

## SALINE STRESS EFFECT ON COWPEA BEANS GROWTH UNDER BIOFERTILIZER CORRECTION

### EFEITO DO ESTRESSE SALINO NO CRESCIMENTO DO FEIJÃO-CAUPI SOB A CORREÇÃO DE BIOFERTILIZANTE

Ednângelo Duarte PEREIRA<sup>1</sup>; Albanise Barbosa MARINHO<sup>2</sup>; Elísia Gomes RAMOS<sup>2</sup>; Christlene Nojosa Dias FERNANDES<sup>3</sup>; Francisca Robevania Medeiros BORGES<sup>2</sup>; Jilson de Nazaré José ADRIANO<sup>2</sup>

1. Mestrando, Departamento de Fitotecnia, Universidade Federal de Viçosa, Campus Universitário, Viçosa, MG, Brasil, ednangeloduarte@gmail.com; 2. Instituto de Desenvolvimento Rural/Universidade da Integração Internacional da Lusofonia Afro-Brasileira, Redenção, CE, Brasil; 3. Doutoranda, Departamento de Engenharia Agrícola, Universidade Federal do Ceará, Campus do Pici, Fortaleza, CE, Brasil.

**ABSTRACT:** New strategies to increase growth and food crops productivity in saline soils are priorities in the research. The experiment was carried out in a pot to investigate the growth of cowpea cv. Canapu in response to saline stress under the correction of a mixed biofertilizer. Five irrigation water salinity levels and three doses of the mixed biofertilizer were tested, with four replicates at each level. The increase in salinity level resulted in reductions in stem diameter, leaves number, shoot biomass and roots. The mixed biofertilizer use was able to minimize the effect of salinity in all analyzed variables, improving the cowpea growth and development, protecting from saline stress negative impacts. In addition, the mixed biofertilizer use may be an option to improve crop growth and productivity in salt affected soils.

**KEYWORDS:** Environmental stress. Water quality. *Vigna unguiculata*.L. Walp.

#### INTRODUCTION

*Vigna unguiculata* L. Walp is one of the main economically planted crops, being a cereal used as food for people and animals, with high nutritional value (QADOS, 2011).

It's considered moderately tolerant to salinity and may be cultivated in arid and semi-arid regions, such as the Brazilian northeast, which is limiting to other species. In these regions, most crops are grown under irrigation and inadequate irrigation management leads to secondary salinization, affecting 20% of irrigated land worldwide (GLICK et al., 2007; SHRIVASTAVA; KUMAR, 2015).

Salinity may affect plants in various ways such as drought stress, ionic toxicity, nutritional disorders, oxidative stress, altered metabolic processes, membrane disorganization, and reduced cell division and expansion (ZHU, 2007; SIDARI et al., 2008; MUSCOLO et al., 2013).

A saline soil is defined when the electrical conductivity (EC) of the saturation extract (CEe) in the root zone exceeds 4 dS m<sup>-1</sup> (approximately 40 mM NaCl) at 25 °C and exchangeable sodium of 15%, reducing yield of most of the plants grown in this CEe (MUNNS, 2005; JAMIL et al., 2011).

About 20 percent of total cultivated and 33 percent of irrigated farmland worldwide is affected

by high salinity (SHAHBAZ; ASHRAF, 2013). Salinized areas increase at a rate of 10% a year due to low rainfall, high surface evaporation, rocks weathering, saltwater irrigation and poor cultural practices, which can result in more than 50% salinized arable land by 2050 (JAMIL et al., 2011).

A biofertilizer is any substance or microorganism applied to plants with the aim of improve nutritional efficiency, tolerance to abiotic stress and/or quality characteristics of the crop, regardless of its nutrient content (DU JARDIM, 2015). This definition represents the clearest and most concise way to define biofertilizers. Currently, there are several biofertilizers, both industrial produced and those produced from waste. The latter are prepared with animal, vegetable and agroindustrial residues applied via soil, via irrigation systems or spread on the plants (YAKHIN et al., 2017).

The relevance of the liquid biofertilizers use as management practice in the form of simple or enriched microbial ferments is in the quantitative, diversity and in the nutrients availability by the biological activity, since these substances increase the efficiency of the nutrients use, reduce fertilizer consumption, stimulate plant development and growth (CALVO et al., 2014; HALPERN et al., 2015) and counterbalance stress factors, eventually

increasing crop quality and productivity (VAN OOSTEN et al., 2017).

The biofertilizer, when applied via soil in liquid form, improves the water infiltration rate and releases humic substances in the soil, increasing the plants osmotic adjustment by the accumulation of these substances, facilitating the water and nutrients absorption in saline environments (AYDIN et al., 2012; SOUTO et al., 2013).

Limited freshwater, growing food needs, the climate change pressure, and the land available reduction for cultivation are threats to agricultural sustainability (SHAHBAZ; ASHRAF, 2013). Practices for managing the saline water resources use may be an important resource for water scarcity (ZAHIR et al., 2008).

The aim of this study was to evaluate the salinity effect on the *V. unguiculata* vegetative growth, with and without mixed biofertilizer applications in saline soils.

## MATERIAL AND METHODS

The work was carried out at the Piroás Experimental Farm, belonging to the Universidade da Integração Internacional da Lusofonia Afro-Brasileira (UNILAB), Piroás/ Redenção – Ceará, Brazil (04°15'55"S, 38°79'37"W; 240 m). The region climate is hot, dry semi-arid, with low cloudiness, high evaporation pattern and strong insolation (KÖPPEN, 1928). Rainfall is concentrated between January and April with average precipitation between 380 and 760 mm, distributed irregularly during the year (CELENTANO et al., 2017).

*V. unguiculata* cv. Canapu was cultivated in open-air in pots of 39.5 L in single rows with spacing of 0.5 x 0.5 m and planting density of 28,570 plants ha<sup>-1</sup>. The vessels were filled with a 0.05 m layer of gravel and a substrate composed of sand and local soil in a 2:1 ratio.

The mean of irrigation water salinity was 0.4 dS m<sup>-1</sup>, being considered the control treatment. Other concentrations were obtained by the addition of the NaCl, CaCl<sub>2</sub>.2H<sub>2</sub>O and MgCl<sub>2</sub>.6H<sub>2</sub>O salts, in the ratio 7:2:1 (RHOADES et al., 2000). Irrigation was started 15 days after emergence (DAE) and maintained until the end of the cycle by drip irrigation. The irrigation shift was of two days, whose water amount to be applied was calculated according to the evaporation of the class "A" tank.

The experimental design was in randomized blocks in the subdivided plots scheme, with four blocks. The plots were composed of water with five levels of salinity, 0.4; 1.4, 2.4, 3.4 and 4.4 dS m<sup>-1</sup>, respectively, without leaching fraction and the

subplots consisted of three doses of mixed biofertilizer equivalent to 0, 400 and 800 mL plant<sup>-1</sup>.

Mixed biofertilizer application was manually through graduated containers and divided twice a week, starting at the same irrigation application period. The biofertilizer was prepared with 100 L of fresh bovine manure, 30 L of chicken manure and 5 L of wood ash, diluted in 270 L of fresh water. The biofertilizer decomposition time was 30 days and monitored daily with EC, pH and temperature measurements. Cultural dealings followed the recommendations for the culture.

The biofertilizer after 30 days of decomposition had the following characteristics: N = 1.20; P = 0.38; K = 0.03; Ca<sup>2+</sup> = 2.82; Mg<sup>2+</sup> = 0.63 and S = 0.01 g L<sup>-1</sup>. Na = 611; Fe = 91.54; Mn = 10, 56; Zn = 6.4 and Cu = 3.06 mg L<sup>-1</sup>.

The stem diameter (DS) and leaves number (NL) were determined every 7 days between 15 and 57 DAE. The stem diameter was measured with digital caliper at the base of the stem 2 cm from the soil surface. The leaves number was quantified through weekly direct counting. The harvest occurred between 59 and 103 DAE and, at the cycle end, the fresh and dry mass of the underground and aboveground parts were evaluated.

Fresh mass of aerial part (FMAP) and root (FRM) were determined with precision scale (model S1002 - BEL). Dry mass of aerial part (DMAP) and root (DRM) were obtained by drying in full sun until constant weight.

Statistical analysis, for all observed variables, was performed by analysis of variance (F test, 5% significance). When significant, regression analysis was performed. In the regression analysis, the equations that best fit the data were chosen based on the significance of the regression coefficients at the significance level of 5% (\*) by the F test, and the highest coefficient of determination (R<sup>2</sup>). The software used was ASSISTAT - Statistical Assistance, version 7.7 beta (2016).

## RESULTS AND DISCUSSION

The treatments effects on stem diameter (SD) and leaf number (NL) of cowpea beans as a evaluation times function, salinity levels and mixed biofertilizer doses indicated a significant effect ( $p \leq 0.05$ ) on the two analyzed variables, with exception of the leaves number that was not significantly affected by the biofertilizer doses. The interaction between the factors was significant ( $p \leq 0.05$ ) among all the treatments applied, except for the leaves number where the interaction times and

mixed biofertilizer doses were not significant (Table 1).

The treatments effect on SD and NL of common bean as a evaluation times function,

salinity levels and mixed biofertilizer doses suggests that plant growth was variable according to season, salinity level and mixed biofertilizer doses as reported for culture (SOUZA et al., 2014).

**Table 1.** Summary of variance analysis for stem diameter (SD) and leaves number (NL) of cowpea plants submitted to irrigation with different salinized water levels and mixed biofertilizer doses

| Source of Variation   | df  | Mean squares |                        |
|-----------------------|-----|--------------|------------------------|
|                       |     | SD           | NL                     |
| Blocks                | 3   | 2.60210*     | 43.11609*              |
| Epoch (a)             | 6   | 80.82617*    | 908.62057*             |
| Error - a             | 18  | 0.13123      | 8.44498                |
| Salinity (b)          | 4   | 4.91688*     | 57.80803*              |
| Interaction a x b     | 24  | 1.35045*     | 39.96648*              |
| Error - b             | 84  | 0.34759      | 6.20613                |
| Biofertilizer (c)     | 2   | 8.36000*     | 11.19299 <sup>ns</sup> |
| Interaction a x c     | 12  | 0.74877*     | 5.90504 <sup>ns</sup>  |
| Interaction b x c     | 8   | 1.21299*     | 18.76311*              |
| Interaction a x b x c | 48  | 0.15352*     | 5.12509 <sup>ns</sup>  |
| Error - c             | 210 | 0.41337      | 6.55610                |
| Total                 | 419 | -            | -                      |

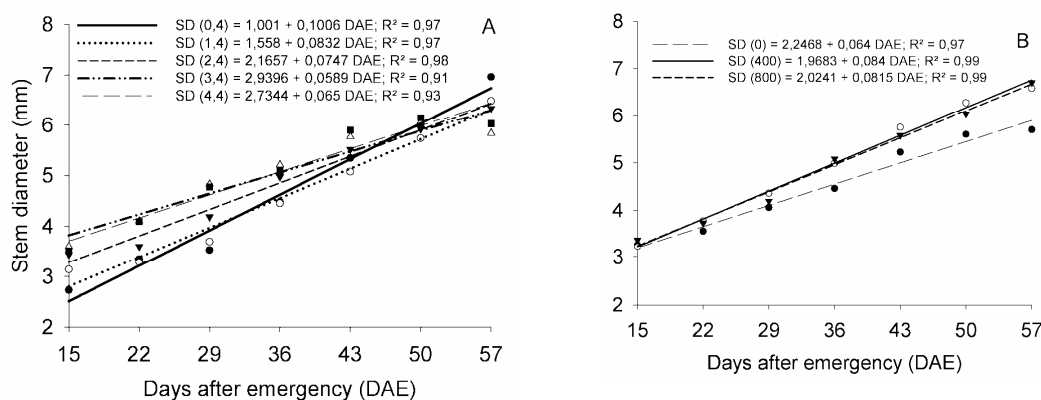
\* significant at 5% by the F test; (ns) not significant by the F test; df - Degree of freedom.

The stem diameter as a function of days after emergence, for each salinity and mixed biofertilizer level doses were adjusted to the increasing linear model. The largest stem diameter (6.74 mm) occurred at 57 DAE at 0.4 dS m<sup>-1</sup> and decreased with increasing salinity (Figure 1A-B).

The largest stem diameter at 0.4 dS m<sup>-1</sup> is due to the potentially toxic ions (Na<sup>+</sup> and Cl<sup>-</sup>) low concentration in water and foliar tissues, without affecting the photosynthetic system and ensuring the CO<sub>2</sub> diffusion into the cells and consequently not harming the plant growth (LACERDA et al., 2003). The *V. unguiculata* reduction stem diameter occurred when irrigated with water at 1.55 dS m<sup>-1</sup> (PRAZERES et al., 2015). *V. unguiculata*,

'Quarentinha' cultivar, had a decline in stem diameter under saline stress at 35 days after sowing (ANDRADE et al., 2013).

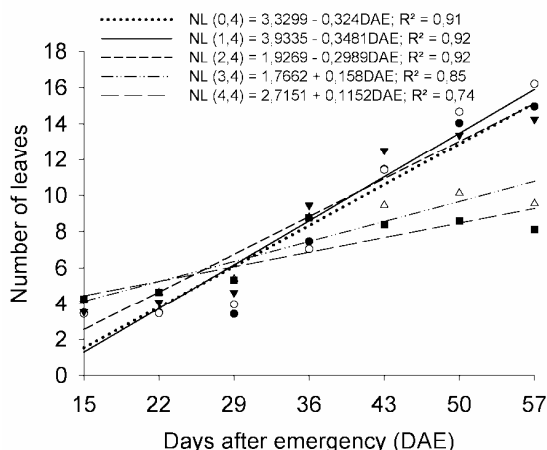
The highest stem diameters obtained with 400 and 800 mL plant<sup>-1</sup> week<sup>-1</sup> mixed biofertilizer doses are due to the nutritional availability of this fertilizer for the crop. The microbial and bioactive activity present in this input has the capacity to provide protection and resistance to the plant (GROVER et al., 2011). The stem growth increase provided by biofertilizers in saline environments was also observed in *Arachis hypogaea* L. and *Passiflora edulis f. flavicarpa* (SOUSA et al., 2012, OLIVEIRA et al., 2015).



**Figure 1.** Regression analysis for stem diameter (mm) of cowpea plants submitted to irrigation with different levels of salinized water (A) and biofertilizer doses (B).

The leaves number reduced with increasing of saline water irrigation. The maximum

leaves number was 15.91 with 1.4 dS m<sup>-1</sup> (Table 1, Figure 2).



**Figure 2.** Number of cowpea leaves submitted to irrigation with different levels of salinized water.

The leaves number decrease with increasing irrigation water salinity probably occurred due to the sodium chloride accumulation in the cell walls and in the older leaves cytoplasm. At the same time, the vacuole sap can not accumulate more salt, reducing the concentration inside the cells, leading to rapid death and leaf fall (MUNNS, 2002). The leaf numbers reduction by saline stress impairs plant growth and development due to lower leaf area and net photosynthesis rate (GOMES et al., 2011; NUNES et al., 2012). The leaves number decreased in *V. unguiculata* plants irrigated with salt water, in soil with common bovine biofertilizer and enriched crab biofertilizer (SOUSA et al., 2014). The biofertilizer excess reduce plant biomass by toxicity could be the direct consequence of the chlorophyll synthesis inhibition and photosynthesis (KONG et al., 2017). Excessive salt and nitrogen amount from biofertilizer may cause decreased uptake of nutrient elements, inhibition of various enzyme activities,

induction of oxidative stress including alterations in enzymes of the antioxidant defense system and leaves fall (WINGLER; ROITSCH, 2008; ZIMMERMANN; ZENTGRAF, 2015). In addition to genetic control, senescence is regulated by joined actions of external (e.g., N availability, light) and internal (e.g., regulating metabolites, C/N ratio) signals (BAO et al., 2015; KONG et al., 2017) Sugar regulation of leaf senescence is also dependent on plant N status and soil N content. Therefore, excessive N and the desynchronization of N application with crop demand will accelerate leaves fall.

The FMAP, DMAP, FRM and DRM were significantly influenced by irrigation water salinity and mixed biofertilizer, except for DRM, which was significantly influenced by the mixed biofertilizer. There was no interaction between the factors (Table 2).

**Table 2.** Summary of variance analysis for FMAP, DMAP, FMR, and DRM of cowpea plants submitted to irrigation with different levels of saline water and mixed biofertilizer doses

| Source of variation | GL | Mean squares             |                       |                       |                       |
|---------------------|----|--------------------------|-----------------------|-----------------------|-----------------------|
|                     |    | FMAP                     | DMAP                  | FRM                   | DRM                   |
| Blocks              | 3  | 1276.90796 <sup>ns</sup> | 9.86095 <sup>ns</sup> | 9.04229 <sup>ns</sup> | 0.99187 <sup>ns</sup> |
| Salinity (a)        | 4  | 4518.95910*              | 77.29303*             | 10.12492*             | 0.53663 <sup>ns</sup> |
| Error-a             | 12 | 389.67810                | 15.08145              | 2.98314               | 0.44052               |
| Biofertilizer (b)   | 2  | 3867.97993*              | 80.71499*             | 26.41949*             | 1.28499*              |
| Interaction - a x b | 8  | 361.12130 <sup>ns</sup>  | 9.16537 <sup>ns</sup> | 4.49979 <sup>ns</sup> | 0.08366 <sup>ns</sup> |
| Error-b             | 30 | 441.26835                | 9.91480               | 4.02946               | 0.15394               |
| Total               | 59 | -                        | -                     | -                     | -                     |

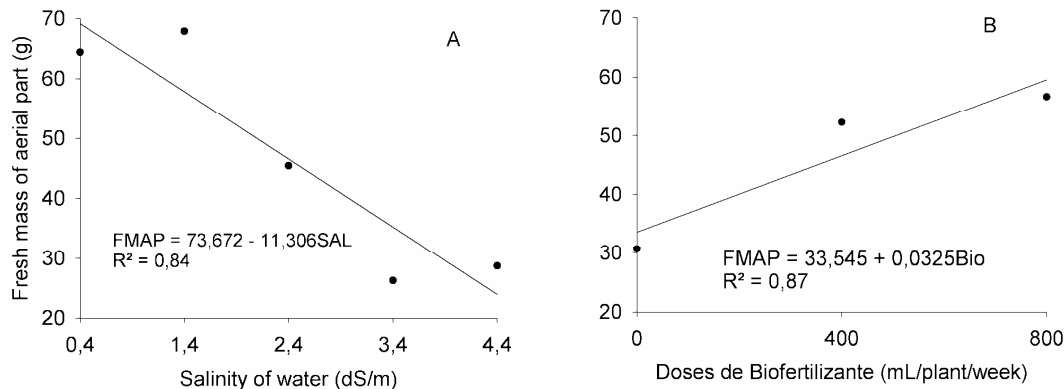
\* significant at 5% by the F test; (ns) not significant by the F test; GL - Degree of freedom.

FMAP decreased linearly with increasing saline concentration of irrigation water (Figure 3A). The highest value of FMAP (69.15 g) was obtained in the control (0.4 dS m<sup>-1</sup>) and the lowest value

(23.93 g) in the highest salinity level was 4.4 dS m<sup>-1</sup>. The FMAP increased linearly with the mixed biofertilizer doses (R<sup>2</sup> 0.87), with higher value

(59.55 g) obtained for the dose of 800 mL plant<sup>-1</sup>

week<sup>-1</sup> (Figure 3B).

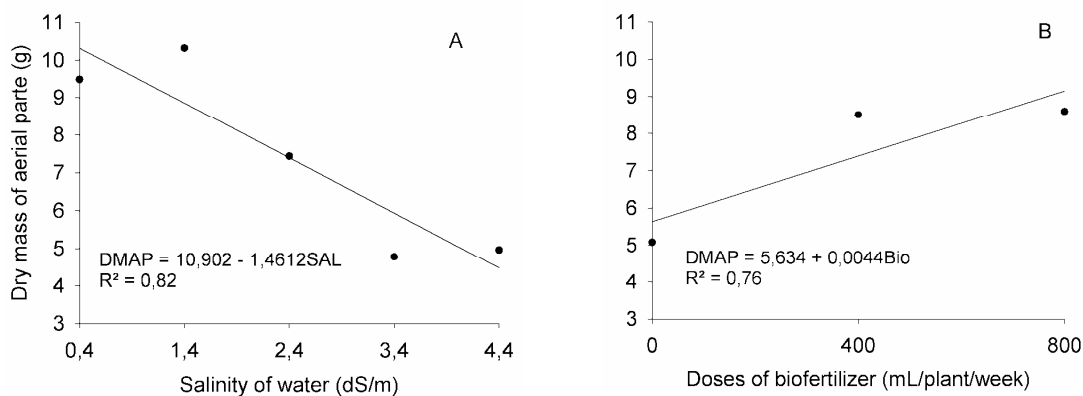


**Figure 3.** Fresh mass of aerial part (g) of cowpea plants submitted to irrigation with esulto f levels of salinized water (A) and mixed doses of biofertilizer (B).

The decrease in *V. unguiculata* plant growth is an energy metabolic cost reflection associated with an attempt to adapt the plant to salinity (DANTAS et al., 2002; MUNNS, 2002). The common bean lowest growth of an adaptive mechanism to the saline condition, so the plant maintains its vital activities, even to a limited extent (COELHO et al., 2013). The biofertilizer positive effect on the aerial part may be associated with the

root growth stimulation by the phytohormones production, which allows the water roots uptake, improving plant nutrition and increasing nutrient availability (JOSKO et al., 2013).

DMAP decreased linearly with increasing irrigation water salinity ( $R^2 = 0.82$ ) and increased linearly as a result of the increase in mixed biofertilizer doses ( $R^2 = 0.76$ ) (Figure 4A-B).



**Figure 4.** Dry mass of aerial part (g) of cowpea plants submitted to irrigation with different levels of saline water (A) and mixed doses of biofertilizer (B).

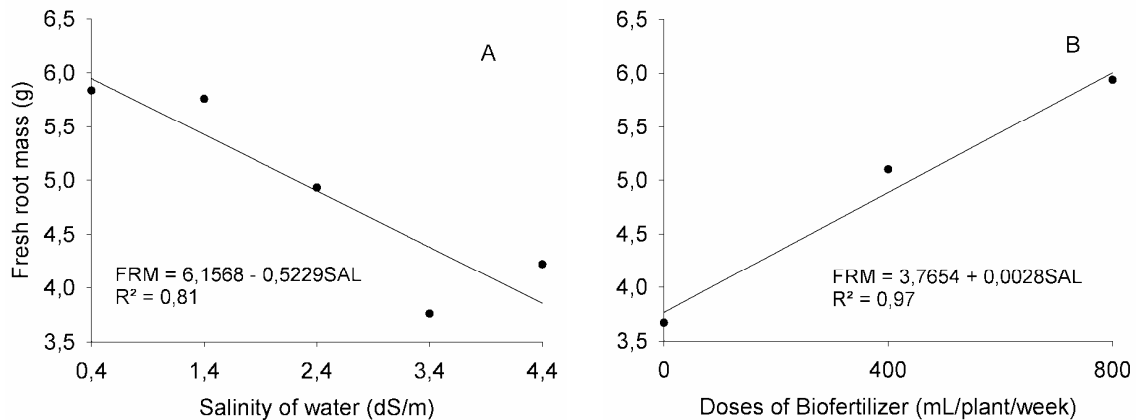
The saline concentration increase in the soil solution causes a decrease in dry mass, which consequently increases the osmotic pressure, decreasing the absorption and the water supply, which are one of the main factors for a dry matter or biomass production (MACHADO; SERRALHEIRO, 2017). The reduction in dry matter may be due to the stomatal closure caused by salinity that limits CO<sub>2</sub> assimilation and inhibits leaf expansion, accelerating the mature leaves senescence, reducing the area destined to the photosynthetic process and the photoassimilates

production (LACERDA et al., 2003). The dry mass increase of the aerial part with the use of mixed biofertilizer prove the attenuation effect of this compound in *V. unguiculata* plants under salt stress due to slow nutrients release of the biofertilizer (FACHINI et al., 2004), besides promoting the soil aggregation and porosity, retention and water infiltration in the soil (RODRIGUES et al., 2013). The dry aerial part biomass of vigna bean cultivated in two soils classes (French-sandy and loamy clay) artificially salinized reduced 64.5% and 60.7% (COELHO et al., 2013), similar to the data found in

this research. The cultivar Quarentinha of *V. unguiculata* also had its dry biomass aerial part reduced by 66.94% with the increase of soil salinity irrigated with water at 5.0 dS m<sup>-1</sup> (LIMA et al., 2007), similar to the data found in this research.

The fresh root mass decreased with increasing salinity. The maximum fresh root mass

value was 5.95 g for the salinity level 0.4 dS m<sup>-1</sup> (Figure 5A). The fresh root mass increased linearly with the mixed biofertilizer doses, reaching a maximum value of 6.01 g with the dose of 800 ml plant<sup>-1</sup> week<sup>-1</sup> (Figure 5B).

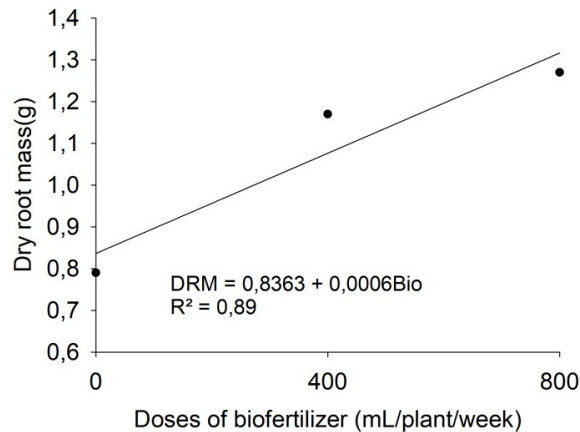


**Figure 5.** Fresh root mass (g) of cowpea plants submitted to irrigation with different levels of saline water (A) and mixed doses of biofertilizer (B).

The fresh root mass decrease by saline stress is associated with the ions accumulation that concentrate in the soil (SHRIVASTAVA; KUMAR, 2015). The soil salt concentration increase by irrigation reduced root growth and consequently the weight of this variable in *V. unguiculata* plants (SILVA et al., 2011). Reduction in root growth and elongation in *Chenopodium quinoa* was caused by high salt levels at various development stages, reducing the supply of photosynthates from the shoot and morphological changes in the root, which should not be considered as a simple growth reduction, but rather as an induced growth reorientation to avoid stress (PANUCCIO et al., 2014).

The fresh mass increase of the roots with the mixed biofertilizer application of demonstrates the input effect attenuation and may be considered an important management strategy for *V. unguiculata* plants cultivated in saline environment. The application of biofertilizer in the soil can induce an increase in the osmotic adjustment to the plants by the organic solutes accumulation, promoting the water and nutrients absorption under saline stress, improving the conditions for emergence, vegetative growth and biomass production of plants (BAALOUSHA et al., 2006).

Dry mass of the root increased linearly with the mixed biofertilizer application, with a maximum value of 1.32 g in the dose 800 mL plant<sup>-1</sup> week<sup>-1</sup> (Figure 6).



**Figure 6.** Dry mass of the root (g) of cowpea plants submitted to biofertilizer different doses.

The increased linearly of root dry mass with the mixed biofertilizer application on *V. unguiculata* irrigated with saline water levels, is related to the humic substances increase of, improving soil quality, providing a source of nutrients and increasing cation exchange capacity and nutrient availability (JINDO et al., 2012). Biofertilizers improve soil structure and water retention (OLMO et al., 2016), while improving biological properties favoring plant-microorganism interactions, increasing water and nutrient uptake by plants (HAMMER et al., 2014). Although it was not included in the scope of the present study, a larger nodules number of nitrogen-fixing bacteria and with a higher finer roots number were observed in the plants root system that received the mixed biofertilizer.

## CONCLUSIONS

*V. unguiculata* cv. Canapu proved to be very sensitive to saline stress, its growth and development being affected from 1.4 ds.m<sup>-1</sup>.

The use of mixed biofertilizer was able to minimize the effect of salinity in all analyzed variables, improving the growth and development of cowpea cv. Canapu, protecting from negative impacts of saline stress. The incorporation of mixed biofertilizer may be a promising strategy to improve crop growth and productivity in salt affected soils.

## ACKNOWLEDGMENTS

To Conselho Nacional de Desenvolvimento Científico e Tecnológico (CNPq) and the Fundação Cearense de Apoio ao Desenvolvimento Científico e Tecnológico (FUNCAP) for financial resources.

**RESUMO:** Novas estratégias para aumentar o crescimento e a produtividade de plantas cultivadas em solos salinos representam prioridades na pesquisa. O experimento foi conduzido em vaso, para investigar o crescimento do Feijão-Caupi em resposta ao estresse salino sob a correção de um biofertilizante misto. Cinco níveis de salinidade da água de irrigação e três doses do biofertilizante misto foram testados, com quatro repetições cada nível. O biofertilizante misto foi capaz de minimizar o estresse salino. O aumento do nível de salinidade resultou em reduções no diâmetro do caule, número de folhas, biomassa da parte aérea e das raízes. O uso do biofertilizante misto minimizou o efeito da salinidade em todas as variáveis analisadas, melhorando o crescimento e desenvolvimento do feijão caupi, protegendo-o dos impactos negativos do estresse salino. O uso de biofertilizante misto pode ser uma opção para melhorar o crescimento da cultura e a produtividade em solos afetados por sais.

**PALAVRAS-CHAVE:** Estresse ambiental. Qualidade da água. *Vigna unguiculata* L. Walp.

## REFERENCES

- ANDRADE, J. R.; MAIA JUNIOR, S. O.; SILVA, P. F.; BARBOSA, J. W. S.; NASCIMENTO, R.; SOUSA, J. S. Crescimento inicial de genótipos de feijão caupi submetidos a diferentes níveis de água salina. **Agropecuária Científica no Semiárido**, Campina Grande, v. 9, n. 4, p. 38-43, 2013. <http://dx.doi.org/10.30969/acsa.v9i4.430>
- AYDIN, A.; KANT, C.; TURAN, M. Humic acid application alleviate salinity stress of bean (*Phaseolus vulgaris* L.) plants decreasing membrane leakage. **African Journal of Agricultural Research**, Nairobi, v. 7, n. 7, p. 1073-1086, 2012. <https://doi.org/10.5897/AJAR10.274>
- BAALOUSHA, M.; HEINO, M. M.; LE COUSTOMER, B. K. Conformation and size of humic substances: effects of major cation concentration and type, pH, salinity, and residence time. **Colloids and surfaces A: physicochemical and engineering aspects**, Amsterdam, v. 272, n. 1, p. 48-55, 2006. <https://doi.org/10.1016/j.colsurfa.2005.07.010>
- BAO, A.; ZHAO, Z.; DING, G.; SHI, L.; XU, F.; CAI, H. The stable level of glutamine synthetase 2 plays an important role in rice growth and in carbon-nitrogen metabolic balance. **International Journal of Molecular Sciences**, Basel, v. 16, n. 6, p. 12713-12736, 2015. <https://doi.org/10.3390/ijms160612713>
- CALVO, P.; NELSON, L.; KLOEPPER, J. W. Agricultural uses of plant biostimulants. **Plant and Soil**, Dordrecht, v. 383, n. 1-2, p. 3-41, 2014. <https://doi.org/10.1007/s11104-014-2131-8>
- CELENTANO, A.; BORGES, F. R. M.; MARINHO, A. B.; BEZERRA, F. M. L.; RODRIGUES, J. P. M.; PEREIRA, E. D. Parâmetros produtivos do girassol submetido à lâminas de irrigação na região do maciço de Baturité – CE. **Revista Brasileira de Agricultura Irrigada**, Fortaleza, v. 11, n. 1, p. 1213-1222, 2017. <http://dx.doi.org/10.7127/rbai.v11n100570>
- COELHO, J. B. M.; BARROS, M. F. C.; BEZERRA NETO, E.; CORREA, M. M. Comportamento hídrico e crescimento do feijão vigna cultivado em solos salinizados. **Revista Brasileira de Engenharia Agrícola e Ambiental**, Campina Grande, v. 17, n. 4, p. 379-385, 2013. <http://dx.doi.org/10.1590/S1415-43662013000400004>
- DANTAS, J. P.; MARINHO, F. J. L.; FERREIRA, M. M. M.; AMORIM, M. S. N.; ANDRADE, S. I. O.; SALES, A. E. Avaliação de genótipos de feijão-de-corda sob salinidade. **Revista Brasileira de Engenharia Agrícola e Ambiental**, Campina Grande, v. 6, n. 3, p. 425-430, 2002. <https://doi.org/10.1590/S1415-43662002000300008>
- DU JARDIN, P. Plant biostimulants: definition, concept, main categories and regulation. **Scientia horticulturae**, Amsterdam, v. 196, p. 3-14, 2015. <https://doi.org/10.1016/j.scienta.2015.09.021>
- FACHINI, E.; GALBIATTI, J. A.; PAVANI, L. C. Níveis de irrigação e de compostos de lixo orgânico na formação de mudas cítricas em casa de vegetação. **Engenharia Agrícola**, Jaboticabal, v. 24, n. 3, p. 578-588, 2004. <http://dx.doi.org/10.1590/S0100-69162004000300010>
- GLICK B. R.; CHENG Z.; CZARNY J.; DUAN J. Promotion of plant growth by ACC deaminase-producing soil bacteria. **European Journal of Plant Pathology**, Dordrecht, v. 119, n. 3, p. 329-339, 2007. <https://doi.org/10.1007/s10658-007-9162-4>
- GOMES, K. R.; AMORIM, A. V.; FERREIRA, F. J.; FILHO, F. L.; LACERDA, C. F.; GOMES FILHO, E. Respostas de crescimento e fisiologia do milho submetido a estresse salino com diferentes espaçamentos de cultivo. **Revista Brasileira de Engenharia Agrícola e Ambiental**, Campina Grande, v. 15, n. 4, p. 365-370, 2011. <https://doi.org/10.1590/S1415-43662011000400006>



- GROVER, M.; ALI S. K. Z.; SANDHYA, V.; RASUL, A.; VENKATESWARLU, B. Role of microorganisms in adaptation of agriculture crops to abiotic stresses. **World Journal of Microbiology and Biotechnology**, Dordrecht, v. 27, n. 5, p. 1231-1240, 2011. <https://doi.org/10.1007/s11274-010-0572-7>
- HALPERN, M.; BAR-TAL, A.; OFEK, M.; MINZ, D.; MULLER, T.; YERMIYAHU, U. The use of biostimulants for enhancing nutrient uptake. D.L. Sparks (Ed.). **Advances in Agronomy**, San Diego, v. 129, p. 141-174, 2015. <https://doi.org/10.1016/bs.agron.2014.10.001>
- HAMMER, E. C.; BALOGH-BRUNSTAD, Z.; JAKOBSEN, I.; OLSSON, P. A.; STIPP, S. S.; RILLIG, M. C. A mycorrhizal fungus grows on biochar and captures phosphorus from its surfaces. **Soil Biology and Biochemistry**, Oxford, v. 77, p. 252-260, 2014. <https://doi.org/10.1016/j.soilbio.2014.06.012>
- JAMIL, A.; RIAZ, S.; ASHRAF, M.; FOOLAD, M. R. Gene expression profiling of plants under salt stress. **Critical Reviews Plant Sciences**, London, v. 30, n. 5, p. 435-458, 2011. <https://doi.org/10.1080/07352689.2011.605739>
- JINDO, K.; MARTIM, A. S.; NAVARRO, E. C.; PÉREZ-ALFOCEA, F.; HERNANDEZ, T.; GARCIA, C.; AGUIAR, N. O.; CANELLAS, L. P. Root growth promotion by humic acids from composted and non-composted urban organic wastes. **Plant and Soil**, Dordrecht, v. 353, n. 1-2, p. 209-220, 2012. <http://dx.doi.org/10.1007/s11104-011-1024-3>
- JOSKO, I.; OLESZCZUK, P.; PRANAGAL, J.; LEHMANN, J.; XING, B.; CORNELISSEN, G. Effect of biochars, activated carbon and multiwalled carbon nanotubes on phytotoxicity of sediment contaminated by inorganic and organic pollutants. **Ecological Engineering**, Oxford, v. 60, p. 50-59, 2013. <https://doi.org/10.1016/j.ecoleng.2013.07.064>
- KONG, L.; XIE, Y.; HU, L.; SI, J.; WANG, Z. Excessive nitrogen application dampens antioxidant capacity and grain filling in wheat as revealed by metabolic and physiological analyses. **Scientific Reports**, Oxford, v. 7, n. 43363, p. 1-14, 2017. <https://doi.org/10.1038/srep43363>
- KÖPPEN, W.; GEIGER, R. **Klimate der Erde**. Gotha: Verlag Justus Perthes. 1928.
- LACERDA, C. F.; CAMBRAIA, J.; CANO, M. A. O.; RUIZ, H. A.; PRISCO, J. T. Solute accumulation and distribution during shoot and leaf development in two sorghum genotypes under salt stress. **Environmental and Experimental Botany**, Paris, v. 49, n. 2, p. 107-120, 2003. [https://doi.org/10.1016/S0098-8472\(02\)00064-3](https://doi.org/10.1016/S0098-8472(02)00064-3)
- LIMA, C. J. G. S.; OLIVEIRA, F. A.; MEDEIROS, J. F.; OLIVEIRA, M. K. T.; ALMEIDA JÚNIOR, A. B. Resposta do feijão caupi a salinidade da água de Irrigação. **Revista Verde de Agroecologia e Desenvolvimento Sustentável**, Mossoró, v. 2, n. 2, p. 79-86, 2007.
- MACHADO, R. M. A.; SERRALHEIRO, R. P. Soil Salinity: Effect on Vegetable Crop Growth. Management Practices to Prevent and Mitigate Soil Salinization. **Horticulturae**, Basel, v. 3 n. 30, p. 1-13, 2017. <https://doi.org/10.3390/horticulturae3020030>
- MUNNS R. Tansley review: genes and salt tolerance: bringing them together. **New Phytologist**, Cambridge, v. 167, p. 645-663, 2005. <http://dx.doi.org/10.1111/j.1469-8137.2005.01487.x>
- MUNNS, R. Comparative physiology of salt and water stress. **Plant, Cell and Environment**, New York, v. 25, n. 2, p. 239-250, 2002. <https://doi.org/10.1046/j.0016-8025.2001.00808.x>
- MUSCOLO, A.; PANUCCIO, M. R.; HESHEL, A. Ecophysiology of *Pennisetum clandestinum*: a valuable salt tolerant grass. **Environmental and Experimental Botany**, England, v. 92, p. 55-63, 2013. <https://doi.org/10.1016/j.envexpbot.2012.07.009>

NUNES, J. C.; CAVALCANTE, L. F.; LIMA NETO, A. J.; REBEQUI, A. M.; DINIZ, B. L. M. T.; GHEYI, H. R. Comportamento de mudas de nim à salinidade da água em solo não salino com biofertilizante. **Revista Brasileira de Engenharia Agrícola e Ambiental**, Campina Grande, v. 16, n. 11, p. 1152-1158, 2012. <http://dx.doi.org/10.1590/S1415-43662012001100002>

OLIVEIRA, F. A.; LOPES, M. Â. C.; SILVA SÁ, F. V.; NOBRE, R. G.; MOREIRA, R. C. L.; DE ANDRADE SILVA, L.; PAIVA, E. P. Interação salinidade da água de irrigação e substratos na produção de mudas de maracujazeiro amarelo. **Comunicata Scientiae**, Bom Jesus, v. 6, n. 4, p. 471-478, 2015. <http://dx.doi.org/10.14295/cs.v6i4.982>

OLMO, M.; VILLAR, R.; SALAZAR, P.; ALBURQUERQUE, J. A. Changes in soil nutrient availability explain biochar's impact on wheat root development. **Plant and soil**, Dordrecht, v. 399, n.1-2, p. 333-343, 2016. <http://dx.doi.org/10.1007/s11104-015-2700-5>

PANUCCIO, M. R.; JACOBSEN, S. E.; AKHTAR, S. S.; MUSCOLO, A. Effect of saline water on seed germination and early seedling growth of the halophyte quinoa. **AoB Plants**, Oxford, v. 6[plu047]. <https://doi.org/10.1093/aobpla/plu047>

PRAZERES, S. S.; LACERDA, C. F.; BARBOSA, F. E. L.; AMORIM, A.V.; ARAUJO, I. C. S.; CAVALCANTE, L. F. Crescimento e trocas gasosas de plantas de feijão-caupi sob irrigação salina e doses de potássio. **Revista Agro@mbiente On-line**, Boa Vista, v. 9, n. 2, p. 111-118, 2015. <http://dx.doi.org/10.18227/1982-8470ragro.v9i2.2161>

QADOS, A. M. S. A. Effect of salt stress on plant growth and metabolism of bean plant *Vicia faba* (L.). **Journal of the Saudi Society of Agricultural Sciences**, Saudi Arabia, v. 10, n. 1, p. 7-15, 2011. <https://doi.org/10.1016/j.jssas.2010.06.002>

RHOADES, J. D.; KANDIAH, A.; MASHALI, A. M. Uso de águas salinas para produção agrícola. (**Estudos da FAO - Irrigação e Drenagem, 48, Revisado**), Campina Grande: UFPB. 2000. 117 p.

RODRIGUES, J. F.; REIS, J. M. R.; REIS, M. A. Utilização de esterco em substituição a adubação mineral na cultura do rabanete. **Revista Trópica: Ciências Agrárias e Biológicas**, Chapadinha, v. 7, n. 2, p.160-168, 2013.

SHAHBAZ, M.; ASHRAF, M. Improving salinity tolerance in cereals. **Critical Reviews in Plant Sciences**, Boca Raton, v. 32, n. 4, p. 237-249, 2013. <http://dx.doi.org/10.1080/07352689.2013.758544>

SHRIVASTAVA, P.; KUMAR, R. Soil salinity: a serious environmental issue and plant growth promoting bacteria as one of the tools for its alleviation. **Saudi Journal of Biological Sciences**, Saudi Arabia, v. 22, n. 2, p. 123-131, 2015. <https://doi.org/10.1016/j.sjbs.2014.12.001>

SIDARI, M.; SANTONOCETO, C.; ANASTASI, U.; PREITI, G.; MUSCOLO, A. Variations in four genotypes of lentil under NaCl-salinity stress. **American Journal of Agricultural and Biological Science**, Dubai, v. 3, n.1, p. 410-416, 2008. <http://dx.doi.org/10.3844/ajabssp.2008.410.416>

SILVA, F. L. B.; LACERDA, C. F.; SOUSA, G. G.; NEVES, A. L. R.; SILVA, G. L.; SOUSA, C. H. C. Interação entre salinidade e biofertilizante bovino na cultura do feijão-de-corda. **Revista Brasileira de Engenharia Agrícola e Ambiental**, Campina Grande, v. 15, n. 4, p. 383-389, 2011. <https://doi.org/10.1590/S1415-43662011000400009>

SOUSA, G. G.; AZEVEDO, B. M.; MESQUITA, J. B. R.; VIANA, T. V. A. Características agrônômicas do amendoineiro sob irrigação com águas salinas em solo com biofertilizantes. **Revista Agro@mbiente On-line**, Boa Vista, v. 6, n. 2, p. 124-132, 2012. <http://dx.doi.org/10.18227/1982-8470ragro.v6i2.708>

SOUSA, G. G.; VIANA, T. V. A.; LACERDA, C. F.; AZEVEDO, B. M.; SILVA, G. L.; COSTA, F. R. B. Estresse salino em plantas de feijão-caupi em solo com fertilizantes orgânicos. **Revista Agro@ambiente Online**, Boa Vista, v. 8, n. 3, p. 359-367, 2014. <http://dx.doi.org/10.18227/1982-8470ragro.v8i3.1824>

SOUTO, A. G. L.; CAVALCANTE, L. F.; NASCIMENTO, J. A. M.; MESQUITA, F. O.; LIMA NETO, A. J. Comportamento do noni à salinidade da água de irrigação em solo com biofertilizante bovino. **Irriga**, Botucatu, v. 18, n. 3, p. 442-453, 2013. <http://dx.doi.org/10.15809/irriga.2013v18n3p442>

VAN OOSTEN, M. J.; PEPE, O.; DE PASCALE, S.; SILLETTI, S.; MAGGIO, A. The role of biostimulants and bioeffectors as alleviators of abiotic stress in crop plants. **Chemical and Biological Technologies in Agriculture**, Portici, v. 4, n. 5, p. 1-12, 2017. <https://doi.org/10.1186/s40538-017-0089-5>.

WINGLER, A.; ROITSCH, T. Metabolic regulation of leaf senescence: interactions of sugar signalling with biotic and abiotic stress response. **Plant Biology**, Amsterdam, v. 10, n. 1, p. 50-62, 2008. <https://doi.org/10.1111/j.1438-8677.2008.00086.x>

YAKHIN, O.; LUBYANOV, A. A.; YAKHIN, I. A.; BROWN, P. Biostimulants in plant science: a global perspective. **Frontiers in Plant Science**, Rockville Pike, v. 7, n. 2049, p. 1-32, 2017. <https://doi.org/10.3389/fpls.2016.02049>

ZAHIR, Z. A.; MUNIR, A.; ASGHAR, H. N.; ARSHAD, M.; SHAHAROONA, B. Effectiveness of rhizobacteria containing ACC-deaminase for growth promotion of peas (*Pisum sativum*) under drought conditions. **Journal of Microbiology and Biotechnology**, Dordrecht, v. 18, n. 5, p. 958-963, 2008.

ZHU, J. K. Plant salt stress. In: O'Daly A, editor. **Encyclopedia of life sciences**. Chichester: John Wiley & Sons, p.1-3, 2007. <http://dx.doi.org/10.1002/9780470015902.a0001300.pub2>.

ZIMMERMANN, P.; ZENTGRAF, U. The correlation between oxidative stress and leaf senescence during plant development. **Cellular & Molecular Biology Letters**, Wrocław, v. 10, n. 3, p. 515-534, 2005.