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# Determination of valproic acid and 3-heptanone in plasma using air-assisted liquid-liquid microextraction with the assistance of vortex: Application in the real samples

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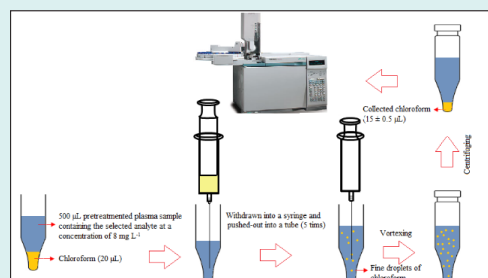
## Abstract

**Introduction:** Valproic acid (VPA) is an antiepileptic drug used to treat epilepsy and bipolar disorder. Adverse effects of VPA were studied in many reports, however, a dose-response relationship between VPA and its metabolites in epilepsy patients are extremely limited. In this paper, a high efficient method was developed for the preconcentration and determination of VPA and its main metabolite in plasma.

**Methods:** For the extraction and preconcentration of the selected analytes, a volume of an extractant was placed at the bottom of the microtube containing pretreated plasma. The mixture was repeatedly withdrawn from the microtube and pushed-out into it using a 1.0-mL glass syringe and resulted in a cloudy mixture. For further turbidity, the mixture was shaken on a vortex agitator. This procedure was used to analyze the plasma samples of patients with epilepsy (n = 70).

**Results:** The results revealed that in most patients with a low level of VPA relative to its expected level, 3-heptanone concentrations were high. The limits of quantification of 3-heptanone and VPA were 0.04 mg L<sup>-1</sup> and 0.2 mg L<sup>-1</sup>, respectively. A suitable precision at a concentration of 2 mg L<sup>-1</sup> for each analyte was obtained (relative standard deviation ≤ 9%).

**Conclusion:** The obtained results indicated that this procedure is easy, sensitive, and reliable, and can be used for the analysis of the selected analytes in the plasma samples of patients with epilepsy.



## Introduction

Pharmacotherapy with anti-epileptic drugs (AEDs) is the most common method for the treatment of epilepsy. However, in 10%-20% of epileptic children, drug-resistant epilepsy (DRE) is observed<sup>1</sup> and therefore, “pseudo” refractiveness should be considered in the diagnosis of DRE. Adequate dosage<sup>2,3</sup> and appropriate combination of AEDs are important issues. Refractory epilepsy is defined as uncontrolled epilepsy despite the administration of an appropriate AED with adjusted dosage and duration.<sup>4</sup>

The failure in treatment could be due to choosing the AED, its administered dose, any variations in its absorption following oral administration, any change in its metabolism, and drug resistance. Monitoring plasma levels of AEDs could provide some evidence for better management of epileptic patients.

2-Propylpentanoic acid or valproic acid (VPA) is the most well-known AED that was used to treat epilepsy and bipolar disorder.<sup>5</sup> The therapeutic dosage of VPA was in the range of 50 to 100 mg mL<sup>-1</sup>.<sup>6,7</sup> Most amount of VPA

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is eliminated by metabolism and a small amount of an administered dosage appears unchanged in urine (less than 4%).<sup>8</sup> Some of the metabolites possess the anticonvulsant activity and some have been suspected to be associated with serious hepatotoxicity.<sup>9</sup> VPA is metabolized to unsaturated compounds by hydrogenation, to hydroxyl metabolites by oxidation, and to carbonyl metabolites by their further oxidation.<sup>10</sup> One of the main carbonyl metabolites of VPA is 3-heptanone.  $\beta$ -oxidation leads to 3-oxo-VPA and after spontaneous decarboxylation leads to 3-heptanone.<sup>11</sup> Study of concentrations of VPA and its main metabolite in plasma and their relationship can be used for dosage adjustment of VPA especially in children with refractory epilepsy. Consequently, there has been considerable interest to develop a sensitive method to determine VPA and 3-heptanone dosages in biological samples and study their relationship in patients with epilepsy.

The available analytical methods for monitoring various AEDs have been reviewed in previous work.<sup>12</sup> Different bioanalytical procedures have been developed to analyze VPA in biological samples such as a gas chromatography (GC) with flame ionization detector (FID)<sup>13-16</sup> and mass spectrometry (MS),<sup>17,18</sup> high performance liquid chromatography with ultraviolet detector,<sup>19</sup> fluorescence detector,<sup>20</sup> MS,<sup>21</sup> and capillary electrophoresis (CE).<sup>22</sup> However, among the applied procedures to determine VPA, GC-FID is a simple and fast procedure.

Although the above-mentioned techniques are sensitive, direct analysis of trace levels of analytes is almost impossible. Therefore, prior to detection of the analytes with the instrument, sample pretreatment methods are needed. Conventional sample pretreatment methods such as solid phase extraction and liquid-liquid extraction are typically time-consuming and boring, as well as using large volumes of toxic solvent.<sup>23-27</sup> Hence, microextraction procedures like solid-phase microextraction (SPME)<sup>14,17</sup> and liquid phase microextraction (LPME)<sup>13,28</sup> were very much considered as alternatives. But, SPME fibers are relatively costly, and commonly fragile, as well as suffering from sample carryover problems. In order to solve abovementioned problems, dispersive liquid-liquid microextraction (DLLME) has been developed as an effective extraction and enrichment procedure.<sup>29</sup> It has been widely used to analyze drugs in urine and serum samples.<sup>16,30,31</sup> In DLLME, a mixture of extraction solvent (low soluble in aqueous phase) and disperser solvent (soluble in both extraction solvent and aqueous phase) was rapidly injected into an aqueous solution containing analytes using a syringe. By this action, extractant was dispersed into aqueous solution as tiny droplets by disperser solvent. After centrifugation, extractant containing the enriched analytes was collected. The use of a disperser solvent in DLLME decreases the sample polarity which leads to the increase of analyte solubility in the sample solution and the decrease of extraction

performance especially in a low volume of biological samples. To improve extraction performance of DLLME, disperser solvent-free procedures were developed for extraction and enrichment of target analytes.<sup>32-34</sup>

Free fraction of drugs is responsible for their biological (either pharmacological or toxicological) activities. The free fraction of VPA could be affected by albumin levels<sup>35</sup> and some co-administered drugs. 3-Heptanone concentrations in exhaled breath of patients receiving VPA was proposed as a biomarker for its monitoring and pharmacokinetics.<sup>11,35</sup> There was a positive correlation between 3-heptanone concentration in exhaled breath and VPA dose administered to the patients.<sup>11</sup> However, no significant correlation was observed between serum concentration of VPA and exhaled breath concentration of 3-heptanone.<sup>11</sup> A number of technical points should be considered in practical applications of analyte determinations in exhaled breath as described in our recent report.<sup>36</sup> These points could be a possible reason for such non-significant correlations. In addition to these efforts to improve the outcomes of pharmacotherapy, new approaches including optogenetics, transcranial magnetic stimulation, responsive neurostimulation, cerebral cooling, local AED perfusion to epileptic foci, gene therapy and stem cell therapy<sup>37,38</sup> are under investigation to provide better management for refractory epilepsy.

The aims of this work were, a) to develop a simple, fast, and efficient microextraction method in small scale for the extraction and quantification of VPA and 3-heptanone in human plasma using GC-FID, b) to evaluate the correlations of the plasma VPA and 3-heptanone concentrations in children with epilepsy.

## Materials and Methods

### Reagents and solutions

3-Heptanone was provided from Sigma-Aldrich (St. Louis, USA). Sodium valproate powder was a gift from Rouz Darou Pharmaceutical Co. (Tehran, Iran). Methanol was also supplied from Sigma-Aldrich. Carbon tetrachloride, trifluoroacetic acid, 1,1,2,2-tetrachloroethane (1,1,2,2-TCE), 1,2-dichloroethane (1,2-DCE), and chloroform were provided from Merck (Darmstadt, Germany). Deionized water was provided from Ghazi Pharmaceutical Company (Tabriz, Iran). Appropriate amounts of the target analytes were dissolved in methanol and stock solutions ( $C=1000 \text{ mg L}^{-1}$ , each analyte) were obtained. For the preparation of working standard solution, the appropriate volume of stock solution was spiked into drug-free plasma.

### Apparatus

Agilent 7890A gas chromatograph equipped with FID (Agilent Technologies, CA, USA) was used to analyze the target analytes. The temperature of the inlet was set at 300°C. Splitless mode with a sampling time of 1 minute and a split ratio of 1:10 was applied. Nitrogen was used as the

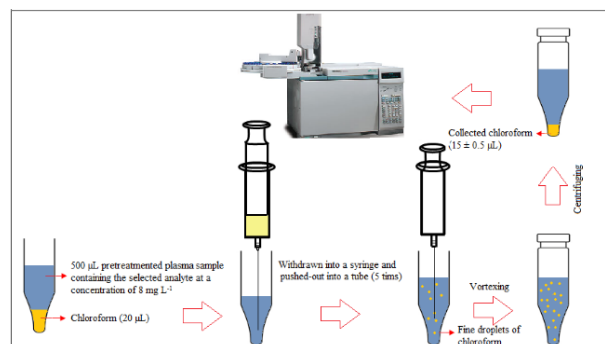
make-up gas and carrier gas with flow rates of 25 mL min<sup>-1</sup> and 1 mL min<sup>-1</sup>, respectively. The target analytes were separated on an HP-5 capillary column (30 m × 0.32 mm i.d. with a 0.25 µm stationary film thickness) (Hewlett-Packard, Santa Clara, USA). The temperature of the oven was set at 70 °C for 2 minutes increased at a rate of 15°C min<sup>-1</sup> to 200°C, and then increased at a ratio of 20°C min<sup>-1</sup> to 300°C to clean up the column. A 1 µL microsyringe (zero dead volume, Hamilton, Switzerland) was applied to introduce samples into GC. Injection volume was 1 µL. The temperature of FID was 300°C. Hydrogen for FID was produced by the hydrogen generator (GLAIND-2200, Dani, Italy). Air with a flow rate of 400 mL min<sup>-1</sup> was used for FID. A vortex from Labtron Company (Tehran, Iran) was applied in extraction steps and sample preparation. To measure pH, a pH meter model 744 (Herisau, Switzerland) was applied. Sigma centrifuge (Osterode, Germany) was applied for extraction steps and protein precipitation.

### Real samples

Seventy epileptic patients were studied (50 females and 20 males) with the mean (ranges) age of 7.9 years (1–18 years old) and mean body weight of 26.4 kg (5–91), respectively. The collected blood samples were centrifuged (8000 rpm for 5 minutes), and the supernatant was removed and kept at -20°C. All sample donors' parents were informed on the details of the sample collection and a written consent form was signed. Drug-free (blank) human plasma samples to optimize the presented procedure were obtained from healthy donors (Iranian Blood Transfusion Research Center, Tabriz, Iran) and kept at -20 °C.

### Procedure

Frozen human plasma samples were placed into the water bath (temperature = 25 °C) to melt and then homogenized using a vortex. An aliquot of plasma (500 µL) was transferred into a microtube (1.5 mL) and fortified with 3-heptanone and sodium valproate (C = 8 mg L<sup>-1</sup>). To precipitate proteins of plasma, 50 µL trifluoroacetic acid was added into the mentioned solution and shaken by hand. After centrifugation at 10000 rpm for 5 minutes, the upper phase was removed and transferred into another microtube with conical bottom. For the isolation and enrichment of the target analytes, 20 µL chloroform was placed at the bottom of the microtube containing the pretreated plasma. The mixture was repeatedly (5 times) withdrawn from the microtube and pushed-out into it using 1.0-mL glass syringe, resulting in a cloudy mixture. For the further turbidity, the mixture was shaken on a vortex agitator at 6000 rpm for 1 minute. The cloudy mixture was formed, and the selected analytes were extracted into the tiny droplets of chloroform. The tube was then centrifuged at 6000 rpm for 2 minutes. After centrifugation, 15 ± 0.5 µL chloroform was collected at the bottom of the microtube. For quantitative analysis, 1 µL of



**Scheme 1.** Microextraction and preconcentration procedure.

the sediment phase was withdrawn by a 1 µL microsyringe and then introduced into GC-FID system. The presented method is schematically shown in Scheme 1.

### Calculation of extraction recovery and enrichment factor

To evaluate the extraction efficiency, extraction recovery (ER) and enrichment factor (EF) were used and calculated using equations 1 and 2:

$$EF = \frac{C_{Sed}}{C_0} \quad (1)$$

$$ER \% = \frac{n_{Sed}}{n_0} \times 100 = \frac{C_{Sed} \times V_{Sed}}{C_0 \times V_{Aq}} \times 100 \quad (2)$$

$$= EF \times \frac{V_{Sed}}{V_{Aq}} \times 100$$

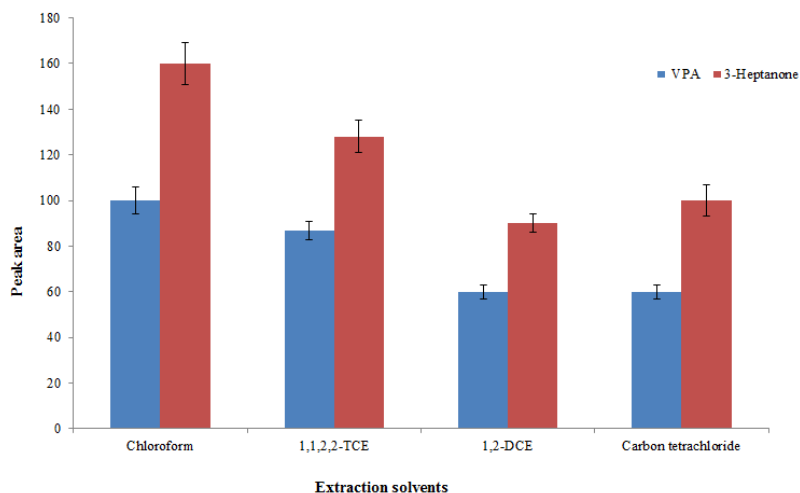
Where  $C_{Sed}$ ,  $V_{Sed}$ ,  $C_0$ , and  $V_0$  are the analyte concentration in the settled phase, the settled phase volume, the initial analyte concentration in the human plasma sample, and the plasma sample volume, respectively.  $n_0$  and  $n_{Sed}$  are the initial and the extracted analyte amounts, respectively.

### Results

To achieve high extraction performance, several parameters affecting the efficiency of the presented procedure including type and volume of extractant, extraction number, pH, and vortex time were investigated.

#### Type of extractant

Selecting a suitable extractant is the crucial step in this procedure since the quality of an extractant determines its efficiency to extract the analytes. The requirements for the extractant in this method are higher density than water, high extraction performance to extract the selected analytes, immiscible with the aqueous phase, and good chromatographic behavior. Based on mentioned requirements, 1,2-DCE, 1,1,2,2-TCE, chloroform, and carbon tetrachloride were evaluated. Comparison of analytical signals obtained with the use of the selected extractants is given in Fig. 1. Chloroform, providing the best performance for the selected analytes, was used for the next experiments.



**Fig. 1.** Selection of extraction solvent. Conditions: Sample volume, 500  $\mu\text{L}$  human plasma spiked with 8  $\text{mg L}^{-1}$  of each analyte; protein precipitation agent, 50  $\mu\text{L}$  trifluoroacetic acid; extraction number, 5; pH, 3; vortex time, 5 min. The error bars indicate the minimum and maximum of three determinations.

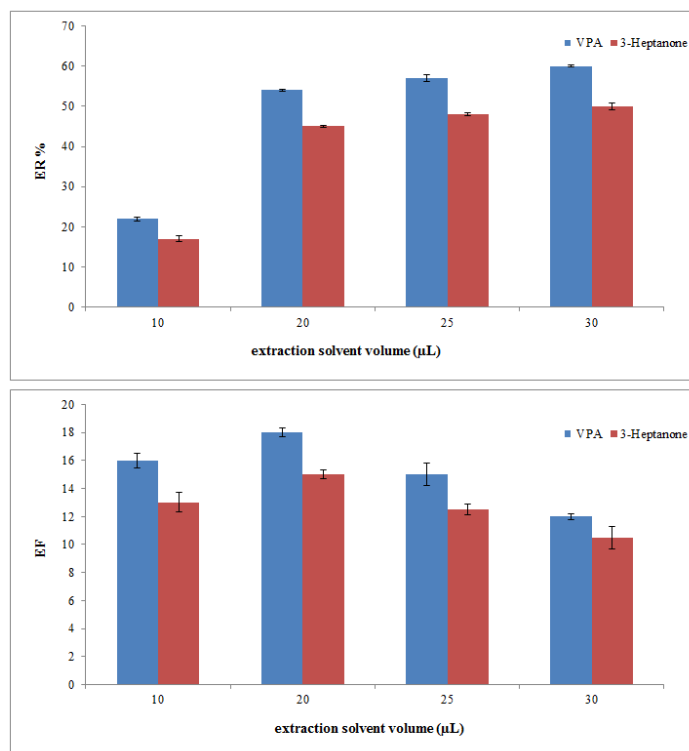
**Selection of extractant volume**

Extractant volume is a very crucial parameter that affects extraction performance in the proposed method. The high volume of extraction solvent leads to an increase in extraction recovery and a decrease in EFs due to the dilution effect. For this reason, the volume of chloroform should be optimized. To optimize extractant volume, this method was performed using different chloroform volumes (10, 20, 25, and 30  $\mu\text{L}$ ). The graphs of ERs and EFs of analytes versus volume of chloroform (Fig. 2) showed that by increasing the volume of chloroform, the

concentration of analyte in the sedimentary chloroform was reduced, resulting in a decreased EF. When the volume of sedimentary phase is increased, ER is enhanced by increasing the volume of chloroform. A suitable ER and best EF could be achieved at a volume of 20  $\mu\text{L}$  of chloroform. Therefore, 20  $\mu\text{L}$  of chloroform was applied in the following experiment.

**Study of the sample pH**

Sample solution is a main factor affecting the extraction performance of the presented procedure. Therefore, the



**Fig 2.** Selection of extraction solvent volume. Conditions: The same as used in Fig. 1, except that chloroform was used as the extraction solvent.

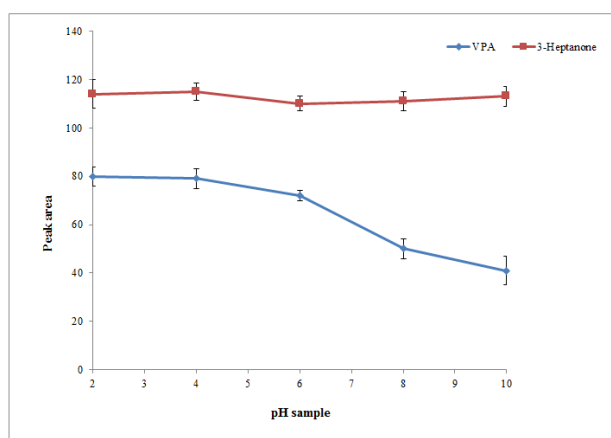
procedure was performed in the plasma sample with different pH values over the range of 2–10, adjusted by adding 0.1 M sodium hydroxide or hydrochloric acid solution. The results showed that in alkaline condition, the analytical signal of VPA decreased, whereas the analytical signal of 3-heptanone was not changed significantly. This can be attributed to the acidic properties of VPA. Indeed, in acidic condition, VPA is changed to ionic form, whereas 3-heptanone is not influenced by changing pH. Hence, the pH ranges of 2 to 4 are suitable. It is noted that in this method pH values of samples were approximately 3 due to the usage of trifluoroacetic acid as a protein precipitation agent. Therefore, for the following studies, the pH of samples was not changed (Fig. 3).

#### Extraction number

In this procedure, number of times that mixture of chloroform and plasma sample was withdrawn from a microtube into a 1-mL syringe and pushed-out into microtube was defined as the extraction number. It is predicted that the extraction efficiency of the presented method increases by increasing the extraction number. For this purpose, the extraction number was evaluated in the range of 1–5 times. As illustrated in Fig. 4, by increasing the extraction numbers till 4, the EF of the presented method increased and then stayed constant. Hence, 5 was selected as an extraction number for further experiments.

#### Effect of vortex time

Vortex time is defined as the minimum time necessary to obtain equilibrium condition between chloroform and plasma sample. By increasing vortex time, degree of solution turbidity increases resulting in higher extraction efficiency. Hence, to achieve equilibrium condition, evaluation of vortex time was essential. The obtained results showed that the equilibrium condition could be obtained only after 2 minutes of vortexing at a rate of 6000 rpm (Fig. 5).



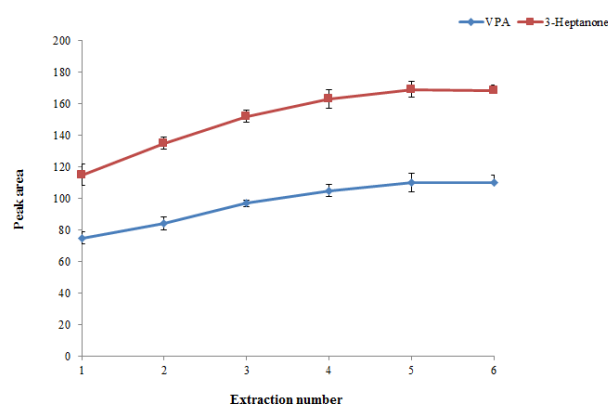
**Fig 3.** Study of pH values. Conditions: The same as used in Fig. 2, except that 20  $\mu$ L chloroform was used as the extraction solvent volume.

#### Analytical characteristics of the procedure

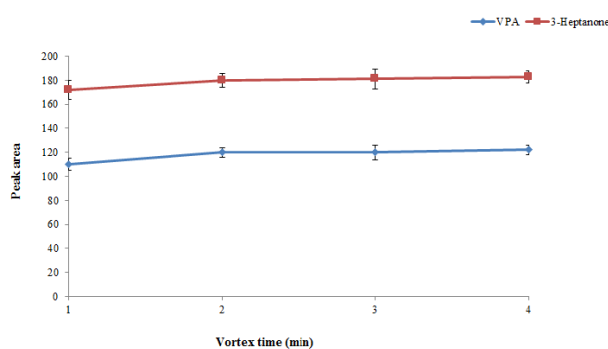
To investigate the performance of the presented procedure, some parameters, including detection limits (LODs), quantification limits (LOQs), linear range (LR), and relative standard deviation (RSD) of repeatability were determined under the optimized conditions. The obtained results are shown in Table 1. Acceptable linearity was achieved for the selected analytes with correlation coefficients ( $r$ ) higher than 0.997. The LODs and LOQs based on a signal-to-noise ( $S/N$ ) of 3 and 10, respectively, were determined to be 0.010 and 0.057  $\text{mg L}^{-1}$  for 3-heptanone and 0.04 and 0.2  $\text{mg L}^{-1}$  for VPA, respectively. The EFs of  $56 \pm 3$  and  $51 \pm 2$  and the ERs of  $19 \pm 1\%$  and  $17 \pm 0.5\%$  were obtained for VPA and 3-heptanone, respectively. The inter-day and intra-day precisions were obtained from four consecutive days and six replicates, respectively ( $C = 2 \text{ mg L}^{-1}$ ). The inter-day and intra-day RSDs were less than 9%.

#### Analysis of epileptic patients' samples

The established procedure was used to analyze 3-heptanone and VPA in plasma samples of 70 children with epilepsy after receiving valproate and for some cases a number of



**Fig 4.** Effect of extraction number on the extraction efficiency of the method. Conditions: The same as used in Fig. 3, except that the pH of the sample was adjusted to 3.



**Fig 5.** Effect of vortex time on the extraction efficiency of the method. Conditions: The same as used in Fig. 4, except that extraction number was 5.



**Table 1.** Quantitative features of the proposed method for the selected analytes

Analyte	LOD <sup>a</sup>	LOQ <sup>b</sup>	LR <sup>c</sup>	r <sup>2</sup> <sup>d</sup>	RSD % <sup>e</sup>	
					Intra-day	Inter-day
VPA	0.057	0.20	0.20 – 100	0.998	8	9
3-Heptanone	0.010	0.040	0.04 – 100	0.997	5	5

<sup>a</sup>Limit of detection (S/N=3) (mg L<sup>-1</sup>).

<sup>b</sup>Limit of quantification (S/N=10) (mg L<sup>-1</sup>).

<sup>c</sup>Linear range (mg L<sup>-1</sup>).

<sup>d</sup>Coefficient of determination.

<sup>e</sup>Relative standard deviation (n=6, C= 2 mg L<sup>-1</sup> of each analyte) for intra–day and (n=4, C= 2 mg L<sup>-1</sup> of each analyte) for inter-day precisions.

co-administered AEDs. The summary of the results of analyses is listed in Table 2. More details are presented as supplementary information (Table 1S). The levels of VPA and 3-heptanone for patients #18 and #57 were lower than LOQ values of the analytes, *i.e.* < 0.2 mg L<sup>-1</sup> for VPA and < 0.04 mg L<sup>-1</sup> for 3-heptanone, respectively. Therefore, we did not report the data and listed them as not determined (ND). No intake of the drug by these patients could be a possible reason for this observation. For patients #4, #11, #23, and #29, VPA levels were 28,

15, 27, and 38 mg L<sup>-1</sup>, respectively, whereas 3-heptanone levels were not determined to reveal the levels of < 0.04 mg L<sup>-1</sup> possibly due to a very low metabolism of VPA to 3-heptanone in these patients. Level of VPA in plasma samples of patients #37, #54, and #60 were less than 2.0 mg L<sup>-1</sup>, whereas relatively high levels of 3-heptanone were found for these samples, *i.e.* 3.0, 1.1, and 1.8 mg L<sup>-1</sup>, revealing fast metabolism of VPA in these patients. As further evidence for this hypothesis, VPA levels for all samples with 3-heptanone > 2.0 mg L<sup>-1</sup> were very low. There was a poor correlation (R = 0.29) between VPA and 3-heptanone plasma concentrations of the investigated cases as illustrated in Fig. 6.

**Table 2.** Descriptive data and characteristics of the studied population (N=70)

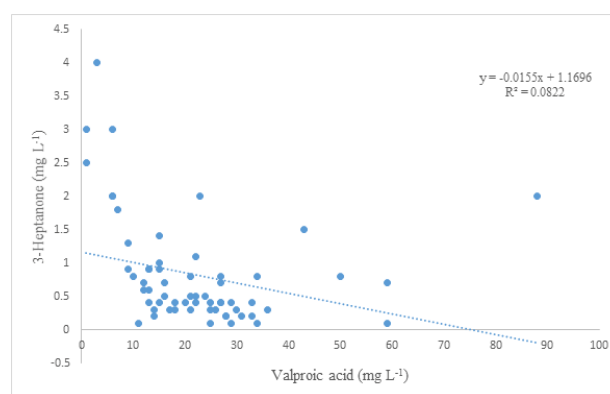
Variable	Mean (±SD) or number
Age (year)	7.95 ± 4.21
≥15	2
10-15	22
5-10	20
0-5	17
NR	9
Weight (kg)	26.55 ± 13.78
≤15	13
15-20	12
20-25	7
25-30	6
≥30	23
NR	9
VPA concentration (mg/L)	22.69 ± 14.61
≤10	10
10-15	14
15-20	6
20-25	11
25-30	11
≥30	13
ND	5
3-Heptanone concentration (mg/L)	0.87 ± 0.84
0.1-0.4	18
0.4-0.8	20
0.8-1.2	12
≥1.2	14
ND	6
Co-administrated AEDs	-
Pb	50
Ph	15
CBZ	17

NR: not recorded; ND: not detected; Pb: Phenobarbital; Ph: Phenytoin; CBZ: Carbamazepine.

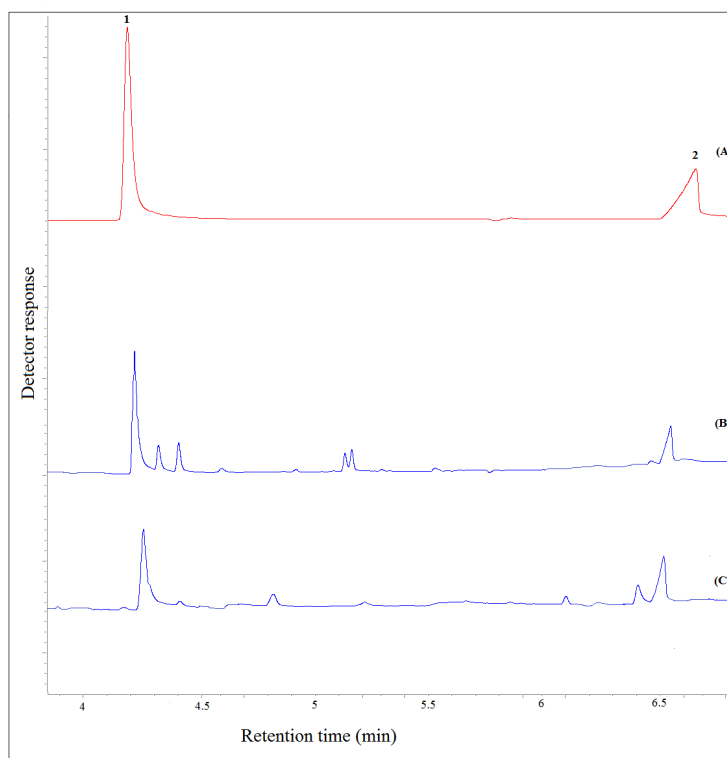
Fig. 7 presents typical chromatograms obtained by GC-FID for a standard solution (C= 250 mg L<sup>-1</sup>, each analyte), a free-drug plasma sample fortified with the target analytes (C=8.0 mg L<sup>-1</sup>, each analyte), and the patient plasma after taking valproate. Considering the obtained results, there is no interfering peak in retention times of VPA and 3-heptanone, proving that the proposed procedure is suitable for clinical studies.

**Discussion**

A comparative study of the analytical properties of the proposed method and some published methods in terms of LOD, RSD, extraction time (ER), and LR for the extraction and determination of the selected analytes was carried out, and the results obtained were tabulated in Table 3. The difference of the presented method with our previous method is in the first step of the procedure. In this method,



**Fig 6.** Relationship of plasma concentration of VPA and 3-heptanone.



**Fig. 7.** Typical GC–FID chromatograms of: (A) a standard solution prepared in chloroform ( $250 \text{ mg L}^{-1}$ , each analyte), (B) plasma sample spiked with the analytes at a concentration of  $8 \text{ mg L}^{-1}$  (each analyte), and (C) patient plasma after performing the proposed method, except for chromatogram (A) in which direct injection was used. Peaks identification: 1, 3-Heptanone; and 2, VPA.

a disperser less method was applied for the extraction and preconcentration of the selected analytes which is more efficient than DLLME (our previous method) due to the elimination of disperser solvent, hence this method provides lower LOD, LOQ, and higher ER. As listed in Table 2, the LR and LOD of the presented procedure are suitable compared to other procedures. The RSD values are comparable with the RSDs reported by other studies. The ER of the presented procedure is shorter than that of other published procedures. Considering the analytical performance of the proposed method compared to other methods, it is a more simple, sensitive, and reliable method

for analyzing the selected analytes in plasma samples.

### Conclusion

Our study reported a new, simple, and efficient disperser-less microextraction procedure combined with GC-FID for isolation, enrichment, and determination of 3-heptanone and VPA in plasma samples. The obtained acceptable results proved that this procedure is sensitive, reliable, and cost-effective and could be used as a suitable alternative to previously reported procedures. Finally, this procedure has been successfully applied to analyze VPA and 3-heptanone in 70 patients with epilepsy.

**Table 3.** Comparison of the presented method with other methods used in preconcentration and determination of the VPA

Sample	RSD% <sup>a</sup>	Extraction time (min)	LR <sup>b</sup> ( $\mu\text{g mL}^{-1}$ )	LOD <sup>c</sup>	Method	Ref.
Human serum	8.8	20	0.5-100	NR <sup>h</sup>	HS-SPME-GC-MS <sup>d</sup>	39
Human serum	13.2	20	2-20	0.8	HS-LPME-GC-FID <sup>e</sup>	13
Human serum	7	10	2-100	1.7	HS-SPME-GC-FID <sup>f</sup>	14
Human serum	7	15	0.25-100	0.065	DLLME-VADLLME- GC-FID <sup>g</sup>	40
Human plasma	9	< 15	0.2-100	0.057	Presented method	-

<sup>a</sup> Relative standard deviation

<sup>b</sup> Linear range.

<sup>c</sup> Limit of detection ( $\text{mg L}^{-1}$ ).

<sup>d</sup> Headspace -solid-phase microextraction-gas chromatography-mass spectrometry.

<sup>e</sup> Headspace-liquid phase microextraction-gas chromatography-flame ionization detector.

<sup>f</sup> Headspace-solid phase microextraction- gas chromatography-flame ionization detector.

<sup>g</sup> Dispersive liquid-liquid microextraction- vortex assisted dispersive liquid-liquid microextraction- gas chromatography-flame ionization detector.

<sup>h</sup> Not reported.

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**Ethical statement**

Volunteers signed a consent form which was approved by the Ethics Committee of Tabriz University of Medical Sciences (approval No. of IR.TBZMED.REC.13953836).

**Competing interests**

The authors declare no conflict of interests.

**Authors contribution**

BF: Developing the analytical method, data analysis, drafting the manuscript. MB: Supervision of the thesis project. ShSa: Plasma sample collection. ShSh: Physical examination of the sample donors. MK: Conceptualization, data analysis, and interpretation. AJ: Team leading, reviewing, and finalizing the manuscript.

**Supplementary Materials**

Supplementary file 1 contains Table S1.

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**Research Highlights****What is current knowledge?**

- ✓ Valproic acid and 3-heptanone were analyzed in plasma samples of patients.
- ✓ This method can be used in clinical studies.

**What is new here?**

- ✓ In this paper, a new, simple, and highly efficient disperser-less microextraction procedure was established for the extraction and preconcentration of VPA and 3-heptanone in patient plasma samples.

- Preparations By Headspace Liquid-Phase Microextraction Gas Chromatography-Flame Ionization Detection Without Prior Derivatization. *J Chromatogr B* 2007; 850: 128-33. Doi: 10.1016/J.jchromb.2006.11.013
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