

Levitsky A. P., Mudrik L. M., Khromagina L. N., Sevostianova T. A., Selivanskaia I. A. Lipopolysaccharide increases lysozyme adhesion to chitin. *Journal of Education, Health and Sport*. 2019;9(11):240-247. eISSN 2391-8306. DOI <http://dx.doi.org/10.5281/zenodo.3558921>
<https://apcz.umk.pl/czasopisma/index.php/JEHS/article/view/JEHS.2019.09.11.021>

The journal has had 5 points in Ministry of Science and Higher Education parametric evaluation. § 8. 2) and § 12. 1. 2) 22.02.2019.

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The authors declare that there is no conflict of interests regarding the publication of this paper.

Received: 06.11.2019. Revised: 14.11.2019. Accepted: 29.11.2019.

UDC 616.153:577.152:616.633:612.31

LIPOPOLYSACCHARIDE INCREASES LYSOZYME ADHESION TO CHITIN

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Abstract

Background. To study the effect of lipopolysaccharide on the adsorption of lysozyme on chitin.

Methods. Egg white lysozyme, lipopolysaccharide and chitin were used. The lyophilized cells of *M. lyzodeikticus* were used as a lysozyme substrate. Lysozyme activity was determined by the bacteriolytic method using different concentrations (8-512 µg/ml). Lysozyme adsorption on chitin was determined by the loss of lysozyme from solution after shaking with chitin. The effect of lipopolysaccharide (0.6-40 µg/ml) on lysozyme adsorption was evaluated by the loss of lysozyme from the solution after shaking with 50 mg of chitin.

Results. In determining the activity of lysozyme by the bacteriolytic method, the linear nature of the dependence of activity on the concentration of the enzyme is observed only at low concentrations (8-100 µg / ml). Lipopolysaccharide does not affect the bacteriolytic

activity of lysozyme, however, dose-dependently increases the adsorption of lysozyme on chitin.

Conclusion. Lipopolysaccharide increases the adsorption of lysozyme on chitin.

Keywords: lysozyme, lipopolysaccharide; chitin; bacteriolysis; adsorption of lysozyme on chitin.

INTRODUCTION

Lysozyme, a hydrolytic enzyme (EC 3.2.1.17), which cleaves the β - (1,4) -glucoside bond between N-acetylglucosamine and N-acetylmuramic acid in the cell wall glycopeptides of microorganisms [1-3].

Lysozyme is one of the important factors of nonspecific immunity in the human body and many animals in connection with its ability to lyse a number of bacteria [4]. Lysozyme most easily hydrolyzes streptococcal cells, one of which (*Micrococcus lysodeikticus*) is even used as a substrate to determine lysozyme activity [2].

However, in addition to bacteriolytic activity, lysozyme affects many biochemical and physiological processes in the body. So, it increases the chemotaxis of leukocytes [1], stimulates the lymphocytic system of specific immunity [5], and has hepatoprotective and mucosoprotective effects [6, 7].

More recently, a group of Moscow biochemists [8] found that lysozyme binds to the intestinal endotoxin lipopolysaccharide, which is secreted by gram-negative bacteria and has a very strong pro-inflammatory effect [9]. On this basis, they proposed the use of immobilized lysozyme for sorbtion of lipopolysaccharide from the blood of patients with sepsis [8].

We have previously shown that lipopolysaccharide (LPS) inhibits lysozyme activity in many body tissues in animal experiments *in vivo*, and is hundreds of times stronger than any other toxins [10]. At the same time, in experiments *in vitro* we were not able to establish a direct inhibitory effect of LPS on lysozyme [2].

All this determined the aim of this study, which is to study the possible effect of LPS on some other properties of lysozyme, for example, on its ability to adsorb on a substrate such as chitin, which is even used as a substrate for determining the lysozyme content in biological media [11, 12].

MATERIAL AND RESEARCH METHODS

In the work was used the lysozyme preparation “Mayozyme 9000 L”, obtained from egg white, manufactured by MAYASAN Gıda Sanayi ve Ticaret Anonim Şirketi, Turkey.

Lysozyme activity was determined by the Gorin bacteriolytic method in the modification [2]. As the substrate used the preparation of lyophilized cells *Micrococcus lysodeikticus* ATCC No. 4698, manufacturer "Sigma", USA.

As a source of lipopolysaccharide used the drug "Pyrogenal", manufactured "Medgamal" N.F. Gamaleya Federal Research Center Epidemiology & Microbiology. RF.

All lysozyme solutions (from 8 µg/ml to 512 µg/ml) were prepared from the initial preparation with a protein concentration of 416 mg/ml by appropriate dilution in 0.1 M phosphate buffer pH 6.2.

The corresponding dilutions of lipopolysaccharide were prepared on the same buffer, taking into account the concentration of the latter in the "Pyrogenal" preparation, equal to 100 µg/ml.

As a specific sorbent for lysozyme, the chitin preparation "Chitin from shrimp shells", manufactured by Sigma, USA, was used. It was preliminarily ground in a coffee grinder; the fraction was sieved through a 0.56 mm sieve and washed with a pH 6.2 phosphate buffer.

The adsorption of lysozyme or a mixture of lysozyme with lipopolysaccharide was carried out by mixing 50 mg of chitin with 2 ml of solution for 15 minutes. After centrifugation at 2500 g for 10 minutes, the activity of non-adsorbed lysozyme was determined in the supernatant.

All determinations of lysozyme activity were carried out in 3-5 replicates and presented as average values.

RESULTS AND DISCUSSION

In table 1 and in fig. 1 presents the results of determining the bacteriolytic activity of lysozyme solutions with different protein contents. As can be seen from the data in table 1, the activity of lysozyme increases with increasing protein concentration, however, the specific activity is decreases. In fig. 1 shows that the dependence of the bacteriolytic activity of lysozyme on its content is linear only at very low concentrations of the enzyme (less than 100 µg/ml). This circumstance must be taken into account when determining the level of lysozyme in biological media.

As for the sharp (almost 10 times) decrease in the specific activity of lysozyme with an increase in its concentration, this circumstance can be explained by the ability of lysozyme molecules to form dimers, tetramers, even octamers when certain concentrations are reached [13, 14].

Table 1

Protein concentration, $\mu\text{g/ml}$	ml Activity, u/ml	Specific activity, u/mg protein
8,0	0,038	47,5
16,0	0,045	28,1
32,0	0,081	25,3
64,0	0,146	22,8
128,0	0,208	16,3
256,0	0,241	9,4
512,0	0,249	4,9

Bacteriolytic activity of lysozyme depending on protein content

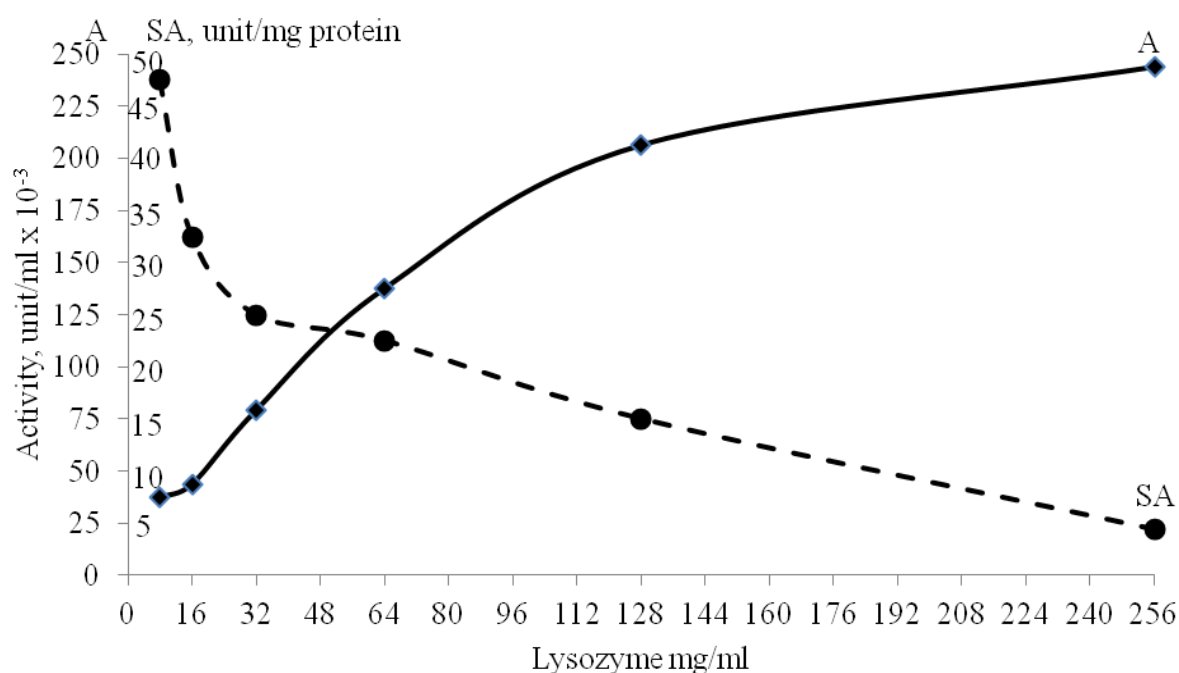


Fig. 1. The dependence of lysozyme activity on concentration

Table 2 presents the results of determining the effect of lipopolysaccharide on lysozyme activity. A mixture of lysozyme and LPS solutions was kept at room temperature for 15 minutes and then lysozyme activity was determined. As can be seen from the data of two experiments with different concentrations of lysozyme, lipopolysaccharide does not

affect the bacteriolytic activity of lysozyme. This circumstance remains incomprehensible in light of the data on the ability of lipopolysaccharide to effectively reduce lysozyme activity in animal tissues *in vivo* [10].

Table 2

The effect of LPS on lysozyme activity

Indicators	Lysozyme, units / ml	LPS, $\mu\text{g} / \text{ml}$	Lysozyme + LPS
<u>Experience 1</u>			
The content of components, mcg / ml	404	40	404 and 40
Lysozyme activity, units / ml	$0,217 \pm 0,012$	0	$0,220 \pm 0,015$ $p > 0,8$
<u>Experience 2</u>			
The content of components, mcg / ml	202	40	202 and 40
Lysozyme activity, units / ml	$0,128 \pm 0,008$	0	$0,126 \pm 0,009$ $p > 0,7$

In addition to bacteriolytic activity, lysozyme has one more specific feature - it is selectively adsorbed on chitin [13, 14]. We decided to check how LPS affects the adsorption of lysozyme on chitin. The results of this check are presented in table 3 and in fig. 2.

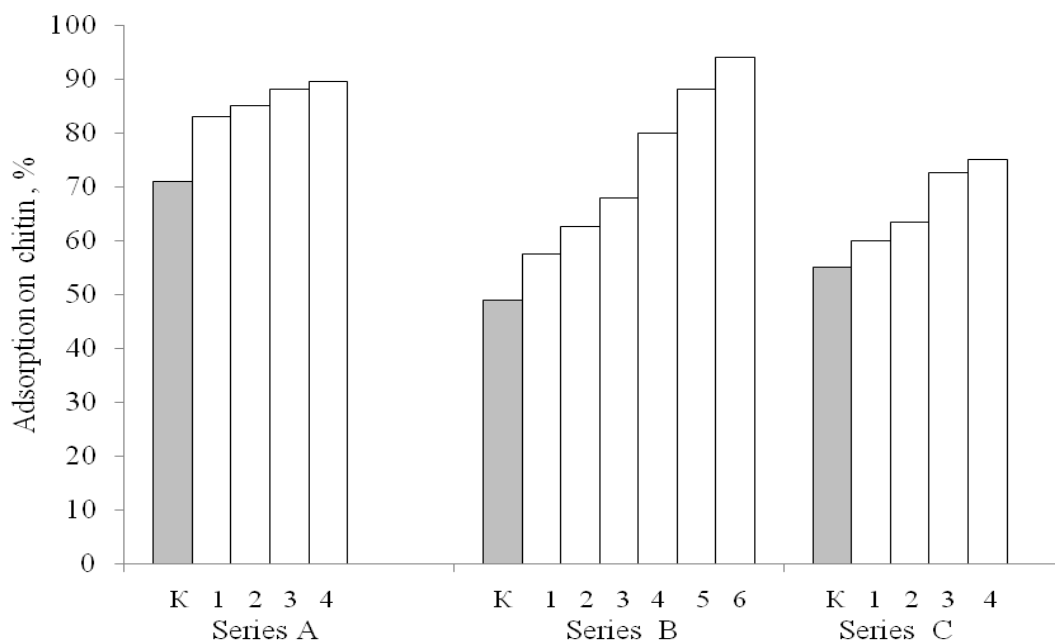


Fig. 2. The effect of LPS on the adsorption of lysozyme on chitin: Series A, 35 $\mu\text{g}/\text{ml}$ lysozyme: 1 – LPS – 0.6 $\mu\text{g}/\text{ml}$, 2 – 1.25 $\mu\text{g}/\text{ml}$, 3 – 8.5 $\mu\text{g}/\text{ml}$, 4 – 10 $\mu\text{g}/\text{ml}$.
Series B, 52 $\mu\text{g}/\text{ml}$ lysozyme: 5 – LPS – 20 $\mu\text{g}/\text{ml}$, 6 – 40 $\mu\text{g}/\text{ml}$.
Series C, 104 $\mu\text{g}/\text{ml}$ lysozyme

Table 3

The effect of LPS on the adsorption of lysozyme on chitin (50 mg)

Series	Lysozyme, mcg / ml. Activity, units / ml	LPS, mcg / ml	Activity after adsorption on chitin
A	35,0 Activity 0,115	0	0,034
		0,6	0,030
		1,25	0,029
		2,5	0,022
		5,0	0,020
		10,0	0,024
B	52,0 Activity 0,192	0	0,131
		0,6	0,089
		1,25	0,088
		2,5	0,070
		5,0	0,037
		10,0	0,032
C	52,0 Activity 0,123	0	0,041
		10,0	0,022
		20,0	0,015
		40,0	0,007
D	104,0 Activity 0,263	0	0,117
		0,6	0,115
		1,25	0,095
		2,5	0,081
		5,0	0,070
		10,0	0,072

The effect of different LPS concentrations (from 0.6 to 40 $\mu\text{g}/\text{ml}$) was studied in 3 lysozyme solutions containing 35 $\mu\text{g}/\text{ml}$, 52 $\mu\text{g}/\text{ml}$ and 104 $\mu\text{g}/\text{ml}$ of the enzyme. The results presented in table 3 indicate that the addition of LPS dose-dependently enhances the adsorption of lysozyme on chitin, and the activating effect of LPS is already apparent from very low concentrations - 0.6 $\mu\text{g}/\text{ml}$. The most sensitive to the action of LPS was the concentration of lysozyme, equal to 52 $\mu\text{g}/\text{ml}$. Based on these data, the degrees of lysozyme adsorption on chitin were calculated, shown in Fig. 2. As can be seen from the figure, the most clear dose dependence of the activating effect on the adsorption of lysozyme LPS is at a concentration of lysozyme 52 $\mu\text{g}/\text{ml}$ (series of experiments B and C).

To explain the activating effect of LPS on the adsorption of lysozyme, we assume that LPS promotes depolymerization of lysozyme aggregates, converting di-, tetra- and octomeres to the monomeric state, which more easily interacts with chitin.

However, the biological role of the activating effect of LPS on the adsorption of lysozyme still needs to be clarified.

CONCLUSIONS

1. Dose-dependent bacteriolytic activity of lysozyme appears only at very low concentrations (less than 100 µg/ml).
2. LPS does not inhibit the bacteriolytic activity of lysozyme *in vitro*.
3. LPS dose-dependently enhances the adsorption of lysozyme on chitin, starting from a concentration of 0.6 µg/ml.

REFERENCES

1. Barabash AD, Levitsky AP. Enzymatic mechanisms of antimicrobe defence of at oral cavity. Questions of medical chemistry. 1978; 24(3): 291-310. (in Russian)
2. Levitsky AP. Lysozyme instead of antibiotics. Odessa, KP OGT, 2005: 74. (in Russian)
3. Sariri R, Gafuri Kh. Proteins of tears: normal, with eye disease and when wearing contact lenses. Biochemistry. 2008; 73(4): 469-482. (in Russian)
4. Bukharin OV, Usvetsov BYa, Matiash IN [and others]. The reference of lysozyme antimicrobe activity. Antibiotics. 1976; 9: 805-808. (in Russian)
5. Dobrotina NA, Kazatskaia ZhA, Emelianova TIu. Lysozyme as immunological modulator. Questions of medical chemistry. 1988; 33: 66-69. (in Russian)
6. Maianskii DN, Kutina SN, Shcherbakova EG. Influence of lysozyme on hepatocytes resistance. Antibiotics and chemotherapy. 1988; 33(2): 128-131. (in Russian)
7. Kutina SN, Zubakhin AA. Liver resistance to damage of CCl₄ at stimulation of macrophages with the preparations of different categories. BEBIM. 2000; 129(6): 620-622. (in Russian)
8. Levashov PA, Matolygina DA, Ovchinnikova ED [and others]. New sorbent based on covalently immobilized lysozyme to remove bacterial lipopolysaccharide (endotoxin) from biological fluids. Biochemistry. 2019; 84(1): 100-108. (in Russian)
9. Yakovlev MYu. The elements of endotoxin theory of human physiology and pathology. Physiology of man. 2003; 29(4): 98-109. (in Russian)
10. Levitsky AP, Ostafiichuk MA, Uspenskii OE [and others]. The influence of different pathogens on the lysozyme activity into tissues of rat oral cavity. Journal of Education, Health and Sport. 2017; 7(8): 1070-1081.
11. Cherkasov IA, Kravchenko NA. An improved method for the isolation of lysozymes by enzyme-substrate chromatography. Biochemistry. 1969; 34(5): 1089-1091. (in Russian)

12. Cherkasov IA, Kravchenko NA. Copyright certificate 178771 USSR, IPC C12K. A method for producing enzyme preparations of lysozymes from their raw materials. Application number 943666/28-13. Date of filing: 17.11.1965. Publ.: 19.11.1966. Bul. № 4. (in Russian)

13. Lakhtin V, Kostanova E, Arbatskii N. Purification and characterization of multiple forms of papaya latex lysozyme. Applied biochemistry and microbiology. 1995; 2: 247-254. (in Russian)

14. Islam R, Kite J, Baker AS [and others]. Affinity purification of hen egg lysozyme using sephadex G75. African J. of Biotechnology. 2006; 5(20): 1902-1908.