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Synthesis of Arylidene- β -lactams via Exo-Selective Matsuda-Heck Arylation of Methylene- β -lactams

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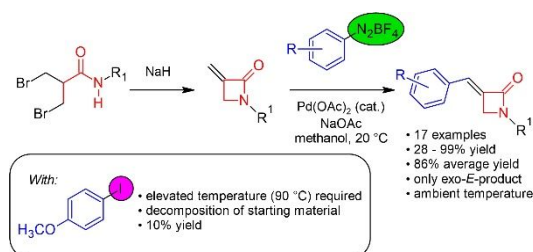
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Abstract:

Exo-methylene- β -lactams were synthesized in two steps from commercially available 3-bromo-2-(bromomethyl)propionic acid and reacted with arene diazonium salts in a Heck-type arylation in the presence of catalytic amounts of Pd(OAc)₂ under ligand-free conditions. The products, arylidene- β -lactams, were obtained in high yields as single isomers. The β -hydride elimination step of the Pd-catalyzed coupling reaction proceeds with high *exo*-regio- and *E*-stereoselectivity. With aryl iodides, triflates, or bromides the coupling products were isolated only in low yields, due to extensive decomposition of the starting material at elevated temperatures. This underlines that arene diazonium salts can be superior arylating reagents in

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3 Heck-type reactions and yield coupling products in synthetically useful yields and selectivities
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5 when conventional conditions fail.
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10 **Introduction**

11
12 Azetidin-2-ones, also referred to as β -lactams, are the pharmacophore unit of β -lactam
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14 antibiotics, such as penicillines, cephalosporins, carbapenems and monobactams. They act by
15
16 deactivating transpeptidases via an acylation of a serine residue at the catalytically active site,
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18 which hinders the final step of bacterial cell wall synthesis.¹ The main driving force for the
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20 acylation is the relief of ring strain, as the azetidin-2-one ring is opened in the process. For β -
21
22 lactam antibiotics resistance mainly occurs through the expression of β -lactamases, which
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24 catalyze the hydrolytic ring opening of the antibiotics and thus deactivate them.¹ A strategy to
25
26 overcome resistance toward β -lactam antibiotics relies on the co-administration of β -lactamase
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28 inhibitors, such as tazobactam, which is clinically combined with piperacillin.² However, some
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30 β -lactamases became in turn resistant against established, clinically used inhibitors. For
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32 instance, some variants of TEM-1 and SHV-1- β -lactamases, which are commonly produced by
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34 *Escherichia coli* and *Klebsiella pneumoniae* (bacteria responsible for infections of the urinary
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36 and respiratory tract and the bloodstream) developed a resistance against clavulanic acid.³ As a
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38 consequence, tackling antibiotic resistance does not only involve the constant search for new
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40 antibiotics, but also the development of novel β -lactamase inhibitors^{1,2} and fluorogenic probes
41
42 for detecting β -lactamases.⁴ Apart from diazabicyclooctanes (DBO's) and cyclic boronic acids
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44 substituted β -lactams, such as the 3-arylidene-azetidin-2-ones (**1**), have been intensively
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46 investigated as inhibitors of β -lactamases (**Figure 1**).
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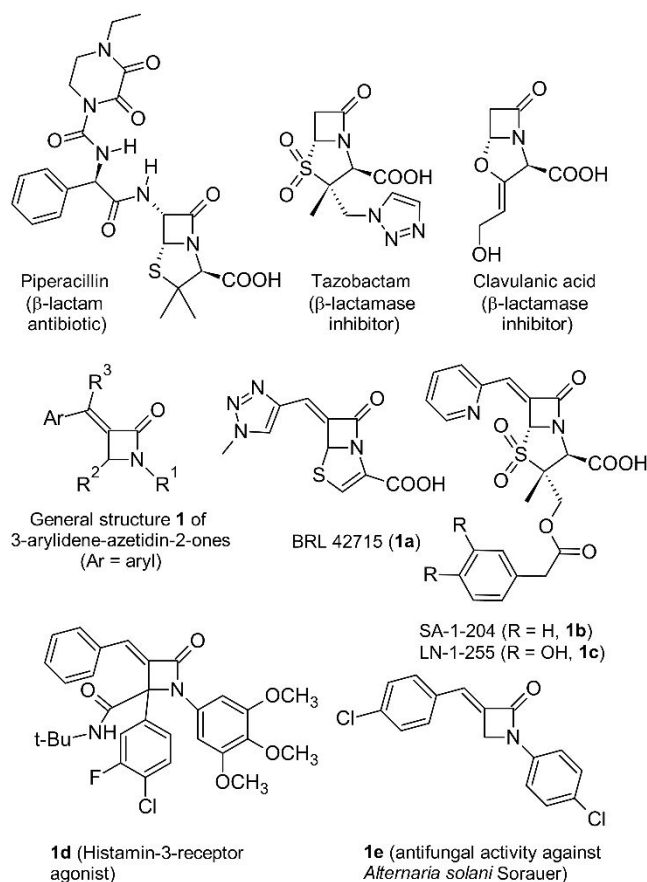


Figure 1. Structures of biologically active azetidin-2-ones.

For example, the penem BRL 42715 (**1a**) was found to inhibit certain cephalosporinases 10^4 to 10^6 times stronger than clavulanic acid.^{5,6} A few years later Buynak et al. started to investigate pyridyl-alkylidene substituted penam sulfones,⁷ e. g. SA-1-204 (**1b**) and LN-1-255 (**1c**), as inhibitors of β-lactamases. Compound SA-1-204 (**1b**) inhibits class A and class D β-lactamases efficiently,⁸ and LN-1-255 (**1c**) combined with piperacillin was found to be more active against *Escherichia coli*-DH10B strains containing extended spectrum and inhibitor-resistant β-lactamases than the clinically used combination of piperacillin and tazobactam.⁹ The mechanism of action of these β-lactamase inhibitors was elucidated by crystallographic and spectroscopic studies.¹⁰⁻¹² Very recently, derivatives of LN-1-255 (**1c**) that are substituted at the pyridine ring were synthesized and successfully tested against multidrug-resistant *Acinetobacter baumannii* in combination with the β-lactam antibiotic imipenem.¹³ Biological

1
2
3 activities of arylidene- β -lactams are, however, not limited to β -lactamase inhibition: several
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5 compounds with this structural pattern (e. g. **1e**) are active against the fungal plant pathogen
6
7 *Alternaria solani* Sorauer, the causal agent of early blight. This disease mainly affects potato
8
9 and tomato plants and is responsible for severe economic losses.¹⁴ Compound **1d** is an effective
10
11 histamin-3-receptor-(H3R)-agonist at nanomolar concentrations. Therapeutic potential of H3R-
12
13 agonists for the treatment of myocardial ischemia has been demonstrated.^{15,16}

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15
16 From a synthetic point of view arylidene-substituted β -lactams with the general formula **1** offer
17
18 the opportunity to connect other pharmacophores to the azetidin-2-one core through a covalent
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20 bond, e. g. by conjugate addition. This concept is known as pharmacophore hybridization and
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22 has been recognized as an emerging strategy for drug discovery.^{17,18} Following this concept β -
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24 lactams have been combined with purine nucleobases in search for novel antiviral agents.¹⁹

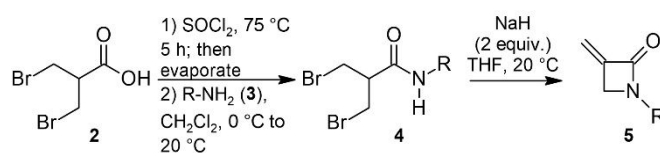
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26
27 Previously reported syntheses of arylidene- β -lactams **1**, as for example the β -lactamase
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29 inhibitors SA-1-204 (**1b**) or LN-1-255 (**1c**), rely on a Wittig-olefination of 6-oxo-penicillanic
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31 acid derivatives, which were in turn synthesized from 6-aminopenicillanic acid in a multistep
32
33 synthesis.¹³ Alternative strategies (as e. g. used for the synthesis of **1e**) proceed via a late-stage
34
35 β -lactamization of 2-aminomethyl cinnamates, which are accessible via a sequence of Baylis-
36
37 Hillman reaction, dehydrative bromination and nucleophilic substitution.^{14,20,21}

38
39
40 In continuation of previous studies from our group^{22,23} on Heck-reactions of *exo*-methylene
41
42 substituted heterocycles with arene diazonium salts (often named Matsuda-Heck-reactions²⁴⁻²⁷)
43
44 we investigated the feasibility of this approach for the regio- and stereoselective synthesis of
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46 arylidene- β -lactams. The results are disclosed herein.
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Results and discussion

Synthesis of α -methylene- β -lactam starting materials. Six α -methylene- β -lactams **5a-f** were synthesized by an adaptation of a previously published route that starts from commercially available 3-bromo-2-bromomethyl propionic acid (**2**).²⁸⁻³⁰ Carboxylic acid **2** was converted to its acid chloride by heating in thionyl chloride, which was then treated with anilines **3a-f** without prior purification. Apart from the acetamide substituted derivative **4c** all amides were isolated in fair to good yields. The yield of **4c** could not be improved by using a dimethylformamide catalyzed synthesis of acid chlorides;³¹ these conditions led to the formation of several unidentified byproducts. The resulting amides **4a-f** underwent a base-mediated intramolecular nucleophilic substitution/elimination to furnish the α -methylene- β -lactams **5a-f** in moderate overall yields upon treatment with NaH (**Table 1**).

Table 1. Synthesis of α -methylene- β -lactams **5**.



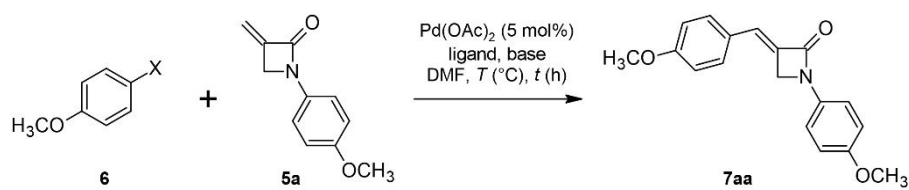
entry	3	R	4^a	yield (%) ^b	5^a	yield (%)
1	3a	4-(OCH ₃)C ₆ H ₄	4a ²⁸	50	5a ^{32,33}	92
2	3b	4-ClC ₆ H ₄	4b	52	5b ³²	63
3	3c	4-(NHAc)C ₆ H ₄	4c	13	5c	39
4	3d	3-BrC ₆ H ₄	4d	87	5d	62
5	3e	3,4-Cl ₂ C ₆ H ₃	4e ²⁸	74	5e ²⁸	74
6	3f	3-(CF ₃)-4-ClC ₆ H ₃	4f	62	5f	Quant.

^aReferences reporting characterization data. ^bYield over two steps from **2**.

Heck-reaction of α -methylene- β -lactam **5a with aryl halides and triflates.** Aryl iodides, bromides and triflates are the most commonly used electrophilic coupling partners in Mizoroki-

Heck reactions.³⁴ Conditions that have been successfully used for the arylation of electron deficient *exo*-methylene heterocycles, e. g. α -methylene- γ -butyrolactones, involve Pd(OAc)₂ as a precatalyst, DMF as a solvent and heating at elevated temperatures for up to 24 hours.³⁵ In particular electron-rich aryl halides often react slowly in Heck-reactions and do not give the desired coupling products in acceptable yields. It has been shown that in these cases the addition of tri-*ortho*-tolyl phosphine in a precatalyst-to-ligand ratio of 1 : 2 reliably accelerates the reaction and that the coupling products can be obtained in synthetically useful yields.^{36,37} We first investigated the coupling of α -methylene- β -lactam **5a** with iodo-4-methoxybenzene (**6a**) (Table 2).

Table 2. Mizoroki-Heck coupling of α -methylene- β -lactam **5a** with aryl halides.



entry	6	X	Ligand (mol %)	base (equiv.)	T (°C)	t (h)	conv. (%)	yield of 7aa (%) ^a
1	6a	I	--	NEt ₃ (3)	90	18	> 95	13
2	6a	I	P(<i>o</i> -tol) ₃ (10)	NEt ₃ (3)	20	18	< 5	n. d.
3	6a	I	P(<i>o</i> -tol) ₃ (10)	NEt ₃ (3)	90	18	> 95	10
4	6b	OTf	P(<i>o</i> -tol) ₃ (10)	NEt ₃ (3)	90	18	> 95	10
5	6c	Br	P(<i>o</i> -tol) ₃ (10)	NaOAc (3)	140	1	> 95	n. d. ^b

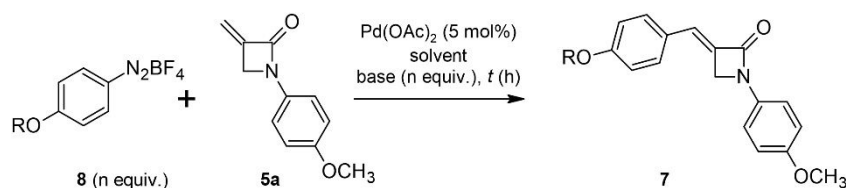
^an. d. not determined. ^bNo product detected.

Under conditions that have previously been successfully applied to Heck-reactions of electron rich aryl iodides and α -methylene- γ -butyrolactones³⁵ or α -methylenesuccinimides²² we observed full conversion of **5a**, but very low yields of coupling product **7aa** (entry 1).

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2
3 Presumably, heating the reaction mixtures at high temperatures over longer periods of time
4 causes extensive decomposition of the β -lactam starting materials due to their high ring strain.
5
6 We reasoned that activating ligands would allow a lower reaction temperature to be used.
7
8 Therefore, P(*o*-tol)₃ was added and the reaction was conducted at ambient temperature, which
9
10 led to the complete recovery of unreacted **5a** (entry 2). We then tested whether the addition of
11
12 P(*o*-tol)₃ would enhance the rate of the Heck coupling at elevated temperature to such an extent
13
14 that it could compete with the decomposition reaction, but the result was virtually identical to
15
16 that observed without any activating ligand (entry 3). An even lower yield was obtained with
17
18 the triflate **6b**³⁸ (entry 4) and no coupling product at all could be detected with bromo-4-
19
20 methoxybenzene (**6c**) (entry 5). In both cases complete consumption of the starting material **5a**
21
22 was observed.
23
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26
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28 **Optimization of coupling conditions for arene diazonium salts 8.** In contrast to Heck-type
29
30 reactions with aryl halides the addition of phosphine ligands has normally detrimental effects
31
32 when arene diazonium salts are used. It is therefore generally advisable to avoid such ligands
33
34 with these electrophilic coupling partners. However, in recent work it was demonstrated that
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36 N,N-chelating or pyridine ligands enable the control of enantioselectivity³⁹⁻⁴¹ or the use of
37
38 allylic alcohols as substrates.⁴² A more contentious issue is the role of the base in these
39
40 reactions. Examples for beneficial as well as deleterious effects of bases in Pd-catalyzed
41
42 couplings with arene diazonium salts have been reported. As Felpin et al. state, there is an
43
44 apparent correlation between the solvent used and the effect of added base.²⁵ While in
45
46 acetonitrile a base is almost always required to obtain useful conversions,⁴³ it should be avoided
47
48 if alcohols are the preferred solvents.²² It is sometimes difficult to predict whether acetonitrile
49
50 or methanol is the solvent of choice for a Matsuda-Heck reaction. This depends on the structure
51
52 of the olefin and the diazonium salt, and apart from stability issues (that take the nucleophilic
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54 nature of methanol and the sensitivity of the diazonium salt into account) the solubility of the
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56 reactants in the respective solvent plays a decisive role. For these reasons, routinely screening
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3 Matsuda-Heck couplings with hitherto unexplored olefins in acetonitrile and methanol under
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5 both basic and base-free conditions has proved successful for identifying optimized reaction
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7 conditions. We investigated two test reactions: the coupling of **5a** with 4-
8
9 methoxybenzenediazonium tetrafluoroborate (**8a**) and with 4-hydroxy-benzenediazonium
10
11 tetrafluoroborate (**8b**). We know from previous investigations that these diazonium salts often
12
13 require orthogonal reaction conditions; for example, the Pd-catalyzed coupling of **8a** with
14
15 methyl acrylate gives higher yields in the absence of a base, while the addition of NaOAc is
16
17 mandatory for phenol diazonium salt **8b**.⁴⁴ For the coupling of both diazonium salts acetonitrile
18
19 turned out to be an unsuitable solvent, because neither basic nor base-free conditions led to a
20
21 notable conversion of the starting material **5a** (entries 1,2, 6 and 7). The same result was
22
23 observed for **8a** in methanol under base-free conditions (entry 3). Upon addition of NaOAc
24
25 most of the starting material **5a** was consumed and the coupling product **7aa** was isolated in
26
27 72% yield (entry 4). Quantitative conversion and isolation of the product **7aa** in a nearly
28
29 quantitative yield was accomplished by using a slight excess of the diazonium salt (1.2 equiv.,
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31 entry 5).
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Table 3. Optimization of conditions for the coupling of **5a** and diazonium salts **8a,b**.

entry	8 (equiv.)	R	solvent	base (equiv.)	t (h)	conv.	7	yield (%)
1	8a (1.0)	CH ₃	CH ₃ CN	--	3	< 5 ^a	7aa	n. d.
2	8a (1.0)	CH ₃	CH ₃ CN	NaOAc (4.0)	3	< 5 ^a	7aa	n. d.
3	8a (1.0)	CH ₃	CH ₃ OH	--	3	< 5 ^a	7aa	n. d.
4	8a (1.0)	CH ₃	CH ₃ OH	NaOAc (4.0)	16	n. d. ^b	7aa	72
5	8a (1.2)	CH ₃	CH ₃ OH	NaOAc (4.0)	16	> 95 ^c	7aa	94 ^f
6	8b (1.0)	H	CH ₃ CN	--	3	< 5 ^a	7ab	n. d.
7	8b (1.0)	H	CH ₃ CN	NaOAc (4.0)	3	< 5 ^a	7ab	n. d.
8	8b (1.0)	H	CH ₃ OH	--	3	> 95 ^{c,d}	7ab	n. d.
9	8b (1.0)	H	CH ₃ OH	NaOAc (4.0)	3	n. d. ^b	7ab	n. d.
10 ^e	8b (1.0)	H	CH ₃ OH	--	3	> 95 ^{c,d}	7ab	n. d.
11	8b (1.2)	H	CH ₃ OH	--	12	> 95 ^{c,d}	7ab	n. d.
12	8b (1.2)	H	CH ₃ OH	NaOAc (4.0)	16	> 95 ^c	7ab	quant. ^f

^aNo product observed in ¹H NMR spectrum of crude reaction mixture. ^bIncomplete conversion; qualitatively observed by TLC. ^cNo starting material **5a** observed by TLC and ¹H NMR of crude reaction mixture. ^dFormation of unidentified side products. ^eReaction mixture cooled to 0 °C. ^fIsolated yields on a 0.25 mmol scale.

In contrast to the methoxy-substituted diazonium salt **8a** (entry 3) we observed full conversion of the starting material **5a** with phenol diazonium salt **8b** in methanol under base-free conditions (entry 8). The NMR-spectra of the crude reaction mixture show that the arylidene β-lactam **7ab** is the major product, but that at least one byproduct is formed (ca. 30%, as estimated from the integrals of the OH-protons), which could not be identified due to similar polarity and

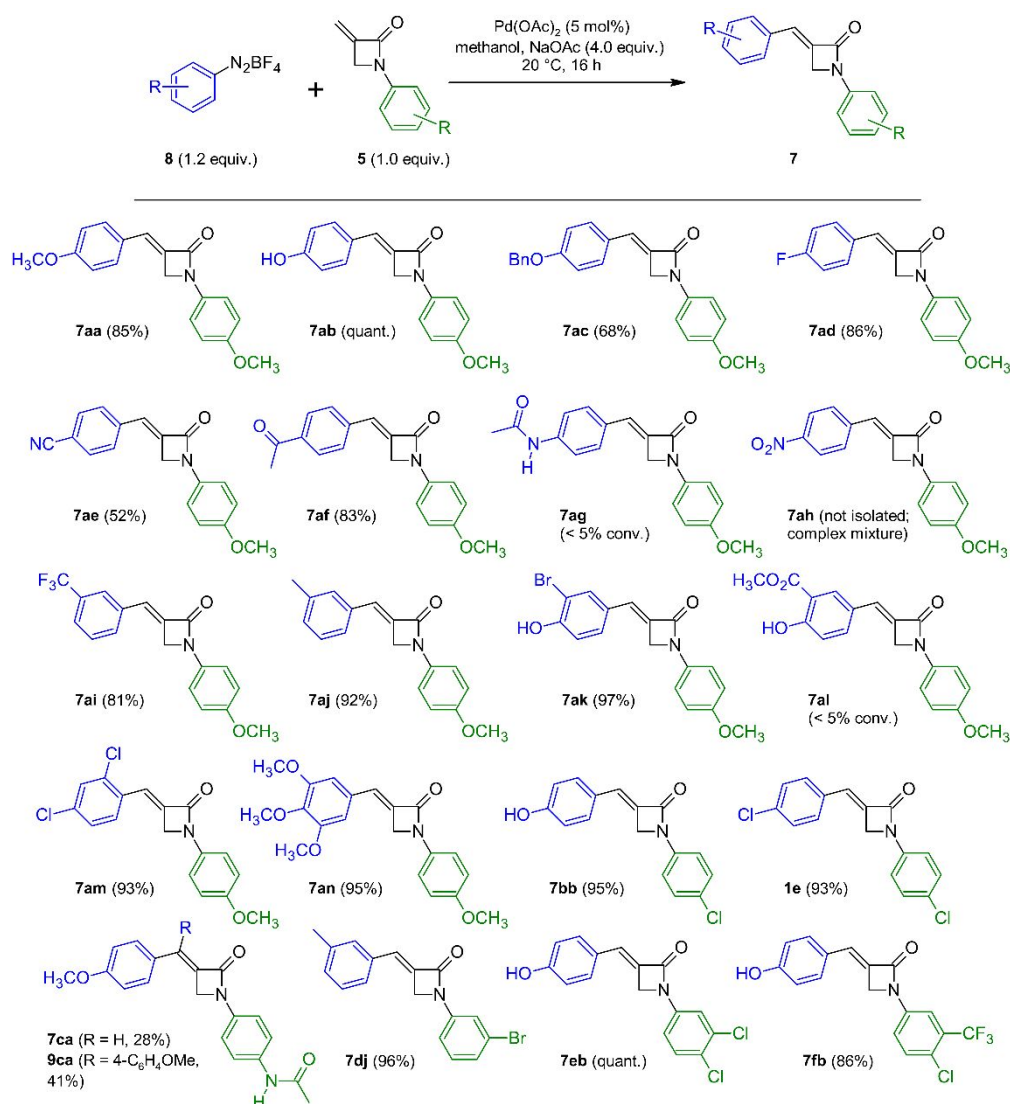
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3 overlapping signals in the aromatic and olefinic region of the ^1H NMR spectrum. Possible side
4 products are the *Z*-stereoisomer or the *endo*-regioisomer of **7ab**, but an acid-catalyzed ring
5 opening by trace amounts of water present in the reaction mixture is also conceivable, because
6 one equivalent of HBF_4 is formed during the reaction, which is not neutralized in the absence
7 of a base. Indeed, no side products were observed if the reaction was run in the presence of
8 NaOAc, but the conversion was incomplete with equimolar amounts of reactants (entry 9).
9
10 Running the reaction at lower temperature (0 °C, entry 10) or with an excess of diazonium salt
11 (1.2 equiv., entry 11) again resulted in full conversion, but also in the formation of the same
12 byproducts as observed under the conditions listed in entry 8. Quantitative conversion and high
13 selectivity were observed with added NaOAc and a slight excess of diazonium salt. Under these
14 conditions the coupling product **7ab** was obtained in quantitative isolated yield (entry 12).

15
16 The optimized conditions for the Matsuda-Heck arylation of α -methylene- β -lactams are similar
17 to those identified earlier for itaconimides²² or α -methylene- γ - or δ -lactones and lactams.²³
18
19 However, with these substrates base-free conditions are preferred. The reason why α -
20 methylene- β -lactams require added base is most likely the high tendency of the strained four-
21 membered rings to undergo hydrolytic ring opening in the presence of an acid. In contrast to
22 Heck-type arylations with aryl iodides the coupling with the corresponding diazonium salts
23 proceeds at ambient temperature, which is important to avoid decomposition of either the
24 starting β -lactams or the products under the reaction conditions.

25
26 **Substrate scope and limitations.** We applied the optimized conditions (Table 3, entries 5 and
27 12) for the coupling of **5a** and diazonium salts **8a,b** to other diazonium salts **8c-o** (see **Table 4**
28 for an overview of diazonium salts used in this study) and α -methylene- β -lactams **5b-e** (see
29 **table 1**). The results are summarized in **Table 5**.

Table 4. Arene diazonium salts used in this study.

No	R ¹	R ²	R ³	R ⁴	R ⁵	Ref.
8a	H	H	OCH ₃	H	H	45
8b	H	H	OH	H	H	44
8c	H	H	OBn	H	H	45
8d	H	H	F	H	H	46
8e	H	H	CN	H	H	47
8f	H	H	C(O)CH ₃	H	H	48
8g	H	H	NHAc	H	H	49
8h	H	H	NO ₂	H	H	50
8i	H	CF ₃	H	H	H	51
8j	H	CH ₃	H	H	H	52
8k	H	Br	OH	H	H	44
8l	H	CO ₂ CH ₃	OH	H	H	44
8m	Cl	H	Cl	H	H	22
8n	H	OCH ₃	OCH ₃	OCH ₃	H	53
8o	H	H	Cl	H	H	47

Table 5. Scope of the Matsuda-Heck arylation of α -methylene- β -lactams **5**.^{a,b}

^aThe first letter of the compound number **7xy** refers to the β -lactam **5**, the second letter refers to the diazonium salt **8** used for its synthesis. ^bCoupling of **5a** and **8a** to **7aa** was performed on a 1.0 mmol scale.

In most cases the desired coupling products were obtained in good to excellent yields as single isomers, with the following exceptions: (i) Compound **7ae**, resulting from the coupling of **5a** and 4-cyanobenzene diazonium salt **8e**, was isolated in a moderate yield of 52%, which can probably be explained by partial catalyst deactivation due to coordination of the nitrile to Pd.⁵⁴ (ii) With 4-acetamido benzene diazonium salt **8g** and 3-(methylcarboxylate) phenol diazonium salt **8l** conversions to **7ag** and **7al**, respectively, remained below 5%. We have previously obtained high yields for the coupling of both diazonium salts with methyl acrylate^{44,49} but only

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3 under base-free conditions, which are not tolerated with α -methylene- β -lactams **5**. (iii) All
4 attempts to couple **5a** with the electron deficient 4-nitrobenzene diazonium salt **8h** resulted in
5 full conversion of the starting material, but led to the formation of a complex mixture of
6 products. We have previously observed sluggish couplings with highly electron deficient
7 diazonium salts, mainly due to uncontrollable addition of the solvent methanol to the C-C-
8 double bond and hydrodediazonation of the diazonium cation.⁴⁴ (iv) In one case (the coupling
9 of **5c** with diazonium salt **8a**) we observed the formation of a mixture of coupling products
10 under standard conditions. The expected product **7ca** was isolated in a low yield of 28%,
11 whereas the double arylated product **9ca** was obtained in a somewhat higher yield of 41% even
12 though only 1.2 equiv. of diazonium salt was used. Although geminal Matsuda-Heck-
13 diarylations have occasionally been reported as side reactions^{55,56} examples for the intentional
14 double arylation are scarce and normally require special conditions or catalysts^{57,58} or at least a
15 twofold excess of diazonium salt.²² Currently, it remains unclear why the double arylated
16 compound **9ca** is the major product in this case.

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Compound **1e** (see **Figure 1** and introduction), obtained in 93% yield from the coupling of *p*-
chlorophenyl substituted β -lactam **5b** and *p*-chlorobenzene diazonium salt **8o**, has previously
been synthesized in four steps via Baylis-Hilman-reaction and intramolecular nucleophilic
substitution. In an antifungal activity assay **1e** was found to be the most active out of 28
compounds tested for their activity against the plant pathogen *Alternaria solani* Sorauer.¹⁴ All
other coupling products **7** shown in **Table 5** have not been described in the literature so far.

Structure elucidation and assignment of *exo-E*-configuration for the coupling products **7.**

Several 3-arylidene-azetidin-2-ones with the general structure **1** (**Figure 1**) have previously
been synthesized via one of the routes mentioned in the introduction. In almost all cases an *exo*-
E-structure was assigned to the reaction products.^{14,21,59-61} In a few reports describing the
synthesis of 3-arylidene-2-azetidin-2-ones via intramolecular nucleophilic substitution the *exo*-
E-structure was deduced from the double bond configuration of the acyclic precursors, but

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3 evidence for their configurational assignment was not provided.^{20,29} Mixtures of *E*- and *Z*-
4 isomers of 3-arylidene-azetidin-2-ones and 3-alkylidene-azetidin-2-ones result from olefin
5 cross metathesis reactions of α -methylene- β -lactams **5** with styrenes and 1-alkenes,
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7
8 respectively.⁶² The stereoisomers were separated and individually characterized by 1D-NMR
9
10
11 methods, but direct spectroscopic or other analytical evidence for the assigned stereochemistry
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13
14 was not given. However, in a related study on the cross metathesis of substituted α -methylene-
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16
17 β -lactones the observed *Z*-stereochemistry of the products was determined by NOESY-
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19
20 experiments, which may allow to some extent conclusions by analogy for the β -lactam cross
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22
23 metathesis products.⁶³ Examples of ring closing metathesis reactions of α -methylene- β -lactams
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26 leading to *Z*-isomers have been reported and the products have been fully characterized by
27
28
29 single crystal X-ray structure analysis.⁶⁴ This lack of direct spectroscopic and analytical proof
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31
32 for the *exo-E*-structures assigned to 3-arylidene-azetidin-2-ones prompted us to perform a
33
34
35 comprehensive NMR-spectroscopic investigation using 2D-methods for two 3-arylidene-
36
37
38 azetidin-2-ones, compounds **7ab** and **7bb**, and for the double arylated product **9ca**. A
39
40
41 combination of H,H-COSY, HSQC- and HMBC experiments allowed a full signal assignment
42
43
44 for compounds **7ab** and **7bb**, and an assignment of the most relevant signals of **9ca**. The
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46
47 numbering scheme refers to the signal assignment used in the experimental section and in the
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50 discussion (**Figure 2**).
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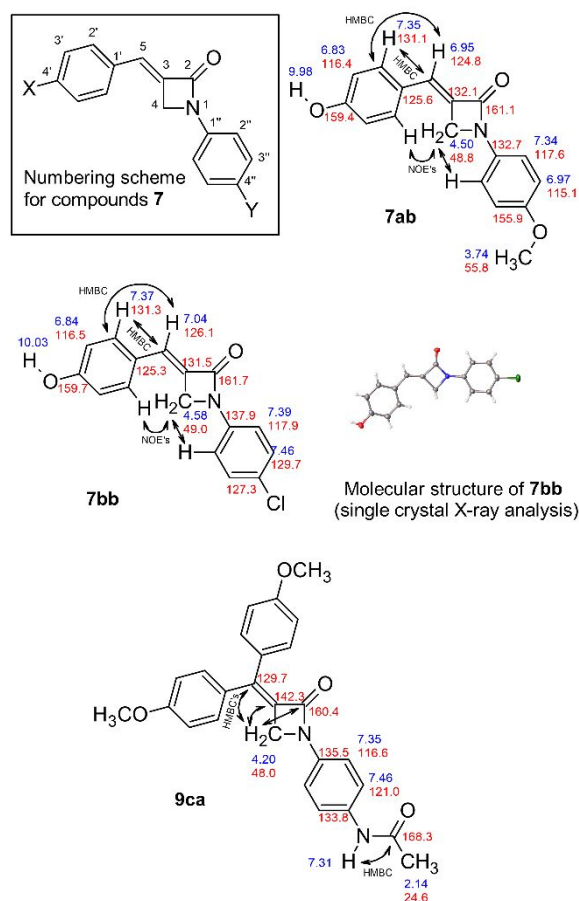


Figure 2. Numbering scheme for compounds **7**; full assignment of ^1H and ^{13}C NMR signals for compounds **7ab**, **7bb** and **9ca** and relevant HMBC and NOE interactions; molecular structure of **7bb** (single crystal X-ray analysis).

Proof for the *exo*-arrangement of the double bond in compounds **7** are HMBC interactions between H2' and C5 and C2' and H5, and HMBC interactions between H4 and C2, C3 and C5. In all compounds **7** the proton H5 appears as a triplet with a 4J -value of 1.4 Hz at ca. 7 ppm and the methylene group H4 as a doublet with the same coupling constant at ca. 4.5 to 4.6 ppm. Howell and co-workers reported very similar values for the *E*-configured cross metathesis products of *N*-Boc-protected α -methylene- β -lactam and styrenes and notably lower (ca. 6.5 ppm for H5 and ca. 4.0 ppm for H4) values for the *Z*-isomer.⁶² We observed NOE-interactions between the methylene group of the β -lactam ring (protons H4) and the *ortho*-protons of both aryl groups (protons H2' and H2''). Due to the small chemical shift differences in the aromatic region it was difficult to assign the observed NOE's unambiguously to the relevant *ortho*-

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3 protons H2'. Gratifyingly, single crystals suitable for X-ray crystallographic analysis could be
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5 obtained from a DMSO-solution of compound **7bb**. Single crystal X-ray structure analysis
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7 revealed unambiguously the assigned *exo-E*-structure. One molecule of DMSO is incorporated
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9 in the unit cell; the oxygen atom of the DMSO molecule binds to the phenolic OH-group of
10
11 **7bb** through a hydrogen bond ($d = 183$ pm).
12
13

14 The structure of **9ca** is supported by HMBC interactions between the protons of the β -lactam
15
16 methylene group (H4) and quaternary carbons C2, C3 and C5. The signals for the carbonyl
17
18 group of the β -lactam (C2) and the acetamide group can be distinguished by an HMBC
19
20 interaction between the signals for the amide proton at 7.31 ppm and the acetamide carbon at
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22 168.3.
23
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27

28 **Conclusions**

29
30 In summary, we report that 3-arylidene-azetid-2-ones, which are relevant substructures in β -
31
32 lactamase inhibitors and other bioactive molecules, can be synthesized in high yield and
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34 selectivity from α -methylene- β -lactams and arene diazonium salts in a Heck-type coupling
35
36 reaction, using Pd(OAc)₂ as precatalyst. The coupling proceeds efficiently at ambient
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38 temperature under ligand-free conditions, but requires added base to avoid acid-induced
39
40 decomposition of the β -lactams. Notably, conventional Heck-coupling conditions (aryl iodides,
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42 activating ligands, elevated temperatures) fail with α -methylene- β -lactams due to
43
44 decomposition of the olefinic coupling partner at the required reaction temperature. In contrast
45
46 to olefin cross metathesis reactions, which give 3-arylidene-azetid-2-ones as *E/Z*-mixtures
47
48 from the same starting materials, the Pd-catalyzed coupling with arene diazonium salts is highly
49
50 *E*-stereoselective.
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Experimental Section

General methods. All experiments were conducted in dry reaction vessels under an atmosphere of dry nitrogen. Solvents were purified by standard procedures. Unless otherwise stated, reaction mixtures were heated with silicon oil baths. ^1H NMR spectra were obtained at 300 MHz, 400 MHz or 500 MHz in CDCl_3 with CHCl_3 ($\delta = 7.26$ ppm) as an internal standard. Coupling constants are given in Hz. ^{13}C NMR spectra were recorded at 75 MHz, 101 MHz or 125 MHz in CDCl_3 with CDCl_3 ($\delta = 77.1$ ppm) as an internal standard. ^{19}F NMR spectra were recorded at 376 MHz. Whenever the solubility of the sample was insufficient in CDCl_3 , it was replaced by $\text{DMSO-}d_6$ ($\text{DMSO-}d_5$ as internal standard for ^1H NMR spectroscopy, $\delta = 2.50$ ppm, $\text{DMSO-}d_6$ as internal standard for ^{13}C NMR spectroscopy, $\delta = 39.5$ ppm). In all cases where signal assignments are given for ^1H - and ^{13}C -NMR data, these are based on 2D-NMR-spectra such as H,H-COSY, HSQC, HMBC and NOESY. IR spectra were recorded as ATR-FTIR spectra. Wavenumbers ($\tilde{\nu}$) are rounded to 1 cm^{-1} . The peak intensities are defined as strong (s), medium (m) or weak (w). Low- and high resolution mass spectra were obtained by EI-TOF or ESI-TOF. Although arene diazonium tetrafluoroborates are generally considered to be thermally very stable and we have never experienced any cases of violent decomposition, there have been reports of explosions caused by arene diazonium tetrafluoroborates. We recommend that these compounds are therefore handled with care and that safety measures⁶⁵ are thoroughly obeyed. All arene diazonium salts used in this work were synthesized following previously published literature procedures (**Table 4**).

General procedure for the synthesis of 3-bromo-2-bromomethylpropionic acid anilides **4**.

3-Bromo-2-bromomethylpropanoic acid (**2**, 1.24 g, 5.0 mmol) was heated in SOCl_2 (2.50 mL, 4.10 g, 34.5 mmol) at $75\text{ }^\circ\text{C}$ for 5 h. The mixture was cooled to ambient temperature and all volatiles were evaporated in vacuo. The residue was dissolved in dry and degassed CH_2Cl_2 and cooled to $0\text{ }^\circ\text{C}$. A solution of the respective aniline **3** (10.0 mmol) was added dropwise at $0\text{ }^\circ\text{C}$,

1
2
3 the mixture was allowed to warm to ambient temperature and stirred for 12 h. It was diluted
4 with CH₂Cl₂ (15 mL), washed with HCl (aq.) (2 M, 2•10 mL) and water (10 mL). The organic
5 layer was dried with MgSO₄, filtered and evaporated until the product started to precipitate.
6
7
8 The solution was stored at –18 °C to ensure complete crystallization for 12 h. The products
9
10 were isolated as colourless crystals by removing the supernatant solution. Alternatively, the
11
12 products can be purified by chromatography on silica, using hexanes-ethyl acetate mixtures of
13
14 increasing polarity as eluent.
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18
19 **3-Bromo-2-(bromomethyl)-N-(4-methoxyphenyl)propanamide (4a)**.²⁸ Following the
20
21 general procedure, **3a** (1.23 g, 10.0 mmol, 2.0 equiv.) was converted to **4a** (0.89 g, 2.5 mmol,
22
23 50%); purification by chromatography (hexanes-ethyl acetate mixtures 5 : 1 to 3 : 1 (v/v)):
24
25 colorless solid, mp 166 – 167 °C; ¹H NMR (300 MHz, CDCl₃) δ 7.42 (d, *J* = 9.0 Hz, 2H), 7.36
26
27 (s (br.), 1H), 6.88 (d, *J* = 9.0 Hz, 2H), 3.80 (s, 3H), 3.70 (dd, *J* = 10.2, 8.1 Hz, 2H), 3.58 (dd, *J*
28
29 = 10.3, 5.8 Hz, 2H), 3.09 – 3.06 (m, 1H); ¹³C {¹H} NMR (75 MHz, CDCl₃) δ 168.2, 157.3,
30
31 130.1, 122.6, 114.4, 55.7, 53.1, 31.1; IR (ATR) ν 3282 (m), 1651 (s), 1600 (m), 1542 (s), 1224
32
33 (s), 825 (s); HRMS (EI) *m/z* calcd for C₁₁H₁₃⁷⁹Br₂NO₂ [M]⁺ 348.9308, found 348.9302.
34
35
36

37
38 **3-Bromo-2-(bromomethyl)-N-(4-chlorophenyl)propanamide (4b)**. Following the general
39
40 procedure, **3b** (1.28 g, 10.0 mmol, 2.0 equiv.) was converted to **4b** (0.92 g, 2.6 mmol, 52%);
41
42 purification by chromatography (hexanes-ethyl acetate mixtures 5 : 1 to 3 : 1 (v/v)): colorless
43
44 solid, mp 160 - 161 °C; ¹H NMR (400 MHz, DMSO-*d*₆) δ 10.39 (s (br.), 1H), 7.64 (d, *J* = 8.8
45
46 Hz, 2H), 7.38 (d, *J* = 8.8 Hz, 2H), 3.73 – 3.63 (m, 4H), 3.32 – 3.23 (m, 1H), ; ¹³C {¹H} NMR
47
48 (101 MHz, DMSO-*d*₆) δ 168.4, 137.5, 128.7, 127.3, 120.9, 50.6, 32.0; IR (ATR) ν 3303 (m),
49
50 1656 (s), 1608 (s), 1544 (s), 1489 (s), 1398 (s), 824 (s); HRMS (ESI) *m/z* calcd for
51
52 C₁₀H₁₁⁷⁹Br₂³⁵ClNO [M+H]⁺ 353.8896, found 353.8904.
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57 **N-(4-Acetamidophenyl)-3-bromo-2-(bromomethyl)propanamide (4c)**. Following the
58
59 general procedure, **3c** (1.50 g, 10.0 mmol, 2.0 equiv.) was converted to **4c** (0.24 g, 0.6 mmol,
60
13%); purification by chromatography (hexanes-ethyl acetate mixtures 5 : 1 to 3 : 1 (v/v)):

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3 colorless solid, mp 198 – 200 °C; ¹H NMR (400 MHz, DMSO-*d*₆) δ 10.18 (s (br.), 1H), 9.88 (s
4 (br.), 1H), 7.58 – 7.43 (m, 4H), 3.72 – 3.62 (m, 4H), 3.30 – 3.20 (m, 1H), 2.02 (s, 3H); ¹³C{¹H}
5 NMR (101 MHz, DMSO-*d*₆) δ 168.0, 167.8, 135.2, 133.8, 119.8, 119.3, 50.5, 32.2, 23.9; IR
6 (ATR) ν 3280 (s), 1652 (s), 1549 (s), 1516 (s), 1371 (s), 1312 (m), 1127 (m), 833 (s), 715 (s);
7
8 HRMS (EI) *m/z* calcd for C₁₂H₁₄⁷⁹Br₂N₂O₂ [M]⁺ 375.9417, found 375.9411.
9

10
11
12 **3-Bromo-2-(bromomethyl)-*N*-(3-bromophenyl)propanamide (4d)**. Following the general
13 procedure, **3d** (1.72 g, 10.0 mmol, 2.0 equiv.) was converted to **4d** (1.75 g, 4.4 mmol, 87%);
14 purification by chromatography (hexanes-ethyl acetate mixtures 5 : 1 to 3 : 1 (v/v)): colorless
15 solid, mp 123 – 125 °C; ¹H NMR (400 MHz, DMSO-*d*₆) δ 10.44 (s (br.), 1H), 7.99 – 7.96 (m,
16 1H), 7.50 (ddd, *J* = 7.0, 2.1, 2.1 Hz, 1H), 7.33 – 7.24 (m, 2H), 3.73 – 3.63 (m, 4H), 3.31 – 3.23
17 (m, 1H); ¹³C{¹H} NMR (101 MHz, DMSO-*d*₆) δ 168.7, 140.1, 130.8, 126.3, 121.7, 121.6,
18 118.1, 50.7, 31.9; IR (ATR) ν 3280 (s), 1657 (s), 1589 (s), 1535 (s), 1476 (s), 1403 (m), 1344
19 (m), 1175 (m), 861 (m); HRMS (EI) *m/z* calcd for C₁₀H₁₁⁷⁹Br₃NO [M+H]⁺ 397.8391, found
20 397.8376.
21
22

23
24
25 **3-Bromo-2-(bromomethyl)-*N*-(3,4-dichlorophenyl)propanamide (4e)**.²⁸ Following the
26 general procedure, **3e** (1.62 g, 10.0 mmol, 2.0 equiv.) was converted to **4e** (1.44 g, 3.7 mmol,
27 74%); purification by chromatography (hexanes-ethyl acetate mixtures 5 : 1 to 3 : 1 (v/v)):
28 colorless solid, mp 157 - 160 °C; ¹H NMR (400 MHz, DMSO-*d*₆) δ 10.57 (s(br.), 1H), 8.00 (d,
29 *J* = 2.4 Hz, 1H), 7.59 (d, *J* = 8.8 Hz, 1H), 7.50 (dd, *J* = 8.8, 2.4 Hz, 1H), 3.74 – 3.62 (m, 4H),
30 3.32 – 3.22 (m, 1H); ¹³C{¹H} NMR (101 MHz, DMSO-*d*₆) δ 168.9, 138.6, 131.1, 130.8, 125.2,
31 120.5, 119.5, 50.7, 31.8; IR (ATR) ν 3107 (m), 1661 (s), 1607 (s), 1586 (s), 1531 (s), 1468 (s),
32 1299 (m), 820 (s); HRMS (EI) *m/z* calcd for C₁₀H₁₀⁷⁹Br₂³⁵Cl₂NO [M+H]⁺ 387.8506, found
33 387.8498.
34
35

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37 **3-Bromo-2-(bromomethyl)-*N*-[4-chloro-3-(trifluoromethyl)phenyl]propanamide (4f)**.
38 Following the general procedure, **3f** (1.96 g, 10.0 mmol, 2.0 equiv.) was converted to **4f** (1.30
39 g, 3.1 mmol, 62%); purification by chromatography (hexanes-ethyl acetate mixtures 5 : 1 to 3
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3 : 1 (v/v)): colorless solid, mp 96 - 99 °C; ¹H NMR (400 MHz, DMSO-*d*₆) δ 10.73 (s (br.), 1H),
4 8.20 (d, *J* = 2.8 Hz, 1H), 7.86 (dd, *J* = 8.8, 2.5 Hz, 1H), 7.67 (d, *J* = 8.8 Hz, 2H), 3.74 – 3.65
5 (m, 4H), 3.34 – 3.24 (m, 1H); ¹³C {¹H} NMR (101 MHz, DMSO-*d*₆) δ 169.1, 138.0, 132.3,
6 126.8 (q, *J* = 30.5 Hz), 124.4 (q, *J* = 1.8 Hz), 124.1, 122.6 (q, *J* = 272.8 Hz), 117.9 (q, *J* = 5.6
7 Hz), 50.8, 31.8; ¹⁹F NMR (377 MHz, DMSO-*d*₆) δ –61.7; IR (ATR) ν 1661 (s), 1598 (m), 1543
8 (s), 1480 (s), 1421 (m), 1319 (s), 1175 (m), 1127 (s), 1111 (s); HRMS (EI) *m/z* calcd for
9 C₁₁H₁₀⁷⁹Br₂³⁵ClF₃NO [M+H]⁺ 421.8770, found 421.8762.

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General procedure for the synthesis of α-methylene-β-lactams 5. The corresponding 3-bromo-2-bromomethyl-propionamide **4** (1.00 mmol) was dissolved in dry and degassed THF (10 mL). NaH (60 wt-% dispersion in mineral oil, 80 mg, 2.00 mmol) was added in small portions at ambient temperature and the mixture was stirred for 16 h. The reactions was quenched by addition of satd. aq. NH₄Cl solution (8 mL) and ethyl acetate (30 mL) was added. The organic layer was separated and the aqueous layer was extracted with ethyl acetate (2 x 30 mL). The combined organic layers were washed with brine (20 mL) and dried with MgSO₄, filtered and evaporated. The crude product was purified by column chromatography on silica, using hexanes-ethyl acetate mixtures of increasing polarity as eluent, to furnish the α-methylene-β-lactams **5**.

1-(4-Methoxyphenyl)-3-methyleneazetid-2-one (5a).^{32,33} Following the general procedure, anilide **4a** (4.50 g, 12.8 mmol) was converted to **5a** (2.23 g, 11.8 mmol, 92%); purification by chromatography (hexanes-ethyl acetate mixtures 10 : 1 to 5 : 1 (v/v)): off-white solid, mp 112 – 113 °C (reported in the literature:³³ mp 108 – 109 °C); ¹H NMR (400 MHz, CDCl₃) δ 7.34 (d, *J* = 9.1 Hz, 2H), 6.89 (d, *J* = 9.1 Hz, 2H), 5.83 (q, *J* = 1.7 Hz, 1H), 5.30 (q, *J* = 1.3 Hz, 1H), 4.09 (t, *J* = 1.5 Hz, 2H), 3.79 (s, 3H); ¹³C {¹H} NMR (101 MHz, CDCl₃) δ 159.8, 156.5, 143.8, 132.2, 117.8, 114.7, 110.6, 55.7, 48.0; IR (ATR) ν 1722 (s), 1512 (m), 1384 (m), 1239 (m), 825 (s); HRMS (ESI) *m/z* calcd for for C₁₁H₁₁NNaO₂ [M+Na]⁺ 212.0682, found 212.0684.

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3 **1-(4-Chlorophenyl)-3-methyleneazetid-2-one (5b)**.³² Following the general procedure,
4 anilide **4b** (355 mg, 1.00 mmol) was converted to **5b** (122 mg, 0.63 mmol, 63%); purification
5 by chromatography (hexanes-ethyl acetate mixtures 10 : 1 to 5 : 1 (v/v)): off-white solid, mp
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7
8
9
10 92 – 93 °C; ¹H NMR (300 MHz, CDCl₃) δ 7.37 - 7.29 (m, 4H), 5.90 (q, *J* = 1.7 Hz, 1H), 5.37
11 (q, *J* = 1.3 Hz, 1H), 4.13 (t, *J* = 1.5 Hz, 2H); ¹³C {¹H} NMR (126 MHz, CDCl₃) δ 160.2, 143.3,
12
13 136.9, 129.5, 129.3, 117.7, 112.0, 48.0; IR (ATR) ν 2921 (w), 1725 (s), 1491 (s), 1376 (s), 1131
14
15 (m), 826 (s); HRMS (EI) *m/z* calcd for C₁₀H₈³⁵ClNO [M⁺] 193.0289, found 193.0293.

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17
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19 **N-(4-(3-Methylene-2-oxoazetid-1-yl)phenyl)acetamide (5c)**. Following the general
20 procedure, anilide **4c** (235 mg, 0.62 mmol) was converted to **5c** (51 mg, 0.24 mmol, 39%);
21 purification by chromatography (hexanes-ethyl acetate mixtures 10 : 1 to 5 : 1 (v/v)): off-white
22 solid, mp 175 - 176 °C; ¹H NMR (500 MHz, DMSO-*d*₆) δ 9.96 (s, 1H), 7.59 (d, *J* = 8.9 Hz,
23 2H), 7.33 (d, *J* = 8.9 Hz, 2H), 5.76 (s, 1H), 5.45 (s, 1H), 4.19 (s, 2H), 2.02 (s, 3H); ¹³C {¹H}
24 NMR (126 MHz, DMSO-*d*₆) δ 168.1, 159.3, 143.7, 135.5, 133.5, 119.8, 116.6, 111.3, 47.7,
25 23.9; IR (ATR) ν 3321 (w), 1724 (s), 1677 (m), 1539 (m), 1512 (s), 826 (s); HRMS (ESI) *m/z*
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37 calcd for C₁₂H₁₂N₂O₂ [M⁺] 216.0893, found 216.0887.

38 **1-(3-Bromophenyl)-3-methyleneazetid-2-one (5d)**. Following the general procedure,
39 anilide **4d** (400 mg, 1.00 mmol) was converted to **5d** (148 mg, 0.62 mmol, 62%); purification
40 by chromatography (hexanes-ethyl acetate mixtures 10 : 1 to 5 : 1 (v/v)): colorless solid, mp
41
42
43
44
45 110 - 112 °C; ¹H NMR (600 MHz, CDCl₃) δ 7.53 - 7.48 (m, 1H), 7.36 (dt, *J* = 7.2, 1.9 Hz, 1H),
46 7.24 (dt, *J* = 7.9, 1.7 Hz, 1H), 7.22 (dd, *J* = 7.7, 7.4 Hz, 1H), 5.92 (q, *J* = 1.7 Hz, 1H), 5.39 (q,
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J = 1.4 Hz, 1H), 4.14 (dd, *J* = 1.7, 1.4 Hz, 2H); ¹³C {¹H} NMR (151 MHz, CDCl₃) δ 160.3,
143.2, 139.5, 130.8, 127.2, 123.1, 119.4, 115.2, 112.3, 48.1; IR (ATR) ν 1738 (s), 1596 (m),
1482 (m), 923 (s), 769 (s); HRMS (EI) *m/z* calcd for C₁₀H₈⁷⁹BrNO [M⁺] 236.9784, found
236.9769.

1-(3,4-Dichlorophenyl)-3-methylideneazetid-2-one (5e).²⁸ Following the general
procedure, anilide **4e** (390 mg, 1.00 mmol) was converted to **5e** (169 mg, 0.74 mmol, 74%);

1
2
3 purification by chromatography (hexanes-ethyl acetate mixtures 10 : 1 to 5 : 1 (v/v)): colorless
4
5 solid, mp 122 - 124 °C (reported in the literature: mp 128 – 129 °C)²⁸; ¹H NMR (400 MHz,
6
7 CDCl₃) δ 7.47 (d, *J* = 2.4 Hz, 1H), 7.40 (d, *J* = 8.7 Hz, 1H), 7.27 (dd, *J* = 8.7, 2.4 Hz, 1H), 5.92
8
9 (q, *J* = 1.7 Hz, 1H), 5.40 (q, *J* = 1.7 Hz, 1H), 4.13 (dd, *J* = 1.7, 1.4 Hz, 2H); ¹³C{¹H} NMR (101
10
11 MHz, CDCl₃) δ 160.2, 143.0, 137.7, 133.4, 131.1, 127.4, 118.1, 115.9, 112.7, 48.2; IR (ATR)
12
13 ν 1745 (s), 1730 (s), 1482 (s), 1394 (s), 1483 (s); HRMS (EI) *m/z* calcd for C₁₀H₇³⁵Cl₂NONa
14
15 [M+Na]⁺ 249.9797, found 249.9790.
16
17

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19 **1-[4-Chloro-3-(trifluoromethyl)phenyl]-3-methylideneazetid-2-one (5f)**. Following the
20
21 general procedure, anilide **4e** (305 mg, 0.72 mmol) was converted to **5e** (188 mg, 0.72 mmol,
22
23 quant.); purification by chromatography (hexanes-ethyl acetate mixtures 10 : 1 to 5 : 1 (v/v)):
24
25 colorless solid, mp 100 - 102 °C; ¹H NMR (400 MHz, CDCl₃) δ 7.62 - 7.56 (m, 2H), 7.47 (d, *J*
26
27 = 8.5 Hz, 1H), 5.95 (q, *J* = 1.8 Hz, 1H), 5.44 (q, *J* = 1.8 Hz, 1H), 4.18 (t, *J* = 1.4 Hz, 2H);
28
29 ¹³C{¹H} NMR (101 MHz, CDCl₃) δ 160.3, 143.0, 137.1, 132.5, 129.3 (q, *J* = 31.7 Hz), 125.3
30
31 (d, *J* = 280.5 Hz), 121.2, 120.6, 115.2 (q, *J* = 5.5 Hz), 113.0, 48.2; ¹⁹F NMR (377 MHz, CDCl₃)
32
33 δ -62.9; IR (ATR) ν 1731 (s), 1486 (s), 1436 (s), 1367 (s), 1107 (s), 934 (s), 832 (s), ; HRMS
34
35 (ESI) *m/z* calcd for C₁₁H₈³⁵ClF₃NO [M+H]⁺ 262.0241, found 262.0241.
36
37

38
39 **Heck coupling of β-lactam 5a with 4-iodoanisole (6a) to 7aa**. To a solution of **5a** (47 mg, 0.25
40
41 mmol) and 4-iodoanisole (**6a**, 70 mg, 0.30 mmol) in DMF (2.0 mL) were added NEt₃ (105 μL,
42
43 0.75 mmol), Pd(OAc)₂ (2.8 mg, 5 mol %) and optionally P(*o*-tol)₃ (7.6 mg, 10 mol %). The
44
45 solution was heated to 90 °C for 4 h and then cooled to ambient temperature. The starting
46
47 material **5a** was fully consumed, as indicated by TLC. The mixture was diluted with CH₂Cl₂
48
49 (30 mL) and washed with water (20 mL) and brine (20 mL). The organic extract was dried with
50
51 MgSO₄, filtered and evaporated. The residue was purified by column chromatography on silica,
52
53 using hexanes-ethyl acetate mixture (10 : 1 (v/v)) as eluent, to furnish **7aa** (7 mg, 0.02 mmol,
54
55 10%). Analytical data are identical to those obtained for the product of the coupling of **5a** and
56
57 diazonium salt **8a**.
58
59
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General procedure for the synthesis of α -arylidene- β -lactams **7 by Matsuda-Heck**

coupling. To a solution of the corresponding α -methylene- β -lactam **5** (0.25 mmol) and the corresponding arene diazonium salt **8** (0.30 mmol) in methanol (4 mL) was added NaOAc (82 mg, 1.00 mmol) and Pd(OAc)₂ (2.8 mg, 5 mol %). The mixture was stirred at ambient temperature for 16 h, dry-loaded on silica (by mixing with silica (1 g) and evaporating all volatiles), and purified by column chromatography on silica, using hexanes-ethyl acetate mixtures of increasing polarity as eluents to furnish the coupling products **7**.

(E)-3-(4-Methoxybenzylidene)-1-(4-methoxyphenyl)azetid-2-one (7aa). Following the general procedure, **5a** (189 mg, 1.00 mmol) and **8a** (266 mg, 1.20 mmol) were converted to **7aa** (251 mg, 0.85 mmol, 85%). For the optimization study (**Table 3**) compounds **5a** (47 mg, 0.25 mmol) and **8a** (67 mg, 0.30 mmol) were converted to **7aa** (69 mg, 0.23 mmol, 94%); purification by chromatography (hexanes-ethyl acetate mixture 10 : 1 (v/v)): colorless solid, mp 188 - 189 °C; ¹H NMR (600 MHz, DMSO-*d*₆) δ 7.47 (d, *J* = 8.8 Hz, 2H), 7.36 (d, *J* = 8.8 Hz, 2H), 7.02 (t, *J* = 1.5 Hz, 1H), 7.01 (d, *J* = 8.8 Hz, 2H), 6.99 (d, *J* = 8.8 Hz, 2H), 4.55 (d, *J* = 1.5 Hz, 2H), 3.80 (s, 3H), 3.74 (s, 3H); ¹³C{¹H} NMR (150 MHz, DMSO-*d*₆) δ 160.4, 160.3, 155.4, 132.7, 132.1, 130.5, 126.6, 124.0, 117.2, 114.6, 114.5, 55.3, 55.3, 48.3; IR (ATR) ν 2926 (w), 1720 (s), 1602 (m), 1508 (s), 1241 (s); HRMS (ESI) *m/z* calcd for C₁₈H₁₈NO₃ [M+H]⁺ 296.1287, found 296.1262.

(E)-3-(4-Hydroxybenzylidene)-1-(4-methoxyphenyl)azetid-2-one (7ab). Following the general procedure, **5a** (47 mg, 0.25 mmol) and **8b** (62 mg, 0.30 mmol) were converted to **7ab** (70 mg, 0.25 mmol, quant.); purification by chromatography (hexanes-ethyl acetate mixture 10 : 1 (v/v)): yellow solid, mp 223 - 225 °C; ¹H NMR (400 MHz, DMSO-*d*₆) δ 9.98 (s (br.), 1H, OH), 7.35 (d, *J* = 8.8 Hz, 2H, H2'), 7.34 (d, *J* = 9.1 Hz, 2H, H2''), 6.97 (d, *J* = 9.1 Hz, 2H, H3''), 6.95 (s (br.), 1H, H5), 6.83 (d, *J* = 8.6 Hz, 2H, H3'), 4.50 (s(br.), 2H, H4), 3.74 (s, 3H, OCH₃); ¹³C{¹H} NMR (101 MHz, DMSO-*d*₆) δ 160.7 (C2), 158.9 (C4'), 155.5 (C4''), 132.3 (C1''), 131.6 (C3), 130.7 (C2'), 125.2 (C1'), 124.4 (C5), 117.2 (C2''), 116.0 (C3'), 114.6

(C3''), 55.4 (OCH₃), 48.4 (C4); IR (ATR) ν 3072 (bw), 2923 (w), 1697 (s), 1604 (m), 1581 (m), 1508 (s); HRMS (ESI) m/z calcd for C₁₇H₁₆NO₃ [M+H]⁺ 282.1130, found 282.1109.

(E)-3-(4-(Benzyloxy)benzylidene)-1-(4-methoxyphenyl)azetid-2-one (7ac). Following the general procedure, **5a** (47 mg, 0.25 mmol) and **8c** (89 mg, 0.30 mmol) were converted to **7ac** (63 mg, 0.17 mmol, 68%); purification by chromatography (hexanes-ethyl acetate mixture 10 : 1 (v/v)): off-white solid, mp 209 - 211 °C; ¹H NMR (500 MHz, CDCl₃) δ 7.46 - 7.30 (m, 7H), 7.32 (d, J = 8.9 Hz, 2H), 7.02 (t, J = 1.4 Hz, 1H), 6.99 (dm, J = 8.8 Hz, 2H), 6.91 (dm, J = 9.0 Hz, 2H), 5.10 (s, 2H), 4.41 (d, J = 1.4 Hz, 2H), 3.80 (s, 3H); ¹³C {¹H} NMR (126 MHz, CDCl₃) δ 161.3, 159.9, 156.1, 136.6, 132.5, 132.4, 130.4, 128.8, 128.3, 127.6, 127.4, 124.7, 117.5, 115.6, 114.7, 70.2, 55.7, 48.7; IR (ATR) ν 1719 (s), 1602 (m), 1508 (s), 1381 (s), 1241 (s); HRMS (EI) m/z calcd for C₂₄H₂₁NO₃ [M⁺] 371.1521, found 371.1526.

(E)-3-(4-Fluorobenzylidene)-1-(4-methoxyphenyl)azetid-2-one (7ad). Following the general procedure, **5a** (47 mg, 0.25 mmol) and **8d** (63 mg, 0.30 mmol) were converted to **7ad** (61 mg, 0.22 mmol, 86%); purification by chromatography (hexanes-ethyl acetate mixture 10 : 1 (v/v)): off-white solid, mp 147 - 148 °C; ¹H NMR (300 MHz, CDCl₃) δ 7.41 - 7.33 (m, 4H), 7.10 (dd, J = 8.6, 8.6 Hz, 2H), 7.04 (t, J = 1.5 Hz, 1H), 6.92 (dm, J = 9.0 Hz, 2H), 4.43 (d, J = 1.5 Hz, 2H), 3.81 (s, 3H); ¹³C {¹H} NMR (75 MHz, CDCl₃) δ 163.4 (d, J = 250.9 Hz), 160.8, 156.4, 134.6 (d, J = 2.5 Hz), 132.4, 130.7 (d, J = 3.4 Hz), 130.6 (d, J = 8.5 Hz), 123.9, 117.7, 116.4 (d, J = 21.9 Hz), 114.8, 55.7, 48.7; IR (ATR) ν 1727 (m), 1598 (w), 1505 (m), 1135 (m), 831 (s); HRMS (EI) m/z calcd for C₁₇H₁₄FNO₂ [M⁺] 283.1009, found 283.1002.

(E)-4-((1-(4-Methoxyphenyl)-2-oxoazetid-3-ylidene)methyl)benzonitrile (7ae).

Following the general procedure, **5a** (47 mg, 0.25 mmol) and **8e** (65 mg, 0.30 mmol) were converted to **7ae** (38 mg, 0.13 mmol, 52%); purification by chromatography (hexanes-ethyl acetate mixture 10 : 1 (v/v)): off-white solid, mp 200 °C (dec.); ¹H NMR (400 MHz, DMSO-*d*₆) δ 7.91 (d, J = 8.0 Hz, 2H), 7.70 (d, J = 7.9 Hz, 2H), 7.38 (d, J = 8.7 Hz, 2H), 7.18 (s, 1H), 7.00 (d, J = 8.6 Hz, 2H), 4.64 (s, 2H), 3.75 (s, 3H); ¹³C {¹H} NMR (101 MHz, DMSO-*d*₆) δ

159.9, 156.3, 139.5, 139.2, 133.3, 132.2, 129.8, 123.0, 119.1, 118.0, 115.1, 111.8, 55.8, 49.1; IR (ATR) ν 2223 (m), 1717 (s), 1511 (m), 1242 (m), 1143 (m); HRMS (EI) m/z calcd for $C_{18}H_{14}N_2O_2$ [M^+] 290.1050, found 290.1053.

(E)-3-(4-Acetylbenzylidene)-1-(4-methoxyphenyl)azetidin-2-one (7af). Following the general procedure, **5a** (47 mg, 0.25 mmol) and **8f** (70 mg, 0.30 mmol) were converted to **7af** (64 mg, 0.21 mmol, 83%); purification by chromatography (hexanes-ethyl acetate mixture 10 : 1 (v/v)): yellow solid, mp 207 – 208 °C; 1H NMR (300 MHz, $CDCl_3$) δ 7.98 (dm, $J = 8.4$ Hz, 2H), 7.47 (dm, $J = 8.4$ Hz, 2H), 7.39 (dm, $J = 9.0$ Hz, 2H), 7.10 (t, $J = 1.5$ Hz, 1H), 6.92 (dm, $J = 9.0$ Hz, 2H), 4.48 (d, $J = 1.5$ Hz, 2H), 3.81 (s, 3H), 2.62 (s, 3H); ^{13}C { 1H } NMR (75 MHz, $CDCl_3$) δ 197.3, 160.3, 156.6, 138.9, 137.7, 137.4, 132.2, 129.1, 128.9, 123.8, 117.8, 114.8, 55.7, 48.9, 26.8; IR (ATR) ν 1720 (s), 1675 (s), 1603 (w), 1511 (s), 1244 (s); HRMS (EI) m/z calcd for $C_{19}H_{17}NO_3$ [M^+] 307.1203, found 307.1205.

(E)-1-(4-Methoxyphenyl)-3-(3-(trifluoromethyl)benzylidene)azetidin-2-one (7ai). Following the general procedure, **5a** (47 mg, 0.25 mmol) and **8i** (78 mg, 0.30 mmol) were converted to **7ai** (67 mg, 0.20 mmol, 81%); purification by chromatography (hexanes-ethyl acetate mixture 10 : 1 (v/v)): yellow solid, mp 175 - 177 °C; 1H NMR (300 MHz, $CDCl_3$): δ 7.65 - 7.49 (m, 4H), 7.39 (dm, $J = 9.0$ Hz, 2H), 7.09 (t, $J = 1.5$ Hz, 1H), 6.92 (dm, $J = 9.0$ Hz, 2H), 4.47 (d, $J = 1.5$ Hz, 2H), 3.81 (s, 3H); ^{13}C { 1H } NMR (75 MHz, $CDCl_3$) δ 160.2, 156.5, 136.9, 135.2, 132.0, 131.8, 131.7 (q, $J = 32.0$ Hz) 129.7, 125.9 (q, $J = 3.6$ Hz), 125.1 (q, $J = 3.8$ Hz), 123.9 (q, $J = 273$ Hz), 123.4, 117.7, 114.7, 55.6, 48.7; IR (ATR) ν 1721 (m), 1512 (m), 1322 (m), 1123 (s), 695 (m); HRMS (EI) m/z calcd for $C_{18}H_{14}F_3NO_2$ [M^+] 333.0977, found 333.0969.

(E)-1-(4-Methoxyphenyl)-3-(3-methylbenzylidene)azetidin-2-one (7aj). Following the general procedure, **5a** (47 mg, 0.25 mmol) and **8j** (62 mg, 0.30 mmol) were converted to **7aj** (64 mg, 0.23 mmol, 92%); purification by chromatography (hexanes-ethyl acetate mixture 10 : 1 (v/v)): yellow solid, mp 151 - 152 °C; 1H NMR (600 MHz, $DMSO-d_6$) δ 7.38 (dm, $J = 9.0$

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3 Hz, 2H), 7.35 - 7.30 (m, 3H), 7.22 (dm, $J = 7.2$ Hz, 1H), 7.02 (t, $J = 1.4$ Hz, 1H), 6.99 (dm, $J =$
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5 9.1 Hz, 2H), 4.59 (d, $J = 1.4$ Hz, 2H), 3.74 (s, 3H), 2.34 (s, 3H); $^{13}\text{C}\{^1\text{H}\}$ NMR (151 MHz,
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7 DMSO- d_6) δ 160.2, 155.6, 138.3, 135.3, 134.1, 132.0, 130.2, 129.4, 129.0, 126.0, 124.3, 117.4,
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9 114.6, 55.4, 48.6, 21.0; IR (ATR) ν 1726 (s), 1583 (w), 1510 (s), 1239 (s), 1142 (s); HRMS
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11 (EI) m/z calcd for $\text{C}_{18}\text{H}_{17}\text{NO}_2$ [M^+] 279.1254, found 279.1262.

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15 **(E)-3-(3-Bromo-4-hydroxybenzylidene)-1-(4-methoxyphenyl)azetid-2-one (7ak).**

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17 Following the general procedure, **5a** (47 mg, 0.25 mmol) and **8k** (86 mg, 0.30 mmol) were
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19 converted to **7ak** (87 mg, 0.24 mmol, 97%); purification by chromatography (hexanes-ethyl
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21 acetate mixture 10 : 1 (v/v)): yellow solid, mp 240 - 242 °C (dec.); ^1H NMR (500 MHz, DMSO-
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23 d_6) δ 10.81 (s, 1H), 7.66 (d, $J = 1.8$ Hz, 1H), 7.38 - 7.33 (m, 3H), 7.05 - 6.94 (m, 4H), 4.55 (s,
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25 2H), 3.74 (s, 3H); $^{13}\text{C}\{^1\text{H}\}$ NMR (126 MHz, DMSO- d_6) δ 160.2, 155.5, 155.3, 133.5, 133.2,
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27 132.1, 129.3, 126.8, 123.0, 117.2, 116.7, 114.6, 109.9, 55.3, 48.2; IR (ATR) ν 3158 (bw), 1704
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29 (s), 1602 (m), 1510 (s), 1250 (s), 819 (s); HRMS (EI) m/z calcd for $\text{C}_{17}\text{H}_{14}^{79}\text{BrNO}_3$ [M^+]
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31 359.0152, found: 359.0141.

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35 **(E)-3-(2,4-Dichlorobenzylidene)-1-(4-methoxyphenyl)azetid-2-one (7am).** Following the
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37 general procedure, **5a** (47 mg, 0.25 mmol) and **8m** (78 mg, 0.30 mmol) were converted to **7am**
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39 (78 mg, 0.23 mmol, 93%); purification by chromatography (hexanes-ethyl acetate mixture 10 :
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41 1 (v/v)): off-white solid, mp 217 - 218 °C; ^1H NMR (500 MHz, CDCl_3) δ 7.49 (d, $J = 2.1$ Hz,
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43 1H), 7.45 (t, $J = 1.4$ Hz, 1H), 7.39 (dm, $J = 9.0$ Hz, 2H), 7.35 (d, $J = 8.4$ Hz, 1H), 7.28 (dd, $J =$
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45 8.4, 2.1 Hz, 1H), 6.93 (dm, $J = 9.0$ Hz, 2H), 4.42 (d, $J = 1.5$ Hz, 2H), 3.82 (s, 3H); $^{13}\text{C}\{^1\text{H}\}$
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47 NMR (126 MHz, CDCl_3) δ 160.1, 156.5, 137.6, 135.9, 135.7, 132.1, 131.0, 130.4, 128.8, 127.6,
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49 120.3, 117.8, 114.7, 55.7, 48.6; IR (ATR) ν 2923 (w), 1725 (s), 1709 (s), 1511 (s), 1245 (s),
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51 808 (s); HRMS (EI) m/z calcd for $\text{C}_{17}\text{H}_{13}^{35}\text{Cl}_2\text{NO}_2$ [M^+] 333.0323, found: 333.0335.

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56 **(E)-1-(4-Methoxyphenyl)-3-(3,4,5-trimethoxybenzylidene)azetid-2-one (7an).** Following
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58 the general procedure, **5a** (47 mg, 0.25 mmol) and **8n** (85 mg, 0.30 mmol) were converted to
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60 **7an** (84 mg, 0.24 mmol, 95%); purification by chromatography (hexanes-ethyl acetate mixture

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3 10 : 1 (v/v): off-white solid, mp 150 - 152 °C; ¹H NMR (300 MHz, CDCl₃) δ 7.36 (dm, *J* = 9.1
4 Hz, 2H), 6.95 (t, *J* = 1.4 Hz, 1H), 6.88 (dm, *J* = 9.1 Hz, 2H), 6.57 (s, 2H), 4.41 (d, *J* = 1.4 Hz,
5 2H), 3.89 (s, 6H), 3.88 (s, 3H), 3.78 (s, 3H); ¹³C{¹H} NMR (75 MHz, CDCl₃) δ 160.8, 156.3,
6 153.7, 139.8, 134.1, 132.3, 129.9, 125.1, 117.6, 114.7, 106.3, 61.1, 56.4, 55.6, 48.5; IR (ATR)
7 ν 2948 (m), 1718 (s), 1583 (m), 1505 (s), 1120 (s), 726 (s); HRMS (EI) *m/z* calcd for C₂₀H₂₁NO₅
8 [M⁺] 355.1420, found 355.1430.

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17 **(*E*)-1-(4-Chlorophenyl)-3-(4-hydroxybenzylidene)azetidin-2-one (7bb)**. Following the
18 general procedure, **5b** (48 mg, 0.25 mmol) and **8b** (62 mg, 0.30 mmol) were converted to **7bb**
19 (68 mg, 0.24 mmol, 95%); purification by chromatography (hexanes-ethyl acetate mixture 10 :
20 1 (v/v)): off-white solid, mp 249 - 251 °C; ¹H NMR (400 MHz, DMSO-*d*₆) δ 10.03 (s, 1H, OH),
21 7.45 (d, *J* = 8.9 Hz, 2H, H3''), 7.39 (d, *J* = 8.9 Hz, 2H, H2''), 7.37 (d, *J* = 8.5 Hz, 2H, H2'),
22 7.04 (s, 1H, H5), 6.84 (d, *J* = 8.5 Hz, 2H, H3'), 4.58 (s, 2H, H4); ¹³C{¹H} NMR (101 MHz,
23 DMSO-*d*₆) δ 161.7 (C2), 159.7 (C4'), 137.9 (C1''), 131.5 (C3), 131.3 (C2'), 129.7 (C3''), 127.3
24 (C4''), 126.1 (C5), 125.3 (C1'), 117.9 (C2''), 116.5 (C3'), 49.0 (C4); IR (ATR) ν 3116 (bw),
25 2924 (w), 1705 (s), 1492 (s), 824 (s); HRMS (ESI) *m/z* calcd for C₁₆H₁₃³⁵CINO₂ [M+H]⁺
26 286.0635, found 286.0660.

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40 **(*E*)-3-(4-Chlorobenzylidene)-1-(4-chlorophenyl)azetidin-2-one (1e)**.¹⁴ Following the
41 general procedure, **5b** (48 mg, 0.25 mmol) and **8o** (68 mg, 0.30 mmol) were converted to **1e**
42 (71 mg, 0.24 mmol, 93%); purification by chromatography (hexanes-ethyl acetate mixture 10 :
43 1 (v/v)): off-white solid, mp 140 – 141 °C (reported in the literature¹⁴ mp 139 - 142 °C); ¹H
44 NMR (600 MHz, CDCl₃) δ 7.39 (d, *J* = 8.5 Hz, 2H), 7.37 (d, *J* = 8.9 Hz, 2H), 7.33 (d, *J* = 8.9
45 Hz, 2H), 7.32 (d, *J* = 8.5 Hz, 2H), 7.08 (t, *J* = 1.4 Hz, 1H), 4.46 (d, *J* = 1.5 Hz, 2H); ¹³C{¹H}
46 NMR (126 MHz, CDCl₃) δ 161.0, 137.1, 135.9, 134.9, 132.6, 130.1, 129.6, 129.5, 129.2, 125.0,
47 117.6, 48.7; IR (ATR) ν 1735 (s), 1591 (s), 1492 (m), 1480 (s), 1373 (s), 1123 (m); HRMS (EI)
48 *m/z* calcd for C₁₆H₁₁³⁵Cl₂NO [M⁺] 303.0218, found 303.0224. All analytical data match those
49 reported in the literature.¹⁴
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(E)-N-(4-(3-(4-Methoxybenzylidene)-2-oxoazetidin-1-yl)phenyl)acetamide (7ca) and N-(4-(3-(bis(4-methoxyphenyl)methylene)-2-oxoazetidin-1-yl)phenyl)acetamide (9ca).

Following the general procedure, **5c** (54 mg, 0.25 mmol) and **8a** (67 mg, 0.30 mmol) were converted to a mixture of **7ca** (23 mg, 0.07 mmol, 28%) and **9ca** (44 mg, 0.10 mmol, 41%).

The reaction products were separated by column chromatography on silica (hexanes-ethyl acetate mixtures 10 : 1 to 3 : 1 (v/v)). *Analytical data for 7ca*: off-white solid, mp 234 °C (dec.);

¹H NMR (600 MHz, DMSO-*d*₆) δ 9.96 (s, 1H), 7.59 (dm, *J* = 9.0 Hz, 2H), 7.47 (dm, *J* = 8.8 Hz, 2H), 7.34 (dm, *J* = 8.9 Hz, 2H), 7.04 (t, *J* = 1.4 Hz, 1H), 7.01 (dm, *J* = 8.8 Hz, 2H), 4.56 (d, *J* = 1.4 Hz, 2H), 3.80 (s, 3H), 2.02 (s, 3H); ¹³C{¹H} NMR (151 MHz, DMSO-*d*₆) δ 168.1, 160.6, 160.4, 135.2, 134.0, 132.6, 130.6, 126.7, 124.4, 119.9, 116.3, 114.6, 55.4, 48.3, 24.0; IR (ATR) ν 3310 (bw), 2955 (m), 2913 (m), 1725 (m), 1709 (m), 1600 (m), 1509 (s), 1248 (s), 823 (s); HRMS (EI) *m/z* calcd for C₁₉H₁₈N₂O₃ [M⁺] 322.1317, found 322.1314. *Analytical data for*

9ca: off-white solid, mp 239 – 241 °C; ¹H NMR (600 MHz, CDCl₃) δ 7.50 (dm, *J* = 8.9 Hz, 2H, *Z* or *E*-H3'), 7.46 (dm, *J* = 8.9 Hz, 2H, H3''), 7.35 (dm, *J* = 8.9 Hz, 2H, H2''), 7.31 (s(br.), 1H, NH), 7.23 (dm, *J* = 8.8 Hz, 2H, *Z* or *E*-H3'), 6.92 (dm, *J* = 8.9 Hz, 2H, *Z* or *E*-H2'), 6.90 (dm, *J* = 8.9 Hz, 2H, *Z* or *E*-H2'), 4.20 (s, 2H, H4), 3.85 (s, 3H, *Z* or *E*-OCH₃), 3.83 (s, 3H, *Z* or *E*-OCH₃), 2.14 (s, 3H, C(O)CH₃); ¹³C{¹H} NMR (151 MHz, CDCl₃) δ 168.3 (C(O)CH₃), 160.4 (C2), 160.3 (*Z*- or *E*-C4'), 160.1 (*Z*- or *E*-C4'), 142.3 (C3), 135.5 (C1''), 133.8 (C4''), 131.8 (*Z*- or *E*-C3'), 131.5 (C1'), 130.8 (*Z*- or *E*-C3'), 129.7 (*Z*- or *E*-C1' or C5), 129.6 (*Z*- or *E*-C1' or C5), 121.0 (C3''), 116.6 (C2''), 114.1 (*Z*- or *E*-C3'), 113.5 (*Z*- or *E*-C3'), 55.5 (*Z*- or *E*-OCH₃), 55.4 (*Z*- or *E*-OCH₃), 48.0 (C4), 24.6 (C(O)CH₃); IR (ATR) ν 3307 (bw), 1712 (w), 1667 (w), 1603 (m), 1506 (s), 1246 (s), 828 (s); HRMS (EI) *m/z* calcd for C₂₆H₂₄N₂O₄ [M⁺] 428.1736, found 428.1735.

(E)-1-(3-Bromophenyl)-3-(3-methylbenzylidene)azetidin-2-one (7dj). Following the general procedure, **5d** (60 mg, 0.25 mmol) and **8j** (62 mg, 0.30 mmol) were converted to **7dj** (79 mg, 0.24 mmol, 96%); purification by chromatography (hexanes-ethyl acetate mixture 10 :

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3 1 (v/v): off-white solid, mp 146 – 147 °C; ¹H NMR (300 MHz, CDCl₃) δ 7.56 - 7.52 (m, 1H),
4 7.45 - 7.37 (m, 1H), 7.35 - 7.27 (m, 1H), 7.24 - 7.16 (m, 5H), 7.10 (t, *J* = 1.4 Hz, 1H), 4.47 (d,
5 *J* = 1.5 Hz, 2H), 2.39 (s, 3H); ¹³C{¹H} NMR (75 MHz, CDCl₃) δ 161.5, 139.9, 139.0, 134.0,
6 134.0, 130.8, 130.8, 129.8, 129.2, 126.9, 126.8, 126.1, 123.2, 119.1, 115.1, 48.9, 21.6; IR
7 (ATR) ν 1726 (s), 1590 (s), 1568 (m), 1480 (s), 1371 (s), 773 (s); HRMS (EI) *m/z* calcd for
8 C₁₇H₁₄⁷⁹BrNO [M⁺] 327.0259, found 327.0255.

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17 **(*E*)-1-(3,4-Dichlorophenyl)-3-(4-hydroxybenzylidene)azetidin-2-one (7eb)**. Following the
18 general procedure, **5e** (57 mg, 0.25 mmol) and **8b** (62 mg, 0.30 mmol) were converted to **7eb**
19 (79 mg, 0.25 mmol, quant.); purification by chromatography (hexanes-ethyl acetate mixture 10
20 : 1 (v/v)): off-white solid, mp 216 °C (dec.); ¹H NMR (400 MHz, DMSO-*d*₆) δ 10.03 (s, 1H),
21 7.64 (d, *J* = 8.7 Hz, 1H), 7.59 (s, 1H), 7.45 - 7.31 (m, 3H), 7.07 (s, 1H), 6.84 (d, *J* = 8.0 Hz,
22 2H), 4.59 (s, 2H); ¹³C{¹H} NMR (101 MHz, DMSO-*d*₆) δ 161.8, 159.8, 138.9, 132.2, 131.7,
23 131.4, 131.2, 126.7, 125.2, 125.1, 117.7, 116.6, 116.5, 49.3; IR (ATR) ν 3356 (bw), 1743 (s),
24 1727 (s), 1705 (s), 1592 (m), 1479 (s), 1134 (s), 812 (s); HRMS (EI) *m/z* calcd for
25 C₁₆H₁₁³⁵Cl₂NO₂ [M⁺] 319.0161, found 319.0162.

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38 **(*E*)-1-(4-Chloro-3-(trifluoromethyl)phenyl)-3-(4-hydroxybenzylidene)azetidin-2-one**
39 **(7fb)**. Following the general procedure, **5f** (65 mg, 0.25 mmol) and **8b** (62 mg, 0.30 mmol)
40 were converted to **7fb** (76 mg, 0.22 mmol, 86%); purification by chromatography (hexanes-
41 ethyl acetate mixture 10 : 1 (v/v)): off-white solid, mp 207 - 208 °C; ¹H NMR (400 MHz,
42 DMSO-*d*₆) δ 10.04 (s, 1H), 7.76 (s, 1H), 7.74 (d, *J* = 8.9 Hz, 1H), 7.65 (d, *J* = 8.9 Hz, 1H), 7.38
43 (d, *J* = 7.9 Hz, 2H), 7.10 (s, 1H), 6.84 (d, *J* = 7.9 Hz, 2H), 4.65 (s, 2H); ¹³C{¹H} NMR (101
44 MHz, DMSO-*d*₆) δ 161.5, 159.4, 137.8, 132.7, 131.0, 130.6, 127.3 (q, *J* = 30.9 Hz), 126.5,
45 124.7, 123.7 (q, *J* = 2.0 Hz), 122.6 (q, *J* = 275 Hz), 120.8, 116.0, 114.6 (q, *J* = 5.6 Hz), 49.3;
46 ¹⁹F NMR (376 MHz, DMSO-*d*₆) δ -61.6; IR (ATR) ν 3289 (bw), 1730 (m), 1722 (s), 1605 (m),
47 1519 (w), 1484 (s), 1108 (s); HRMS (EI) *m/z* calcd for C₁₇H₁₁³⁵ClF₃NO₂ [M⁺] 353.0425, found:
48 353.0418.
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Supporting Information Available statement

The Supporting Information is available free of charge on the ACS Publications website at DOI:

Copies of ^1H and ^{13}C NMR spectra for all compounds; 2D-NMR-spectra for representative compounds.

Accession Codes

CCDC 2068052 contains the supplementary crystallographic data for this paper. These data can be obtained free of charge via www.ccdc.cam.ac.uk/data_request/cif, or by emailing data_request@ccdc.cam.ac.uk, or by contacting The Cambridge Crystallographic Data Centre, 12 Union Road, Cambridge CB2 1EZ, UK; fax: +44 1223 336033.

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