , estimated from the designed geometry of the lattice structure:

$$190 \quad \theta = \frac{t}{t_r^{\text{theo}}} \quad (3)$$

191 This expression can be also used to define an experimental dimensionless residence
192 time, θ_r^{exp} :

$$193 \quad \theta_r^{\text{exp}} = \frac{t_r^{\text{exp}}}{t_r^{\text{theo}}} \quad (4)$$

194 Comparison of the theoretical and experimental residence times was used to assess the
195 quality of the printed lattices and the uniformity of the flow distribution.

196

197 **Results and Discussion**

198

199 Creation of STL models represents the first step in the production of the 3D printed
200 porous columns. Definition of the building elements of the lattice is crucial, in part
201 because this is the attribute that has the most influence on file size and subsequent file
202 handling. Contribution to the final file size arising from column walls, end fittings,
203 and distributor/collector can be neglected. In the initial design, spherical beads and
204 circular channels were considered but the STL file sizes were too large, from both the
205 viewpoints of the speed of rendering during CAD modeling and of the printer file
206 handling capacity. For example, to accurately model a single sphere, irrespective of
207 diameter, our CAD package used approximately 6162 triangles, with a file size of 306
208 kbytes, while an octahedron was described by only 8 triangles, giving a file size of
209 less than 0.5 kbytes, three orders of magnitude smaller. However, file size *per se* is
210 not fundamentally a limiting factor for 3D printing, and could be handled with
211 efficient computational algorithms or compression, particularly with the constantly
212 growing capacity of microprocessors, communications and storage media with time.
213 Furthermore, ordered packing geometries are based on repetitive structures that would
214 lend themselves to iterative printer command sequences. Thus, there is no
215 fundamental reason why spherical elements could not be used, given sufficient
216 software and hardware processing power.

217

218 It is well understood that in 2D image processing, the resolution of a picture is
219 proportional to the number of pixels of which it is comprised. Similarly, the quality of
220 the rendering of a solid shape is proportional to the number of 3D dots used to
221 discretize it. The resolution of a 3D printer is an indication of the size of the smallest
222 feature that is possible to print, i.e. it corresponds to the dimensions of the “3D dots”
223 that make up the printed model. However, the final size and shape of the 3D dots are
224 ultimately determined by a number of uncontrollable variables. In our case, using
225 layer deposition followed by UV curing, examples of these uncontrollable variables
226 are: i) the interfacial forces acting between the ABS polymer and the support material,
227 ii) the local temperature of the ABS polymer during UV curing, iii) venting
228 characteristics of the printing chamber, iv) defects and irregularities during layering of
229 the ABS powder.

230

231 Also, the resolution quoted by a 3D printer manufacturer may well comprise the best
232 possible that can be achieved under ideal conditions but this may not be routinely
233 achievable in normal practice. The 3D printer used in this work had a nominal
234 resolution of 28 μm so the printed octahedral particles or hexagonal cross-section
235 channels were characterized by relatively rounded edges at the micron scale.
236 However, as is shown in the following discussion, the features of the CAD models
237 were conserved in the 3D printed objects, hence microscopic limitations in the
238 resolution do not represent a significant limitation of 3D printed porous media.

239

240 In addition to full operational columns, cross-sectional “cutaway” models of each
241 packing configuration were printed to display the internal structures of the columns,
242 distributors and porous beds. Figure 3 shows that not only the column macrostructures
243 but also the microstructures of the CAD models were reproduced with reasonable
244 fidelity by the printer.

245

246 Inspection of Fig. 3 reveals that the mean bead and channel apothems were $113.6 \pm$
247 3.8 and 148.2 ± 2.0 μm , respectively, while the design values were 115 and 150 μm ,
248 respectively, demonstrating the precise control over packed bed microstructures
249 delivered by our 3D printing approach. Figure 3c shows a magnification of the simple
250 cubic cutaway model, showing that the particles were approximately octahedral and
251 the dimensions of the pores and relative diameters of the beads were consistent with
252 the design compression factor $\alpha = 1.4$. Likewise, magnified images of the straight and
253 herringbone channels (Fig. 3f and 3l) show reasonable fidelity between the CAD
254 models and the printed columns in the cutaway models, revealing that the 3D printer
255 used was able to reproduce the CAD models well. It is reasonable to expect the same
256 fidelity was obtained between the CAD models and the full operational printed
257 columns.

258

259 Residence time distribution (RTD) tests were conducted on all 3D printed columns,
260 first, to highlight differences between the “packing” geometries used and, second, to
261 compare the effectiveness of the two distributor designs.

262

263 Normalized residence time distribution profiles of the printed columns are shown in
264 Fig. 4. We were concerned that the printer might not faithfully reproduce octahedral
265 beads but rather create roughly spherical beads because of limitations in resolution.
266 However, evidence of the high fidelity between the CAD model and the 3D printed
267 columns can be found in the RTD experiments by comparing theoretical and
268 experimental porosity values. At $\alpha = 1.4$, the designed extra-particle porosity of the
269 simple cubic octahedral beads is $\varepsilon = 0.575$. For comparison, a simple cubic
270 configuration of spherical beads with the same overlap would have a theoretical
271 porosity of $\varepsilon = 0.041$, while it would be $\varepsilon = 0.476$ with no overlap. The
272 experimentally determined porosities of $\varepsilon = 0.678$, 0.569 and 0.551 for 1, 1.5 and 2 ml
273 columns are closer to the design porosity for octahedral beads (17.9%, 1.0% and 4.2%
274 differences, respectively) rather than that for spherical particles (minimum difference
275 34.4%), suggesting good control over particle shape at the 3D printer's limiting
276 resolution. The mean normalized residence times for simple cubic bead columns also
277 indicate reasonable consistency between the design (expected) and experimental
278 column porosities.

279

280 The low mean residence times in the straight and herringbone channel cylindrical
281 columns (Fig. 4a) compared with the cubic packing suggest that a substantial
282 proportion of the channels in those columns were not accessed by the fluid when
283 using radial flow distributors. This is a strong indication of the low efficiency of the
284 radial distributor, which was not able to spread the incoming flow uniformly over the
285 entire cross section. The radial distributor primarily conveys the flow through the
286 central channel, while there is no reason for flow to move radially in the distributor
287 unless there is an axial flow resistance in the bed. In the case of cubic packing, the
288 interconnected network of beads allows for both radial and axial dispersion, so it is
289 the packing itself that assists in the uniform distribution of the flow across the cross-
290 section, giving an experimental dimensionless residence time, θ_r^{exp} , close to unity
291 (Fig. 4 and Table 2). In contrast, because radial dispersion is absent throughout the
292 parallel and herringbone channel columns, the fluid would have followed only the
293 channels into which it initially entered. It is likely, therefore that the performance of
294 these latter columns was limited by inadequate radial flow distribution at entry,
295 resulting in $\theta_r^{exp} < 1.0$ in the RTDs shown in Figure 4a.

296

297 In an attempt to improve uniform flow distribution over the entire cross-section, a
298 fractal distributor was designed. For both the radial (circular column cross-section)
299 and fractal (square column cross-section) designs, we printed shortened columns
300 containing no bed but with the inlet flow distributor and outlet flow collector placed
301 immediately adjacent to one another and compared their residence time distributions.
302 This approach may also be useful to measure extra-bed dispersion but in this case we
303 simply compared the average residence times of the two distributor designs. The
304 mean residence times for the radial and fractal distributors were $\theta_r^{\text{exp}} = 0.32$ and 0.93 ,
305 respectively, indicating that flow was not well distributed in the radial flow
306 distributor, while it was relatively uniformly distributed in the fractal design.

307

308 Figure 5 compares the normalized RTD curves in 2 ml PC (Fig. 5a) and SC (Fig. 5b)
309 columns containing the two distributor designs. Note that there is a significant
310 difference between the mean elution times for the two distributor designs in the PC
311 column, in which there was no radial dispersion within the bed itself, while in the
312 simple cubic bead column, where the bed itself provides radial distribution, there was
313 little difference between the RTDs for the two distributor designs. Thus, for bed pore
314 geometries that do not promote radial flow dispersion, careful design of the fluid
315 distributor is required.

316

317 Note also, in Fig. 5b, that the fractal distributor system for the SC column resulted in
318 a longer tailing in the RTD curve. This possibly occurs because the flow rate in the
319 corners of the square cross-section column with the fractal distributor may be slower
320 for the SC packing than the mean flow rates in the rest of the bed, leading to greater
321 axial dispersion than that in the cylindrical cross-section column. These differences in
322 tailing are not seen between the two PC columns (Fig. 5a) because the axial
323 dispersion is affected only by the flow through the independent channels, which have
324 uniform geometry, regardless of the overall column cross-sectional geometry.

325

326 Pressure-flow measurements of the printed columns were found to be
327 indistinguishable from control measurements using just the FPLC system in by-pass
328 mode (i.e. with no column attached, data not shown). This result is consistent with

329 expectations because short columns were used, resulting in low column
330 backpressures. No external structural defects nor leaks were observed at superficial
331 velocities of up to 594 cm.hr⁻¹ (corresponding to 20 ml.min⁻¹), demonstrating the
332 structural robustness of the all-in-one-piece 3D printed columns.

333

334 Our choice of materials for this work was constrained to those that were readily
335 available for rapid prototyping, so we used a non-porous material and focused on
336 demonstrating that we could achieve control over packing geometry, at least within
337 the resolution of the particular printer used here. Clearly, an ideal chromatographic
338 media would comprise finer-resolution elements to minimize the theoretical plate
339 height, a functionalizable surface chemistry to enable ligand attachment for reversible
340 adsorption, and porous materials to maximize adsorption capacity. We have not
341 demonstrated these ideal characteristics in this paper but see no fundamental reasons
342 why all of these ideal characteristics could not be achieved. We also limited the
343 volumes of our columns to those that were convenient to handle in the laboratory.
344 However, even with the printer used here, there is no particular reason why we could
345 not have produced columns with significantly greater dimensions, at the same fine
346 resolution but with a diameter and length of 30 cm or more i.e. preparative scale. The
347 materials used here were low-cost and in general, the use of materials in 3D printing
348 is very efficient, using only the amount necessary to produce the specific features of
349 the CAD model. Thus, we believe our approach is scalable and will enable not only
350 fundamental studies of flow, mass transfer and adsorption through structured porous
351 media but perhaps, in time, commercial column production.

352

353 Our approach can be applied not only to chromatography but to any application
354 requiring fluid-solid contacting, including filtration and catalytic or other reaction
355 applications. One could create precise replicates of randomly packed beds, to enable
356 experimental validation of computational models. Furthermore, the ability to orientate
357 the individual particles means we can go beyond using spheres and conceive beds
358 comprising unusual and complex particle shapes, while maintaining uniform porosity
359 throughout. There is no particular need for all elements within the bed to be uniform
360 with regard to size or shape and one could imagine porous media with a wide range of
361 controlled geometry elements throughout the bed could be designed and printed.
362 There are currently printers on the market capable of printing multiple materials at

363 once (rather like a color inkjet printer) so the various column and bed components
364 could, in principle, be printed using different materials, each suited to its particular
365 purpose e.g. porous, functional bed materials with non-porous, inert column walls,
366 flow connectors and distributors.

367

368 Thus, there is enormous potential for using additive manufacturing to produce
369 versatile monolithic porous media with designed geometries not only for the beds
370 themselves but for the entire columns.

371

372 **Conclusions**

373

374 We have shown, for the first time, that 3D printing can be used to precisely replicate
375 the fine structure of CAD models of porous media, comprising both ordered particle
376 packing and monoliths with internal channels. Residence time distributions measured
377 in the printed columns were consistent with predicted porosities and designed
378 geometric structures, indicating that the CAD features were reproduced with good
379 fidelity at the scales attempted here.

380

381 Furthermore, we have demonstrated that not only the porous beds themselves but
382 entire columns can be printed as single physical artifacts, meaning that flow
383 connectors, flow distributors and internal column packing can be printed within a
384 single, complete column. Fractal flow distributors are capable of distributing the flow
385 across the entire column cross-section, even when there is no radial dispersion across
386 the flow channels of monolithic beds. This distributor design enabled the creation of
387 square cross-section monolithic columns with good flow distribution and residence
388 time distributions through parallel channels that were independent of the column
389 cross-sectional shape.

390

391 3D printing frees us from the constraints of previous manufacturing methods and
392 enables the creation of porous media characterized by a combination of fine precision,
393 scalability and versatility, at low cost. We expect this approach to column design will
394 revolutionize the production of packed bed columns and monoliths across a wide
395 range of applications, not only in chromatography but also filtration, catalysis,
396 adsorption and other applications where intimate fluid-solid contact is desired.

397

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431

432 **Figure Legends**

433

434

435 **Figure 1.** The three bed geometric designs: a) simple cubic beads, b) straight

436 channels, c) herringbone channels

437

438 **Figure 2.** Illustration of the flow distributor templates: a) radial flow distributor, b)

439 fractal flow distributor (Note: for clarity, only the first 64 of the 1024 nodes are

440 shown)

441

442 **Figure 3.** CAD designs versus printed cutaway columns a) SC CAD model b) SC

443 printed model c) 20X magnification of SC beads d) PC CAD model e) PC printed

444 model f) 20X magnification of parallel channels g) HC CAD model h) HC printed

445 model i) 20X magnification of herringbone channels

446

447 **Figure 4.** Residence time distribution profiles of a) the three packing geometries in

448 1.5 ml cylindrical columns, b) SC bead columns at three different column volumes.

449

450 **Figure 5.** Comparison of 2 ml columns with radial and fractal flow distributors for a)

451 PC columns b) SC columns

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Table 1. Specifications of 3D Columns

Column Cross-Section	Flow Distributor	Column Volumes (ml)	Packing Configuration	Theoretical Porosity
Circular	Radial	1.0, 1.5, 2.0	SC	0.575
Circular	Radial	1.0, 1.5, 2.0	PC	0.334
Circular	Radial	1.0, 1.5, 2.0	HC	0.334
Square	Fractal	2.0	SC	0.575
Square	Fractal	2.0	PC	0.393

Table 2. Theoretical and experimental mean residence times of SC columns

Column Volume (ml)	t_r^{theo} (min)	t_r^{exp} (min)	θ_r^{exp} (-)
1.0	0.705	0.804	1.14
1.5	0.993	0.993	1.00
2.0	1.280	1.229	0.96

Figure 1

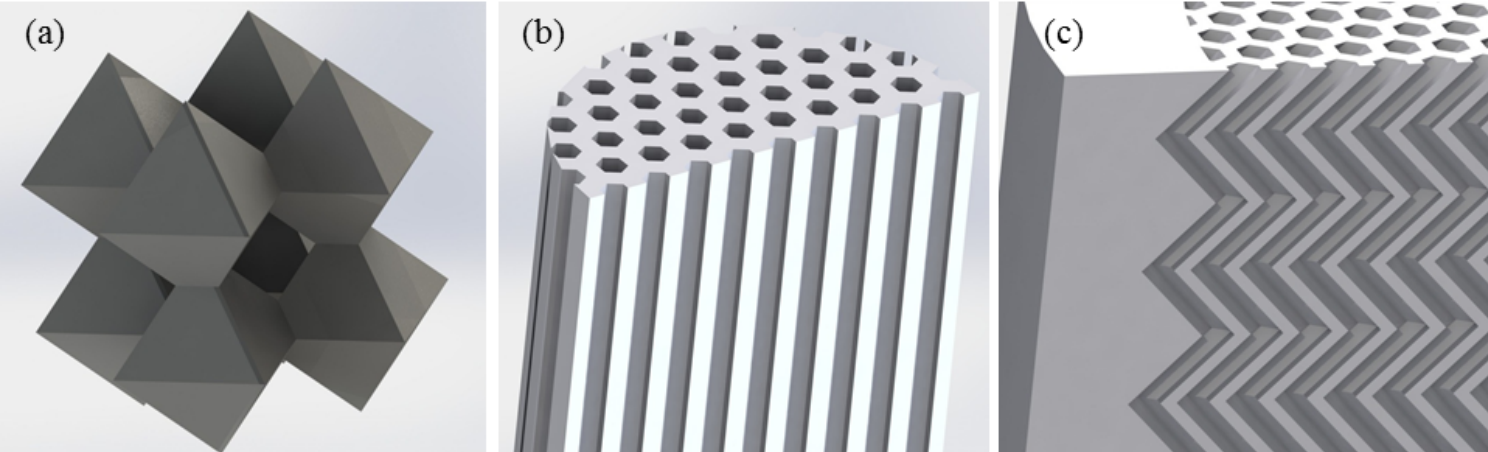


Figure 2

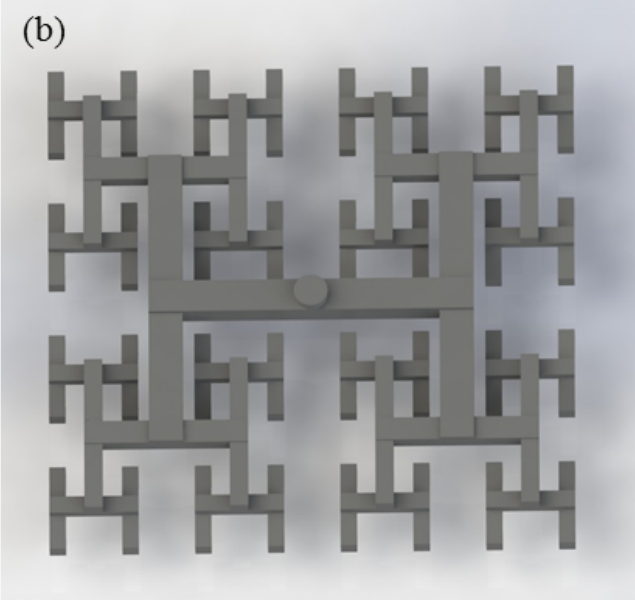
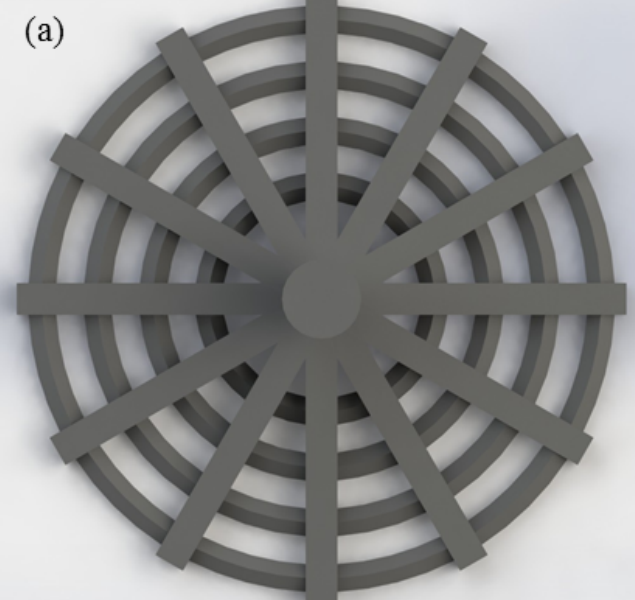


Figure 3

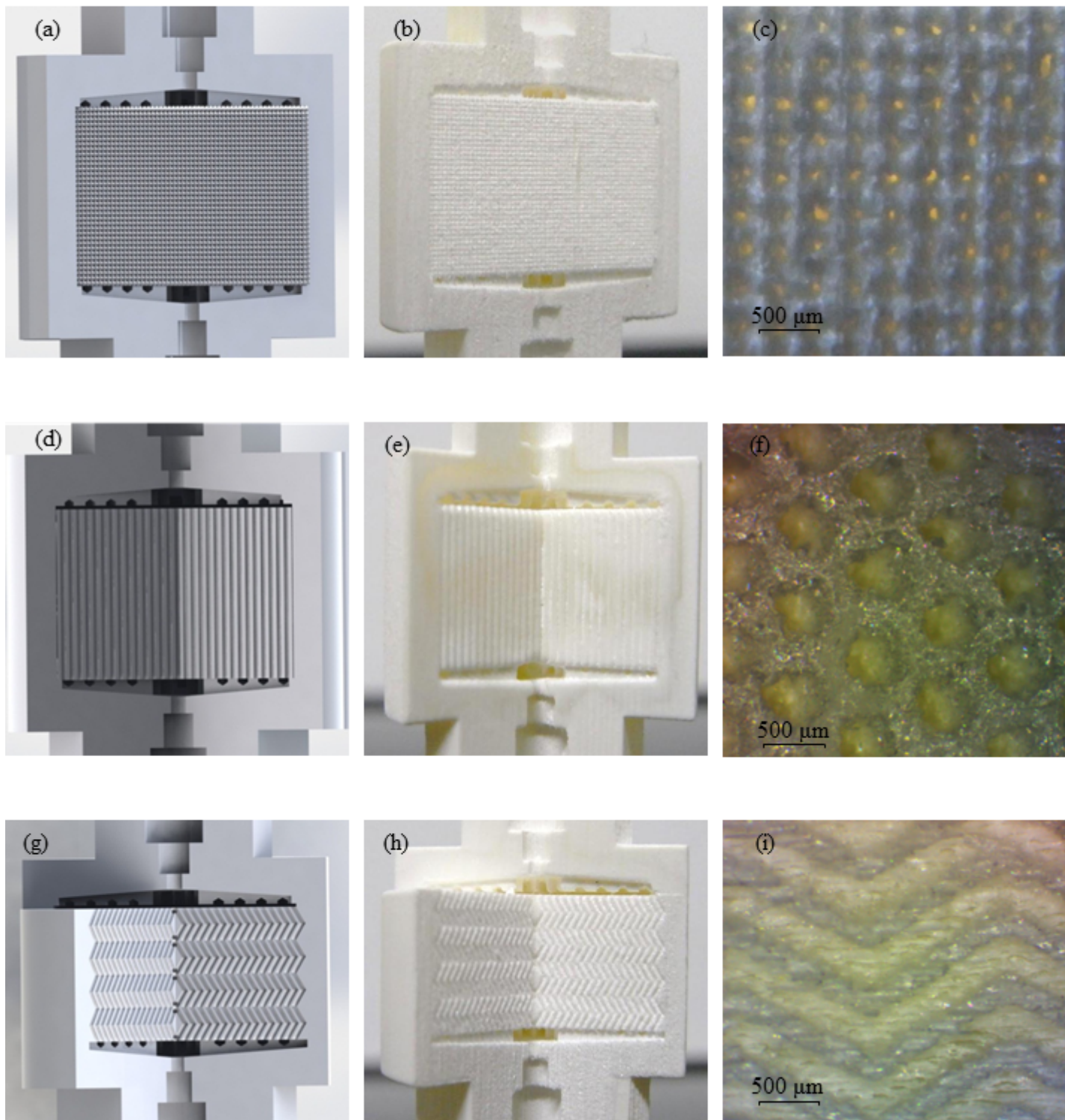


Figure 4

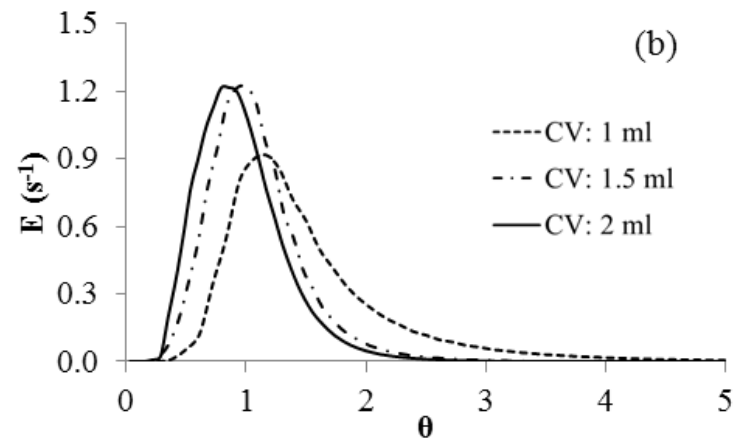
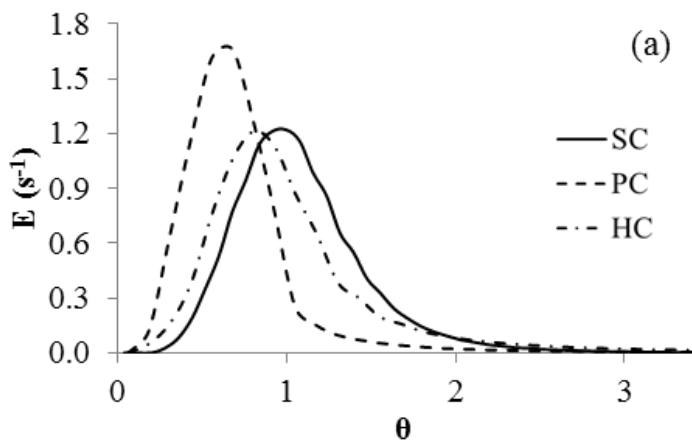


Figure 5

