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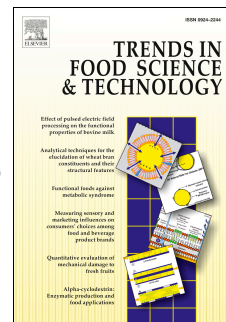
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Starch-based food matrices containing protein: Recent understanding of morphology, structure, and properties

Binjia Zhang, Dongling Qiao, Siming Zhao, Qinlu Lin, Jing Wang, Fengwei Xie



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1 Starch-based food matrices containing protein: Recent
2 understanding of morphology, structure, and properties

3

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49 **Abbreviations**

50	CLSM	Confocal laser scanning microscopy
51	DSC	Differential scanning calorimetry
52	FTIR	Fourier-transform infrared
53	G'	Storage modulus
54	G''	Loss modulus
55	HMT	Heat-moisture treatment
56	pI	Isoelectric point
57	RS	Resistant starch
58	RVA	Rapid visco-analyzer
59	SAXS	Small-angle X-ray scattering
60	SEM	Scanning electron microscopy
61	TGase	Transglutaminase
62	T_p	Peak temperature
63	WHC	Water-holding capacity
64	XRD	X-ray diffraction
65	ΔH	Enthalpy change

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67 Abstract

68 *Background:* Starches and proteins are two major types of biopolymer components, especially in
69 many flour (starch)-based foods consumed worldwide, which provide energy and nutrition needed by
70 the human body. In many such starch-based matrices (the main structural component of such foods),
71 proteins and their interactions with starches greatly influence the matrix structure and properties.

72 Studying the different roles played by proteins (endogenous and exogenous) in various starch-based
73 food systems can provide a frame of reference for the design and production of improved
74 starch-based food products with tailored properties and desirable nutritional functions.

75 *Scope and approach:* Significant efforts have recently been made to tailor the morphology, structure,
76 and properties of many starch-based food systems, and thus to design various starch-based food
77 products with satisfactory attributes. This review surveys the latest literature on starch-based
78 matrices containing proteins. Discussed are the influences of proteins and their interactions with
79 starches on the morphologies and structures (e.g. short- and long-range orders) of starch-based
80 matrices, as well as on their pasting, thermal, rheological, textural, sensory, and digestive properties.
81 Also, current understandings of structure–property links are presented, along with their implications
82 on the production of various starchy foods (e.g. pastas, breads, cakes, and biscuits), including
83 gluten-free versions.

84 *Key findings and conclusions:* Proteins in many starchy food matrices can encapsulate the starch
85 phase (or be adsorbed on its surfaces) on a micron scale, and thereby interact with starch chains via
86 both non-covalent (e.g. hydrogen bonding, hydrophobic, and electrostatic) and covalent bonds (e.g.

87 via Maillard reactions). These facts and protein features (e.g. hydration and gelation abilities) can
88 play major roles in inhibiting starch retrogradation (the reassembly of cooked starch chains into
89 ordered structures) and in regulating various other properties of such starch-based matrices,
90 including viscosity, transition temperatures, moduli, hardness, sensory, digestibility, and shelf-life.
91 Despite the fact that the current literature presents considerable information on the structure–property
92 relationships of many different starch-based matrices and their applications in the processing of
93 various starchy foods (e.g. pastas, noodles, and biscuits), it is still highly necessary to define more
94 comprehensive correlations among starch–protein interactions, starch-protein matrix structures, and
95 the resulting properties of such food products.

96

97 Keywords: Starch–protein interactions; Starch-based food processing; Starch structure; Food textural
98 properties; Food sensory properties; Starch digestibility; Gluten-free products

99 **1. Introduction**

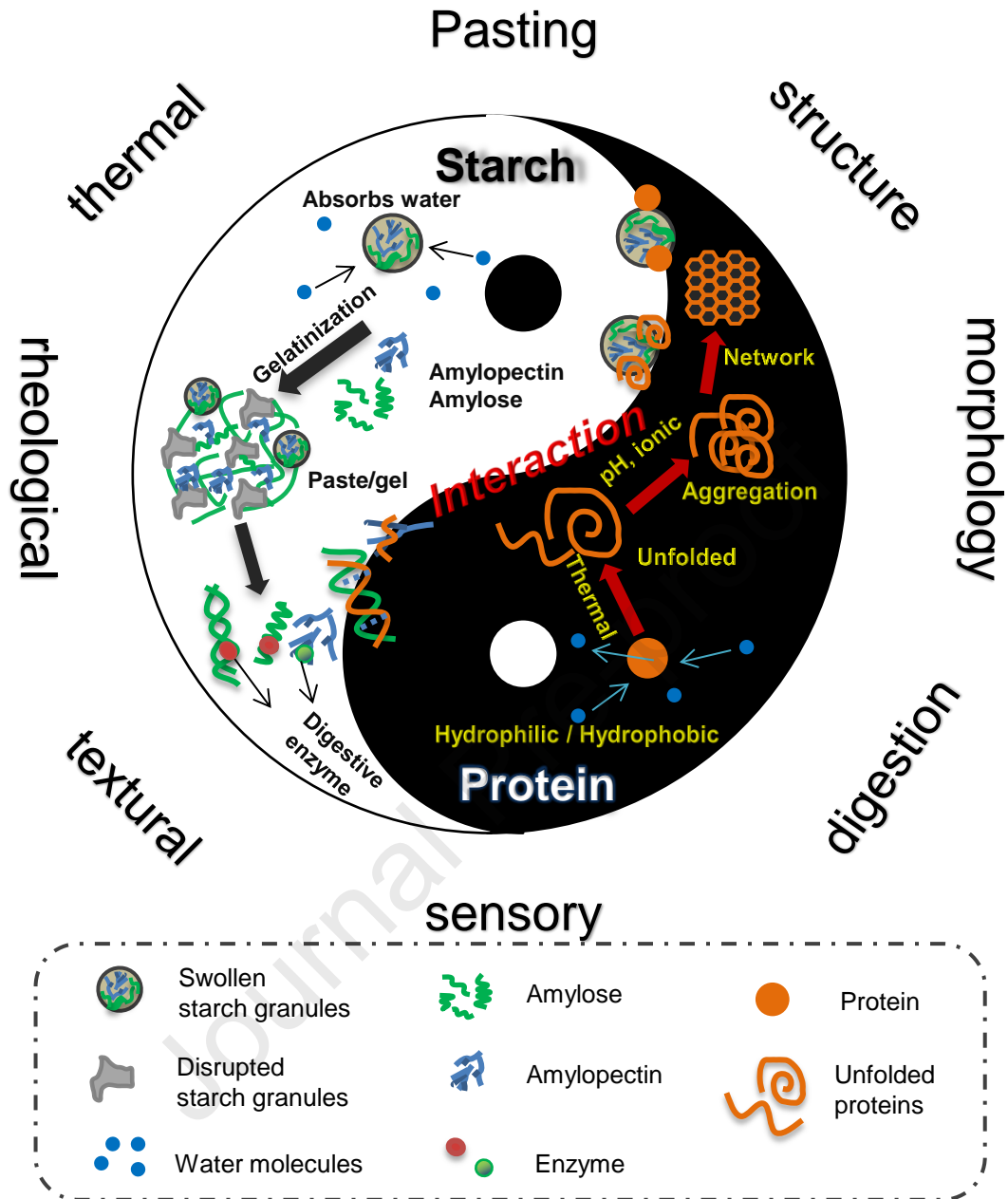
100 Many foods, especially flour (starch)-based ones, contain mainly two types of biopolymers,
101 namely, starches and proteins. Such foods, primarily starch-based matrices containing protein, are
102 consumed worldwide, serving as vital sources of energy and nutrition that maintain the health and
103 functioning of the human body (Wang, Zhang, Wang, Ai, & Xiong, 2020; Yang, Zhong, Douglas
104 Goff, & Li, 2019). Traditionally, such foods are prepared with relatively common ingredients
105 containing starches and endogenous proteins (e.g. wheat or other cereal flour), through different
106 types of processing. Among these products, bread, pasta, and pizza are enjoyed by Western people,
107 while steamed bread, noodles, and rice are consumed by people in the East (Li, Zhu, Guo, Brijs, &
108 Zhou, 2014). To meet consumer demand for improving diet quality and food functions, many
109 high-quality flours and supplements (e.g. proteins, food gums, and fatty acids) have been developed
110 and used in starchy foods (Li, Zhu, et al., 2014). Addition of exogenous proteins to improve the
111 quality of starch-based products has been widely practiced, due to the safety, health, and numerous
112 sources of these proteins. In this regard, although starches and proteins exist widely in many natural
113 foods, it can be necessary to recombine them or incorporate exogenous proteins during processing, in
114 order to obtain food products with desired properties (Li & Huang, 2015). For example, the
115 incorporation of protein into gluten-free food results in a continuous protein phase and a crosslinked
116 structure, leading to increased elastic modulus (Ronda, Villanueva, & Collar, 2014; Villanueva,
117 Ronda, Moschakis, Lazaridou, & Biliaderis, 2018).

118 In recent years, there has been a research focus on the morphology, structure, and properties of
119 starch-based matrices containing endogenous and/or exogenous proteins, aimed at providing a

120 reference for the rational design and production of high-quality starch-based food products that meet
121 the needs of diverse consumers. Also, researchers e.g. (Bhattarai, Dhital, & Gidley, 2016; Considine
122 et al., 2011; Jekle, Mühlberger, & Becker, 2016; Kumar, Brennan, Mason, Zheng, & Brennan, 2017;
123 López-Barón, Gu, Vasanthan, & Hoover, 2017; Witzcak, Ziobro, Juszczak, & Korus, 2016) have
124 carried out extensive studies on many different starches and proteins, which have provided a large
125 amount of basic and important information for the understanding of mixed starch–protein matrices
126 for food product development.

127 As shown in **Fig. 1**, starches and proteins have different functional properties that largely
128 determine the processing, product quality, and nutritional properties of starch-based food matrices.
129 Additionally, interactions between starches and proteins present in natural or processed food systems
130 are often responsible for the structure and thus the properties and quality of such food products
131 (Quiroga Ledezma, 2018). Therefore, a better understanding of protein inclusion and related
132 interactions with starches can help to enable the achievement of desirable structural, textural, sensory,
133 and digestive properties, and shelf stability, and can enable expanded applications of starch matrices,
134 based on advanced food technologies. At present, the role of proteins in starch-based products has
135 attracted extensive attention, with many studies specifically focused on the effects of proteins
136 (endogenous and exogenous) on the structures and properties of starch-based food matrices.
137 However, the results of such specific effects have not been systematically summarized.

138



139

140 **Fig. 1** Overview of the characteristics of mixed starch–protein matrices

141

142 Therefore, this review provides a survey of the latest developments in starch-based food

143 matrices, with a particular focus on the impact of protein presence and resulting starch–protein

144 interactions on the morphology and structure, as well as the physicochemical and digestive properties

145 of such starch-based systems. Based on that focus, this review further discusses the structure–

146 property relationships and mechanisms of mixed starch–protein systems, as reported in the literature
147 to date. Additionally, some possible hypotheses are proposed to describe the effects of proteins on the
148 different properties common to starches, hypotheses that can guide the processing of such
149 starch-based food systems. Furthermore, we suggest that this review can provide insights into the
150 development of novel food systems based on starches and proteins.

151 **2. Basic aspects of starch and protein**

152 **2.1 Starch**

153 Starch, the major component of starch-based foods (e.g. noodles, pasta, and bread), is a
154 glycemic carbohydrate in the diet (Svihus & Hervik, 2016; Wang & Copeland, 2013). Starch is
155 typically composed of a mixture of amylose and amylopectin polymers (Barak, Mudgil, & Khatkar,
156 2014), and its multi-level structure (i.e. starch granules, crystalline, semi-crystalline, and amorphous
157 lamellar structures) has been described extensively and reviewed in detail (Pérez & Bertoft, 2010;
158 Quiroga Ledezma, 2018; Vamadevan & Bertoft, 2015; Wang & Copeland, 2013). The characteristic
159 behaviors of starch (e.g. gelatinization and retrogradation) greatly affect the properties (e.g. structure,
160 texture, and digestibility) of many starch-based food matrices (Toutounji et al., 2019; Wang &
161 Copeland, 2013).

162 The properties of such starch matrices are often largely affected by hydrothermal treatments.
163 During heating, starch granules absorb water and swell, and some starch molecules leach out,
164 resulting in changes in the viscosity of such starch suspensions (Vamadevan & Bertoft, 2015). This
165 process is called “gelatinization”, the process by which native starch granules lose their natural order

166 and crystalline structure and become amorphous (i.e. water uptake, swelling, crystallite melting, the
167 disruption of molecular order, and starch solubilization) (Wang & Copeland, 2013). With the
168 application of constant heating and shearing, gelatinized starch forms a paste with certain rheological
169 properties (Ai & Jane, 2015). Due to the rupture of starch granules, starch molecules in the starch
170 paste are more easily bound to amylase enzyme, thereby accelerating their hydrolysis. After cooling
171 and storage, some starch pastes can form gels and thereby lose their fluidity at appropriate
172 concentrations, while others can remain more liquid-like (Ai & Jane, 2015). This starch gelation
173 process is called “retrogradation” (aka recrystallization), and is due to chain rearrangement,
174 including the formation of new double helices (Quiroga Ledezma, 2018; Wang, Li, Copeland, Niu, &
175 Wang, 2015). Fu, Chen, Luo, Liu, and Liu (2015) systematically summarized the influencing factors
176 (e.g. proteins, lipids, carbohydrates, and salts) of starch retrogradation. Furthermore, the
177 retrogradation of starch results in considerable changes, such as increases in viscosity, opacity, and
178 gel hardness, and phase separation of the polymers and water, and has a great impact on the texture
179 of many starch-based food systems (Wang & Copeland, 2013). The effects of starch retrogradation
180 on its digestion have been summarized by Toutounji et al. (2019), showing that this reordering of
181 starch chains decreases starch digestibility and can increase the content of resistant starch (RS).

182 Studies have shown that the rate and extent of starch gelatinization and retrogradation depend
183 on multiple intrinsic and extrinsic factors (Toutounji et al., 2019; Wang & Copeland, 2013). Intrinsic
184 factors include starch granule morphology, amylose/amylopectin ratio, and starch molecular structure.
185 Extrinsic factors can include processing operations and conditions (e.g. thermal processing, extrusion
186 cooking, and processing environment) and the presence of other constituents (e.g. proteins, lipids,

187 and polysaccharides) (Singh, Dartois, & Kaur, 2010; Wang & Copeland, 2013; Wang, Li, et al.,
188 2015).

189 **2.2 Protein**

190 Protein (endogenous or exogenous) is typically the second-highest component in many
191 starch-based foods, at about 4–20% by weight (Baik, 2010; Ortolan & Steel, 2017; Storck et al.,
192 2013). Therefore, protein can have an important impact on the quality of such starch-based food
193 matrices. For example, wheat gluten protein can be a major determinant of wheat-based product
194 quality, by affecting the water-holding capacity (WHC), cohesiveness, and viscoelasticity of wheat
195 flour doughs (Wang, Jin, & Xu, 2015). Moreover, the quality and nutritional properties of gluten-free
196 foods are often improved by the addition of exogenous, non-gluten proteins with certain functional
197 properties (e.g. gelling ability and WHC) (Aryee, Agyei, & Udenigwe, 2018; M, 2017; Manoj Kumar
198 et al., 2019; Phongthai, D'Amico, Schoenlechner, Homthawornchoo, & Rawdkuen, 2017; Ribotta &
199 Rosell, 2010). An understanding of the functional properties of proteins is of great significance in
200 their application in developing many starch-based food systems.

201 Gelation is an important behavioral characteristic of proteins, especially related to their
202 elasticity and textural properties, and is often affected in food products (Aryee et al., 2018). Proteins
203 can be induced to undergo gelation by heat, chemical (e.g. pH and salt ions), and/or enzymatic
204 treatments (Nieto-Nieto, Wang, Ozimek, & Chen, 2015; Tarhan, Spotti, Schaffter, Corvalan, &
205 Campanella, 2016). Protein gel formation involves protein unfolding, leading to the exposure of
206 hydrophobic amino acid residues (Foegeding & Davis, 2011). Subsequently, unfolded molecules are

207 irreversibly rearranged, associated, and aggregated by interactions such as disulfide interactions,
208 hydrophobic interactions, hydrogen bonding, and van der Waals forces (Foegeding & Davis, 2011).
209 If the protein concentration is high enough, a three-dimensional gel network can be formed
210 (Foegeding & Davis, 2011). Furthermore, various studies have shown that proteins alone or
211 combined with polysaccharides (e.g. inulin, native starch, or acetylated starch) can produce gel
212 matrices with different microstructures, thereby improving their water-retention, rheological, and
213 texture properties (Nieto-Nieto et al., 2015; Ren, Dong, Yu, Hou, & Cui, 2017; Ren & Wang, 2019;
214 Yu, Ren, Zhao, Cui, & Liu, 2020).

215 WHC is directly related to the interactions between protein molecules and water. The WHC of
216 proteins in starch-based food matrices can affect the distribution of water in such mixed systems,
217 thereby modifying the interactions between other components and water molecules (Aryee & Boye,
218 2017; Pelgrom, Vissers, Boom, & Schutyser, 2013). In addition, protein aggregation is promoted in
219 protein gels, and, as a result, protein–water interactions are limited, leading to a decrease in WHC
220 (Nieto Nieto, Wang, Ozimek, & Chen, 2016). Protein gels that have low WHC may not be able to
221 hold water effectively, thereby leading to low textural stability (Boye, Zare, & Pletch, 2010).
222 Properties such as WHC can be affected by both intrinsic (e.g. protein structure, conformation,
223 amino acid composition, hydrophobicity, and hydrophilicity) and extrinsic (e.g. pH, temperature, and
224 ionic strength) factors (Aryee et al., 2018; Foegeding & Davis, 2011).

225 **2.3 Interactions between starches and proteins**

226 Both covalent-bonding and non-covalent interactions have been reported to exist between
227 starches and proteins (Li, Wang, Chen, Yu, & Feng, 2018; Ren & Wang, 2019; Wang et al., 2021).
228 Such interactions are affected by the intrinsic nature of the polymers (e.g. net charge, solubility, size,
229 and weight ratios), protein/starch ratio, temperature, pH, and ionic strength (Heertje, 2014; Li &
230 Huang, 2015; Warnakulasuriya & Nickerson, 2018; Wei & Huang, 2019). Moreover, the interactions
231 between the starches and proteins present in many natural or processed food materials are
232 responsible for the structures, properties (e.g. physicochemical and digestion), and quality of such
233 foods (Quiroga Ledezma, 2018). Sometimes, these interactions may be more important than the
234 physicochemical properties of the individual components. Therefore, a better understanding of such
235 interactions can help to enable the achievement of desirable textural, sensory, and digestive
236 properties of many starch-based food systems, especially those produced using advanced food
237 processing techniques (Quiroga Ledezma, 2018).

238 **2.3.1 Non-covalent interactions**

239 Non-covalent binding is the most common type of interactions between starches and proteins,
240 involving hydrogen bonding, hydrophobic interactions, electrostatic forces, ionic interactions, and
241 van der Waals force (Li et al., 2018; Quiroga Ledezma, 2018; Wang, Appels, et al., 2017). The main
242 types of interactions between starches and proteins are summarized in **Table 1**. It has been found that
243 the tryptophan (Trp), tyrosine (Tyr), or phenylalanine (Phe) aromatic side chains of proteins can
244 interact with starches by non-covalent binding (Li et al., 2018). Proteins contain many hydrophilic
245 groups (e.g. $-\text{COOH}$, $-\text{NH}_2$, $-\text{OH}$, and $-\text{SH}$), all of which are capable of forming physical

246 crosslinks with starches (Kumar et al., 2017; Zhu et al., 2020). In the past few years, the factors
 247 resulting in starch–protein interactions have been widely studied, and there is often a co-existence of
 248 multiple interactions such as hydrogen bonding, hydrophobic interaction, electrostatic interactions,
 249 and van der Waals forces (Joshi, Aldred, Panozzo, Kasapis, & Adhikari, 2014; Li et al., 2018).

250

251 **Table 1** Overview of starch and protein interactions

System	Processing	Interactions	Reference
Lentil starch, lentil protein	Cooking	Non-covalent interactions (hydrophobic and hydrogen bonding) and covalent bonds	(Joshi et al., 2014)
Wheat starch, hydrolyzed pea protein	Extrusion	Hydrogen bonding	(López-Barón et al., 2018)
Corn starch, whey protein isolate	Cooking	Hydrogen bonding	(Yang et al., 2019)
Rice	Cooking	Hydrogen bonding (weak)	(Zhu et al., 2020)
Wheat flour, soy protein	Mixing	Hydrophobic	(Ryan & Brewer, 2007)
Waxy rice flour	Soaking	Hydrogen bonds and hydrophobic	(Li et al., 2018)
Waxy maize starch, caseinates	Mixing	Hydrophobic	(Kett et al., 2013)

Wheat starch, wheat gluten	Heating	Hydrophobic, hydrogen bonds	(Li et al., 2020)
Wheat starch, soy protein	Cooking	Hydrogen bonds	(Ribotta, Colombo, León, & Añón, 2007)
Damaged cassava starch, wheat gluten protein	Dough formation	Non-covalent bonds	(Liu et al., 2019)
Corn starch, hydrophilic protein	Dehydration	Hydrogen bonds	(Zeng et al., 2010)
Phosphate starch, casein	Heating, mixing	Electrostatic adhesion (main)	(Sun, Liang, Yu, Tan, & Cui, 2016)
Starch granules, soybean peptide	Mixing	Weak electrostatic interactions	(Chen, Luo, et al., 2019)
Starch ester of octenyl succinic, casein	Heating, mixing	Steric stabilization (main)	(Sun et al., 2016)
Potato starch, whey protein isolate	Acidic conditions	Electrostatic interactions	(Chen, Fang, Federici, Campanella, & Jones, 2020)
Indica rice starch (IRS), whey protein	Heating, mixing	Hydrophobic molecular interactions (main)	(Wang et al., 2021)

isolate (WPI), casein
(CS)

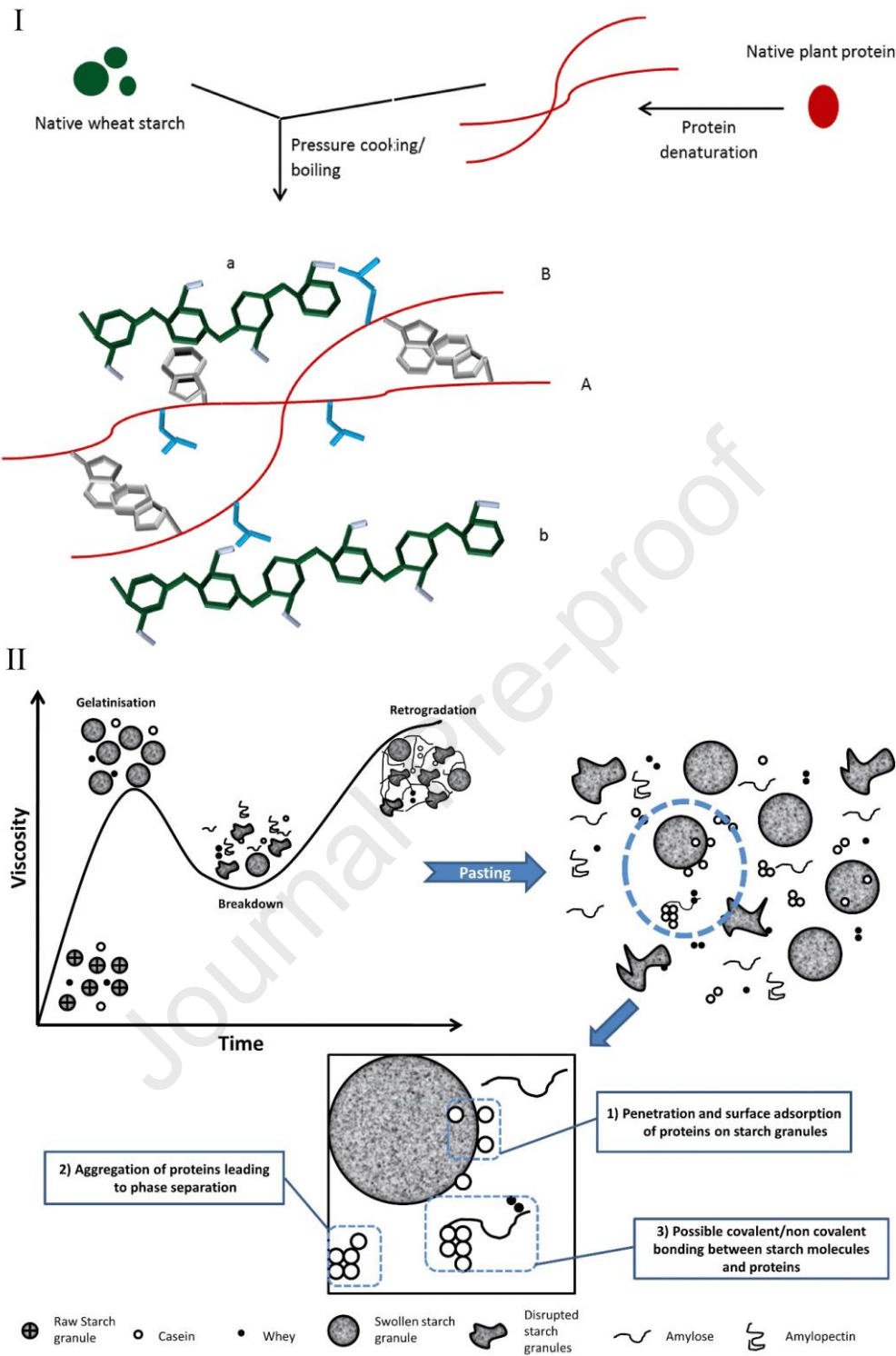
Indica rice starch Heating, Hydrophobic, hydrogen bonding, and (Wang et al.,
(IRS), soy protein mixing electrostatic interactions 2021)
isolate (SPI)

252

253 Hydrogen bonds are formed by the electrostatic attraction of negatively charged atoms to
254 hydrogen atoms (Silverman & Holladay, 2014). The formation of hydrogen bonds must satisfy two
255 basic conditions: a hydrogen donor and a hydrogen acceptor (Silverman & Holladay, 2014).
256 Therefore, in mixed starch–protein matrices, hydrogen bonds are the most prominent hydrophilic
257 interactions, due to the presence of abundant —OH groups in starches (López-Barón et al., 2017).
258 These —OH groups interact not only with protein side chains containing polar residues (e.g. aspartic
259 acid (Asp), glutamic acid (Glu), asparagine (Asn), glutamic acid (Gln), arginine (Arg), and serine
260 (Ser)), but also with protein backbone amine and carbonyl groups (Fernández-Alonso et al., 2012;
261 López-Barón et al., 2017). For example, Yang et al. (2019) have reported, based on a rheological
262 study, that the main interactions between corn starch and whey protein isolate are by hydrogen
263 bonding. Additionally, Fourier-transform infrared (FTIR) analysis of extruded samples has clearly
264 indicated enhanced hydrogen bonding between wheat starch and hydrolyzed pea protein
265 (López-Barón et al., 2018), suggested to be due to hydrolyzed pea protein having more free carboxyl
266 groups, thereby increasing its ability to hydrogen bond with wheat starch (López-Barón et al., 2018).

267 As illustrated in **Fig. 2I**, in addition to hydrogen bonding, there can also be hydrophobic
268 interactions between processed proteins and starches (Li et al., 2020; López-Barón et al., 2017).
269 Hydrophobic interactions result from the tendency of hydrophobic residues to aggregate with each
270 other, thereby avoiding water (Pace et al., 2011). In proteins, hydrophobic interactions provide a
271 major driving force for their folding, leaving hydrophobic residues inside native protein molecules
272 (Aryee et al., 2018). Ryan and Brewer (2007) found that exogenous soy proteins could bind to wheat
273 starch granules through hydrophobic interactions, as there are proteins inherently on the surface of
274 wheat starch granules. If the surface of starch granules is rendered hydrophobic, hydrophobic
275 interactions between the starch and denatured protein can occur. Studies (Li et al., 2020; Wang et al.,
276 2013) have shown that the hydrophilic groups of wheat gliadin protein remain on its surface, while
277 the hydrophobic groups are located inside the gliadin molecules, which may lead to a more stable
278 double-helical structure of the gliadin–starch system, resulting from hydrophobic interactions and
279 hydrogen bonding. Furthermore, heating causes the swelling of starch granules and the leaching of
280 starch chains (amylose and amylopectin) and the polypeptide chains of protein are partially expanded
281 (e.g. exposing the hydrophobic amino acids buried inside native proteins), and then the starch chains
282 can also bind with polypeptide chains through hydrophobic interactions and/or hydrogen bonding
283 (López-Barón et al., 2017; López-Barón et al., 2018).

284



285

286 **Fig. 2 (I)** Polypeptide chains A and B present tryptophan residues (gray) and aspartic acid residues
 287 (blue) as binding sites, which create non-polar and polar interactions with starch chains a and b
 288 (green). Polypeptide chains A and B interact through tryptophan residues (i.e. hydrophobic
 289 interactions), facilitating a coating effect by denatured protein on the surface of the gelatinized starch

290 matrix. Also, polypeptide chains (B) interact with the starch chains (a and b) through aspartic acid
291 residues (blue), thereby connecting the two starch chains. Reprinted from López-Barón et al. (2017),
292 copyright (2017), with permission from Elsevier. **(II)** Graphical representation of starch–milk
293 proteins interactions during continuous-shear heating. Reprinted from Kumar et al. (2017), copyright
294 (2016), with permission from John Wiley and Sons.

295

296 Electrostatic interactions occur mainly between anionic groups on starches and positively
297 charged groups on proteins (Jamilah et al., 2009). Starch molecules are generally considered to be
298 neutral macromolecules, but the amylopectin from potato, root and tuber starches is negatively
299 charged, due to the presence of phosphate groups; and some modified starches (e.g. phosphate starch)
300 are also charged (Quiroga Ledezma, 2018). The charge on a protein surface depends on the protein's
301 *pI* (isoelectric point) and the pH of the system, and the presence of salts can affect the net charge of
302 the protein (Quiroga Ledezma, 2018). Therefore, interaction forces can be affected by the
303 environmental pH, the presence of salts, and the charges on the starch and protein (Warnakulasuriya
304 & Nickerson, 2018). Sun et al. (2016) have reported that the interactions between modified starches
305 (e.g. phosphate starch, hydroxypropyl starch, and octenyl succinic esters of starch) and casein
306 involved electrostatic adhesion and hydrogen bonding, both of which were affected by steric
307 hindrance, and that electrostatic adhesion was the main type of interaction between starch phosphate
308 and casein. Ionic interactions may occur between negatively charged starches and amino acid
309 residues of proteins; however, few studies on such ionic interactions have been reported (Majumdar,
310 Sen, & Ray, 2019).

311 **2.3.2 Covalent interactions**

312 Covalent interactions can be described as strong chemical bonds between specific reactive
313 groups on macromolecules. For example, enzyme conjugation, chemical crosslinking, and the
314 Maillard reaction are common ways of forming covalent bonds between polysaccharides and
315 proteins (de Oliveira, Coimbra, de Oliveira, Zuñiga, & Rojas, 2016; Nicoletti Telis, 2019; Wei &
316 Huang, 2019). The Maillard reaction is safe and convenient as no additional chemicals have been
317 added and the conditions under which it occurs are simple and controllable (de Oliveira et al., 2016).
318 The Maillard reaction conditions for the generation of protein–polysaccharide conjugates, the
319 functional properties of such conjugates, and their applications in food are reviewed by de Oliveira et
320 al. (2016). However, because starch molecules contain terminal reducing groups, the Maillard
321 reaction is considered to be the most common type of covalent binding reaction between starches and
322 proteins in starch-based, processed food systems, which typically produces attractive aromas, colors,
323 and flavors in baked products (Pérez, Matta, Osella, de la Torre, & Sánchez, 2013; Wei & Huang,
324 2019), but condensation reactions between reducing sugars and lysine amino side chains during
325 Maillard reaction result in a severe reduction of lysine availability (Pérez et al., 2013). Additionally,
326 dry-heating and wet-heating are common processing operations used for creating Maillard reactions
327 (Consoli et al., 2018). Covalent bonding can occur between terminal carbonyl reducing groups on
328 starch molecules and amino groups on proteins, and requires the control of reaction temperature and
329 time, system pH, and humidity conditions (de Oliveira et al., 2016).

330 2.3.3 Factors influencing interactions

331 Many factors have been reported to affect the interactions between starches and proteins,
332 especially including processing conditions such as temperature, pH, and ionic strength (Heertje, 2014;
333 Li & Huang, 2015; Warnakulasuriya & Nickerson, 2018; Wei & Huang, 2019).

334 Thermal treatments can affect starch and protein conformations and, consequently, the
335 occurrence of interactions. For example, cooking can result in starch gelatinization and protein
336 denaturation, thus exposing additional sites for reaction, which may facilitate interactions between
337 the starch and protein, ultimately affecting the structural and digestive properties of various food
338 products (López-Barón et al., 2017; Lu, Donner, Yada, & Liu, 2016; Paliwal, Thakur, & Erkinbaev,
339 2019; Petitot, Abecassis, & Micard, 2009). Baking is also highly impactful on the interactions
340 between starches and proteins (Paliwal et al., 2019). During baking, protein unfolding and protein–
341 protein interactions, as well as interactions of protein with starch in a paste, may occur, thus affecting
342 structural properties (Quiroga Ledezma, 2018). During extrusion, biopolymers can undergo many
343 physical and chemical transformations. Specifically, starches can be gelatinized and proteins can
344 become unfolded, realigned, hydrolyzed, and can physically crosslink with starch chains, all of
345 which may ultimately lead to improved product texture and digestibility (Cabrera-Chávez et al.,
346 2012; Heredia - Olea, Contreras - Alvarado, Perez - Carrillo, Rosa - Millán, & Serna - Saldivar,
347 2019; Moisis, Forssell, Partanen, Damerau, & Hill, 2015; Philipp, Buckow, Silcock, & Oey, 2017;
348 Philipp, Oey, Silcock, Beck, & Buckow, 2017; Yu, Ramaswamy, & Boye, 2012). Kumar et al. (2017)
349 have recently reviewed the application of dairy proteins in starch-based extruded products, reporting
350 that starch–protein complexes may be formed during extrusion, and that the formation of such

351 complexes can prevent starch fragmentation during extrusion and thereby affect the viscosity,
352 hardness, water absorption, and water solubility of the resulting extrudates.

353 pH can often affect the charges in mixed starch–protein systems. In particular, the surface
354 charge on a protein depends on the protein's *pI* and the pH of the system, and thus, the behavior of
355 proteins in food processing operations is affected by the pH used (Ghosh & Bandyopadhyay, 2012;
356 Quiroga Ledezma, 2018). For example, Chen et al. (2020) have reported that the elasticity and
357 viscosity of potato starch gel matrices containing added whey protein fibrils were higher at pH 3.5
358 than at pH 6.8, possibly because electrostatic interactions between the starch (with its anionic
359 components) and protein could be promoted, as the pH decreased below the protein's *pI* (Chen et al.,
360 2020; Firoozmand, Murray, & Dickinson, 2012).

361 The addition of salt ions can affect the degree of network formation, through electrostatic
362 shielding of protein molecules, thus affecting interactions with starch (Lambrecht, Rombouts,
363 Nivelles, & Delcour, 2017a, 2017b). At low salt concentrations, salt ions can preferentially interact
364 with protein molecules, and such interactions can interfere with protein–protein and protein–starch
365 interactions. However, higher salt concentrations do not favor protein dissolution, which thus can
366 limit protein interactions with starch (Joshi et al., 2014; Li & Huang, 2015). Therefore, a better
367 understanding of starch–protein interactions, under different pH and salt conditions of processing
368 operations for starch-based foods, can be conducive to improving the quality of many such products.

369 **3. Morphology and structure of starch-based matrices containing protein**

370 **3.1 Morphology**

371 The microstructure of starch-based food matrices is highly dependent on their composition,
372 processing, and post-processing storage, and plays a crucial role in determining the textural, sensory,
373 and digestive properties of such foods (Singh, Kaur, & Singh, 2013).

374 Many studies (Feng, Mu, Zhang, & Ma, 2020; Joshi et al., 2014; Liu et al., 2019; Phongthai et
375 al., 2017; Sun & Xiong, 2014; Wang, Zhang, et al., 2020; Xie et al., 2017; Zhang, Chen, Chen, &
376 Chen, 2019; Zhou, Liu, & Tang, 2018) have used scanning electron microscopy (SEM) to observe
377 the micromorphology of doughs and mixed starch–protein gel systems and found that proteins affect
378 the micromorphology of such mixed systems. Morphological observations of doughs have shown
379 that starch granules are trapped in a protein matrix, and the dough structure can be changed by
380 different dough preparation conditions (i.e. mixing time and pressure) (Gao, Koh, Tay, & Zhou, 2017;
381 Liu et al., 2019; Liu et al., 2015; Sarabhai & Prabhasankar, 2015). Structural properties of starches
382 and proteins, in doughs and close-contact dough fractions, may lead to certain interactions, which
383 consequently change the rheological properties of such doughs and restrict the swelling of starch
384 granules (Feng et al., 2020; Liu et al., 2019). For example, for gluten-free pasta, its structural
385 characteristics depend on the type of exogenous protein added, and the resulting protein network
386 encapsulates starch granules, which can limit the access of water during cooking, thereby affecting
387 the starch's digestive characteristics (Giuberti, Gallo, Cerioli, Fortunati, & Masoero, 2015; Phongthai
388 et al., 2017). Additionally, when different proteins (gluten and egg white protein) were mixed with

389 sweet potato starch, leached amylose and low-molecular-mass amylopectin interacted with denatured
390 protein during gelatinization and formed different network structures, resulting in altered textural and
391 cooking properties of the final sweet potato vermicelli products (Feng et al., 2020). For mixed
392 starch–protein gel matrices, in general, both starch and protein gels can form three-dimensional
393 network structures (Joshi et al., 2014). Starch gels generally have dense and porous network
394 structures, while protein gel structures can differ, depending on a given protein’s gelation ability
395 (Joshi et al., 2014; Yang, Liu, Ashton, Gorczyca, & Kasapis, 2013; Zhang et al., 2019). Some studies
396 (Joshi et al., 2014; Zhang et al., 2019) have reported that gels with protein have a tighter and more
397 homogeneous structure, compared to gels without protein, while other researchers (Sun & Xiong,
398 2014; Wang et al., 2021) have reported that mixed starch–protein gels become looser with an
399 increasing proportion of protein in the blend. Such behavior is thought to be related to the ability of
400 proteins to form gels and to their interactions with starches (Sun & Xiong, 2014; Zhang et al., 2019).

401 The distribution of starch and protein phases in mixed gels can be observed by confocal laser
402 scanning microscopy (CLSM). CLSM observations have shown that proteins tended to adsorb onto
403 the surfaces of starch granules, which resulted in a steric hindrance effect to keep the starch granules
404 separated (López-Barón et al., 2017; Ye et al., 2018; Yu et al., 2018). It has been reported that such
405 mixed gels may be homogeneous or exhibit two-phase separation (such as by protein aggregation)
406 (Noisuwan, Hemar, Wilkinson, & Bronlund, 2009; Vu Dang, Loisel, Desrumaux, & Doublier, 2009).
407 Chanvrier, Colonna, Della Valle, and Lourdin (2005) have reported that, during thermomechanical
408 processing, proteins in a corn starch–zein mixture aggregated, and the blend morphology was
409 affected by the starch/zein ratio, which in turn largely affected the mixture’s mechanical properties.

410 Moreover, added protein hydrolysates, because of their small molecular mass, are said to be able to
411 easily penetrate starch granules and reach the granule's center, and thus, form a network structure
412 with starch chains (Kong, Niu, Sun, Han, & Liu, 2016; Niu, Wu, & Xiao, 2017; Xiao & Zhong,
413 2017).

414 Atomic force microscopy (AFM) can be used to reveal nanoscale structural features of samples,
415 which are considerably smaller than those observable by CLSM (Niu et al., 2017; Xiao, Niu, Wu, Li,
416 & He, 2019; Xiao & Zhong, 2017). AFM studies have demonstrated that the number and height of
417 protrusions on the surfaces of starch–protein hydrolysate mixtures were significantly altered,
418 compared to those of starch-only samples, suggesting that protein hydrolysates can affect the
419 aggregation of amylose and amylopectin on a nanoscale (Kong et al., 2016; Niu et al., 2017; Xiao et
420 al., 2019; Xiao & Zhong, 2017).

421 **3.2 Long-range molecular ordered structures**

422 The long-range ordered structures of starch granules are generally considered to involve
423 crystalline structures, which are believed to form through the parallel packing of left-handed coaxial
424 double helices in extended regular arrays (Wang, Xue, Yousaf, Hu, & Shen, 2020). Native starches
425 possess either an A-type, B-type, or C-type polymorphic structure (Wang & Copeland, 2013). X-ray
426 scattering has been widely used to characterize the long-range ordered structures of starches,
427 including their relative crystallinity (Wang, Li, et al., 2015). The crystalline structure of native
428 starches is destroyed during gelatinization, leading to the disappearance of the XRD peaks
429 representing crystallites (Wang et al., 2021). With increasing storage time, many cooked starch

430 matrices typically manifest diffraction peaks due to recrystallization or retrogradation (Hu et al.,
431 2020; Xiao & Zhong, 2017). XRD analyses have shown that the addition of proteins or protein
432 hydrolysates can reduce the crystallinity of starch gel matrices after long-term storage by effectively
433 inhibiting starch retrogradation (Hu et al., 2020; Niu et al., 2017; Xiao & Zhong, 2017; Zhang et al.,
434 2019). For example, compared to gelatinized rice starch alone, rice starch samples with an increasing
435 ratio of whey protein hydrolysate displayed gradually reduced relative crystallinity after storage for 7
436 and 14 days (Hu et al., 2020). The inhibition of starch retrogradation by proteins could be due to
437 interactions between the protein and starch granules, which could hinder the penetration of sufficient
438 water into the granules, resulting in reduced starch gelatinization (Wang, Zhang, et al., 2020). In this
439 way, the protein together with un-gelatinized starch could prevent starch chain rearrangement. It has
440 also been suggested that the hydrogen bonding between starch and water molecules could be
441 hindered, due to hydrophobic interactions between protein chains, thus inhibiting starch
442 retrogradation (Wang, Zhang, et al., 2020). However, Chen, Wang, Fan, Yang, and Chen (2019) have
443 reported that retrograded samples of mixtures of enzyme-modified wheat protein–potato starch
444 exhibited a typical V-type XRD pattern, and the relative crystallinity of the mixture samples
445 increased with increasing concentration of the enzyme-modified wheat proteins, which could have
446 promoted amylopectin recrystallization. It has been found that the removal of endogenous proteins
447 from rice starch granules did not destroy the crystalline morphology of the starch, but decreased the
448 extent of crystallinity, suggesting either that both types of endogenous rice proteins (i.e. starch
449 granule–channel proteins and starch granule–surface proteins) might be involved in the formation of
450 crystalline structures or that the operative extraction methods partially destroyed starch crystallites

451 (Hu et al., 2017; Zhan et al., 2020). Moreover, processing operations can influence the multi-scale
452 structures of starch–protein systems, possibly by promoting interactions between starches and
453 proteins which lead to specific structures (Chen, Luo, et al., 2019; Chi, Li, Zhang, Chen, & Li, 2018;
454 Li et al., 2018; Qiu et al., 2015; Xiao et al., 2019). For instance, starches (corn and potato)
455 complexed with soybean peptide, subjected to HMT, have been reported to display higher relative
456 crystallinity than their non-HMT counterparts, which was attributed to interactions between starch
457 chains and side-chain groups in the soybean peptide during HMT (Chen, Luo, et al., 2019).

458 Small-angle X-ray scattering (SAXS) can complement wide-angle X-ray scattering (WAXS),
459 in understanding long-range order at nanometer scales (Li, Senesi, & Lee, 2016). Using a SAXS
460 technique, Chi et al. (2018) have reported that the SAXS intensity of rice starch samples varied with
461 the addition of rice protein or protein hydrolysates, thereby revealing changes in long-range starch
462 order. Thus, the ordered structures and amorphous structures of rice starch can be altered by
463 interactions with rice proteins or protein hydrolysates (Chi et al., 2018). This suggests that such
464 structural features of rice starch can be altered simply by complexing with rice protein or protein
465 hydrolysates before cooking (Chi et al., 2018). Despite this earlier work, to date, there have been few
466 other reports on the use of SAXS to study structural properties of starch–protein mixtures. Thus,
467 more work is warranted on the use of SAXS to explore the effects of proteins on starch structures.

468 **3.3 Short-range molecular ordered structures**

469 Short-range ordered structures in starches are said to mainly involve double-helices and V-type
470 single helices, both of which are involved in starch crystallinity (Wang, Xue, et al., 2020). FTIR

471 spectroscopy can provide valuable information about the presence of ordered and amorphous
472 structures in starches (Chávez-Murillo, Veyna-Torres, Cavazos-Tamez, de la Rosa-Millán, &
473 Serna-Saldívar, 2018). FTIR peaks at 1047, 1022, and 995 cm^{-1} have been assigned to short-range
474 order, amorphous content, and hydrated crystallites, respectively, in starches; two FTIR peak
475 intensity ratios, 1047/1022 cm^{-1} and 995/1022 cm^{-1} , have been used to estimate the short-range
476 order in various starches, and increases in their ratios indicate an increase in structural order. (Chen,
477 Wang, et al., 2019; Li et al., 2020; Liu et al., 2015; Lu et al., 2016; Yang et al., 2019). Moreover,
478 since the characteristic peaks in FTIR spectra correspond to bond stretching, possible interactions in
479 starch-based systems can be inferred from the changes in those characteristic peaks (Lu et al., 2016).

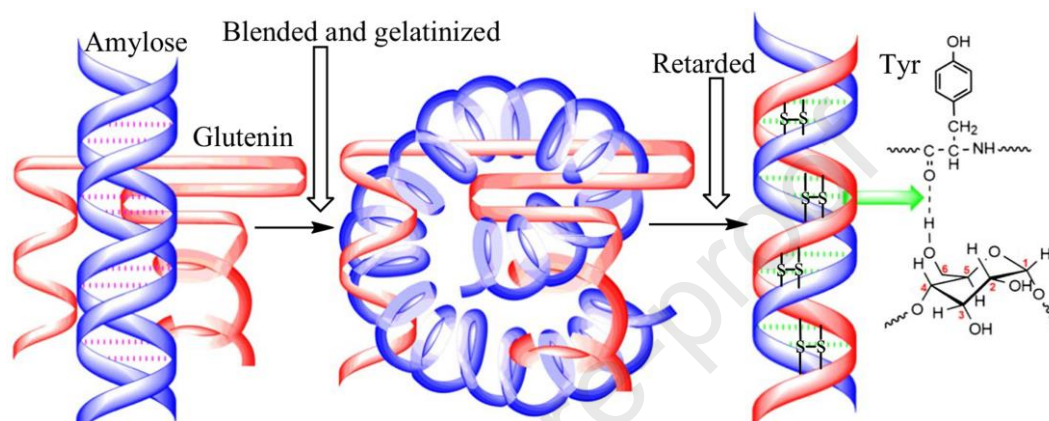
480 The short-range ordered structures in starches are said to be maintained mainly by hydrogen
481 bonding (Lu et al., 2016; Zhan et al., 2020). During gelatinization, hydrogen bonds between starch
482 chains are thought to be broken, due to the entry of water molecules, leading to weakening or even a
483 disappearance of characteristic absorption peaks, especially at 995 cm^{-1} (Li et al., 2020). Then,
484 starch chains can rearrange during cooling and retrogradation, as reflected by increases in the
485 characteristic FTIR peak intensities (Li et al., 2020). Since proteins can interact with starch
486 molecules, especially by hydrogen bonding, the addition of proteins or protein hydrolysates can
487 affect the structural changes of starches at a molecular level (López-Barón et al., 2018). Several
488 studies have shown that proteins or protein hydrolysates can inhibit the hydrogen-bonded
489 crosslinking of starch chains during aging, and thus, the formation of ordered structures (as reflected
490 by a decreased intensity of the 1047/1022 cm^{-1} ratio) with this inhibitory effect dependent on the
491 protein/protein hydrolysate concentration (Guo, Lian, Kang, Gao, & Li, 2016; Hu et al., 2020; Lian,

492 Zhu, Wen, Li, & Zhao, 2013; Xiao et al., 2019; Zhang et al., 2019). Thus, the presence of proteins or
493 protein hydrolysates can be seen to inhibit starch retrogradation. However, other researchers (Chen,
494 Wang, et al., 2019; Xijun, Junjie, Danli, Lin, & Jiaran, 2014; Yang et al., 2019) have found that
495 proteins can promote starch retrogradation, and that more ordered structures can result from the
496 addition of proteins. Such discrepancies might be explained by differences in protein structure and
497 hydration capacity (Chen, Wang, et al., 2019; Wang, Zhang, et al., 2020; Xijun et al., 2014; Zhang et
498 al., 2019). In any event, the reasons behind the different effects of proteins on starch retrogradation
499 are not entirely understood, and therefore warrant further investigation.

500 ^{13}C cross-polarization/magic-angle-spinning nuclear magnetic resonance (^{13}C CP/MAS NMR)
501 has also been used to analyze the short-range order of starches in starch-based systems (Chi et al.,
502 2018; Xijun et al., 2014). Normally, the C4 resonance peak can be correlated with a starch's
503 amorphous fraction, and the C1 resonance peak can be highly correlated with the contents of
504 amylose V-type single-helices and amylopectin double-helices in various starches (Chi et al., 2018;
505 Flores-Morales, Jiménez-Estrada, & Mora-Escobedo, 2012). As shown in **Fig. 3**, Guo et al. (2016)
506 have suggested, based on ^{13}C NMR results, that tyrosine (Tyr) on wheat protein may interact with the
507 amylose in wheat starch, and have described a mechanism for the inhibition of amylose
508 retrogradation by wheat glutenin. Related studies have also reported that the addition of proteins can
509 result in changes in starch–protein interactions (as reflected by changes of the peak signals in ^{13}C
510 CP/MAS NMR spectra), which can subsequently affect the retrogradation and digestive properties of
511 starches (Flores-Morales et al., 2012; Guo et al., 2016; Renzetti et al., 2012; Xijun et al., 2014).
512 Additionally, Chi et al. (2018) have reported that rice starch with added pepsin–pancreatin protein

513 hydrolysates exhibited more amorphous starch, suggesting that the pepsin–pancreatin protein
 514 hydrolysates hindered rice starch aging after gelatinization. In the studies discussed above, the ^{13}C
 515 CP-MAS/NMR results consistently supported the results obtained by FTIR.

516



517

518 **Fig. 3** Mechanism of glutenin retardation of the retrogradation of amylose

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 520 Heidelberg.

521

522 **4. Properties of starch-based matrices containing protein**

523 For mixed starch–protein matrix systems, protein structure–property relationships, including
 524 the hydration ability, thermal properties, and gelation behavior of the proteins, largely influence the
 525 features of the mixed systems. Specific effects reported have included the following: the hydration
 526 ability of proteins affects the interactions of starches with water (López-Barón et al., 2017; Wang et
 527 al., 2021; Yang et al., 2019); due to the thermal properties of proteins, heat-induced conformational
 528 changes of proteins can result in protein–starch interactions (López-Barón et al., 2017; Wang et al.,

529 2021); and the gelling ability of proteins can cause crosslinking or phase separation in starch–protein
530 mixtures during the gelation process (Warnakulasuriya & Nickerson, 2018). Thus, in these ways, the
531 inclusion of proteins can significantly affect the textural, sensory, pasting, thermal, rheological, and
532 digestive properties of starch-based matrices.

533 **4.1 Pasting properties**

534 The pasting behavior of starch is central to many starch-based food matrices and is usually
535 characterized by changes in viscosity on heating, holding, and cooling (Li et al., 2018). Pasting
536 temperature, peak viscosity, final viscosity, breakdown viscosity, and setback viscosity can all be
537 easily determined using a rapid visco-analyzer (RVA) (Gani et al., 2015). Many studies have reported
538 the effects of different proteins on the viscosity of different starch systems, for which viscosity is of
539 great significance to the applications of such biopolymers in various food (Joshi et al., 2014; Kim,
540 Kee, Lee, & Yoo, 2014; Kumar et al., 2017; Li et al., 2018; Sarabhai & Prabhasankar, 2015). For
541 example, **Fig. 2II** shows the viscosity changes for a starch–milk proteins system under
542 continuous-shear and -heating conditions, and the possible interactions between the starch and
543 proteins that can cause such viscosity changes (Kumar et al., 2017). Proteins on the surfaces of starch
544 granules can limit the swelling of the granules, and such interactions between starches and proteins
545 can affect the viscosity parameters of such systems (Chinma, Ariahu, & Abu, 2013; Kett et al., 2013;
546 Qiu et al., 2015; Reddy Surasani, Singh, Gupta, & Sharma, 2019; Ribotta, Colombo, & Rosell, 2012;
547 Ribotta & Rosell, 2010). For instance, Gani et al. (2015) reported that wheat flour supplemented with
548 milk protein hydrolysates exhibited a lower viscosity during cooling than that without

549 supplementation, and the low setback viscosity could be expected to result in softer cookie crumb
550 texture.

551 Pasting temperature corresponds to the minimum temperature required for starch cooking and
552 the temperature at which viscosity increases during heating (Barak et al., 2014). Many studies
553 (Baxter, Blanchard, & Zhao, 2014; Bravo-Núñez, Garzón, Rosell, & Gómez, 2019; Chen, Luo, et al.,
554 2019; Chinma et al., 2013; Joshi et al., 2014; Kong et al., 2016; Likitwattanasade & Hongsprabhas,
555 2010; Sarabhai & Prabhasankar, 2015; Shevkani, Kaur, Kumar, & Singh, 2015; Wang, Zhang, et al.,
556 2020; Xiao et al., 2019; Zhang, Sun, Wang, Wang, & Zhou, 2020) have reported that the pasting
557 temperatures of starch–protein mixtures are higher than those of starch-only systems. For example,
558 removal of protein fractions from rice flour has been found to reduce the pasting temperature of such
559 rice-based systems (Baxter, Zhao, & Blanchard, 2010; Likitwattanasade & Hongsprabhas, 2010).
560 Two explanations for how proteins limit the pasting temperature of starches have been suggested: 1)
561 the absorption of excess water by proteins; and/or 2) the formation of complexes between starches
562 and proteins, which restrict starch–water interactions and starch gelatinization (Joshi et al., 2014;
563 Reddy Surasani et al., 2019; Wang, Zhang, et al., 2020).

564 In contrast, significant decreases in the pasting temperatures for various starch–protein
565 mixtures have also been reported (Qiu et al., 2015; Ribotta et al., 2007; Ribotta et al., 2012; Ribotta
566 & Rosell, 2010). Elsewhere, starch pasting temperatures have been reported to be unchanged by the
567 addition of wheat gliadin and glutenin (Barak et al., 2014; Chen, Zhang, Li, Xie, & Chen, 2018).
568 Thus, proteins can affect starch gelatinization in different ways, depending on their ability to retain

569 water and their interactions with starch granule surfaces and starch molecules (Ribotta et al., 2007;
570 Wang et al., 2021).

571 Addition of proteins to starch-based systems has been shown to reduce the pasting viscosity
572 (Baxter et al., 2014; Chen, Luo, et al., 2019; Joshi et al., 2014; Kim et al., 2014; Kong et al., 2016;
573 Sarabhai & Prabhasankar, 2015; Shevkani et al., 2015; Sun & Xiong, 2014; Xiao et al., 2019), and
574 the degree of viscosity reduction has been found to be related to protein type (Barak et al., 2014;
575 Shin, Gang, & Song, 2010; Storck et al., 2013). For example, Barak et al. (2014) have reported that,
576 compared to glutenins, gliadins were more effective at decreasing the pasting viscosity of dough.
577 Both types of proteins reduced the system viscosity because they competed for the water along with
578 the starch granules. However, glutenins have a β -sheet structure and tend to form an entangled
579 network upon hydration, and thus glutenins formed a network throughout the flour paste, being more
580 resistant to stirring blades than gliadins (Barak et al., 2014). Additionally, viscosity decreased more
581 when protein hydrolysates rather than proteins were added, which was attributed to the fragmentation
582 of proteins during hydrolysis, resulting in a loss of the water entrapment ability of proteins (Gani et
583 al., 2015; Xiao et al., 2019). From other related studies, the following possible explanations have
584 been suggested for the reduction in starch pasting viscosity due to the addition of proteins:

- 585 - Competition between starches and proteins for water decreases the swelling power of starch
586 (Chávez-Murillo et al., 2018; Hu et al., 2020; Li et al., 2020; Shevkani et al., 2015; Sun &
587 Xiong, 2014; Wang, Zhang, et al., 2020; Zhang et al., 2020);

- 588 - Interactions between starches and proteins may reduce the amount of amylose leached out
589 during gelatinization (Bravo-Núñez et al., 2019; Hu et al., 2020; Reddy Surasani et al., 2019;
590 Sun & Xiong, 2014);
- 591 - Protein addition causes starch dilution (Marti et al., 2014; Sarabhai & Prabhasankar, 2015;
592 Sciarini, Ribotta, León, & Pérez, 2010; Shevkani et al., 2015; Zhang et al., 2020);
- 593 - Proteins can act as inert fillers, to hinder hydrogen bonding between starch chains (Kumar,
594 Brennan, Zheng, & Brennan, 2018; Sarabhai & Prabhasankar, 2015; Sciarini et al., 2010;
595 Sopade, Hardin, Fitzpatrick, Desmoe, & Halley, 2006).

596 However, Shin et al. (2010) have reported that added whey protein inhibited the pasting
597 viscosity of rice flour, whereas added TGase enzyme increased the pasting viscosity by crosslinking
598 between the added and rice proteins. Additionally, other studies (Chinma et al., 2013; Qiu et al., 2015;
599 Ribotta et al., 2007; Ribotta et al., 2012; Ribotta & Rosell, 2010) have suggested that certain proteins
600 (e.g. soy protein and pea protein) can increase the overall viscosity of protein–starch pastes, due to
601 enhanced protein–starch interactions during heat treatment. Those interactions might involve
602 crosslinks formed between hydrophilic groups on the protein and starch molecules during heat
603 treatment (Chinma et al., 2013; Qiu et al., 2015; Ribotta et al., 2012; Ribotta & Rosell, 2010).

604 **4.2 Thermal properties**

605 The effect of proteins on the thermal properties of starch–protein blended matrices has been
606 widely reported, as related to the characteristics and quality of various starch-based foods (Chen,
607 Wang, et al., 2019; Jamilah et al., 2009; Li, Wei, Fang, Zhang, & Zhang, 2014; Liu et al., 2019; Lu et

608 al., 2016; Zhu et al., 2020). DSC is most often used to study structural changes in proteins and
609 starches as a function of temperature, and onset temperature (T_o), peak temperature (T_p), conclusion
610 temperature (T_c), and ΔH values have been obtained (Jamilah et al., 2009; Marín, Alemán, Montero,
611 & Gómez-Guillén, 2018; Wan, Liu, & Guo, 2018).

612 DSC heating curves for acetylated potato starch–whey protein blended systems have shown
613 two obvious peaks, reported to represent acetylated potato starch gelatinization (about 61 °C) and
614 whey protein denaturation (about 73 °C) (Ren et al., 2017). However, single endothermic peaks for
615 other starch–protein blends have also been reported in other studies (Lu et al., 2016; Yang et al.,
616 2019). The following two explanations for these latter observations have been suggested: 1) the
617 protein concentration in the mixed system was low ($\leq 20\%$), so that the thermal events for the protein
618 were overwhelmingly overlapped by those for the starch; and/or 2) the protein in the mixed system
619 had already been denatured and thus thermally inactivated (Lu et al., 2016; Sciarini et al., 2010; Yang
620 et al., 2019). Moreover, no new endothermic peaks have been seen to appear in DSC thermal curves
621 for such mixed systems, suggesting that no new substances were generated during thermal treatment
622 (Wang, Zheng, Yu, Wang, & Copeland, 2017). Various studies e.g. (Villanueva, Mauro, Collar, &
623 Ronda, 2015) have reported significant effects of the presence of proteins on starch gelatinization.
624 Often, proteins are found to have an inhibitory effect on the gelatinization of starches, consistent
625 with RVA results (Lu et al., 2016). The extent of the DSC peak shift to higher gelatinization
626 temperature is considered to be closely related to the type and proportion of protein present. For
627 example, Li et al. (2020) have suggested that the higher T_p value for wheat starch gelatinization,
628 obtained for a glutenin–wheat starch mixture as compared to a gliadin–wheat starch mixture, was

629 probably due to the higher hydrophobicity of glutenin than gliadin (Mittal & Best, 2008).
630 López-Barón et al. (2017) have reported that the addition of denatured and hydrolyzed plant proteins
631 to wheat starch increased the T_p value for wheat starch gelatinization more obviously than did the
632 corresponding addition of native plant proteins, possibly due to changes in surface hydrophobicity
633 and WHC of such plant proteins after their heat-induced denaturation or hydrolysis. It has been
634 speculated that the effects of proteins on starch gelatinization may result from a competition for
635 available water between these two types of biopolymer, leading to a water re-distribution between
636 them (Li et al., 2020; Lu et al., 2016; Ren & Wang, 2019; Yang, Luan, Ashton, Gorczyca, & Kasapis,
637 2014; Zhu et al., 2020), or from interactions between starch granules and proteins and/or between
638 material leached out of starch granules and proteins (Chen, Zhou, Yang, & Cui, 2015; Chen, Luo, et
639 al., 2019; Li & Zhu, 2017; Ribotta et al., 2007; Wang et al., 2021; Zhu et al., 2020). In these ways,
640 the starch gelatinization temperature could be significantly affected by the specific type and dosage
641 of protein and/or by various starch–protein interactions.

642 Starch retrogradation is a process in which certain chains of gelatinized starch re-align and
643 re-associate to form crystallites during cooling, which results in significantly influenced textural and
644 sensory characteristics of many starch-based food systems (Chen, Wang, et al., 2019; Lu et al., 2016;
645 Wu, Chen, Li, & Wang, 2010). Thermal analysis can provide information on starch aging in starch–
646 protein mixtures; from DSC analysis, the effects of proteins on the retrogradation properties of starch
647 gels can be better understood (Wang, Li, et al., 2015). It has been reported that the ΔH value
648 measured by DSC mainly reflects the energy required to melt the potato amylopectin crystallites
649 recrystallized during retrogradation (Chen, Wang, et al., 2019). Thus, measured changes in ΔH can

650 reflect whether the aging process of starch matrices is inhibited or promoted. Various studies have
651 found that the measured enthalpies for retrograded starch–protein systems were higher than those for
652 starch-only systems, but could be decreased with even higher amounts of added proteins (Chen,
653 Wang, et al., 2019; Lu et al., 2016; Yang et al., 2019). Moreover, other studies have reported that the
654 addition of rice (or other) protein hydrolysates can significantly reduce the measured ΔH for
655 retrograded rice or wheat starch, indicating that such protein hydrolysates can inhibit the
656 retrogradation of such starches (Niu et al., 2017; Xiao & Zhong, 2017; Zhang et al., 2020). This
657 phenomenon may be due to the active polyhydroxyl groups in protein hydrolysates, which may block
658 or insert into the hydrogen bonds among starch molecules (Niu et al., 2017). For a food product such
659 as Chinese rice vermicelli, starch retrogradation is desired, as it provides preferred food
660 characteristics, including improved textural, sensory, and digestive properties (Karim, Norziah, &
661 Seow, 2000). However, in many other cases, inhibiting starch retrogradation is advantageous for
662 prolonging the shelf-life of various starch-based food products (Chen, Wang, et al., 2019; Niu et al.,
663 2017; Xiao & Zhong, 2017; Zhang et al., 2020), most familiarly exemplified by bread staling. Thus,
664 understanding the effects of proteins on starch retrogradation characteristics can be highly beneficial
665 to the informed development of improved starch-based food products with desired properties.

666 **4.3 Rheological properties**

667 The rheology of various food products plays a significant role in quality control, sensory
668 evaluation, process assessment, and product development (Jamilah et al., 2009). Many studies
669 (Amjid et al., 2013; Brandner, Becker, & Jekle, 2019; Considine et al., 2011; Jamilah et al., 2009;

670 Kumar et al., 2017) have reported the effects of various proteins on the rheological properties of
671 starches and highlighted component interactions. Understanding how proteins affect the rheological
672 characteristics of starch-based food systems can be of great significance for food processing.

673 A temperature sweep technique can be used to study rheological changes during heating and
674 cooling of starch-based systems and to identify their starch gelatinization temperatures (Yang et al.,
675 2013). For example, temperature sweep studies of mixed lentil starch–lentil protein gel matrices have
676 shown that the gel development process in lentil protein-dominated composites was much slower
677 than that in lentil starch-dominated composites, and lentil starch-rich composite gels exhibited more
678 solid-like properties (Joshi et al., 2014). During heating, the storage modulus (G') values for starch–
679 protein mixtures have been found to increase with temperature until reaching peak values (the
680 swelling of starch granules), then to decrease upon further heating (the breakage of swollen starch
681 granules); with increasing protein content, the temperature where G' reached a peak increased, while
682 the G' value at this point decreased, which may be due to the proteins limiting starch swelling
683 (Ghumman, Kaur, & Singh, 2016; Hu et al., 2020; Lu et al., 2016; Xiao & Zhong, 2017).

684 Furthermore, Zhou et al. (2018) have reported that there were obvious differences in the G' and loss
685 modulus (G'') behaviors, as affected by protein addition, between wheat flour–soy protein doughs
686 and wheat flour–whey protein doughs. The difference in modulus changing may be due to the higher
687 water absorption and weaker gelation abilities of soy proteins than whey proteins (Comfort &
688 Howell, 2002). For wheat flour–whey protein doughs, both the G' and G'' peak values first decreased
689 slightly, as the whey protein level was increased from 0 to 10 wt%, and then increased significantly,
690 as the whey protein level was increased further to 30 wt% (Zhou et al., 2018). Regarding this, the

691 crosslinking sites between whey protein and wheat starch increased with increasing protein
692 concentration, subsequently affecting the properties of the dough (Zhou et al., 2018). Thus,
693 differences have been found in the effects of both protein type and protein concentration on the G'
694 and G'' values for different starches (Ghumman et al., 2016). During cooling, gelatinized starch
695 chains can undergo retrogradation, which typically results in increased moduli (Yang et al., 2019).
696 Also, the formation of starch–protein gel networks stabilized by interactions has been reported to be
697 responsible for increasing gel elasticity (Ghumman et al., 2016; Xiao & Zhong, 2017; Yang et al.,
698 2019). From the discussion above, we can conclude that the trend of modulus change determined
699 from temperature sweep studies is consistent with the trend of changing RVA viscosity; that is, the
700 moduli and viscosity of various mixed starch–protein systems first increase and then decrease during
701 heating, while the moduli and viscosity both increase again during subsequent cooling; and the
702 moduli and viscosity of mixed starch–protein systems are lower than those of corresponding
703 starch-only systems.

704 Frequency sweep testing can be used to provide information about the type of gel formed in
705 mixed starch-protein samples; protein addition to starch-based batters has been reported to have a
706 significant effect on moduli (Patraşcu, Banu, Vasilean, & Aprodu, 2016). Generally, protein gels
707 show greater frequency dependence, indicating that they are weaker gels, while starch gels are
708 stronger and more independent of frequency (Joshi et al., 2014). Several studies (Kim et al., 2014;
709 Sang et al., 2018; Wang, Chen, Yang, & Cui, 2017; Yang et al., 2019; Zhou et al., 2018) have
710 reported that the addition of protein decreased the G' and G'' values of various starch-based systems,
711 whereas other studies have reported increased G' and G'' values in a certain frequency range (Chen et

712 al., 2018; Qiu et al., 2015; Ribotta et al., 2012; Ronda, Oliete, Gómez, Caballero, & Pando, 2011;
713 Wang, Huang, Kim, Liu, & Tilley, 2011). A decrease in moduli indicates a weakened starch gel
714 structure, while increased moduli indicate a stronger gel structure (Yang et al., 2019). Moreover, the
715 G' values for mixed corn starch–soy protein pastes have been found to increase significantly after
716 dry-heating (the moisture content was <10%, at 130 °C for 4 h), possibly due to enhanced
717 interactions between the involved starches and proteins (Qiu et al., 2015).

718 Steady-shear, creep-recovery, strain-sweep, stress-sweep, and time-sweep tests, also used to
719 determine the rheological properties of mixed protein–starch systems, have been helpful in
720 understanding the starch–protein interactions in such systems (Chen, Wang, et al., 2019; Feng et al.,
721 2020; Li et al., 2018; Villanueva, De Lamo, Harasym, & Ronda, 2018). It is of obvious importance to
722 establish a food system with stable rheological properties, which would be favorable to food
723 processing operations and the avoidance of collapse of product structure (Larrosa, Lorenzo, Zaritzky,
724 & Califano, 2016; McCann, Le Gall, & Day, 2016; Zhang, Mu, & Sun, 2018).

725 Specifically, from steady-shear tests, it has been observed that addition of proteins can
726 influence the flow behavior of starch-based samples to different extents, depending on the protein
727 concentration (Chen et al., 2018; Kumar et al., 2018; Qiu et al., 2015; Ronda et al., 2011; Vu Dang et
728 al., 2009; Wang, Chen, et al., 2017). For example, an increase in added wheat protein (glutenin
729 treated by protein-glutaminase) concentration from 0.5 wt% to 1.0 wt% has been reported to lead to
730 increased shear stress for a potato starch-based system, while a protein concentration further
731 increased to 1.5 wt% led to reduced shear stress for the same system (Chen et al., 2018). Higher
732 shear stress is said to indicate that the network structure of a sample is more resistant to shear (Chen

733 et al., 2018). Furthermore, replacing starches with proteins has been found to reduce the viscosity of
734 various mixed systems, compared to corresponding starch-alone systems, but to support the same
735 shear-thinning behavior (Kumar et al., 2018; Wang, Chen, et al., 2017). For example, the addition of
736 soy proteins has been found not to change the non-Newtonian shear-thinning behavior of corn starch
737 systems (Qiu et al., 2015).

738 Creep and recovery measurements can provide insights into dough macrostructure (Zhang et al.,
739 2018). It is said to be important to use large-deformation and/or -stress measurements, because the
740 resulting rheological behavior of doughs can be determined under conditions similar to those for
741 actual dough processing (Federici, Jones, Selling, Tagliasco, & Campanella, 2020). Sometimes, the
742 addition of certain proteins can decrease the strength of specific gluten-free doughs and their
743 recovery ability (Feng et al., 2020; Hernández-Estrada, Rayas-Duarte, Figueroa, & Morales-Sánchez,
744 2014; Sarabhai & Prabhasankar, 2015), but the opposite behavior can also result, due to the addition
745 of different proteins (Ronda et al., 2014). Besides, strain-sweep testing has been used to determine
746 the linear viscoelastic range for various starch-based samples, from which the effect of protein
747 addition on the moduli of such starch-based systems in this range can be observed (Feng et al., 2020;
748 Villanueva, Ronda, et al., 2018; Yu et al., 2020).

749 The following possible explanations for changes in the rheological properties of starch–protein
750 mixed gel matrices during dynamic sweeping have been proposed:

- 751 - Different WHC characteristics of proteins and starches can result in proteins competing with
752 starches for available water, which, in turn, can affect the rheological properties of various
753 mixed systems (Feng et al., 2020; Joshi et al., 2014; Ronda et al., 2011);
- 754 - Different interactions in mixed starch–protein gels can result in different rheological
755 properties. Interactions can occur between proteins and starch chains (Chen et al., 2018; Joshi
756 et al., 2014; Qiu et al., 2015; Ribotta et al., 2012; Ronda et al., 2011; Ronda et al., 2014; Sang
757 et al., 2018; Yang et al., 2019; Zhang et al., 2018), or between starch chains alone (Chen et al.,
758 2018; Ribotta et al., 2012; Zhang et al., 2018), and can lead to crosslinking in proteins
759 (Phongthai, D’Amico, Schoenlechner, & Rawdkuen, 2016; Singh & Singh, 2013; Zhang et al.,
760 2018), or to self-aggregation of proteins (Ribotta et al., 2012; Ronda et al., 2014; Zhou et al.,
761 2018), while phase separation between starches and proteins may also occur (Chen et al.,
762 2018).

763 In summary, compared to starch-only gel systems, protein addition can strongly affect starch
764 gel moduli, but such an effect can also be linked to many other factors, such as starch or protein type,
765 protein/starch ratio, and treatment and environmental conditions.

766 **4.4 Textural properties**

767 Textural information is important in food product development, quality control, and in
768 determining product shelf-life and evaluating characteristics associated with product sensory analysis
769 (Levine & Finley, 2018; Rodriguez Furlán, Pérez Padilla, & Campderrós, 2015). Levine and Finley
770 (2018) provide a comprehensive introduction to the food texture, including basic definitions of

771 texture, measurement of texture, texture profile, and applications in food. A Texture Analyzer device
 772 is often used to determine the textural properties of food product samples and measure textural
 773 parameters such as hardness, cohesiveness, resilience, springiness, and chewiness (Joshi et al., 2014;
 774 Lu et al., 2016; Yang et al., 2013). Proteins often affect the properties of starch-based gels and
 775 doughs, and thus, the resulting textural characteristics of starch-based products (e.g. breads, cookies,
 776 and pasta). As illustrated in **Table 2**, protein addition can affect the textural properties of many
 777 different types of starch-based food systems, the extent of the effects being dependent on the type
 778 and dosage of protein added, as well as on acid addition (Villanueva et al., 2015).

779

780 **Table 2** Textural properties of starch-based food systems

Test sample	Added protein and content	Textural results	Reference
Rice-based bread	Pea protein isolate (5 wt%)	Increased crumb firmness significantly	(Villanueva et al., 2015)
	Egg albumin (5 and 10 wt%)	Firmness increased at 10% level	
	Calcium caseinate (5 and 10 wt%)	Did not promote any crumb hardening	
Rice-based bread	Soy flour (10 wt%)	Diminished crumb hardness to half the value of rice-alone bread	(Sciarini et al., 2010)
Wheat-based bread	Whey protein and soy protein (0–30 wt%)	Both increased bread hardness, but the addition of whey protein	(Zhou et al., 2018)

		increased it more; gumminess and chewiness showed similarly increasing trends.	
Chinese noodles	Wheat gluten protein (7.0–19.6 wt%)	Noodle hardness increased sharply with gluten protein content >14.3%. Noodle springiness increased remarkably at 19.6% protein content.	(Zhang, Lu, Yang, & Dan Meng, 2011)
Gluten-free pasta	Rice bran protein concentrate (RBPC) and soy protein concentrate (SPC) (6 and 9 wt%, respectively)	Addition of RBPC or SPC significantly reduced the firmness of these rice flour-based gluten-free pasta.	(Phongthai et al., 2017)
Vermicelli	Defatted soy flour (DSF), whey protein concentrate (WPC) (0–15 wt%)	Addition of DSF and WPC improved hardness, springiness, and cohesiveness, but there were differences in the degrees of improvement.	(Lakshminarayan, Rajeswari, & Rao, 2010)
Cookies	Whey and casein protein concentrates and hydrolysates (0, 5, 10, and 15 wt%)	The fracture force of cookies increased significantly with increasing protein content up to 15%. The fracture force was higher with added hydrolysates than with added concentrates.	(Gani et al., 2015)

782 For various starch gel matrices, the textural profile can be significantly changed by the addition
783 of proteins. Compared to a lentil starch gel, a lentil protein gel has been reported to have a lower
784 hardness (Joshi et al., 2014). The hardness of various starch-based gels tends to decrease with the
785 addition of proteins, and the resulting mixed starch–protein gels are reported to be softer
786 (Bravo-Núñez et al., 2019; Hu et al., 2020; Joshi et al., 2014; Lu et al., 2016; Ribotta et al., 2007;
787 Sun & Xiong, 2014; Wang, Zhang, et al., 2020; Xie et al., 2017). However, in contrast, Baxter et al.
788 (2014) have found that increasing glutelin and globulin protein levels in rice starch and flour resulted
789 in linear increases in gel hardness, in agreement with their earlier findings on albumin addition to
790 rice flour (Baxter et al., 2010). The influence of added proteins on the cohesiveness, springiness, and
791 elasticity of starch-based gels is often limited by specific protein type and concentration, resulting in
792 large differences in these parameters for different mixed starch–protein gels (Baxter et al., 2014; Lu
793 et al., 2016; Sun & Xiong, 2014). For example, it has been suggested that interactions between rice
794 starch granules and water largely determine the textural properties of such starch slurries, so that
795 added proteins with strong WHCs may cause incomplete gelatinization of certain starches, thereby
796 impacting mixed gel texture (Baxter et al., 2010). Additionally, the addition of proteins can change
797 the structure of starch-based gels and thus, their texture (Bravo-Núñez et al., 2019; Joshi et al., 2014;
798 Xie et al., 2017). For example, changes in gel structure caused by interactions of proteins with starch
799 chains may affect the formation of hydrogen bonds between amylose and amylopectin or the “inert
800 filler” effect of proteins (i.e. certain proteins can act as inert filler, to hinder hydrogen bonding
801 between starch chains) (Bravo-Núñez et al., 2019; Hu et al., 2020; Xiao & Zhong, 2017).

802 An increased understanding of the texture of doughs can help to optimize dough processing
803 and predict the quality of freshly baked products (Kweon, Slade, Levine, & Gannon, 2014). The
804 effect of protein addition on dough hardness has been shown to be influenced by protein type (Barak
805 et al., 2014; Campbell, Euston, & Ahmed, 2016; Larrosa et al., 2016; Manoj Kumar et al., 2019;
806 Storck et al., 2013). For example, the addition of wheat glutenin protein has been reported to increase
807 the hardness of a control wheat flour dough, while the addition of wheat gliadin protein decreases the
808 control dough's hardness (Barak et al., 2014). Campbell et al. (2016) have reported that a wheat
809 flour-based dough containing added cowpea protein isolate was significantly harder than the
810 corresponding wheat flour-only control dough, while dough hardness was decreased by the addition
811 of thermally denatured cowpea protein isolate and was the softest with added glycated cowpea
812 protein isolate, possibly due to the enhanced WHC of such protein isolates after denaturation and
813 glycation (Campbell et al., 2016). In this way, the magnitude of the effect of protein addition on
814 dough hardness can be influenced by the WHC of such proteins, such that a protein with strong
815 WHC can significantly reduce dough hardness. Additionally, the cohesiveness and adhesiveness of
816 doughs can also be significantly affected by added proteins, with protein type being one of the key
817 factors. For instance, the stickiness of wheat flour-based bread dough has been reported to increase
818 with increasing whey protein content but decrease with the addition of soy protein (Zhou et al., 2018).
819 Elsewhere, Barak et al. (2014) have determined that added wheat gliadins were responsible for the
820 cohesiveness and extensibility of wheat flour-based doughs, while added wheat glutenins made those
821 doughs more rubbery and elastic. It has been suggested that the effect of different proteins on dough
822 texture could likely be due to corresponding differences in the hydration capacities of such proteins,

823 with higher hydration ability leading to doughs being softer and stickier, or else to the structural
824 changes in doughs caused by protein crosslinking (Campbell et al., 2016; Manoj Kumar et al., 2019;
825 Storck et al., 2013; Zhou et al., 2018).

826 Obviously, texture is an important sensory attribute of many food products, in the context of
827 assessing product acceptability (Levine & Finley, 2018; Yu et al., 2020). The textural characteristics
828 of various starch-based food systems, including firmness, springiness, and cohesiveness, are of prime
829 importance, as they can decide consumer acceptance (Shevkani & Singh, 2014). Therefore, a better
830 understanding of the effects of the type and dose of added proteins on the textural properties of
831 starch-based matrices can be highly beneficial to the production of high-quality food products.

832 **4.5 Sensory properties**

833 Sensory evaluations of many different types of starch-based food products are most commonly
834 carried out by human sensory panels (Philipp, Buckow, et al., 2017; Rosa-Sibakov et al., 2016;
835 Sarabhai & Prabhasankar, 2015). Initially, training sessions, attended by groups of sensory panelists,
836 are typically conducted to identify and agree on each relevant descriptive attribute (Philipp, Buckow,
837 et al., 2017). Participants then analyze the different sensory attributes specified, according to the
838 different sensory characteristics of the particular type of product being paneled, and finally evaluate
839 that food product's overall acceptability. Starch-based food products such as breads, cookies, and
840 noodles often contain proteins added for the purpose of adjusting sensory properties (Campbell et al.,
841 2016; Mancebo, Rodriguez, & Gómez, 2016; Manoj Kumar et al., 2019; Phongthai et al., 2016; Wani,
842 Sogi, Singh, Sharma, & Pangal, 2012). Frequently, the type and amount of added protein critically

843 determine the sensory properties of such starch-based food matrix systems(Campbell et al., 2016;
844 Gani et al., 2015; Shin et al., 2010).

845 For various starch-based food products prepared from wheat flour, wheat gluten can have a
846 greater influence than wheat starch on the sensory properties of the system, because gluten proteins
847 can form three-dimensional protein networks that impart unique viscoelasticity to such wheat
848 flour-based doughs (Kweon et al., 2014), and can limit the gelatinization of starch during baking or
849 cooking (Ortolan & Steel, 2017). Ortolan and Steel (2017) have reported that increasing the wheat
850 gluten protein content in bread dough can increase resulting bread volume, improve bread texture (i.e.
851 softness) and uniformity, and give the product a better color (i.e. golden brown). Additionally, adding
852 certain formula amounts of exogenous proteins (e.g. milk proteins, cowpea protein isolates, or
853 watermelon seed protein isolates) can improve the sensory properties of products such as pasta and
854 baked goods such as wheat bread, sponge cake, and cookies (Campbell et al., 2016; Gani et al., 2015;
855 Giménez et al., 2012; Wani et al., 2012). Campbell et al. (2016) have reported that there was a trend
856 toward higher sensory acceptability scores for wheat breads prepared with added glycated cowpea
857 protein isolate, compared to those prepared with added thermally denatured cowpea protein isolate.
858 They also found that, for sponge cakes, replacing 20% of whole egg with glycated cowpea protein
859 isolate did not affect product sensory acceptability (Campbell et al., 2016). Such findings have
860 confirmed that different types of added protein can have different effects on the sensory properties of
861 various starch-based food matrices.

862 Many recent studies have been devoted to the development of various wheat gluten-free food
863 products with desirable nutritional and acceptable sensory properties (Mancebo et al., 2016; Manoj

864 Kumar et al., 2019; Miñarro, Normahomed, Guamis, & Capellas, 2010; Phongthai et al., 2016;
865 Sarabhai & Prabhasankar, 2015; Shin et al., 2010; Villanueva et al., 2015). Most gluten-free food
866 products are based on the use of different gluten-free flours, such as from rice, buckwheat, and
867 chestnut (Mancebo et al., 2016). Because the proteins endogenous to such non-wheat flours lack the
868 ability to form viscoelastic networks unique to wheat gluten (Kweon et al., 2014), the addition of
869 exogenous proteins is often needed to improve the sensory properties of such gluten-free products.
870 For example, Sarabhai and Prabhasankar (2015) have reported that the addition of an optimal amount
871 of whey protein concentrate to cookie dough improved the color, appearance, taste, and overall
872 acceptability of the resulting gluten-free cookies, whereas the overall acceptability of those cookies
873 decreased, when the level of added protein differed from the optimized amount. Mancebo et al.
874 (2016) have found that gluten-free cookies made from mixtures of rice flour and pea protein had
875 lower hardness, darker baked color (similar to that for control cookies made from wheat flour), and
876 higher acceptability. Phongthai et al. (2016) have reported that substituting rice bran protein
877 concentrate for added egg albumin can improve the quality of gluten-free bread. However, while
878 added proteins can often improve the appearance, color, and texture of various starch-based food
879 matrices, the unpleasant odors of certain proteins can adversely affect the overall acceptability of
880 such products (Villanueva et al., 2015). Nevertheless, the use of added proteins along with TGase has
881 been reported to be able to improve the quality of some gluten-free products, such as pasta and bread
882 (Manoj Kumar et al., 2019; Shin et al., 2010). For example, Manoj Kumar et al. (2019) have found
883 that the hardness, integrity, and overall acceptability of gluten-free pasta were increased by mixing
884 pearl millet flour with milk proteins and then treating the mixtures with TGase.

885 Extrusion is one of the most widely used snack production processes, for which ingredient
886 materials that provide adequate product expansion and texture need to be selected (Witczak et al.,
887 2016). Studies have demonstrated that starch and protein types and their ratios play an important role
888 in expansion, and that the interactions between starches and proteins during extrusion can affect the
889 sensory properties of final products (Philipp, Buckow, et al., 2017; Witczak et al., 2016). For
890 example, extrudates with higher contents of added pea protein isolate have been reported to exhibit
891 higher pea flavor intensity, harder and more brittle texture, darker color, and less uniform shape and
892 surface appearance than those with higher proportions of rice flour to pea protein (Philipp, Buckow,
893 et al., 2017). In summary, it can once again be seen to be important to identify the best type and
894 proportion of added protein for use in the development of starch-based food products with
895 satisfactory sensory properties.

896 **4.6 Starch digestibility**

897 Starch digestion properties can be shown by the amount of glucose released and the contents of
898 rapidly digestible starch (RDS), slowly digestible starch (SDS), and RS (López-Barón et al., 2017).
899 Meanwhile, a first-order kinetic model and an associated logarithm of the slope (LOS) plot have
900 been applied to characterize the reaction rate of starch amyolysis (Zou, Sissons, Gidley, Gilbert, &
901 Warren, 2015; Zou, Sissons, Warren, Gidley, & Gilbert, 2016). With increasing demand for
902 low-glycemic-index foods, starch-based foods with reduced digestibility have attracted wide interest
903 (Singh et al., 2010). Starch digestibility has been determined to be significantly influenced by the
904 presence of proteins in cereal-based food systems; typically, proteins have been found to inhibit

905 starch digestion (e.g. increasing RS content and reducing digestion rate) (Bhattarai et al., 2016; Chen,
906 Wang, et al., 2019; Petitot et al., 2009; Singh et al., 2010; Toutounji et al., 2019). For example, *in*
907 *vitro* digestion experiments on pasta samples — i.e. spaghetti and powdered pasta were prepared
908 from different varieties of durum wheat semolina, and starch was purified from each variety — have
909 shown that the embedding of gluten and the compact microstructure of the pasta reduced starch
910 digestion rates (Zou et al., 2015). Oñate Narciso and Brennan (2018) have found that the amount of
911 glucose released from glutinous rice decreased with increasing content of added proteins, and that
912 both pea protein isolate and whey protein concentrate affect the amount of glucose released in a
913 similar manner. Lu et al. (2016) have reported that the RS content of processed potato starch–protein
914 mixtures was significantly affected by processing, in the order cooled after cooking > just cooked \approx
915 reheated after cooking and cooling. Furthermore, HMT can promote interactions between starches
916 and proteins, which can restrict starch hydrolysis (Chen, He, Fu, & Huang, 2015; Chen, Luo, et al.,
917 2019; Vu, Bean, Hsieh, & Shi, 2017). Thus, the digestibility of starches in starch–protein mixtures
918 can be influenced by many different factors, such as the types of starch and protein, the starch–
919 protein mixing ratio, and processing conditions (e.g. heat treatment methods).

920 To date, the following four possible mechanisms, by which proteins can affect starch digestion
921 properties, have been proposed:

- 922 - Proteins as physical barriers impact the digestion of starches (López-Barón et al., 2018; Lu et
923 al., 2016; Oñate Narciso & Brennan, 2018; Rosa-Sibakov et al., 2016; Ye et al., 2018);

- 924 - Proteins interact with starches, thereby blocking the binding sites of amylase enzyme, so that
925 starch molecules are not easily bound by amylase (Chen, He, et al., 2015; Hu et al., 2020;
926 López-Barón et al., 2017; Yang et al., 2013);
- 927 - The binding of proteins to amylase results in the inhibition of amylase activity or prevents the
928 otherwise typical binding of amylase to starch molecules (Bhattarai et al., 2016; Chen, He, et
929 al., 2019; Oñate Narciso & Brennan, 2018; Yu et al., 2018; Zou et al., 2016);
- 930 - Gelatinized starches and proteins rapidly form compact protein–amylose aggregates (Chen,
931 Wang, et al., 2019; Chi et al., 2018; Xiao et al., 2019).

932 With regard to the first mechanism, proteins can interact with starch granules and thus
933 encapsulate them, thereby acting as physical barriers that effectively hinder digestion of the starch. It
934 can be seen from CLSM images (**Fig. 4I**) that proteins can actually enwrap starch granules to form a
935 physical barrier (López-Barón et al., 2017). Along the same lines, it has been reported that the
936 inhibition of starch digestion by endogenous proteins is often due to the latter's physical barrier
937 effect that limits the contact of starch with amylase (Hu et al., 2017; Ye et al., 2018). Moreover,
938 protein networks can encapsulate starches, thereby limiting starch swelling and gelatinization and
939 starch's contact with digestive enzymes (Chen, Wang, et al., 2019; Chen, He, et al., 2015; Feng et al.,
940 2020; Yang et al., 2019).

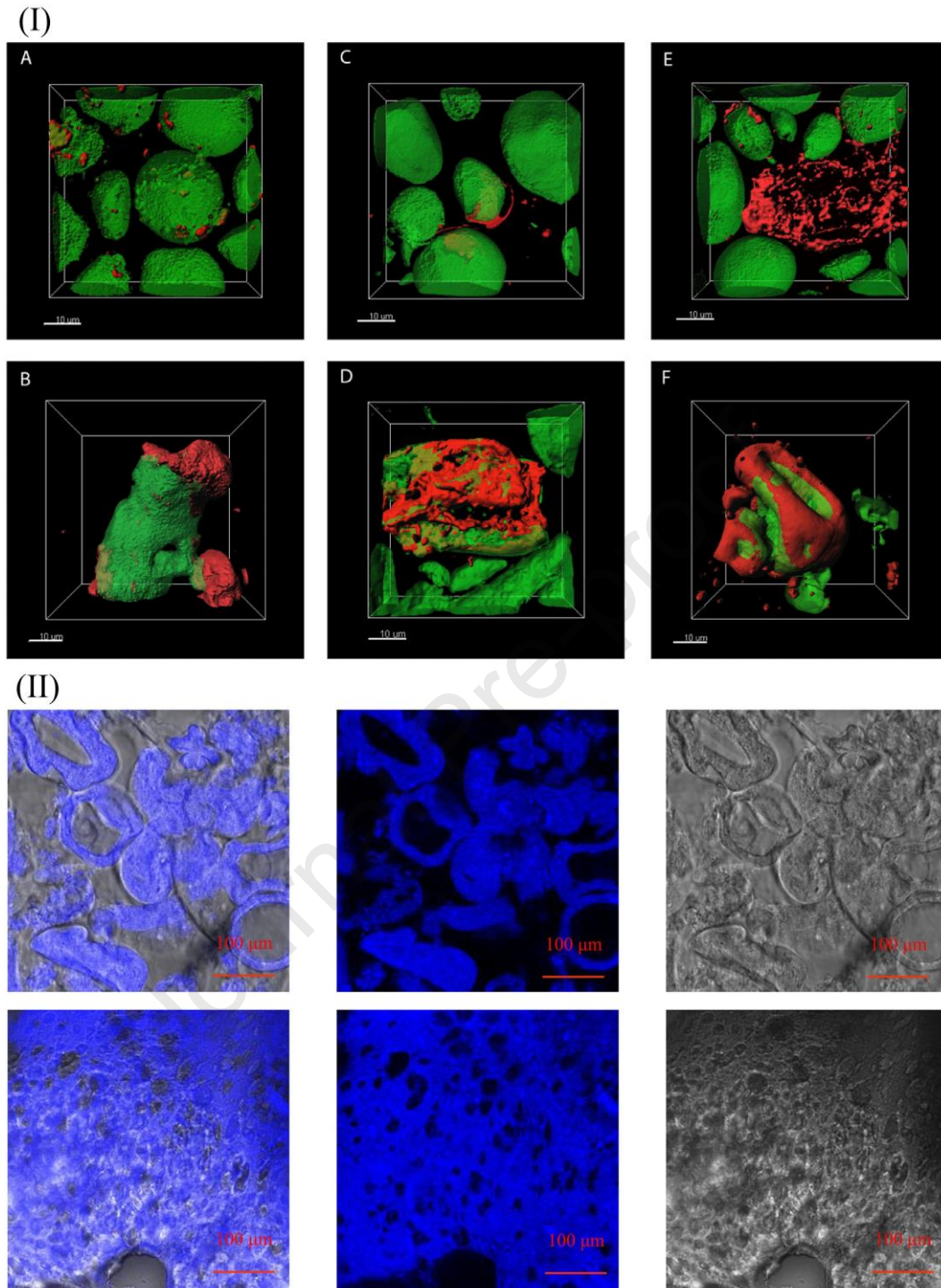
941 With regard to the second mechanism, Yang et al. (2013) have suggested that combinings
942 between whey protein and wheat starch chains may reduce the number of available action sites for
943 enzymes on the starch. Starch–protein interactions can play an important role in this mechanism,

944 wherein such interactions depend on the molecular configurations of proteins (López-Barón et al.,
945 2017).

946 With regard to the third mechanism, CLSM images of FITC (fluorescein isothiocyanate)
947 -amylase conjugates in the presence of cooked protein (**Fig. 4II**) suggest that the interactions
948 between α -amylase and proteins were sufficiently strong to cause the starch to compete with the
949 protein to bind digestive enzymes, thereby affecting the digestive properties of the starch (Chen, He,
950 et al., 2019).

951 With regard to the fourth mechanism, Chi et al. (2018) have presented a schematic diagram
952 illustrating the effect of rice protein and its hydrolysates on starch digestibility, indicating that some
953 starch–rice protein samples had higher double-helix content and more compact, aggregated structures
954 with less amylose leaching during short-term cooling, thus inhibiting the otherwise usual attack by
955 digestive enzymes.

956



957

958 **Fig. 4 (I)** Confocal laser scanning microscopy (CLSM) images of wheat starch in the presence of
 959 purified plant proteins. Wheat starch and cellulose were mixed with (A) pea protein, no cooking, (B)
 960 denatured pea protein, pressure-cooked, (C) rice protein, no cooking (D) hydrolyzed rice protein,
 961 pressure-cooked, (E) soybean protein, no cooking, (F) hydrolyzed soybean protein, pressure-cooked.

962 Reprinted from López-Barón et al. (2017), copyright (2017), with permission from Elsevier. **(II)**
963 Representative CLSM images of FITC (fluorescein isothiocyanate)-amylase conjugates bound onto
964 cooked protein surfaces after two successive washings: soy protein isolate (upper) and wheat gluten
965 protein (bottom). Reprinted from Chen, He, et al. (2019), copyright (2019), with permission from
966 Elsevier.

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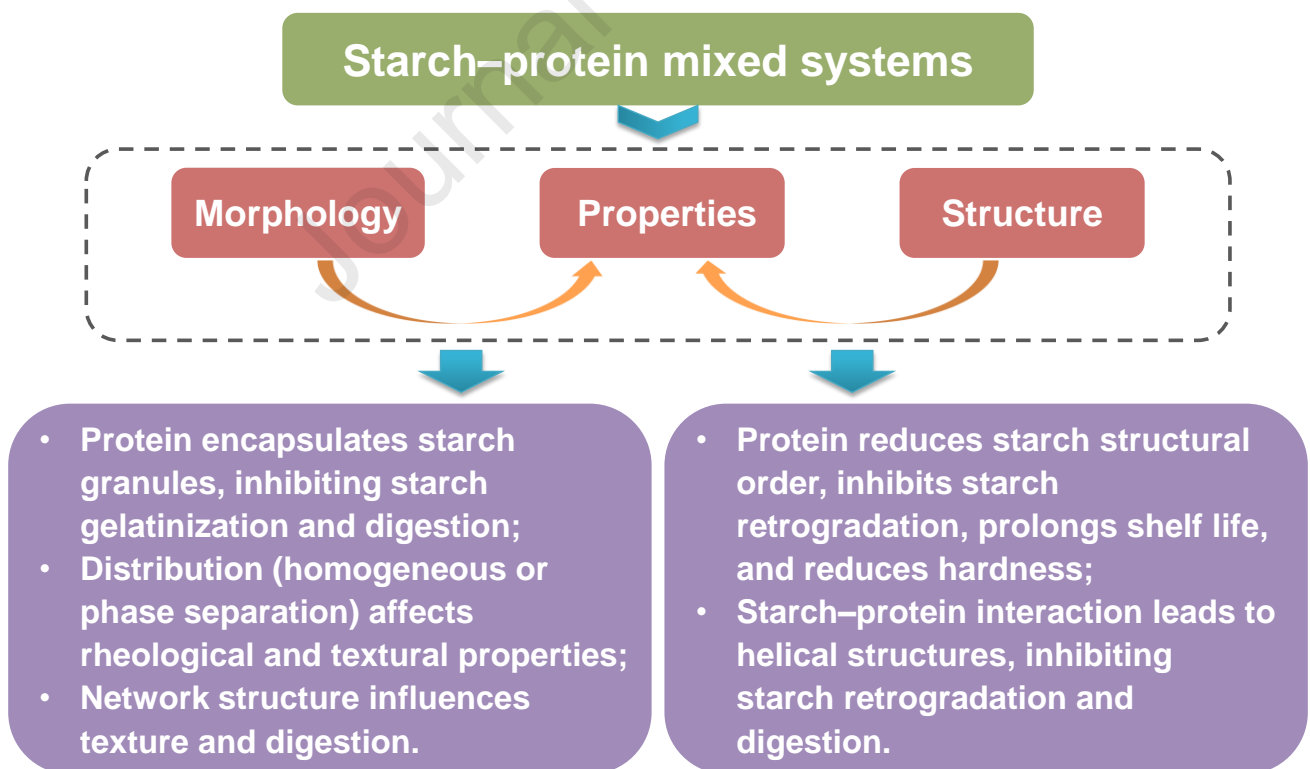
968 **4.7 Morphology/structure–property relationships**

969 Although the structure and physicochemical properties of many starch–protein mixtures have
970 been extensively studied, the structural reasons for changes in physicochemical properties are still
971 not fully understood. To date, structure–property relationships for various starch–protein systems
972 have been established to only a limited extent, and these generalized relationships are described in
973 **Fig. 5.** Microscopic observations have revealed that proteins can be adsorbed on the surfaces of
974 starch granules, thus hindering water molecules from entering the granules and limiting contacts
975 between the starch and amylase, thereby ultimately inhibiting starch gelatinization and digestion
976 (López-Barón et al., 2017; Phongthai et al., 2017; Yang et al., 2019). Protein networks formed in
977 certain starch-based doughs can encapsulate starch granules, ultimately with a similar effect,
978 especially for product applications involving pastas with reduced digestibility (Feng et al., 2020;
979 Giuberti et al., 2015). Porous starch–protein gel networks can be responsible for endowing such gels
980 with elastic/spongy properties and promoting water retention, so the network structures of such gels
981 can provide them with particular textural properties (Joshi et al., 2014). For instance, Sun and Xiong
982 (2014) have reported that the network structure formed by a pea starch–peanut protein mixed gel was
983 significantly different from the three-dimensional network structure of a pea starch-alone gel matrix,

984 and the network structure of the former was affected by the type and proportion of added protein,
 985 which ultimately influenced the textural properties of the mixed gel. In another example, Joshi et al.
 986 (2014) have reported that the hardness of lentil starch–lentil protein composite gels was reduced, due
 987 to the physical separation of protein-rich domains, which may have been related to the failure of such
 988 weak lentil protein networks to penetrate such strong lentil starch gel networks. Wang, Zhang, et al.
 989 (2020) have suggested that the presence of protein can make the network structure of a mixed rice
 990 flour-based gel somewhat more compact, because filling by the protein can make the resulting pore
 991 size of the gel smaller, which would be conducive to greater softness of the gel.

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995 **Fig. 5** Overview of the relationships between the morphology/structure and properties of starch–

996 protein mixed systems

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998 The consequences of long-range and short-range ordered structures have been suggested to
999 include the fact that the presence of added protein can generally reduce the extent of starch structural
1000 order during retrogradation, thus indicating that protein can inhibit starch retrogradation and thereby
1001 help to prolong the shelf-life of various starch-based foods (Xiao & Zhong, 2017; Zhang et al., 2019).
1002 Hu et al. (2020) have observed that the significant decreases in rice starch gel hardness and water
1003 mobility after the addition of whey protein hydrolysate were related to the protein's retarding effect
1004 on starch retrogradation. However, in other instances, the addition of rice proteins has been found to
1005 enhance the ordered structure of rice starch, possibly due to interactions between the starch and
1006 protein to form more stable helical structures, which significantly increased the hardness of the
1007 resulting starch–protein gels and thereby decreased starch digestibility (Chi et al., 2018; Wang,
1008 Zhang, et al., 2020). The RS content of various processed starch–protein mixtures has been
1009 positively correlated with starch short-range order; in other words, the short-range ordered structures
1010 of such mixtures were able to be correlated with starch digestive characteristics (Chen, Wang, et al.,
1011 2019; Lu et al., 2016; Xiao et al., 2019). Thus, in the ways just described, the gelatinization,
1012 retrogradation, textural, and digestion characteristics of various starch–protein mixtures are closely
1013 related to their structural changes. Further investigations need to be conducted to more firmly
1014 strengthen the morphology/structure–property links for such starch-based food matrices, and thereby
1015 to facilitate the production of starch-based food products with desirable qualities and functions.

5. Applications in food processing and production

Pastas, noodles, breads, steamed breads, biscuits, cakes and many other starch-based food products are popular with consumers worldwide. Their textural, sensory, and nutritional properties have attracted considerable research attention. Such starch-based food products are most commonly prepared from wheat flours, of which the gluten proteins can often form continuous three-dimensional network structures that can encapsulate wheat starch granules, and which are vital to the formation of elastic doughs, from which, through extrusion or baking processes, specific products with certain desirable textural and sensory properties can be produced (Kim et al., 2008; Kweon et al., 2014; Levine & Finley, 2018; Slade, Kweon, & Levine, 2021; Wang, Guo, & Zhu, 2016). Studies (Mohammed, Ahmed, & Senge, 2012; Zhou et al., 2018) have revealed that the addition of certain exogenous proteins can disrupt the continuity of wheat gluten networks, leading to poor texture and other quality defects of baked starch-based products such as breads, while the color of such starch-based matrix products has been found to be closely related to the color of the added proteins. The effects of added proteins on various types of starch-based food products are summarized in **Table 3**.

Table 3 Influence of proteins on properties of starch-based food products

Application	Major components	Characteristics	Reference
Pasta	Durum wheat semolina (68.6% starch, 11.9%	- Protein network encapsulating starch granules can delay starch digestion; - Mixing or sheeting processes	(Kim et al., 2008)

	protein)	produced elastic doughs;	
		- Increased sheeting might reduce starch–protein interactions, thereby increasing starch accessibility to α -amylase.	
Gluten-free pasta	Rice flour, egg albumen (EB, 74.7% protein), rice bran protein concentrate (RBPC, 68.1% protein)	- EB-rich pasta showed a compact and homogenous structure, reduced cooking loss, and improved hardness; - RBPC-rich pasta showed cracked and non-continuous surfaces, resulting in high cooking loss and low firmness.	(Phongthai et al., 2017)
Chinese steamed bread	Wheat flour (11.2% protein)	- Formation of a continuous and three-dimensional gluten network; - The starch granules were embedded in the protein network; - Gluten polymerization was conducive to retaining gas and restricting starch swelling.	(Wang et al., 2016)
Bread	Wheat flour (63.5% starch, 11.9% protein), chickpea flour (51.2% starch, 25.5% protein)	- Addition of chickpea flour increased dough development time, stability, and tensile properties; - Addition of chickpea flour significantly increased bread crumb hardness; - Bread crust color became darker with increased chickpea flour level.	(Mohammed et al., 2012)
Bread	Wheat flour (13.2% protein), whey protein	- Addition of whey protein improved gas retention ability during baking	(Zhou et

	(76.4% protein), soy protein (86.6% protein)	and produced better specific bread loaf volume; - Weak gel formed by soy protein and its poor heat preservation ability led to a decrease in bread volume and poor bread quality.	al., 2018)
Gluten-free bread	Rice flour (90.6% starch, 8.1% protein), corn flour (86.1% starch, 6.9% protein), soy flour (30.6% starch, 55.0% protein)	- High water affinity of soy protein and starch–protein interactions influenced starch gelatinization; - Soy flour incorporation increased bread volume, possibly due to increased batter consistency.	(Sciarini et al., 2010)
Gluten-free cookies	Rice flour (74.4% starch, 8.0% protein), pea protein (80% protein)	- Protein incorporation improved hydration properties of the mixtures and dough consistency; - Cookies with higher protein content showed higher acceptability.	(Mancebo et al., 2016)

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For various gluten-free food products, rice flour is typically used along with added exogenous proteins to obtain products comparable to traditional wheat-based counterparts, but there are still product issues related to sensory properties, nutritional value, and consumer acceptance (Rodriguez Furlán et al., 2015; Shevkani et al., 2015; Storck et al., 2013; Villanueva, Ronda, et al., 2018). Thus, gluten-free formulations have been optimized for greater consumer acceptability, for example, by the selection of appropriate individual proteins or protein combinations or by the addition of other components such as polysaccharides (Gularte, Gómez, & Rosell, 2012; Phongthai et al., 2017;

1041 Rodriguez Furlán et al., 2015; Sarabhai & Prabhasankar, 2015; Sciarini et al., 2010). Since such
1042 gluten-free starch-based products are mainly composed of starch and protein, it should be intuitively
1043 obvious that the characteristics of the particular starches and proteins and their interactions can affect
1044 the sensory and nutritional properties of the final food products. For instance, researchers have found
1045 that replacing rice flour with pea protein or maize starch could help to change the gluten free cookie
1046 characteristics (Mancebo et al., 2016). The inclusion of protein in the formulation reduced cookie
1047 size (thickness and width), resulting in cookies with lower hardness values and darker color, while
1048 the addition of starch increased cookie size without affecting texture or color (Mancebo et al., 2016).
1049 However, considering that protein addition modifies cookie dough rheology, resulting in a more
1050 consistent dough, the problem of cookie lamination and formation can be solved if the dough is too
1051 soft (Mancebo et al., 2016).

1052 Many pastas and noodles are typically made by mixing wheat flours or semolinas with water to
1053 form unfermented doughs, followed by extrusion (Baik, 2010; Li, Zhu, et al., 2014). The quality of
1054 cooked pastas or noodles is known to be influenced by gluten protein network formation during
1055 dough mixing and wheat starch gelatinization during cooking (Bonomi et al., 2012; Bruneel, Pareyt,
1056 Brijs, & Delcour, 2010). When a given gluten network lacks elasticity and compactness, starch
1057 granules tend to swell more during cooking, resulting in a greater loss of soluble solids (Bonomi et
1058 al., 2012). However, excessive addition of exogenous wheat gluten can make a dough too strong to
1059 be easily and efficiently handled during subsequent rolling or extrusion (Li, Zhu, et al., 2014). In
1060 contrast, certain exogenous proteins can positively affect the formation of protein networks in
1061 doughs, thereby enhancing the structures of pasta and noodle doughs and improving the sensory

1062 properties of the final products (Li, Zhu, et al., 2014; Marti et al., 2014; Susanna & Prabhasankar,
1063 2013). For example, the addition of egg albumin to rice flour-based gluten-free pasta dough can lead
1064 to the product having less tendency toward structural disintegration, thereby reducing cooking losses
1065 (Phongthai et al., 2017). It has been said that starch chains, especially amylose, in cooked noodles
1066 can interact with wheat gluten proteins through hydrogen bonding (Baik, 2010; Liu et al., 2019). In
1067 this way, amylose chains exuded during cooking may cause rigidity of cooked noodles by interacting
1068 with gluten proteins (Baik, 2010). The compact structure and the interactions between starches and
1069 proteins can also lead to reduced starch digestibility of pastas (Kim et al., 2008; Zou et al., 2016).

1070 For bread doughs, high elasticity and extensibility are typically required during fermentation
1071 and baking, in order to enable dough expansion and retention of leavening gases (Liu et al., 2019).
1072 Wheat gluten is well-known to play a key role in determining the processing and baking quality of
1073 wheat flours, by imparting WHC, viscosity, cohesiveness, and elasticity to bread doughs
1074 (Hernández-Estrada et al., 2014; Zhou et al., 2018). Hence, the absence of wheat gluten can often
1075 result in a paste rather than a dough, which, in turn, may result in low loaf volume, poor texture, and
1076 other post-baking quality defects of baked breads (Phongthai et al., 2016; Witczak et al., 2016; Zhou
1077 et al., 2018). However, it has been reported that gluten-free bread doughs can be rendered capable of
1078 forming protein network structures, similar to those of gluten networks, by the addition of certain
1079 exogenous proteins (e.g. soybean, egg and pea proteins) and TGase (Nozawa, Ito, & Arai, 2016;
1080 Sciarini et al., 2010; Shin et al., 2010). For example, Shin et al. (2010) have reported that added
1081 whey protein and caseinate reduced the paste viscosity of rice flour, whereas TGase increased the
1082 viscosity by crosslinking between rice protein and the added proteins, and the resulting crosslinked

1083 protein network contributed to gas retention during baking. Exogenous proteins are typically
1084 incorporated into gluten-free systems to increase elasticity by crosslinking, and to enhance color and
1085 flavor development by Maillard reactions, improve structures by gelation, and support foaming
1086 (Campbell et al., 2016; Phongthai et al., 2016). Protein unfolding and protein–protein interactions, as
1087 well as protein interactions with starches, can occur during baking, leading to improved textural
1088 properties (Phongthai et al., 2016). For instance, the springiness of a model-system bread has been
1089 reported to result from a combination of gluten protein aggregation, interactions between cassava
1090 starch and gluten, and water retention (Liu et al., 2019). Moreover, starch granules embedded in a
1091 gluten-free protein matrix produced by baking can result in hindered starch retrogradation (Phongthai
1092 et al., 2016). The high WHC of soybean protein and its established interactions with amylopectin can
1093 also hinder the starch retrogradation process (Sciarini et al., 2010).

1094 Other baked products such as cookies, crackers, and cakes have been reviewed by several
1095 researchers with a focus on their major ingredients, formulas, processes, and effect on quality
1096 characteristics (Kweon et al., 2014; Slade et al., 2021). Kweon et al. (2014) also reported that gluten
1097 development was promoted in lower-sugar cracker doughs during mixing and sheeting, which was a
1098 key factor affecting the quality of baked-crackers. Besides, further studies reported that for these
1099 baked products, the protein structures in pastes or doughs and the distribution and interactions of
1100 starches with proteins can affect the textural and sensory properties of the final products (Gularte et
1101 al., 2012; Kweon et al., 2014; Mancebo et al., 2016; Sarabhai & Prabhasankar, 2015; Slade et al.,
1102 2021). It has been reported that a gelatinized starch gel contributes to initial cake crumb firmness
1103 (Slade et al., 2021), while the aggregated gluten proteins in cake crumb are responsible for crumb

1104 elasticity (Wilderjans, Luyts, Goesaert, Brijs, & Delcour, 2010). A large collection of literature has
1105 reported the effects of various starch–protein mixtures on the properties of many such final baked
1106 products (Gularte et al., 2012; Kim et al., 2008; Mancebo et al., 2016; Phongthai et al., 2017; Sciarini
1107 et al., 2010; Wang et al., 2016; Zhou et al., 2018). Different starches and proteins have been added to
1108 model dough systems, in order to prepare doughs with unique microstructures and textural properties
1109 (Liu et al., 2019; Sarabhai & Prabhasankar, 2015; Zhou et al., 2018). In such ways, starch-based
1110 products with particular sensory properties have been obtained by extrusion, cooking, or baking
1111 processes (Kumar et al., 2017; Reddy Surasani et al., 2019; Sarabhai & Prabhasankar, 2015).

1112 **6. Conclusions and future perspectives**

1113 Many starch-based foods, consisting mainly of starch matrices containing proteins, are
1114 important energy and nutrient sources for people. In such starchy foods, the presence of endogenous
1115 and exogenous proteins has been shown to have a great impact on their textural, sensory, and
1116 digestive properties. Based on descriptions of the structural characteristics of various starch–protein
1117 mixtures on different scales (i.e. micromorphology, and long-range and short-range ordered
1118 structures), the effects of different proteins on the structural characteristics of different starches have
1119 been demonstrated. In particular, proteins with stronger gelation ability and interaction with starch
1120 molecules could lead to starch–protein systems with a more compact network structure and being
1121 more homogeneous; however, some proteins interact with starch molecules to hinder the formation
1122 of a dense network structure. Usually, the interaction of proteins with starch inhibits starch molecular
1123 rearrangement during cooling, leading to decreased amounts of long-range crystallites and

1124 short-range helices. But under certain conditions, proteins interact with starch strongly, increasing
1125 the extent of crystallinity, and protein adsorption on the surface of starch granules inhibits starch
1126 gelatinization, enhancing short-range order. At the same time, the presence of different proteins in
1127 starch-based food systems has been shown to significantly affect the viscosity, thermal, rheological,
1128 textural, and digestive properties of such mixed systems. Due to the WHC, dilution and hindrance
1129 effects of proteins and to starch–protein interactions, starch–protein systems generally showed a
1130 decrease in viscosity, an increase in the starch gelatinization temperature, and alterations in the
1131 rheological and textural properties. Starch digestion could be inhibited due to the physical barrier
1132 effect of proteins, starch–protein interactions, protein binding to amylase, and the formation of
1133 starch–protein aggregates. However, the currently available information, in terms of structure–
1134 property relationships, is still limited. Additionally, various types of starch–protein interactions have
1135 been shown to play key roles in the quality and functional properties of many starch-based foods.
1136 Therefore, a thorough understanding of such starch–protein interactions in many different
1137 starch-based food systems can be of great significance to the development of products with stable
1138 thermal and rheological properties, and can guide the processing of such foods, aimed at creating
1139 desirable structures and properties.

1140 Looking ahead, starch and protein source materials can be reconstituted, in order to enable the
1141 development of starch-based products with particular textural, sensory, and nutritional properties,
1142 through specific processing techniques. There is still a strong need to define more comprehensive
1143 relationships among biopolymer interactions, food product structures, and product properties.
1144 Innovative techniques for the manipulation of starch–protein interactions could open a new

1145 dimension for health and nutrition platforms. Furthermore, while many recent studies have reported
1146 that the addition of proteins can have a significant impact on the texture, taste, and other aspects of
1147 various starch-based products (e.g. breads, cookies, cakes, and pastas), there is still a need for more
1148 systematic studies on how the addition of proteins affects the many characteristics of such final
1149 products. It is of paramount importance to better understand how the above knowledge can impact
1150 starch-based food products and processes, and thus the food industry as a whole.

1151 **Conflict of Interest**

1152 Declarations of interest: none

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1157 **References**

1158 Ai, Y., & Jane, J.-l. (2015). Gelatinization and rheological properties of starch. *Starch - Stärke*,
1159 67(3-4), 213-224.

1160 Amjid, M., Shehzad, A., Hussain, S., Shabbir, M. A., Khan, M. R., & Shoaib, M. (2013). A
1161 comprehensive review on wheat flour dough rheology. *Pakistan Journal of Food Sciences*, 23,
1162 105-123.

- 1163 Aryee, A. N. A., Agyei, D., & Udenigwe, C. C. (2018). Impact of processing on the chemistry and
1164 functionality of food proteins. In R. Y. Yada (Ed.), *Proteins in Food Processing (Second*
1165 *Edition)* (pp. 27-45): Woodhead Publishing/Elsevier.
- 1166 Aryee, A. N. A., & Boye, J. I. (2017). Comparative study of the effects of processing on the
1167 nutritional, physicochemical and functional properties of lentil. *Journal of Food Processing*
1168 *and Preservation*, 41(1), e12824.
- 1169 Baik, B.-K. (2010). Effects of flour protein and starch on noodle quality. In G. G. Hou (Ed.), *Asian*
1170 *Noodles: Science, Technology, and Processing* (pp. 261-283). Hoboken, NJ, USA: John Wiley
1171 & Sons, Inc.
- 1172 Barak, S., Mudgil, D., & Khatkar, B. S. (2014). Influence of gliadin and glutenin fractions on
1173 rheological, pasting, and textural properties of dough. *International Journal of Food Properties*,
1174 17(7), 1428-1438.
- 1175 Baxter, G., Blanchard, C., & Zhao, J. (2014). Effects of glutelin and globulin on the physicochemical
1176 properties of rice starch and flour. *Journal of Cereal Science*, 60(2), 414-420.
- 1177 Baxter, G., Zhao, J., & Blanchard, C. (2010). Albumin significantly affects pasting and textural
1178 characteristics of rice flour. *Cereal Chemistry*, 87(3), 250-255.
- 1179 Bhattarai, R. R., Dhital, S., & Gidley, M. J. (2016). Interactions among macronutrients in wheat flour
1180 determine their enzymic susceptibility. *Food Hydrocolloids*, 61, 415-425.
- 1181 Bonomi, F., D'Egidio, M. G., Iametti, S., Marengo, M., Marti, A., Pagani, M. A., & Ragg, E. M.
1182 (2012). Structure–quality relationship in commercial pasta: A molecular glimpse. *Food*
1183 *Chemistry*, 135(2), 348-355.

- 1184 Boye, J., Zare, F., & Pletch, A. (2010). Pulse proteins: Processing, characterization, functional
1185 properties and applications in food and feed. *Food Research International*, 43(2), 414-431.
- 1186 Brandner, S., Becker, T., & Jekle, M. (2019). Classification of starch-gluten networks into a
1187 viscoelastic liquid or solid, based on rheological aspects — A review. *International Journal of*
1188 *Biological Macromolecules*, 136, 1018-1025.
- 1189 Bravo-Núñez, Á., Garzón, R., Rosell, M. C., & Gómez, M. (2019). Evaluation of starch–protein
1190 interactions as a function of pH. *Foods*, 8(5), 155.
- 1191 Bruneel, C., Pareyt, B., Brijs, K., & Delcour, J. A. (2010). The impact of the protein network on the
1192 pasting and cooking properties of dry pasta products. *Food Chemistry*, 120(2), 371-378.
- 1193 Cabrera-Chávez, F., Calderón de la Barca, A. M., Islas-Rubio, A. R., Marti, A., Marengo, M., Pagani,
1194 M. A., Bonomi, F., & Iametti, S. (2012). Molecular rearrangements in extrusion processes for
1195 the production of amaranth-enriched, gluten-free rice pasta. *LWT - Food Science and*
1196 *Technology*, 47(2), 421-426.
- 1197 Campbell, L., Euston, S. R., & Ahmed, M. A. (2016). Effect of addition of thermally modified
1198 cowpea protein on sensory acceptability and textural properties of wheat bread and sponge
1199 cake. *Food Chemistry*, 194, 1230-1237.
- 1200 Chanvrier, H., Colonna, P., Della Valle, G., & Lourdin, D. (2005). Structure and mechanical
1201 behaviour of corn flour and starch–zein based materials in the glassy state. *Carbohydrate*
1202 *Polymers*, 59(1), 109-119.
- 1203 Chávez-Murillo, C. E., Veyna-Torres, J. I., Cavazos-Tamez, L. M., de la Rosa-Millán, J., &
1204 Serna-Saldívar, S. O. (2018). Physicochemical characteristics, ATR-FTIR molecular

- 1205 interactions and in vitro starch and protein digestion of thermally-treated whole pulse flours.
1206 *Food Research International*, 105, 371-383.
- 1207 Chen, B., Wang, Y.-R., Fan, J.-L., Yang, Q., & Chen, H.-Q. (2019). Effect of glutenin and gliadin
1208 modified by protein-glutaminase on retrogradation properties and digestibility of potato starch.
1209 *Food Chemistry*, 301, 125226.
- 1210 Chen, B., Zhang, B., Li, M.-N., Xie, Y., & Chen, H.-Q. (2018). Effects of glutenin and gliadin
1211 modified by protein-glutaminase on pasting, rheological properties and microstructure of
1212 potato starch. *Food Chemistry*, 253, 148-155.
- 1213 Chen, D., Fang, F., Federici, E., Campanella, O., & Jones, O. G. (2020). Rheology, microstructure
1214 and phase behavior of potato starch-protein fibril mixed gel. *Carbohydrate Polymers*, 239,
1215 116247.
- 1216 Chen, W., Zhou, H., Yang, H., & Cui, M. (2015). Effects of charge-carrying amino acids on the
1217 gelatinization and retrogradation properties of potato starch. *Food Chemistry*, 167, 180-184.
- 1218 Chen, X., He, X., Fu, X., & Huang, Q. (2015). In vitro digestion and physicochemical properties of
1219 wheat starch/flour modified by heat-moisture treatment. *Journal of Cereal Science*, 63,
1220 109-115.
- 1221 Chen, X., He, X., Zhang, B., Sun, L., Liang, Z., & Huang, Q. (2019). Wheat gluten protein inhibits
1222 α -amylase activity more strongly than a soy protein isolate based on kinetic analysis.
1223 *International Journal of Biological Macromolecules*, 129, 433-441.

- 1224 Chen, X., Luo, J., Fu, L., Cai, D., Lu, X., Liang, Z., Zhu, J., & Li, L. (2019). Structural,
1225 physicochemical, and digestibility properties of starch-soybean peptide complex subjected to
1226 heat moisture treatment. *Food Chemistry*, 297, 124957.
- 1227 Chi, C., Li, X., Zhang, Y., Chen, L., & Li, L. (2018). Understanding the mechanism of starch
1228 digestion mitigation by rice protein and its enzymatic hydrolysates. *Food Hydrocolloids*, 84,
1229 473-480.
- 1230 Chinma, C. E., Ariahu, C. C., & Abu, J. O. (2013). Chemical composition, functional and pasting
1231 properties of cassava starch and soy protein concentrate blends. *Journal of Food Science and
1232 Technology*, 50(6), 1179-1185.
- 1233 Comfort, S., & Howell, N. K. (2002). Gelation properties of soya and whey protein isolate mixtures.
1234 *Food Hydrocolloids*, 16(6), 661-672.
- 1235 Considine, T., Noisuwan, A., Hemar, Y., Wilkinson, B., Bronlund, J., & Kasapis, S. (2011).
1236 Rheological investigations of the interactions between starch and milk proteins in model dairy
1237 systems: A review. *Food Hydrocolloids*, 25(8), 2008-2017.
- 1238 Consoli, L., Dias, R. A. O., Rabelo, R. S., Furtado, G. F., Sussulini, A., Cunha, R. L., & Hubinger, M.
1239 D. (2018). Sodium caseinate-corn starch hydrolysates conjugates obtained through the Maillard
1240 reaction as stabilizing agents in resveratrol-loaded emulsions. *Food Hydrocolloids*, 84,
1241 458-472.
- 1242 de Oliveira, F. C., Coimbra, J. S. d. R., de Oliveira, E. B., Zuñiga, A. D. G., & Rojas, E. E. G. (2016).
1243 Food protein-polysaccharide conjugates obtained via the maillard reaction: A review. *Critical
1244 reviews in food science and nutrition*, 56(7), 1108-1125.

- 1245 Federici, E., Jones, O. G., Selling, G. W., Tagliasco, M., & Campanella, O. H. (2020). Effect of zein
1246 extrusion and starch type on the rheological behavior of gluten-free dough. *Journal of Cereal*
1247 *Science*, *91*, 102866.
- 1248 Feng, Y.-Y., Mu, T.-H., Zhang, M., & Ma, M.-M. (2020). Effects of different polysaccharides and
1249 proteins on dough rheological properties, texture, structure and in vitro starch digestibility of
1250 wet sweet potato vermicelli. *International Journal of Biological Macromolecules*, *148*, 1-10.
- 1251 Fernández-Alonso, M. d. C., Díaz, D., Berbis, M. Á., Marcelo, F., Cañada, J., & Jiménez-Barbero, J.
1252 (2012). Protein-carbohydrate interactions studied by NMR: From molecular recognition to
1253 drug design. *Current protein and peptide science*, *13*(8), 816-830.
- 1254 Firoozmand, H., Murray, B. S., & Dickinson, E. (2012). Microstructure and elastic modulus of mixed
1255 gels of gelatin+oxidized starch: Effect of pH. *Food Hydrocolloids*, *26*(1), 286-292.
- 1256 Flores-Morales, A., Jiménez-Estrada, M., & Mora-Escobedo, R. (2012). Determination of the
1257 structural changes by FT-IR, Raman, and CP/MAS ¹³C NMR spectroscopy on retrograded
1258 starch of maize tortillas. *Carbohydrate Polymers*, *87*(1), 61-68.
- 1259 Foegeding, E. A., & Davis, J. P. (2011). Food protein functionality: A comprehensive approach. *Food*
1260 *Hydrocolloids*, *25*(8), 1853-1864.
- 1261 Fu, Z., Chen, J., Luo, S.-J., Liu, C.-M., & Liu, W. (2015). Effect of food additives on starch
1262 retrogradation: A review. *Starch - Stärke*, *67*(1-2), 69-78.
- 1263 Gani, A., Broadway, A. A., Ahmad, M., Ashwar, B. A., Wani, A. A., Wani, S. M., Masoodi, F. A., &
1264 Khatkar, B. S. (2015). Effect of whey and casein protein hydrolysates on rheological, textural
1265 and sensory properties of cookies. *Journal of Food Science and Technology*, *52*(9), 5718-5726.

- 1266 Gao, J., Koh, A. H. S., Tay, S. L., & Zhou, W. (2017). Dough and bread made from high- and
1267 low-protein flours by vacuum mixing: Part 1: Gluten network formation. *Journal of Cereal*
1268 *Science*, 74, 288-295.
- 1269 Ghosh, A. K., & Bandyopadhyay, P. (2012). Polysaccharide-protein interactions and their relevance
1270 in food colloids. In D. N. Karunaratne (Ed.), *The Complex World of Polysaccharides* (pp.
1271 395-408): IntechOpen.
- 1272 Ghumman, A., Kaur, A., & Singh, N. (2016). Functionality and digestibility of albumins and
1273 globulins from lentil and horse gram and their effect on starch rheology. *Food Hydrocolloids*,
1274 61, 843-850.
- 1275 Giménez, M. A., Drago, S. R., De Greef, D., Gonzalez, R. J., Lobo, M. O., & Samman, N. C. (2012).
1276 Rheological, functional and nutritional properties of wheat/broad bean (*Vicia faba*) flour blends
1277 for pasta formulation. *Food Chemistry*, 134(1), 200-206.
- 1278 Giuberti, G., Gallo, A., Cerioli, C., Fortunati, P., & Masoero, F. (2015). Cooking quality and starch
1279 digestibility of gluten free pasta using new bean flour. *Food Chemistry*, 175, 43-49.
- 1280 Gularte, M. A., Gómez, M., & Rosell, C. M. (2012). Impact of legume flours on quality and in vitro
1281 digestibility of starch and protein from gluten-free cakes. *Food and Bioprocess Technology*,
1282 5(8), 3142-3150.
- 1283 Guo, J., Lian, X., Kang, H., Gao, K., & Li, L. (2016). Effects of glutenin in wheat gluten on
1284 retrogradation of wheat starch. *European Food Research and Technology*, 242(9), 1485-1494.
- 1285 Heertje, I. (2014). Structure and function of food products: A review. *Food Structure*, 1(1), 3-23.

- 1286 Heredia - Olea, E., Contreras - Alvarado, M. D., Perez - Carrillo, E., Rosa - Millón, J. D. I., &
1287 Serna - Saldivar, S. O. (2019). Assessment of the techno - functionality, starch digestion rates
1288 and protein quality of rice flour - whey protein instant powders produced in a twin extruder.
1289 *International Journal of Food Science and Technology*, 55(2), 878-890.
- 1290 Hernández-Estrada, Z. J., Rayas-Duarte, P., Figueroa, J. D. C., & Morales-Sánchez, E. (2014). Creep
1291 recovery tests to measure the effects of wheat glutenins on doughs and the relationships to
1292 rheological and breadmaking properties. *Journal of Food Engineering*, 143, 62-68.
- 1293 Hu, P., Fan, X., Lin, L., Wang, J., Zhang, L., & Wei, C. (2017). Effects of surface proteins and lipids
1294 on molecular structure, thermal properties, and enzymatic hydrolysis of rice starch. *Food
1295 Science and Technology*, 38(1), 84-90.
- 1296 Hu, Y., He, C., Zhang, M., Zhang, L., Xiong, H., & Zhao, Q. (2020). Inhibition from whey protein
1297 hydrolysate on the retrogradation of gelatinized rice starch. *Food Hydrocolloids*, 108, 105840.
- 1298 Jamilah, B., Mohamed, A., Abbas, K. A., Abdul Rahman, R., Karim, R., & Hashim, D. M. (2009).
1299 Protein-starch interaction and their effect on thermal and rheological characteristics of a food
1300 system: A review. *Journal of Food Agriculture and Environment* 7(2), 169-174.
- 1301 Jekle, M., Mühlberger, K., & Becker, T. (2016). Starch–gluten interactions during gelatinization and
1302 its functionality in dough like model systems. *Food Hydrocolloids*, 54, 196-201.
- 1303 Joshi, M., Aldred, P., Panozzo, J. F., Kasapis, S., & Adhikari, B. (2014). Rheological and
1304 microstructural characteristics of lentil starch–lentil protein composite pastes and gels. *Food
1305 Hydrocolloids*, 35, 226-237.

- 1306 Karim, A. A., Norziah, M. H., & Seow, C. C. (2000). Methods for the study of starch retrogradation.
1307 *Food Chemistry*, 71(1), 9-36.
- 1308 Kett, A. P., Chaurin, V., Fitzsimons, S. M., Morris, E. R., O'Mahony, J. A., & Fenelon, M. A. (2013).
1309 Influence of milk proteins on the pasting behaviour and microstructural characteristics of waxy
1310 maize starch. *Food Hydrocolloids*, 30(2), 661-671.
- 1311 Kim, E. H. J., Petrie, J. R., Motoi, L., Morgenstern, M. P., Sutton, K. H., Mishra, S., & Simmons, L.
1312 D. (2008). Effect of structural and physicochemical characteristics of the protein matrix in
1313 pasta on in vitro starch digestibility. *Food Biophysics*, 3(2), 229-234.
- 1314 Kim, Y., Kee, J. I., Lee, S., & Yoo, S.-H. (2014). Quality improvement of rice noodle restructured
1315 with rice protein isolate and transglutaminase. *Food Chemistry*, 145, 409-416.
- 1316 Kong, B., Niu, H., Sun, F., Han, J., & Liu, Q. (2016). Regulatory effect of porcine plasma protein
1317 hydrolysates on pasting and gelatinization action of corn starch. *International Journal of*
1318 *Biological Macromolecules*, 82, 637-644.
- 1319 Kumar, L., Brennan, M., Zheng, H., & Brennan, C. (2018). The effects of dairy ingredients on the
1320 pasting, textural, rheological, freeze-thaw properties and swelling behaviour of oat starch.
1321 *Food Chemistry*, 245, 518-524.
- 1322 Kumar, L., Brennan, M. A., Mason, S. L., Zheng, H., & Brennan, C. S. (2017). Rheological, pasting
1323 and microstructural studies of dairy protein–starch interactions and their application in
1324 extrusion-based products: A review. *Starch - Stärke*, 69(1-2), 1600273.

- 1325 Kweon, M., Slade, L., Levine, H., & Gannon, D. (2014). Cookie- versus cracker-baking—What's the
1326 difference? Flour functionality requirements explored by SRC and alveography. *Critical*
1327 *reviews in food science and nutrition*, 54(1), 115-138.
- 1328 Lakshminarayan, S., Rajeswari, G., & Rao, G. (2010). Influence of defatted soy flour and whey
1329 protein concentrate on dough rheological characteristics and quality of instant vermicelli.
1330 *Journal of Texture Studies*, 42, 72-80.
- 1331 Lambrecht, M. A., Rombouts, I., Nivelles, M. A., & Delcour, J. A. (2017a). The impact of protein
1332 characteristics on the protein network in and properties of fresh and cooked wheat-based
1333 noodles. *Journal of Cereal Science*, 75, 234-242.
- 1334 Lambrecht, M. A., Rombouts, I., Nivelles, M. A., & Delcour, J. A. (2017b). The role of wheat and egg
1335 constituents in the formation of a covalent and non-covalent protein network in fresh and
1336 cooked egg noodles. *Journal of Food Science*, 82(1), 24-35.
- 1337 Larrosa, V., Lorenzo, G., Zaritzky, N., & Califano, A. (2016). Improvement of the texture and quality
1338 of cooked gluten-free pasta. *LWT - Food Science and Technology*, 70, 96-103.
- 1339 Levine, H., & Finley, J. W. (2018). Texture. In J. M. deMan, J. W. Finley, W. J. Hurst & C. Y. Lee
1340 (Eds.), *Principles of Food Chemistry* (pp. 329-363). Cham: Springer International Publishing.
- 1341 Li, G., & Zhu, F. (2017). Physicochemical properties of quinoa flour as affected by starch
1342 interactions. *Food Chemistry*, 221, 1560-1568.
- 1343 Li, M., Yue, Q., Liu, C., Zheng, X., Hong, J., Li, L., & Bian, K. (2020). Effect of gliadin/glutenin
1344 ratio on pasting, thermal, and structural properties of wheat starch. *Journal of Cereal Science*,
1345 93, 102973.

- 1346 Li, M., Zhu, K.-X., Guo, X.-N., Brijs, K., & Zhou, H.-M. (2014). Natural additives in wheat-based
1347 pasta and noodle products: Opportunities for enhanced nutritional and functional properties.
1348 *Comprehensive Reviews in Food Science and Food Safety*, 13(4), 347-357.
- 1349 Li, S., Wei, Y., Fang, Y., Zhang, W., & Zhang, B. (2014). DSC study on the thermal properties of
1350 soybean protein isolates/corn starch mixture. *Journal of Thermal Analysis and Calorimetry*,
1351 115(2), 1633-1638.
- 1352 Li, T., Senesi, A. J., & Lee, B. (2016). Small angle X-ray scattering for nanoparticle research.
1353 *Chemical Reviews*, 116(18), 11128-11180.
- 1354 Li, Y., & Huang, Q. (2015). Protein-polysaccharide complexes for effective delivery of bioactive
1355 functional food ingredients. In C. M. Sabliov, H. Chen & R. Y. Yada (Eds.), *Nanotechnology
1356 and Functional Foods: Effective Delivery of Bioactive Ingredients* (pp. 224-246). Hoboken, NJ,
1357 USA: John Wiley & Sons, Ltd.
- 1358 Li, Z., Wang, L., Chen, Z., Yu, Q., & Feng, W. (2018). Impact of binding interaction characteristics
1359 on physicochemical, structural, and rheological properties of waxy rice flour. *Food Chemistry*,
1360 266, 551-556.
- 1361 Lian, X., Zhu, W., Wen, Y., Li, L., & Zhao, X. (2013). Effects of soy protein hydrolysates on maize
1362 starch retrogradation studied by IR spectra and ESI-MS analysis. *International Journal of
1363 Biological Macromolecules*, 59, 143-150.
- 1364 Likitwattanasade, T., & Hongsprabhas, P. (2010). Effect of storage proteins on pasting properties and
1365 microstructure of Thai rice. *Food Research International*, 43(5), 1402-1409.

- 1366 Liu, R., Sun, W., Zhang, Y., Huang, Z., Hu, H., Zhao, M., & Li, W. (2019). Development of a novel
1367 model dough based on mechanically activated cassava starch and gluten protein: Application in
1368 bread. *Food Chemistry*, *300*, 125196.
- 1369 Liu, R., Xing, Y., Zhang, Y., Zhang, B., Jiang, X., & Wei, Y. (2015). Effect of mixing time on the
1370 structural characteristics of noodle dough under vacuum. *Food Chemistry*, *188*, 328-336.
- 1371 López-Barón, N., Gu, Y., Vasanthan, T., & Hoover, R. (2017). Plant proteins mitigate *in vitro* wheat
1372 starch digestibility. *Food Hydrocolloids*, *69*, 19-27.
- 1373 López-Barón, N., Sagnelli, D., Blennow, A., Hulse, M., Gao, J., Saaby, L., Müllertz, A., Jespersen, B.,
1374 & Vasanthan, T. (2018). Hydrolysed pea proteins mitigate *in vitro* wheat starch digestibility.
1375 *Food Hydrocolloids*, *79*, 117-126.
- 1376 Lu, Z.-H., Donner, E., Yada, R. Y., & Liu, Q. (2016). Physicochemical properties and *in vitro* starch
1377 digestibility of potato starch/protein blends. *Carbohydrate Polymers*, *154*, 214-222.
- 1378 M, V. P. (2017). Soy protein: Introduction, structure and properties relationship. In V. P. M & O.
1379 Nazarenko (Eds.), *Soy Protein - Based Blends, Composites and Nanocomposites* (pp. 23-37).
1380 Beverly, MA, USA: Scrivener Publishing LLC.
- 1381 Majumdar, S., Sen, P., & Ray, R. (2019). Ionic interactions and transport properties in chitosan-starch
1382 based blend solid biopolymer electrolytes. *Materials Today: Proceedings*, *18*, 4913-4920.
- 1383 Mancebo, C. M., Rodriguez, P., & Gómez, M. (2016). Assessing rice flour-starch-protein mixtures to
1384 produce gluten free sugar-snap cookies. *LWT - Food Science and Technology*, *67*, 127-132.
- 1385 Manoj Kumar, C. T., Sabikhi, L., Singh, A. K., Raju, P. N., Kumar, R., & Sharma, R. (2019). Effect
1386 of incorporation of sodium caseinate, whey protein concentrate and transglutaminase on the

- 1387 properties of depigmented pearl millet based gluten free pasta. *LWT - Food Science and*
1388 *Technology*, 103, 19-26.
- 1389 Marín, D., Alemán, A., Montero, P., & Gómez-Guillén, M. C. (2018). Protein aggregation, water
1390 binding and thermal gelation of salt-ground hake muscle in the presence of wet and dried soy
1391 phosphatidylcholine liposomes. *Food Hydrocolloids*, 82, 466-477.
- 1392 Marti, A., Barbiroli, A., Marengo, M., Fongaro, L., Iametti, S., & Pagani, M. A. (2014). Structuring
1393 and texturing gluten-free pasta: egg albumen or whey proteins? *European Food Research and*
1394 *Technology*, 238(2), 217-224.
- 1395 McCann, T. H., Le Gall, M., & Day, L. (2016). Extensional dough rheology – Impact of flour
1396 composition and extension speed. *Journal of Cereal Science*, 69, 228-237.
- 1397 Miñarro, B., Normahomed, I., Guamis, B., & Capellas, M. (2010). Influence of unicellular protein on
1398 gluten-free bread characteristics. *European Food Research and Technology*, 231(2), 171-179.
- 1399 Mittal, J., & Best, R. B. (2008). Thermodynamics and kinetics of protein folding under confinement.
1400 *Proceedings of the National Academy of Sciences*, 105(51), 20233.
- 1401 Mohammed, I., Ahmed, A. R., & Senge, B. (2012). Dough rheology and bread quality of wheat–
1402 chickpea flour blends. *Industrial Crops and Products*, 36(1), 196-202.
- 1403 Moio, T., Forssell, P., Partanen, R., Damerou, A., & Hill, S. E. (2015). Reorganisation of starch,
1404 proteins and lipids in extrusion of oats. *Journal of Cereal Science*, 64, 48-55.
- 1405 Nicoletti Telis, V. R. (2019). O/W emulsions stabilized by interactions between proteins and
1406 polysaccharides. In L. Melton, F. Shahidi & P. Varelis (Eds.), *Encyclopedia of Food Chemistry*
1407 (pp. 494-498). Amsterdam, Netherlands: Elsevier.

- 1408 Nieto-Nieto, T. V., Wang, Y. X., Ozimek, L., & Chen, L. (2015). Inulin at low concentrations
1409 significantly improves the gelling properties of oat protein – A molecular mechanism study.
1410 *Food Hydrocolloids*, 50, 116-127.
- 1411 Nieto Nieto, T. V., Wang, Y., Ozimek, L., & Chen, L. (2016). Improved thermal gelation of oat
1412 protein with the formation of controlled phase-separated networks using dextrin and
1413 carrageenan polysaccharides. *Food Research International*, 82, 95-103.
- 1414 Niu, L., Wu, L., & Xiao, J. (2017). Inhibition of gelatinized rice starch retrogradation by rice bran
1415 protein hydrolysates. *Carbohydrate Polymers*, 175, 311-319.
- 1416 Noisuwan, A., Hemar, Y., Wilkinson, B., & Bronlund, J. E. (2009). Dynamic rheological and
1417 microstructural properties of normal and waxy rice starch gels containing milk protein
1418 ingredients. *Starch - Stärke*, 61(3 - 4), 214-227.
- 1419 Nozawa, M., Ito, S., & Arai, E. (2016). Effect of ovalbumin on the quality of gluten-free rice flour
1420 bread made with soymilk. *LWT - Food Science and Technology*, 66, 598-605.
- 1421 Oñate Narciso, J., & Brennan, C. (2018). Whey and pea protein fortification of rice starches: Effects
1422 on protein and starch digestibility and starch pasting properties. *Starch - Stärke*, 70(9-10),
1423 1700315.
- 1424 Ortolan, F., & Steel, C. J. (2017). Protein characteristics that affect the quality of vital wheat gluten
1425 to be used in baking: A review. *Comprehensive Reviews in Food Science and Food Safety*,
1426 16(3), 369-381.

- 1427 Pace, C. N., Fu, H., Fryar, K. L., Landua, J., Trevino, S. R., Shirley, B. A., Hendricks, M. M., Iimura,
1428 S., Gajiwala, K., Scholtz, J. M., & Grimsley, G. R. (2011). Contribution of Hydrophobic
1429 Interactions to Protein Stability. *Journal of Molecular Biology*, 408(3), 514-528.
- 1430 Paliwal, J., Thakur, S., & Erkinbaev, C. (2019). Protein-starch interactions in cereal grains and pulses.
1431 In L. Melton, F. Shahidi & P. Varelis (Eds.), *Encyclopedia of Food Chemistry* (pp. 446-452).
1432 Oxford: Academic Press.
- 1433 Patraşcu, L., Banu, I., Vasilean, I., & Aprodu, I. (2016). Rheological and thermo-mechanical
1434 characterization of starch – protein mixtures. *Agriculture and Agricultural Science Procedia*,
1435 10, 280-288.
- 1436 Pelgrom, P. J. M., Vissers, A. M., Boom, R. M., & Schutyser, M. A. I. (2013). Dry fractionation for
1437 production of functional pea protein concentrates. *Food Research International*, 53(1),
1438 232-239.
- 1439 Pérez, S., & Bertoft, E. (2010). The molecular structures of starch components and their contribution
1440 to the architecture of starch granules: A comprehensive review. *Starch - Stärke*, 62(8), 389-420.
- 1441 Pérez, S., Matta, E., Osella, C., de la Torre, M., & Sánchez, H. D. (2013). Effect of soy flour and
1442 whey protein concentrate on cookie color. *LWT - Food Science and Technology*, 50(1),
1443 120-125.
- 1444 Petitot, M., Abecassis, J., & Micard, V. (2009). Structuring of pasta components during processing:
1445 Impact on starch and protein digestibility and allergenicity. *Trends in Food Science and*
1446 *Technology*, 20(11), 521-532.

- 1447 Philipp, C., Buckow, R., Silcock, P., & Oey, I. (2017). Instrumental and sensory properties of pea
1448 protein-fortified extruded rice snacks. *Food Research International*, *102*, 658-665.
- 1449 Philipp, C., Oey, I., Silcock, P., Beck, S. M., & Buckow, R. (2017). Impact of protein content on
1450 physical and microstructural properties of extruded rice starch-pea protein snacks. *Journal of*
1451 *Food Engineering*, *212*, 165-173.
- 1452 Phongthai, S., D'Amico, S., Schoenlechner, R., Homthawornchoo, W., & Rawdkuen, S. (2017).
1453 Effects of protein enrichment on the properties of rice flour based gluten-free pasta. *LWT -*
1454 *Food Science and Technology*, *80*, 378-385.
- 1455 Phongthai, S., D'Amico, S., Schoenlechner, R., & Rawdkuen, S. (2016). Comparative study of rice
1456 bran protein concentrate and egg albumin on gluten-free bread properties. *Journal of Cereal*
1457 *Science*, *72*, 38-45.
- 1458 Qiu, C., Li, X., Ji, N., Qin, Y., Sun, Q., & Xiong, L. (2015). Rheological properties and
1459 microstructure characterization of normal and waxy corn starch dry heated with soy protein
1460 isolate. *Food Hydrocolloids*, *48*, 1-7.
- 1461 Quiroga Ledezma, C. C. (2018). Starch interactions with native and added food components. In M.
1462 Sjöö & L. Nilsson (Eds.), *Starch in Food (Second Edition)* (pp. 769-801): Woodhead
1463 Publishing/Elsevier.
- 1464 Reddy Surasani, V. K., Singh, A., Gupta, A., & Sharma, S. (2019). Functionality and cooking
1465 characteristics of pasta supplemented with protein isolate from pangas processing waste. *LWT -*
1466 *Food Science and Technology*, *111*, 443-448.

- 1467 Ren, F., Dong, D., Yu, B., Hou, Z.-h., & Cui, B. (2017). Rheology, thermal properties, and
1468 microstructure of heat-induced gel of whey protein–acetylated potato starch. *Starch - Stärke*,
1469 *69*(9-10), 1600344.
- 1470 Ren, F., & Wang, S. (2019). Effect of modified tapioca starches on the gelling properties of whey
1471 protein isolate. *Food Hydrocolloids*, *93*, 87-91.
- 1472 Renzetti, S., Behr, J., Vogel, R. F., Barbiroli, A., Iametti, S., Bonomi, F., & Arendt, E. K. (2012).
1473 Transglutaminase treatment of brown rice flour: A chromatographic, electrophoretic and
1474 spectroscopic study of protein modifications. *Food Chemistry*, *131*(4), 1076-1085.
- 1475 Ribotta, P. D., Colombo, A., León, A. E., & Añón, M. C. (2007). Effects of soy protein on physical
1476 and rheological properties of wheat starch. *Starch - Stärke*, *59*(12), 614-623.
- 1477 Ribotta, P. D., Colombo, A., & Rosell, C. M. (2012). Enzymatic modifications of pea protein and its
1478 application in protein–cassava and corn starch gels. *Food Hydrocolloids*, *27*(1), 185-190.
- 1479 Ribotta, P. D., & Rosell, C. M. (2010). Effects of enzymatic modification of soybean protein on the
1480 pasting and rheological profile of starch–protein systems. *Starch - Stärke*, *62*(7), 373-383.
- 1481 Rodriguez Furlán, L. T., Pérez Padilla, A., & Campderrós, M. E. (2015). Improvement of gluten-free
1482 bread properties by the incorporation of bovine plasma proteins and different saccharides into
1483 the matrix. *Food Chemistry*, *170*, 257-264.
- 1484 Ronda, F., Oliete, B., Gómez, M., Caballero, P. A., & Pando, V. (2011). Rheological study of layer
1485 cake batters made with soybean protein isolate and different starch sources. *Journal of Food*
1486 *Engineering*, *102*(3), 272-277.

- 1487 Ronda, F., Villanueva, M., & Collar, C. (2014). Influence of acidification on dough viscoelasticity of
1488 gluten-free rice starch-based dough matrices enriched with exogenous protein. *LWT - Food
1489 Science and Technology*, 59(1), 12-20.
- 1490 Rosa-Sibakov, N., Heiniö, R.-L., Cassan, D., Holopainen-Mantila, U., Micard, V., Lantto, R., &
1491 Sozer, N. (2016). Effect of bioprocessing and fractionation on the structural, textural and
1492 sensory properties of gluten-free faba bean pasta. *LWT - Food Science and Technology*, 67,
1493 27-36.
- 1494 Ryan, K. J., & Brewer, M. S. (2007). In situ examination of starch granule-soy protein and wheat
1495 protein interactions. *Food Chemistry*, 104(2), 619-629.
- 1496 Sang, S., Zhang, H., Xu, L., Chen, Y., Xu, X., Jin, Z., Yang, N., Wu, F., & Li, D. (2018).
1497 Functionality of ovalbumin during Chinese steamed bread-making processing. *Food Chemistry*,
1498 253, 203-210.
- 1499 Sarabhai, S., & Prabhasankar, P. (2015). Influence of whey protein concentrate and potato starch on
1500 rheological properties and baking performance of Indian water chestnut flour based gluten free
1501 cookie dough. *LWT - Food Science and Technology*, 63(2), 1301-1308.
- 1502 Sciarini, L. S., Ribotta, P. D., León, A. E., & Pérez, G. T. (2010). Influence of gluten-free flours and
1503 their mixtures on batter properties and bread quality. *Food and Bioprocess Technology*, 3(4),
1504 577-585.
- 1505 Shevkani, K., Kaur, A., Kumar, S., & Singh, N. (2015). Cowpea protein isolates: Functional
1506 properties and application in gluten-free rice muffins. *LWT - Food Science and Technology*,
1507 63(2), 927-933.

- 1508 Shevkani, K., & Singh, N. (2014). Influence of kidney bean, field pea and amaranth protein isolates
1509 on the characteristics of starch-based gluten-free muffins. *International Journal of Food*
1510 *Science and Technology*, 49(10), 2237-2244.
- 1511 Shin, M., Gang, D.-O., & Song, J.-Y. (2010). Effects of protein and transglutaminase on the
1512 preparation of gluten-free rice bread. *Food Science and Biotechnology*, 19(4), 951-956.
- 1513 Silverman, R. B., & Holladay, M. W. (2014). Chapter 3 - Receptors. In R. B. Silverman & M. W.
1514 Holladay (Eds.), *The Organic Chemistry of Drug Design and Drug Action (Third Edition)* (pp.
1515 123-163). Boston: Academic Press.
- 1516 Singh, J., Dartois, A., & Kaur, L. (2010). Starch digestibility in food matrix: a review. *Trends in Food*
1517 *Science and Technology*, 21(4), 168-180.
- 1518 Singh, J., Kaur, L., & Singh, H. (2013). Food microstructure and starch digestion. In J. Henry (Ed.),
1519 *Advances in Food and Nutrition Research* (Vol. 70, pp. 137-179). Waltham, MA, USA:
1520 Academic Press/Elsevier.
- 1521 Singh, S., & Singh, N. (2013). Relationship of polymeric proteins and empirical dough rheology with
1522 dynamic rheology of dough and gluten from different wheat varieties. *Food Hydrocolloids*,
1523 33(2), 342-348.
- 1524 Slade, L., Kweon, M., & Levine, H. (2021). Exploration of the functionality of sugars in cake-baking,
1525 and effects on cake quality. *Critical reviews in food science and nutrition*, 61(2), 283-311.
- 1526 Sopade, P., Hardin, M., Fitzpatrick, P., Desmee, H., & Halley, P. (2006). Macromolecular interactions
1527 during gelatinisation and retrogradation in starch-whey systems as studied by rapid
1528 visco-analyser. *International Journal of Food Engineering* 2(4), Article 7.

- 1529 Storck, C. R., da Rosa Zavareze, E., Gularte, M. A., Elias, M. C., Rosell, C. M., & Guerra Dias, A. R.
1530 (2013). Protein enrichment and its effects on gluten-free bread characteristics. *LWT - Food*
1531 *Science and Technology*, 53(1), 346-354.
- 1532 Sun, N.-x., Liang, Y., Yu, B., Tan, C.-p., & Cui, B. (2016). Interaction of starch and casein. *Food*
1533 *Hydrocolloids*, 60, 572-579.
- 1534 Sun, Q., & Xiong, C. S. L. (2014). Functional and pasting properties of pea starch and peanut protein
1535 isolate blends. *Carbohydrate Polymers*, 101, 1134-1139.
- 1536 Susanna, S., & Prabhasankar, P. (2013). A study on development of gluten free pasta and its
1537 biochemical and immunological validation. *LWT - Food Science and Technology*, 50(2),
1538 613-621.
- 1539 Svihus, B., & Hervik, A. K. (2016). Digestion and metabolic fates of starch, and its relation to major
1540 nutrition-related health problems: A review. *Starch - Stärke*, 68(3-4), 302-313.
- 1541 Tarhan, O., Spotti, M. J., Schaffter, S., Corvalan, C. M., & Campanella, O. H. (2016). Rheological
1542 and structural characterization of whey protein gelation induced by enzymatic hydrolysis. *Food*
1543 *Hydrocolloids*, 61, 211-220.
- 1544 Toutounji, M. R., Farahnaky, A., Santhakumar, A. B., Oli, P., Butardo, V. M., & Blanchard, C. L.
1545 (2019). Intrinsic and extrinsic factors affecting rice starch digestibility. *Trends in Food Science*
1546 *and Technology*, 88, 10-22.
- 1547 Vamadevan, V., & Bertoft, E. (2015). Structure-function relationships of starch components. *Starch -*
1548 *Stärke*, 67(1-2), 55-68.

- 1549 Villanueva, M., De Lamo, B., Harasym, J., & Ronda, F. (2018). Microwave radiation and protein
1550 addition modulate hydration, pasting and gel rheological characteristics of rice and potato
1551 starches. *Carbohydrate Polymers*, 201, 374-381.
- 1552 Villanueva, M., Mauro, R. R., Collar, C., & Ronda, F. (2015). Acidification of protein-enriched rice
1553 starch doughs: effects on breadmaking. *European Food Research and Technology*, 240(4),
1554 783-794.
- 1555 Villanueva, M., Ronda, F., Moschakis, T., Lazaridou, A., & Biliaderis, C. G. (2018). Impact of
1556 acidification and protein fortification on thermal properties of rice, potato and tapioca starches
1557 and rheological behaviour of their gels. *Food Hydrocolloids*, 79, 20-29.
- 1558 Vu Dang, H., Loisel, C., Desrumaux, A., & Doublier, J. L. (2009). Rheology and microstructure of
1559 cross-linked waxy maize starch/whey protein suspensions. *Food Hydrocolloids*, 23(7),
1560 1678-1686.
- 1561 Vu, T.-H., Bean, S., Hsieh, C.-F., & Shi, Y.-C. (2017). Changes in protein and starch digestibility in
1562 sorghum flour during heat–moisture treatments. *Journal of the Science of Food and Agriculture*,
1563 97(14), 4770-4779.
- 1564 Wan, Y., Liu, J., & Guo, S. (2018). Effects of succinylation on the structure and thermal aggregation
1565 of soy protein isolate. *Food Chemistry*, 245, 542-550.
- 1566 Wang, C., Xue, Y., Yousaf, L., Hu, J., & Shen, Q. (2020). Effects of high hydrostatic pressure on the
1567 ordered structure including double helices and V-type single helices of rice starch.
1568 *International Journal of Biological Macromolecules*, 144, 1034-1042.

- 1569 Wang, F., Huang, W., Kim, Y., Liu, R., & Tilley, M. (2011). Effects of transglutaminase on the
1570 rheological and noodle-making characteristics of oat dough containing vital wheat gluten or
1571 egg albumin. *Journal of Cereal Science*, *54*(1), 53-59.
- 1572 Wang, J., Zhao, S., Min, G., Qiao, D., Zhang, B., Niu, M., Jia, C., Xu, Y., & Lin, Q. (2021).
1573 Starch-protein interplay varies the multi-scale structures of starch undergoing thermal
1574 processing. *International Journal of Biological Macromolecules*, *175*, 179-187.
- 1575 Wang, L., Zhang, L., Wang, H., Ai, L., & Xiong, W. (2020). Insight into protein-starch ratio on the
1576 gelatinization and retrogradation characteristics of reconstituted rice flour. *International*
1577 *Journal of Biological Macromolecules*, *146*, 524-529.
- 1578 Wang, P., Jin, Z., & Xu, X. (2015). Physicochemical alterations of wheat gluten proteins upon dough
1579 formation and frozen storage – A review from gluten, glutenin and gliadin perspectives. *Trends*
1580 *in Food Science & Technology*, *46*(2, Part A), 189-198.
- 1581 Wang, P., Xu, L., Nikoo, M., Ocen, D., Wu, F., Na, Y., Jin, Z., & Xu, X. (2013). Effect of frozen
1582 storage on the conformational, thermal and microscopic properties of gluten: Comparative
1583 studies on gluten-, glutenin- and gliadin-rich fractions. *Food Hydrocolloids*, *35*, 238-246.
- 1584 Wang, S., & Copeland, L. (2013). Molecular disassembly of starch granules during gelatinization and
1585 its effect on starch digestibility: A review. *Food and Function*, *4*(11), 1564-1580.
- 1586 Wang, S., Li, C., Copeland, L., Niu, Q., & Wang, S. (2015). Starch retrogradation: A comprehensive
1587 review. *Comprehensive Reviews in Food Science and Food Safety*, *14*(5), 568-585.
- 1588 Wang, S., Zheng, M., Yu, J., Wang, S., & Copeland, L. (2017). Insights into the formation and
1589 structures of starch protein lipid complexes. *Journal of Agricultural and Food Chemistry*, *65*.

- 1590 Wang, W., Chen, W., Yang, H., & Cui, M. (2017). Textural and rheological properties of potato starch
1591 as affected by amino acids. *International Journal of Food Properties*, 20(sup3), S3123-S3134.
- 1592 Wang, X.-Y., Guo, X.-N., & Zhu, K.-X. (2016). Polymerization of wheat gluten and the changes of
1593 glutenin macropolymer (GMP) during the production of Chinese steamed bread. *Food*
1594 *Chemistry*, 201, 275-283.
- 1595 Wang, X., Appels, R., Zhang, X., Diepeveen, D., Torok, K., Tomoskozi, S., Bekes, F., Ma, W., Sharp,
1596 P., & Islam, S. (2017). Protein interactions during flour mixing using wheat flour with altered
1597 starch. *Food Chemistry*, 231, 247-257.
- 1598 Wani, A. A., Sogi, D. S., Singh, P., Sharma, P., & Pangal, A. (2012). Dough-handling and
1599 cookie-making properties of wheat flour–watermelon protein isolate blends. *Food and*
1600 *Bioprocess Technology*, 5(5), 1612-1621.
- 1601 Warnakulasuriya, S. N., & Nickerson, M. T. (2018). Review on plant protein–polysaccharide
1602 complex coacervation, and the functionality and applicability of formed complexes. *Journal of*
1603 *the Science of Food and Agriculture*, 98(15), 5559-5571.
- 1604 Wei, Z., & Huang, Q. (2019). Assembly of protein–polysaccharide complexes for delivery of
1605 bioactive ingredients: A perspective paper. *Journal of Agricultural and Food Chemistry*, 67(5),
1606 1344-1352.
- 1607 Wilderjans, E., Luyts, A., Goesaert, H., Brijs, K., & Delcour, J. A. (2010). A model approach to
1608 starch and protein functionality in a pound cake system. *Food Chemistry*, 120(1), 44-51.
- 1609 Witczak, M., Ziobro, R., Juszczak, L., & Korus, J. (2016). Starch and starch derivatives in
1610 gluten-free systems – A review. *Journal of Cereal Science*, 67, 46-57.

- 1611 Wu, Y., Chen, Z., Li, X., & Wang, Z. (2010). Retrogradation properties of high amylose rice flour
1612 and rice starch by physical modification. *LWT - Food Science and Technology*, 43(3), 492-497.
- 1613 Xiao, J., Niu, L., Wu, L., Li, D., & He, H. (2019). Preparation of an in vitro low-digestible rice starch
1614 by addition of grass carp protein hydrolysates and its possible mechanisms. *Starch - Stärke*,
1615 71(1-2), 1800159.
- 1616 Xiao, J., & Zhong, Q. (2017). Suppression of retrogradation of gelatinized rice starch by anti-listerial
1617 grass carp protein hydrolysate. *Food Hydrocolloids*, 72, 338-345.
- 1618 Xie, D., Liu, X., Zhang, H., Xia, W., Huang, X., Bi, D., & Pan, S. (2017). Textural properties and
1619 morphology of soy 7S globulin-corn starch (amylose, amylopectin). *International Journal of*
1620 *Food Properties*, 20(10), 2197-2205.
- 1621 Xijun, L., Junjie, G., Danli, W., Lin, L., & Jiaran, Z. (2014). Effects of protein in wheat flour on
1622 retrogradation of wheat starch. *Journal of Food Science*, 79(8), C1505-C1511.
- 1623 Yang, C., Zhong, F., Douglas Goff, H., & Li, Y. (2019). Study on starch-protein interactions and their
1624 effects on physicochemical and digestible properties of the blends. *Food Chemistry*, 280,
1625 51-58.
- 1626 Yang, N., Liu, Y., Ashton, J., Gorczyca, E., & Kasapis, S. (2013). Phase behaviour and in vitro
1627 hydrolysis of wheat starch in mixture with whey protein. *Food Chemistry*, 137(1), 76-82.
- 1628 Yang, N., Luan, J., Ashton, J., Gorczyca, E., & Kasapis, S. (2014). Effect of calcium chloride on the
1629 structure and in vitro hydrolysis of heat induced whey protein and wheat starch composite gels.
1630 *Food Hydrocolloids*, 42, 260-268.

- 1631 Ye, J., Hu, X., Luo, S., McClements, D. J., Liang, L., & Liu, C. (2018). Effect of endogenous
1632 proteins and lipids on starch digestibility in rice flour. *Food Research International*, *106*,
1633 404-409.
- 1634 Yu, B., Ren, F., Zhao, H., Cui, B., & Liu, P. (2020). Effects of native starch and modified starches on
1635 the textural, rheological and microstructural characteristics of soybean protein gel.
1636 *International Journal of Biological Macromolecules*, *142*, 237-243.
- 1637 Yu, L., Ramaswamy, H. S., & Boye, J. (2012). Twin-screw extrusion of corn flour and soy protein
1638 isolate (SPI) blends: A response surface analysis. *Food and Bioprocess Technology*, *5*(2),
1639 485-497.
- 1640 Yu, W., Zou, W., Dhital, S., Wu, P., Gidley, M. J., Fox, G. P., & Gilbert, R. G. (2018). The adsorption
1641 of α -amylase on barley proteins affects the in vitro digestion of starch in barley flour. *Food*
1642 *Chemistry*, *241*, 493-501.
- 1643 Zeng, M., Wu, Y., Gao, H., Fan, L., Mangavel, C., & Lourdin, D. (2010). Influence of dehydration
1644 treatment on intermolecular interaction and morphology of pills prepared from proteins and
1645 corn starch. *Science of Advanced Materials*, *2*(4), 514-521.
- 1646 Zhan, Q., Ye, X., Zhang, Y., Kong, X., Bao, J., Corke, H., & Sui, Z. (2020). Starch
1647 granule-associated proteins affect the physicochemical properties of rice starch. *Food*
1648 *Hydrocolloids*, *101*, 105504.
- 1649 Zhang, D., Mu, T., & Sun, H. (2018). Effects of starch from five different botanical sources on the
1650 rheological and structural properties of starch–gluten model doughs. *Food Research*
1651 *International*, *103*, 156-162.

- 1652 Zhang, M., Sun, C., Wang, X., Wang, N., & Zhou, Y. (2020). Effect of rice protein hydrolysates on
1653 the short-term and long-term retrogradation of wheat starch. *International Journal of*
1654 *Biological Macromolecules*, *155*, 1169-1175.
- 1655 Zhang, S. B., Lu, Q. Y., Yang, H., & Dan Meng, D. (2011). Effects of protein content,
1656 glutenin-to-gliadin ratio, amylose content, and starch damage on textural properties of chinese
1657 fresh white noodles. *Cereal Chemistry*, *88*(3), 296-301.
- 1658 Zhang, Y., Chen, C., Chen, Y., & Chen, Y. (2019). Effect of rice protein on the water mobility, water
1659 migration and microstructure of rice starch during retrogradation. *Food Hydrocolloids*, *91*,
1660 136-142.
- 1661 Zhou, J., Liu, J., & Tang, X. (2018). Effects of whey and soy protein addition on bread rheological
1662 property of wheat flour. *Journal of Texture Studies*, *49*(1), 38-46.
- 1663 Zhu, L., Wu, G., Cheng, L., Zhang, H., Wang, L., Qian, H., & Qi, X. (2020). Investigation on
1664 molecular and morphology changes of protein and starch in rice kernel during cooking. *Food*
1665 *Chemistry*, *316*, 126262.
- 1666 Zou, W., Sissons, M., Gidley, M. J., Gilbert, R. G., & Warren, F. J. (2015). Combined techniques for
1667 characterising pasta structure reveals how the gluten network slows enzymic digestion rate.
1668 *Food Chemistry*, *188*, 559-568.
- 1669 Zou, W., Sissons, M., Warren, F. J., Gidley, M. J., & Gilbert, R. G. (2016). Compact structure and
1670 proteins of pasta retard in vitro digestive evolution of branched starch molecular structure.
1671 *Carbohydrate Polymers*, *152*, 441-449.
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1673 **Highlights**

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Journal Pre-proof

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