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# New perspectives on rare connective tissue calcifying diseases

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Connective tissue calcifying diseases (CTCs) are characterized by abnormal calcium deposition in connective tissues. CTCs are caused by multiple factors including chronic diseases (Type II diabetes mellitus, chronic kidney disease), the use of pharmaceuticals (e.g. warfarin, glucocorticoids) and inherited rare genetic diseases such as pseudoxanthoma elasticum (PXE), generalized arterial calcification in infancy (GACI) and Keutel syndrome (KTLS). This review explores our current knowledge of these rare inherited CTCs, and highlights the most promising avenues for pharmaceutical intervention. Advancing our understanding of rare inherited forms of CTC is not only essential for the development of therapeutic strategies for patients suffering from these diseases, but also fundamental to delineating the mechanisms underpinning acquired chronic forms of CTC.

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## Introduction

A disease or disorder is defined as rare in Europe when it affects less than 1 in 2000 people. In the EU, as many as 30 million people alone may be affected by one of over 6000 existing rare diseases (<http://www.eurordis.org/about-rare-diseases>). A number of rare inherited forms of

Connective Tissue Calcifying diseases (CTCs) have been identified, and are characterized by abnormal calcium mineral deposition in connective tissues. Although all tissue has the potential to undergo calcification, several tissues have a higher propensity to calcify including skin, kidney, blood vessels and cardiac valves [1]. There are multiple mechanisms by which connective tissue calcification can progress, however these mechanisms are not exclusive and multiple pathologies can concurrently promote aberrant calcification. Common causes of connective tissue calcification include aging, as well as diseases such as atherosclerosis, chronic kidney disease (CKD) and Type II diabetes mellitus. Connective tissue calcification is also the result of specific rare congenital diseases such as generalized arterial calcification of infancy (GACI), pseudoxanthoma elasticum (PXE), Hutchinson–Gilford progeria syndrome (HGPS), arterial calcification due to deficiency of CD73 (ACDC) and Keutel syndrome (KTLS) [2]. Despite scarcity of cases, these diseases provide significant insight into the complex biological processes underpinning connective tissue calcification. The single gene deficiencies of rare inherited forms of CTC have allowed the identification of specific targets and the development of novel animal models to further study the process of connective tissue calcification. Furthermore, data from patients and animal models has resulted in the elucidation of pathways involved in both the promotion and inhibition of connective tissue calcification.

## Basic mechanisms of bone mineralization

In order to understand more fully the mechanisms underpinning connective tissue calcification, it is important to appreciate the physiological process of bone mineralization, which occurs through the deposition of hydroxyapatite (HA) onto a collagenous extracellular matrix (ECM). HA crystal formation is regulated by matrix vesicles (MVs), which maintain calcium ( $\text{Ca}^{2+}$ ) and inorganic phosphate ( $\text{P}_i$ ) concentrations at levels optimal for HA nucleation. As the HA crystals grow they disrupt the MV and deposit onto the ECM where they continue to grow [3]. The transport of  $\text{Ca}^{2+}$  into MVs is primarily controlled by annexin channels, whereas  $\text{P}_i$  is transported into the MV by the type III sodium-dependent  $\text{P}_i$  co-transporter-1 ( $\text{PiT-1}$ ) [4]. Intracellular to extracellular channelling of pyrophosphate ( $\text{PP}_i$ ) is mediated by ANK [5].

Whilst  $\text{P}_i$  acts to promote HA crystal formation,  $\text{PP}_i$ , generated by ecto-nucleotide pyrophosphatase/phosphodiesterase 1 (ENPP1), has a dual role as an inhibitor of HA

generation and as a precursor to  $P_i$  [6]. The ratio of  $P_i$  to  $PP_i$  is controlled by a complex interaction between the regulatory phosphatases tissue non-specific alkaline phosphatase (TNAP) and phosphoethanolamine/phosphocholine phosphatase (PHOSPHO1). TNAP hydrolyses  $PP_i$  in the ECM to release  $P_i$  and PHOSPHO1 hydrolyses phosphocholine and phosphoethanolamine to produce  $P_i$  inside the MVs. Together these phosphatases control the  $P_i/PP_i$  balance during the mineralization process [7]. Further feedback signalling allows modulation of mineralization; inorganic pyrophosphatase stimulates mineralization without reducing  $PP_i$  levels [8]. Both exogenous  $P_i$  and  $PP_i$  upregulate the bone sialoprotein osteopontin (OPN), which in turn inhibits mineralization through restricting HA crystal formation and growth [9]. Intriguingly, a clear dissociation in the hierarchical roles of  $PP_i$  and OPN has recently been highlighted [10\*].

### Overview of mechanisms of connective tissue calcification

The control and regulation of connective tissue calcification is a multifactorial process which shares many similarities with that of the physiological matrix mineralization during skeletal development described previously. A wealth of knowledge emanates from research into vascular calcification, a strong and independent predictor of morbidity and mortality in cardiovascular disease [2]. Indeed normal vascular smooth muscle cell (VSMC) populations contain cells that undergo phenotypic transition to osteocytic, osteoblastic and chondrocytic cells in a calcified environment [11]. In VSMCs, MVs have been shown to nucleate hydroxyapatite crystals that contain calcium and inorganic phosphate [12\*\*] forming the first nidus for calcification. This nucleation occurs via a tightly controlled

balance of inhibitors and inducers comparable to that seen in bone, with PHOSPHO1, sphingomyelinase 3 (SMPD3), TNAP, annexins, ANK and ENPP1 playing key regulatory roles [2,12\*\*]. Furthermore, MVs derived from VSMCs have been shown to contain negative regulators of hydroxyapatite crystal nucleation and growth, such as fetuin-A and matrix gla protein (MGP) [13\*]. In cooperation with local mediators such as  $PP_i$  [6], these molecules protect the arteries from mineral deposition and growth. In the absence of these inhibitors, or following the stimulation of apoptotic processes [14\*], together with the osteogenic activity of VSMCs, vascular calcification readily proceeds.

### A key role for pyrophosphate ( $PP_i$ ) in rare CTCs

It has recently been established that connective tissue calcification is contingent on circulating  $PP_i$  levels rather than local  $PP_i$  production [15]. As previously highlighted,  $PP_i$  not only acts as a potent inhibitor of connective tissue calcification, but also contributes directly to the calcification process. Intriguingly, one of the identified sources of systemic  $PP_i$  is through ATP binding cassette sub-family C member 6 (ABCC6)-mediated ATP release from hepatocytes [16]. Still within the vasculature of the liver, released ATP is rapidly converted to  $PP_i$  and AMP by ENPP1, which are in turn distributed throughout the body via the circulation. In connective tissues the metabolite AMP is further hydrolysed into  $P_i$  and adenosine by ecto-5'-nucleotidase (CD73). Adenosine in turn inhibits TNAP transcription, thus decreasing  $P_i$  production and more importantly maintaining  $PP_i$  levels [17]. Different perturbations in this mechanism can contribute to several rare CTCs (Table 1) which whilst varying in degree of severity and phenotype, show notable overlap.

**Table 1**

#### The cause and phenotype of significant rare inherited CTCs

CTC disease	Cause	Phenotype
Pseudoxanthoma elasticum (PXE)	ABCC6 deficiency	Elastic fibre mineralization in skin, eyes, and arteries.
Generalized arterial calcification in infancy (GACI)	ENPP1 deficiency	Widespread mineralization of arteries, and to a lesser extent joints.
Arterial calcification due to deficiency of CD73 (ACDC)	CD73 deficiency	Mineralization of arteries and joints in the extremities.
Hutchinson–Gilford progeria syndrome (HGPS)	Progerin (lamin A mutant)	Premature aging, atherosclerosis and calcification of blood vessels and the aortic valve.
Keutel syndrome (KTLS)	MGP deficiency	Facial abnormalities, calcification of the larynx trachea and bronchi, along with auricular, nasal and rib cartilage.
Fibrodysplasia ossificans progressiva (FOP)	ACVR1 gain of function	Progressive heterotopic endochondral ossification of skeletal muscle, fascia, tendons, and ligaments.
Coeliac disease with epilepsy and cerebral calcifications (CEC)	Unknown	Occipital epilepsy, with bilateral occipital calcifications and coeliac disease.
Idiopathic basal ganglia calcification (IBGC)	PIT-2 and/or PDGFR-B deficiency?	Calcification of the basal ganglia as well as the thalamus and cerebellum.

## Disease models of connective tissue calcification

### Pseudoxanthoma elasticum (PXE)

A defect in the *ABCC6* gene is responsible for pseudoxanthoma elasticum (PXE) (#OMIM 264800), an inherited autosomal recessive multisystem disorder affecting connective tissues in humans. The *ABCC6* gene carried by chromosome 16p13.11 encodes a trans-membrane ATP-binding cassette transporter, subfamily C, member 6 [15], which is primarily expressed in the hepatocyte and to a lesser extent in the proximal tubule cells in the kidney. The phenotypic expression of the disease is characterized by the fragmentation and mineralization of elastic fibres in the skin, retina and arterial wall [16–18]. The symptoms are represented by unaesthetic skin folds, central blindness and adverse cardiovascular events (Figure 1).

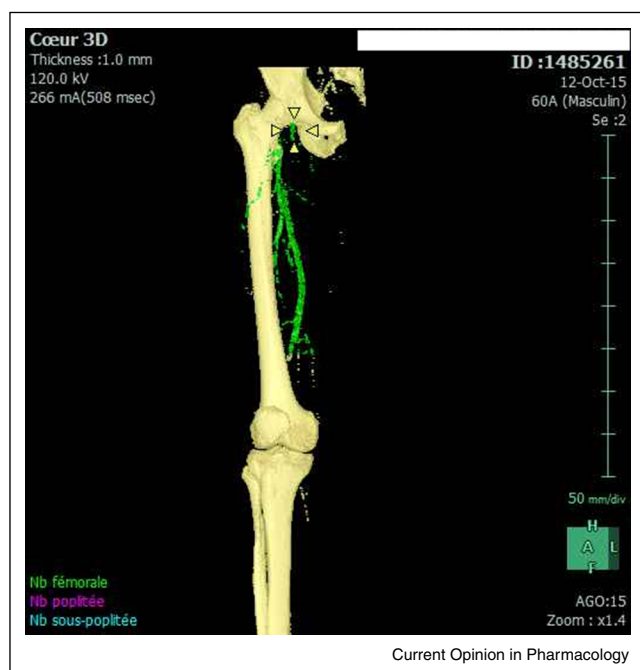
To date, the role of *ABCC6* and the nature of its substrate(s) in the PXE disease remain unknown and are presently subject to extensive research efforts worldwide. After several inconclusive hypotheses, recent seminal studies disclosed that *ABCC6* participates in the release of intracellular nucleotides (such as ATP) by a yet unknown pathway in the liver [19\*\*]. This results in a decrease in the extracellular ATP available for degradation by hepatic ecto-nucleotidase enzymes (such as ENPP1), which ultimately leads to less circulating  $PP_i$ , a powerful tissue and circulating anti-calcifying factor (as highlighted above) [20]. This important study places the liver, and to a lesser extent the kidney, as the metabolic

control centre of PXE associated calcification, with almost 40% of the overall systemic  $PP_i$  production generated from the liver [21\*\*], which cannot be compensated for by the local release of  $PP_i$  through alternative ectonucleotidic pathways (i.e. ENPP1 or TNAP).

Additional confounding factors of genetic [22] and/or metabolic origin [23–25] highlight the complex direct and remote interactions in PXE. Further clinical manifestations of PXE, such as elevated thrombotic susceptibility, increased myogenic tone and vascular malformations [26,27] may be a result of altered purinergic signalling, which unequivocally mediates complex autocrine and paracrine cellular signalling pathways within various tissues [28–31].

PXE represents a prototypic metabolic disease that shares some of the features of calcifying vascular diseases of acquired metabolic origin such as type II diabetes mellitus and CKD [2]. The discovery of the role of *ABCC6* raises new challenging questions on the central role of the hepato-renal axis in the connective-tissue calcifying process. From a clinical point of view, the current absence of an efficient targeted therapy requires the tight control and management of the typical cardiovascular risk factors associated with PXE, in addition to the limitation of pro-calcifying conditions. Promising new perspectives for etiologic treatments are however in progress, and may shortly yield exciting new treatment options for patients with PXE.

Figure 1



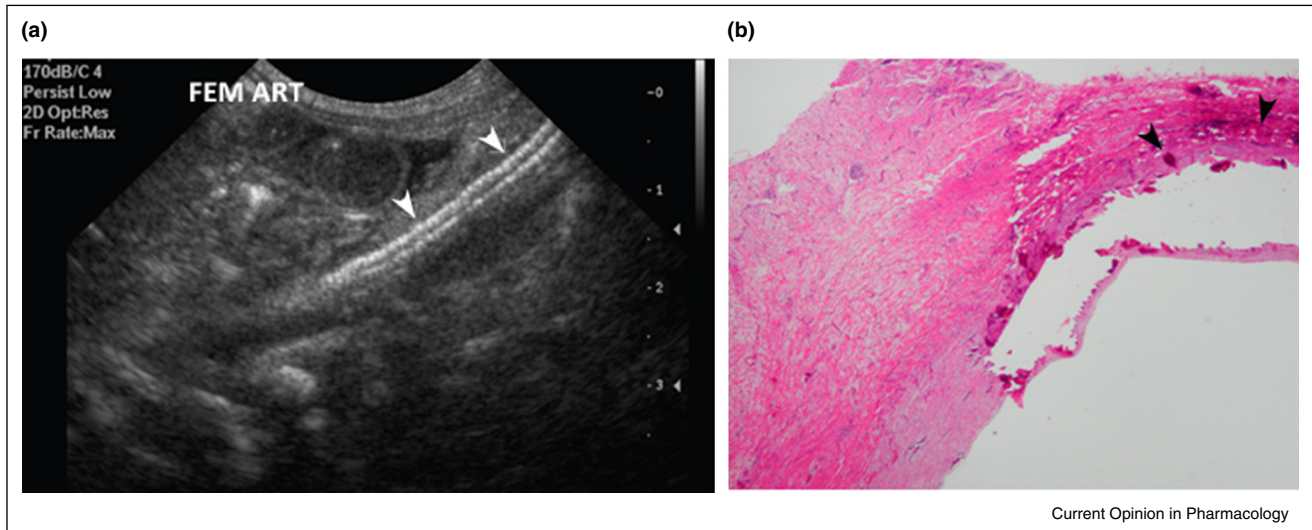
Diffuse arterial calcifications visualized by CT-scan in the lower limb arteries of a patient with pseudoxanthoma elasticum (PXE).

### Generalized arterial calcification in infancy (GACI)

Mutations in *ENPP1* are known to cause a rare human disease phenotype, namely generalized arterial calcification in infancy (GACI, MIM# 208000), formerly known as idiopathic infantile arterial calcification (IIAC) [32]. In GACI, calcification of the media of large and medium-sized arteries (Figure 2a) is associated with intimal proliferation (Figure 2b) leading to arterial stenosis. Depending on the severity and the local distribution of the calcific stenoses, affected infants can present with neonatal heart failure, arterial hypertension and death within the first six months of life [33]. To date, more than 40 different causative mutations in *ENPP1* have been identified in GACI patients accounting for approximately 70% of the affected cases [33]. Based on the number of reported pathogenic variants in NHLBI ESP6500 (16 carriers in 6021 individuals) a carrier frequency of one in 376 individuals (0.27%), and a disease frequency of one in 566,000 individuals can be estimated [34\*\*].

The cardiovascular phenotype of the disease is quite variable [35] and can even vary to a great extent between siblings carrying the same mutations: of two Taiwanese siblings with identical genotype, one developed extensive arterial calcification and severe hypertension, and died of heart failure at the age of 6 weeks, while the other

Figure 2



(a) Sonography of the femoral artery (Fem Art, arrows) of an infant with Generalized Arterial Calcification of Infancy (GACI). Note the increased echogenicity of the arterial wall. (b) Histology section of the aorta from an infant who died from GACI. Note the deposition of crystallized material (arrows) at the level of the lamina elastica interna and myointimal proliferation (H&E staining).

sibling had an uncomplicated clinical course [36]. The phenotype of the disease is recapitulated in the so-called tiptoe-walking (*ttw/ttw*) mouse associated with articular cartilage and peri-spinal ligament calcification and also with aortic calcification [37,38]. In these mice, connective tissue calcification progresses to hyperostotic joints and spine ankylosis leading to the ‘tiptoe-walking’ phenotype. A comparable phenotype can be found in *asj/asj* mice, which carry the V246D missense mutation [39,40] and in *Enpp1* knockout mice [41,42,43<sup>\*\*</sup>]. Intriguingly, deficiency of ENPP1 has also been shown to exert protective effects against obesity and diabetes in mice [44], however, the metabolic phenotype has yet to be assessed in GACI patients, and warrants future investigation.

Hypophosphataemia can compensate for the GACI phenotype and this might reflect a physiologic compensation mechanism rather than a primary defect [33]. However, several mutations in the *ENPP1* gene can result in the phenotype of autosomal-recessive hypophosphatemic rickets (ARHR) without any arterial calcification [45,46], suggesting a different pathway involved in the generation of ARHR linked to direct renal  $P_i$ -handling functions of ENPP1. Treatment with synthetic analogues of pyrophosphate, namely bisphosphonates seems to significantly increase survival in patients with GACI [33], however, spontaneous regression of the calcifications can occur [47], and studies on the exact long term natural history of the disease are pending. Based on the finding that mutations in *ENPP1* can also cause PXE and that mutations in *ABCC6* can also cause GACI it has become obvious that an overlap of genotype and phenotype in

GACI and PXE exists [48]. This has led to the hypothesis of a shared pathogenic principle in GACI and PXE [49], which finally held to be true. Most recently, subcutaneous administration of an ENPP1-Fc fusion protein was shown to prevent the mortality, vascular calcifications and sequelae of disease in the *asj/asj* mouse model of GACI [50<sup>\*\*</sup>]. This very promising preclinical study may pave the way to clinical trials with enzyme replacement therapy in patients with this rare disorder carrying mutations in *ENPP1*.

#### Arterial calcification due to deficiency of CD73 (ACDC)

Calcification of joints and arteries (arterial calcification due to deficiency of CD73 (ACDC); OMIM# 211800), as originally described by Magnus-Levy [51] was recently shown to be caused by deficiency of CD73, encoded by the *NT5E* gene [52,53]. Clinical features include calcified large vessels and periarticular calcifications in the joints of the hands and feet. CD73 has 5' exonucleotidase activity that converts AMP to adenosine and  $P_i$  [54]. Receptor binding of adenosine triggers a downstream intracellular signalling cascade that results in inhibition of TNAP activity [55]. Thus, it has been proposed that increased TNAP activity is central to the mechanism underpinning ACDC.

#### Hutchinson–Gilford progeria syndrome

The rare premature aging disorder Hutchinson–Gilford progeria syndrome (HGPS) is characterized by excessive atherosclerosis and both blood vessel and aortic valve calcification [56–58]. HGPS patients express progerin, a mutant form of lamin A. Progerin expression results in

abnormal nuclear membrane architecture causing abnormal higher-order chromatin organization [59,60]. Knock-in mice expressing progerin exhibit reduced circulating PP<sub>i</sub> levels similar to mice lacking ENPP1 [61]. Interestingly this low PP<sub>i</sub> is the result of increased TNAP activity and decreased extracellular ATP due to mitochondrial dysfunction. Consistent with this observation of suppressed PP<sub>i</sub> levels, these mice show excessive aortic calcification which is ameliorated by exogenous PP<sub>i</sub> administration [61]. These findings suggest connective tissue calcification in HGPS shares key features of PXE (decreased ATP), GACI (low PP<sub>i</sub>) and ACDC (increased TNAP activity), further demonstrating the overlap between these CTCs.

### Keutel syndrome

Keutel syndrome (KTLS) (OM#245150) is an extremely rare autosomal recessive disorder that manifests during the early childhood of patients predominantly from the Middle East. Since its first description in 1971 by Keutel and colleagues in two consanguineous siblings [62], less than 30 patients have been reported to date.

Foremost clinical characteristics of KTLS include abnormal calcification in laryngeal, tracheobronchial, auricular, nasal and rib cartilage, brachytelephalangism and facial abnormalities such as mid-facial hypoplasia, depressed nasal bridge and reduced alae nasi [62–67]. Further symptoms include mild to severe unilateral or bilateral hearing loss, multiple peripheral pulmonary artery stenosis, mental retardation and respiratory conditions, including dyspnoea and cough, that lead to hospitalization and incidentally to diagnosis [62–68]. Additionally, long-term follow-up studies have revealed that, KTLS patients develop skin lesions, typically after 30 years of age [69] and suffer chronic and progressive respiratory disease caused by gradual laryngotracheobronchial calcification and stenosis [65,69]. Subsequent post-mortem examination of the youngest sibling originally described has also uncovered calcification of pulmonary, coronary, hepatic, renal, meningeal and cerebral arteries [65–68].

It is well established that KTLS is due to loss-of-function mutations in the *MGP* gene [66,67,70–72], encoding MGP, a potent local mineralization inhibitor predominantly expressed by chondrocytes and VSMCs [73]. Post-translational modifications of MGP, such as vitamin K-dependent glutamate carboxylation and serine phosphorylation, have been shown to trigger these inhibitory properties [74]. Mice lacking *Mgp* develop abnormal cartilage calcification and extensive vascular calcification that leads to premature death due to aortic dissection [73]. Although all of the seven mutations reported in humans predict absent or non-functional MGP, they result in variable phenotypes with cardinal and secondary features of variable penetrance [66,67,70–72]. Moreover, in contrast to *Mgp*-deficient mice, with the exception of one

clinical case [65,67], vascular calcification has not been observed in KTLS patients. Interestingly, measurements of circulating MGP species have highlighted high levels of phosphorylated MGP [67]. This suggests that phosphorylation-dependent residual MGP activity may contribute to the absence of arterial calcification and more generally, that the variable phenotypic features observed clinically in KTLS could be explained by altered levels of the different MGP species.

Systemic hypertension is frequently found in KTLS patients before adulthood. Therefore, under the control of standard anti-hypertensive medication [69], KTLS has a good prognosis. Nevertheless, life expectancy of patients primarily depends on the severity of the associated respiratory complications. Indeed, symptomatic treatment with corticosteroids or bronchodilators is not always effective [65,67,69]. Interestingly, a recent study reported the efficacy of BMP inhibitors in reducing vascular calcification and improving survival in *Mgp*<sup>-/-</sup> mice [75\*\*]. Besides targeting vascular calcification, which is very rare in KTLS, BMP inhibition strategies could also be employed as a pharmaceutical approach to reduce abnormal cartilage calcification. This would have particularly high therapeutic value if successfully developed to treat tracheobronchial tree calcification, which initiates respiratory distress and complications, and ultimately determines the quality and duration of life of KTLS patients.

### Fibrodysplasia ossificans progressiva (FOP)

Fibrodysplasia ossificans progressiva (FOP; OMIM# 135100) is a devastating rare disease, characterized by the progressive heterotopic endochondral ossification (HOE) of skeletal muscle, fascia, tendons, and ligaments [76]. The range of joint motion in FOP patients becomes gradually and progressively limited by HOE. Indeed, this ossification is so diffuse that it is commonly referred to as a second skeleton [76]. HOE occurs in flare ups and is most commonly triggered by muscle injury, as also observed in *Abcc6*<sup>-/-</sup> mice [77], and viral infection. Unlike other CTCs discussed in this review, FOP is not the result of a deficiency but rather a gain of function mutation. All patients with the typical FOP phenotype have a substitution of the Arg206 residue for a His residue in Activin A receptor type I (ACVR1) which is a type 1 Bone Morphogenetic Protein Receptor (BMPR) [78]. This A206H ACVR1 exhibits ligand free activation of its BMP signalling pathways and a greatly enhanced response to BMP [79,80]. Studies suggest that FKBP1A (a protein that negatively regulates BMP type I R) has reduced binding capabilities to A206H ACVR1, preventing it from modulating its activity [79–81]. Knock-in mice that express the A206H ACVR1 recapitulate the phenotype of FOP patients [82], with data from these mice and other models suggesting that Tie2<sup>+</sup> mesenchymal cells are the progenitors of the HOE through their response to

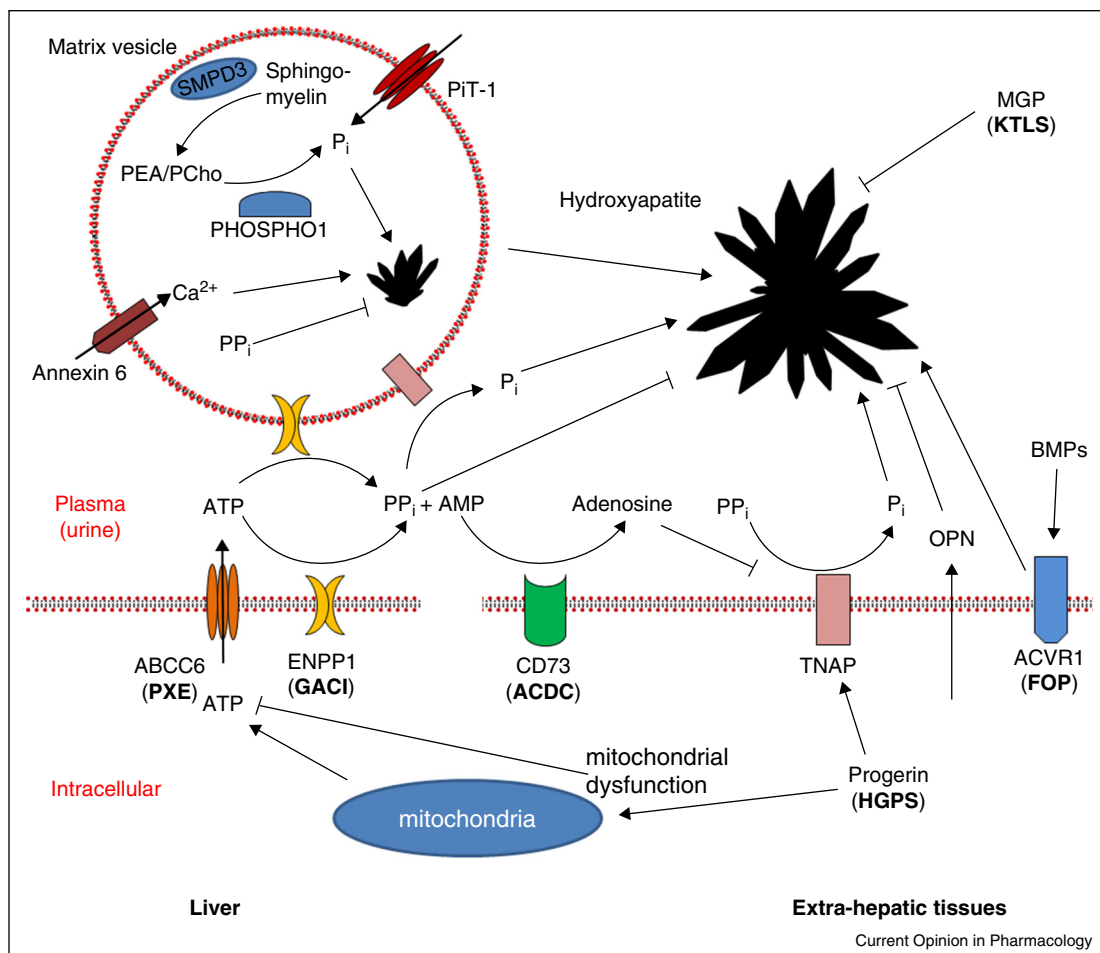
tissue inflammation [83–85]. Currently, management of FOP symptoms is limited to glucocorticoids and non-steroidal anti-inflammatories to minimize flare ups and pain [76], with no therapeutic strategy currently available to inhibit or prevent the HOE associated with this rare CTC. Given the positive effects of BMP inhibitors in *Mgp*-deficient mice, it would be beneficial to recapitulate these studies in the FOP mouse model as a first step to a pharmaceutical approach.

### Rare CTCs of the brain

There are a number of disorders demonstrating connective tissue calcification in the brain, including coeliac disease with Epilepsy and Cerebral calcifications (CEC; OMIM# 226810). To date, less than 200 CEC

patients have been reported in the literature [86]. The suspected Mendelian basis of this disease is characterized by three primary pathologies. The first is epilepsy, most commonly occipital epilepsy. Drug resistance is also common for this epilepsy with evolution towards epileptic encephalopathy. The second pathology is cerebral calcification, with bilateral occipital calcifications most frequently observed. The third pathology is coeliac disease, although in older patients (>2 years) bowel symptoms are less common [87,88]. Whilst folate deficiency has been proposed as the cause of CEC [89], recent studies suggest a possible autoimmune component to the disease with the identification of autoantibodies to transglutaminase isoenzyme 6 in the serum of a CEC patient [90]. Current CEC treatment regimens

Figure 3



Model of a functional network revealed by rare CTCs (GACI due to ENPP1 deficiency, PXE due to ABCC6 deficiency, KTLS due to MGP deficiency, ACDC due to CD73 deficiency, HGPS due to abnormal progerin expression and FOP due to ACVR1 gain of function). This model describes how ENPP1, ABCC6, MGP, CD73, progerin and ACVR1 serve as components in a network of factors, concurrent with established mechanisms of matrix vesicle regulated HA crystal formation, consequentially exerting balanced effects to promote and suppress connective tissue calcification. ENPP1 generates AMP and PP<sub>i</sub> from ATP, CD73 (ecto-5'-nucleotidase) hydrolyses AMP to generate adenosine and P<sub>i</sub>, TNAP hydrolyses PP<sub>i</sub> into two P<sub>i</sub> molecules. PP<sub>i</sub> suppresses hydroxyapatite deposition and inhibits connective tissue calcification. Adenosine signalling suppresses TNAP expression. P<sub>i</sub> is a component of hydroxyapatite crystal deposition. A206H ACVR1 exhibits an enhanced response to BMP signalling. The roles of ABCC6 and MGP have yet to be fully defined. Progerin causes increased TNAP activity and decreased ATP concentrations due to mitochondrial dysfunction, combined these effects of progerin result in decreased PP<sub>i</sub>.

include folate supplementation and gluten free diet, the efficacy of which has been shown to directly correlate with how early treatment is implemented [87,88].

Idiopathic Basal Ganglia Calcification (IBGC; OMIM# 213600) is characterized by calcification of the basal ganglia as well as the thalamus and cerebellum [91]. Patients display a range of neuropsychiatric and movement disorders including dementia, psychosis, Parkinsonism, dystonia, and migraine. Mutations in the gene for type III sodium-dependent  $P_i$  co-transporter 2 (PiT-2) leading to impaired or loss of function have been identified in several patient cohorts [92–96]. Direct evidence for a role for PiT-2 in IBGC was first provided by studies investigating the phenotype of mice lacking PiT-2. These mice develop calcification predominantly in the thalamus but also in the basal ganglia and brain cortex [97]. Of particular interest, histological analyses of these mice suggest that calcification initiates in or around the vasculature which closely mimics the phenotype observed in IBGC patients [91]. Mutations in the gene for platelet derived growth factor receptor B (PDGFRB) have also been described in IBGC patients [98]. PDGFRB regulates PiT-1 in VSMCs [99,100] and a role for this molecule in IBGC would be consistent with a vascular origin of the calcification. Considering IBGC patients have normal circulating  $P_i$  levels [91] any role PiT-2 and/or PDGFRB may play in calcification would be local to the affected tissue. This highlights the heterogeneity of  $P_i$  regulation in different tissues which should be an important consideration when planning pharmaceutical interventions for CTCs.

### Future directions

Much of our understanding of the potential mechanisms underpinning CTCs (Figure 3) has arisen through the use of rodent models. However, significant differences between physiology, anatomy, and pathology exist between mice and men. In contrast, large animal models can show markedly greater similarity to humans [101\*]. The recent explosion of precise and efficient genome editing techniques through CRISPR/Cas9 technology permits the generation of tailored models for translational research [102\*\*]. These novel systems provide huge potential for large animal models for future investigations into the regulatory factors and molecular pathways that contribute to rare inherited forms of CTC *in vivo*.

At present, only very limited pharmaceutical strategies exist to inhibit connective tissue calcification. Further pre-clinical and clinical studies are required to examine new approaches such as targeting mechanisms common to different CTCs (e.g.  $PP_i$  regulation) and/or enzyme replacement therapy (e.g. ENPP1). These investigations may bring to fruition the first comprehensive treatment for both inherited and acquired CTCs.

### Conflict of interest statement

Nothing declared.

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### References and recommended reading

Papers of particular interest, published within the period of review, have been highlighted as:

- of special interest
- of outstanding interest

1. Anderson HC, Morris DC: **Mineralization. Physiology and Pharmacology of Bone. Handbook of Experimental Pharmacology.** Berlin Heidelberg: Springer; 1993, 267-298.
2. Zhu D, Mackenzie NCW, Farquharson C, Macrae VE: **Mechanisms and clinical consequences of vascular calcification.** *Front Endocrinol* 2012, **3**.
3. Anderson HC: **Matrix vesicles and calcification.** *Curr Rheumatol Rep* 2003, **5**:222-226.
4. Wuthier RE, Lipscomb GF: **Matrix vesicles: structure, composition, formation and function in calcification.** *Front Biosci* 2011, **16**:2812-2902.
5. Ho AM, Johnson MD, Kingsley DM: **Role of the mouse ank gene in control of tissue calcification and arthritis.** *Science* 2000, **289**:265-270.
6. Mackenzie NCW, Huesa C, Rutsch F, MacRae VE: **New insights into NPP1 function: lessons from clinical and animal studies.** *Bone* 2012, **51**:961-968.
7. Yadav MC, Simão AMS, Narisawa S, Huesa C, McKee MD, Farquharson C, Millán JL: **Loss of skeletal mineralization by the simultaneous ablation of PHOSPHO1 and alkaline phosphatase function: a unified model of the mechanisms of initiation of skeletal calcification.** *J Bone Miner Res* 2011, **26**:286-297.
8. Polewski MD, Johnson KA, Foster M, Millán JL, Terkeltaub R: **Inorganic pyrophosphatase induces type I collagen in osteoblasts.** *Bone* 2010, **46**:81.
9. Harmey D, Hesse L, Narisawa S, Johnson KA, Terkeltaub R, Millán JL: **Concerted regulation of inorganic pyrophosphate and osteopontin by Akp2, Enpp1, and Ank: an integrated model of the pathogenesis of mineralization disorders.** *Am J Pathol* 2004, **164**:1199-1209.
10. Yadav MC, Huesa C, Narisawa S, Hoylaerts MF, Moreau A, Farquharson C, Millán JL: **Ablation of osteopontin improves the skeletal phenotype of Phospho1<sup>-/-</sup> mice.** *J Bone Miner Res* 2014, **29**:2369-2381.
- This study demonstrates that osteopontin (Opn) is elevated in *Phospho1* knock out mice and that concurrent knockout of *Opn* with *Phospho1* rescues the skeletal phenotype.
11. Zhu D, Mackenzie NCW, Millán JL, Farquharson C, MacRae VE: **The appearance and modulation of osteocyte marker expression during calcification of vascular smooth muscle cells.** *PLoS One* 2011, **6**:e19595.
12. Kapustin AN, Chatrou MLL, Drozdov I, Zheng Y, Davidson SM, Soong D, Furmanik M, Sanchis P, De Rosales RTM, Alvarez-Hernandez D *et al.*: **Vascular smooth muscle cell calcification is mediated by regulated exosome secretion.** *Circ Res* 2015, **116**:1312-1323.
- This study identifies matrix vesicles as exosomes and demonstrates that factors that can increase exosome release promote vascular calcification.



13. Cui L, Houston DA, Farquharson C, MacRae VE: **Characterisation of matrix vesicles in skeletal and soft tissue calcification.** *Bone* 2016. in press.  
This study identifies matrix vesicles as exosomes and demonstrates that factors that can increase exosome release promote vascular calcification.
14. Dai XY, Zhao MM, Cai Y, Guan QC, Zhao Y, Guan Y, Kong W, Zhu WG, Xu MJ, Wang X: **Phosphate-induced autophagy counteracts vascular calcification by reducing matrix vesicle release.** *Kidney Int* 2013, **83**:1042-1051.  
This paper demonstrates that autophagy is dependent on oxidative stress and that inhibiting it increases calcification, whilst inducing autophagy decreases calcification.
15. Le Saux O, Urban Z, Tschuch C, Csiszar K, Bacchelli B, Quaglini D, Pasquali-Ronchetti I, Pope FM, Richards A, Terry S et al.: **Mutations in a gene encoding an ABC transporter cause pseudoxanthoma elasticum.** *Nat Genet* 2000, **25**:223-227.
16. Hu X, Plomp AS, van Soest S, Wijnholds J, de Jong PT, Bergen AA: **Pseudoxanthoma elasticum: a clinical, histopathological, and molecular update.** *Surv Ophthalmol* 2003, **48**:424-438.
17. Uitto J, Bercovitch L, Terry SF, Terry PF: **Pseudoxanthoma elasticum: progress in diagnostics and research towards treatment: summary of the 2010 PXE International Research Meeting.** *Am J Med Genet A* 2011, **155**:1517-1526.
18. Neldner KH: **Pseudoxanthoma elasticum.** *Clin Dermatol* 1988, **6**:1-159.
19. Jansen RS, Kucukosmanoglu A, de Haas M, Sapthu S, Otero JA, Hegman IE, Bergen AA, Gorgels TG, Borst P, van de Wetering K: **ABCC6 prevents ectopic mineralization seen in pseudoxanthoma elasticum by inducing cellular nucleotide release.** *Proc Natl Acad Sci U S A* 2013, **110**:20206-20211.  
This study reveals that the factor that normally prevents PXE is pyrophosphate, which is provided to the circulation in the form of nucleoside triphosphates.
20. Terkeltaub RA: **Inorganic pyrophosphate generation and disposition in pathophysiology.** *Am J Physiol Cell Physiol* 2001, **281**:C1-C11.
21. Jansen RS, Duijst S, Mahakena S, Sommer D, Szeri F, Varadi A, Plomp A, Bergen AA, Oude Elferink RP, Borst P et al.: **ABCC6-mediated ATP secretion by the liver is the main source of the mineralization inhibitor inorganic pyrophosphate in the systemic circulation-brief report.** *Arterioscler Thromb Vasc Biol* 2014, **34**:1985-1989.  
This is the first report to demonstrate that the liver is the main source of circulating ATP and that this is mediated by ABCC6.
22. Hosen MJ, Van Nieuwerburgh F, Steyaert W, Deforce D, Martin L, Leftheriotis G, De Paepe A, Coucke PJ, Vanakker OM: **Efficiency of exome sequencing for the molecular diagnosis of pseudoxanthoma elasticum.** *J Invest Dermatol* 2015, **135**:992-998.
23. Kuzaj P, Kuhn J, Michalek RD, Karoly ED, Faust I, Dabisch-Ruthe M, Knabbe C, Hendig D: **Large-scaled metabolic profiling of human dermal fibroblasts derived from pseudoxanthoma elasticum patients and healthy controls.** *PLoS One* 2014, **9**:e108336.
24. Boraldi F, Bartolomeo A, Li Q, Uitto J, Quaglini D: **Changes in dermal fibroblasts from Abcc6(-/-) mice are present before and after the onset of ectopic tissue mineralization.** *J Invest Dermatol* 2014, **134**:1855-1861.
25. Boraldi F, Annovi G, Guerra D, Paolinelli Devincenzi C, Garcia-Fernandez MI, Panico F, De Santis G, Tiozzo R, Ronchetti I, Quaglini D: **Fibroblast protein profile analysis highlights the role of oxidative stress and vitamin K recycling in the pathogenesis of pseudoxanthoma elasticum.** *Proteomics Clin Appl* 2009, **3**:1084-1098.
26. Kauffenstein G, Pizard A, Le Corre Y, Vessieres E, Grimaud L, Toutain B, Labat C, Mauras Y, Gorgels TG, Bergen AA et al.: **Disseminated arterial calcification and enhanced myogenic response are associated with abcc6 deficiency in a mouse model of pseudoxanthoma elasticum.** *Arterioscler Thromb Vasc Biol* 2014, **34**:1045-1056.
27. Vasseur M, Carsin-Nicol B, Ebran JM, Willoteaux S, Martin L, Leftheriotis G, Angers PXECCG: **Carotid rete mirabile and pseudoxanthoma elasticum: an accidental association?** *Eur J Vasc Endovasc Surg* 2011, **42**:292-294.
28. Burnstock G, Vaughn B, Robson SC: **Purinergic signalling in the liver in health and disease.** *Purinergic Signal* 2014, **10**:51-70.
29. Burnstock G, Evans LC, Bailey MA: **Purinergic signalling in the kidney in health and disease.** *Purinergic Signal* 2014, **10**:71-101.
30. Burnstock G, Ralevic V: **Purinergic signaling and blood vessels in health and disease.** *Pharmacol Rev* 2014, **66**:102-192.
31. Orriss IR: **The role of purinergic signalling in the musculoskeletal system.** *Auton Neurosci* 2015, **191**:124-134.
32. Rutsch F, Ruf N, Vaingankar S, Toliat MR, Suk A, Hohne W, Schauer G, Lehmann M, Roscioli T, Schnabel D et al.: **Mutations in ENPP1 are associated with 'idiopathic' infantile arterial calcification.** *Nat Genet* 2003, **34**:379-381.
33. Rutsch F, Boyer P, Nitschke Y, Ruf N, Lorenz-Depierieux B, Wittkamp T, Weissen-Plenz G, Fischer RJ, Mughal Z, Gregory JW et al.: **Hypophosphatemia, hyperphosphaturia, and bisphosphonate treatment are associated with survival beyond infancy in generalized arterial calcification of infancy.** *Circ Cardiovasc Genet* 2008, **1**:133-140.
34. Ferreira C, Ziegler S, Gahl W: **Generalized arterial calcification of infancy.** In *GeneReviews(R)*. Edited by Pagon RA, Adam MP, Ardinger HH, Wallace SE, Amemiya A, Bean LH, Bird TD, Fong CT, Mefford HC, Smith RJ.. Seattle: University of Washington; 2014.  
This paper describes the clinical manifestations and epidemiology of GACI.
35. Dlamini N, Splitt M, Durkan A, Siddiqui A, Padayachee S, Hobbins S, Rutsch F, Wraige E: **Generalized arterial calcification of infancy: phenotypic spectrum among three siblings including one case without obvious arterial calcifications.** *Am J Med Genet A* 2009, **149A**:456-460.
36. Cheng KS, Chen MR, Ruf N, Lin SP, Rutsch F: **Generalized arterial calcification of infancy: different clinical courses in two affected siblings.** *Am J Med Genet A* 2005, **136**:210-213.
37. Okawa A, Nakamura I, Goto S, Moriya H, Nakamura Y, Ikegawa S: **Mutation in Npps in a mouse model of ossification of the posterior longitudinal ligament of the spine.** *Nat Genet* 1998, **19**:271-273.
38. Hosoda Y, Yoshimura Y, Higaki S: **A new breed of mouse showing multiple osteochondral lesions – twy mouse.** *Ryumachi* 1981, **21 Suppl**:157-164.
39. Li Q, Guo H, Chou DW, Berndt A, Sundberg JP, Uitto J: **Mutant Enpp1(asj) mice as a model for generalized arterial calcification of infancy.** *Dis Model Mech* 2013, **6**:1227-1235.
40. Li Q, Pratt CH, Dionne LA, Fairfield H, Karst SY, Sundberg JP, Uitto J: **Spontaneous asj-2J mutant mouse as a model for generalized arterial calcification of infancy: a large deletion/insertion mutation in the Enpp1 gene.** *PLoS One* 2014, **9**:e113542.
41. Johnson K, Polewski M, van Etten D, Terkeltaub R: **Chondrogenesis mediated by PPI depletion promotes spontaneous aortic calcification in NPP1-/- mice.** *Arterioscler Thromb Vasc Biol* 2005, **25**:686-691.
42. Mackenzie NCW, Zhu D, Milne EM, van 't Hof R, Martin A, Quarles DL, Millán JL, Farquharson C, MacRae VE: **Altered bone development and an increase in FGF-23 expression in Enpp1-/- mice.** *PLoS One* 2012, **7**:e32177.
43. Hajjawi MOR, MacRae VE, Huesa C, Boyde A, Millán JL, Arnett TR, Orriss IR: **Mineralisation of collagen rich soft tissues and osteocyte lacunae in Enpp1(-/-) mice.** *Bone* 2014, **69C**:139-147.  
This study demonstrated for the first time that ENPP1 is expressed both in osteocytes and osteoclasts and further characterizes the ENPP1<sup>-/-</sup> mouse phenotype.
44. Huesa C, Zhu D, Glover JD, Ferron M, Karsenty G, Milne EM, Millán JL, Ahmed SF, Farquharson C, Morton NM et al.: **Deficiency of the bone mineralization inhibitor NPP1 protects**

- against obesity and diabetes. *Dis Model Mech* 2014, **7**: 1341-1350.
45. Lorenz-Depiereux B, Schnabel D, Tiosano D, Hausler G, Strom TM: **Loss-of-function ENPP1 mutations cause both generalized arterial calcification of infancy and autosomal-recessive hypophosphatemic rickets.** *Am J Hum Genet* 2010, **86**:267-272.
  46. Levy-Litan V, Hershkovitz E, Avizov L, Leventhal N, Bercovich D, Chalifa-Caspi V, Manor E, Buriakovsky S, Hadad Y, Goding J *et al.*: **Autosomal-recessive hypophosphatemic rickets is associated with an inactivation mutation in the ENPP1 gene.** *Am J Hum Genet* 2010, **86**:273-278.
  47. Sholler GF, Yu JS, Bale PM, Hawker RE, Celermajer JM, Kozlowski K: **Generalized arterial calcification of infancy: three case reports, including spontaneous regression with long-term survival.** *J Pediatr* 1984, **105**:257-260.
  48. Nitschke Y, Baujat G, Botschen U, Wittkamp T, du Moulin M, Stella J, Le Merrer M, Guest G, Lambot K, Tazarourte-Pinturier MF *et al.*: **Generalized arterial calcification of infancy and pseudoxanthoma elasticum can be caused by mutations in either ENPP1 or ABCC6.** *Am J Hum Genet* 2012, **90**:25-39.
  49. Nitschke Y, Rutsch F: **Generalized arterial calcification of infancy and pseudoxanthoma elasticum: two sides of the same coin.** *Front Genet* 2012, **3**:302.
  50. Albright RA, Stabach P, Cao W, Kavanagh D, Mullen I, Braddock AA, Covo MS, Tehan M, Yang G, Cheng Z *et al.*: **ENPP1-Fc prevents mortality and vascular calcifications in rodent model of generalized arterial calcification of infancy.** *Nat Commun* 2015, **6**:10006.
- This is the first report to demonstrate the efficacy of exogenous recombinant ENPP1 to treat GACI.
51. Von Magnus-Levy A: **Ueber ungewöhnliche Verkalkung der Arterien.** *Dtsch Med Wochenschr* 1914, **40**:1305-1309.
  52. St. Hilaire C, Ziegler SG, Markello TC, Brusco A, Groden C, Gill F, Carlson-Donohoe H, Lederman RJ, Chen MY, Yang D *et al.*: **NT5E mutations and arterial calcifications.** *N Engl J Med* 2011, **364**:432-442.
  53. Zhang Z, He JW, Fu WZ, Zhang CQ, Zhang ZL: **Calcification of joints and arteries: second report with novel NT5E mutations and expansion of the phenotype.** *J Hum Genet* 2015, **60**:561-564.
  54. Yegutkin GG: **Nucleotide-and nucleoside-converting ectoenzymes: important modulators of purinergic signalling cascade.** *BBA Mol Cell Res* 2008, **1783**:673-694.
  55. Markello TC, Pak LK, St. Hilaire C, Dorward H, Ziegler SG, Chen MY, Chaganti K, Nussbaum RL, Boehm M, Gahl WA: **Vascular pathology of medial arterial calcifications in NT5E deficiency: implications for the role of adenosine in pseudoxanthoma elasticum.** *Mol Genet Metab* 2011, **103**:44-50.
  56. Nair K, Ramachandran P, Krishnamoorthy KM, Dora S, Achuthan TJ: **Hutchinson-Gilford progeria syndrome with severe calcific aortic valve stenosis and calcific mitral valve.** *J Heart Valve Dis* 2004, **13**:866-869.
  57. Salamat M, Dhar PK, Neagu DL, Lyon JB: **Aortic calcification in a patient with Hutchinson-Gilford progeria syndrome.** *Pediatr Cardiol* 2010, **31**:925-926.
  58. Hanumanthappa NB, Madhusudan G, Mahimarangaiha J, Manjunath CN: **Hutchinson-Gilford progeria syndrome with severe calcific aortic valve stenosis.** *Ann Pediatr Cardiol* 2011, **4**:204.
  59. Eriksson M, Brown WT, Gordon LB, Glynn MW, Singer J, Scott L, Erdos MR, Robbins CM, Moses TY, Berglund P: **Recurrent de novo point mutations in lamin A cause Hutchinson-Gilford progeria syndrome.** *Nature* 2003, **423**:293-298.
  60. De Sandre-Giovannoli A, Bernard R, Cau P, Navarro C, Amiel J, Boccaccio I, Lyonnet S, Stewart CL, Munnich A, Le Merrer M *et al.*: **Lamin A truncation in Hutchinson-Gilford progeria.** *Science* 2003, **300**:2055.
  61. Villa-Bellosta R, Rivera-Torres J, Osorio FG, Acín-Pérez R, Enriquez JA, López-Otín C, Andrés V: **Defective extracellular pyrophosphate metabolism promotes vascular calcification in a mouse model of Hutchinson-Gilford progeria syndrome that is ameliorated on pyrophosphate treatment.** *Circulation* 2013, **127**:2442-2451.
  62. Keutel J, Jorgensen G, Gabriel P: **A new autosomal-recessive hereditary syndrome. Multiple peripheral pulmonary stenosis, brachytelephalangia, inner-ear deafness, ossification or calcification of cartilages.** *Dtsch Med Wochenschr* 1971, **96**:1676-1681 passim.
  63. Khosroshahi HE, Uluoglu O, Olgunturk R, Basaklar C: **Keutel syndrome: a report of four cases.** *Eur J Pediatr* 1989, **149**:188-191.
  64. Ziereisen F, De Munter C, Perlmutter N: **The Keutel syndrome. Report of a case and review of the literature.** *Pediatr Radiol* 1993, **23**:314-315.
  65. Meier M, Weng LP, Alexandrakis E, Ruschoff J, Goeckenjan G: **Tracheobronchial stenosis in Keutel syndrome.** *Eur Respir J* 2001, **17**:566-569.
  66. Hur DJ, Raymond GV, Kahler SG, Riegert-Johnson DL, Cohen BA, Boyadiev SA: **A novel MGP mutation in a consanguineous family: review of the clinical and molecular characteristics of Keutel syndrome.** *Am J Med Genet A* 2005, **135**:36-40.
  67. Cranenburg EC, Vans-Z KY, Bonafe L, Mittaz Crettol L, Rodiger LA, Dikkers FG, Vane AJ, Superti-Furga A, Alexandrakis E, Vermeer C *et al.*: **Circulating matrix gamma-carboxyglutamate protein (MGP) species are refractory to vitamin K treatment in a new case of Keutel syndrome.** *J Thromb Haemost* 2011, **9**:1225-1235.
  68. Sun LF, Chen X: **Tracheobronchial stenosis in Keutel syndrome.** *Indian Pediatr* 2012, **49**:759.
  69. Khosroshahi HE, Sahin SC, Akyuz Y, Ede H: **Long term follow-up of four patients with Keutel syndrome.** *Am J Med Genet A* 2014, **164A**:2849-2856.
  70. Munroe PB, Olgunturk RO, Fryns J-P, Maldergem LV, Ziereisen F, Yuksel B, Gardiner RM, Chung E: **Mutations in the gene encoding the human matrix Gla protein cause Keutel syndrome.** *Nat Genet* 1999, **21**:142-144.
  71. Bayramoglu A, Saritemur M, Tasdemir S, Omeroglu M, Erdem HB, Sahin I: **A rare cause of dyspnea in emergency medicine: Keutel syndrome.** *Am J Emerg Med* 2015.
  72. Weaver KN, El Hallek M, Hopkin RJ, Sund KL, Henrickson M, Del Gaudio D, Yuksel A, Acar GO, Bober MB, Kim J *et al.*: **Keutel syndrome: report of two novel MGP mutations and discussion of clinical overlap with arylsulfatase E deficiency and relapsing polychondritis.** *Am J Med Genet A* 2014, **164A**:1062-1068.
  73. Luo G, Ducey P, McKee MD, Pinero GJ, Loyer E, Behringer RR, Karsenty G: **Spontaneous calcification of arteries and cartilage in mice lacking matrix GLA protein.** *Nature* 1997, **386**:78-81.
  74. Schurgers LJ, Spronk HM, Skepper JN, Hackeng TM, Shanahan CM, Vermeer C, Weissberg PL, Proudfoot D: **Post-translational modifications regulate matrix Gla protein function: importance for inhibition of vascular smooth muscle cell calcification.** *J Thromb Haemost* 2007, **5**:2503-2511.
  75. Malhotra R, Burke MF, Martyn T, Shakartz HR, Thayer TE, O'Rourke C, Li P, Derwall M, Spagnoli E, Kolodziej SA *et al.*: **Inhibition of bone morphogenetic protein signal transduction prevents the medial vascular calcification associated with matrix Gla protein deficiency.** *PLoS One* 2015, **10**:e0117098.
- This study demonstrates that inhibiting BMP signalling decreases vascular calcification in MGP<sup>-/-</sup> mice and that some of the features of MGP<sup>-/-</sup> are dependent on BMP signalling.
76. Kaplan FS, Chakkalakal SA, Shore EM: **Fibrodysplasia ossificans progressiva: mechanisms and models of skeletal metamorphosis.** *Dis Model Mech* 2012, **5**:756-762.
  77. Brampton C, Aherrahrou Z, Chen LH, Martin L, Bergen AA, Gorgels TG, Erdmann J, Schunkert H, Szabo Z, Varadi A *et al.*: **The level of hepatic ABCC6 expression determines the severity of calcification after cardiac injury.** *Am J Pathol* 2014, **184**:159-170.

78. Shore EM, Xu M, Feldman GJ, Fenstermacher DA, Cho T-J, Choi IH, Connor JM, Delai P, Glaser DL, LeMerrer M et al.: **A recurrent mutation in the BMP type I receptor ACVR1 causes inherited and sporadic fibrodysplasia ossificans progressiva.** *Nat Genet* 2006, **38**:525-527.
79. Song G-A, Kim H-J, Woo K-M, Baek J-H, Kim G-S, Choi J-Y, Ryoo H-M: **Molecular consequences of the ACVR1R206H mutation of fibrodysplasia ossificans progressiva.** *J Biol Chem* 2010, **285**:22542-22553.
80. van Dinther M, Visser N, de Gorter DJ, Doorn J, Goumans MJ, de Boer J, ten Dijke P: **ALK2 R206H mutation linked to fibrodysplasia ossificans progressiva confers constitutive activity to the BMP type I receptor and sensitizes mesenchymal cells to BMP-induced osteoblast differentiation and bone formation.** *J Bone Miner Res* 2010, **25**:1208-1215.
81. Groppe JC, Wu J, Shore EM, Kaplan FS: **In vitro analyses of the dysregulated R206H ALK2 kinase-FKBP12 interaction associated with heterotopic ossification in FOP.** *Cells Tissues Organs* 2011, **194**:291-295.
82. Chakkalakal SA, Zhang D, Culbert AL, Convente MR, Caron RJ, Wright AC, Maidment AD, Kaplan FS, Shore EM: **An Acvr1 R206H knock-in mouse has fibrodysplasia ossificans progressiva.** *J Bone Miner Res* 2012, **27**:1746-1756.
83. Lounev VY, Ramachandran R, Wosczyzna MN, Yamamoto M, Maidment AD, Shore EM, Glaser DL, Goldhamer DJ, Kaplan FS: **Identification of progenitor cells that contribute to heterotopic skeletogenesis.** *J Bone Joint Surg* 2009, **91**:652-663.
84. Medici D, Shore EM, Lounev VY, Kaplan FS, Kalluri R, Olsen BR: **Conversion of vascular endothelial cells into multipotent stem-like cells.** *Nat med* 2010, **16**:1400-1406.
85. Wosczyzna MN, Biswas AA, Cogswell CA, Goldhamer DJ: **Multipotent progenitors resident in the skeletal muscle interstitium exhibit robust BMP-dependent osteogenic activity and mediate heterotopic ossification.** *J Bone Miner Res* 2012, **27**:1004-1017.
86. Gobbi G: *Orphanet: Celiac Disease, Epilepsy and Cerebral Calcification Syndrome.* 2012.
87. Gobbi G: **Coeliac disease, epilepsy and cerebral calcifications.** *Brain Dev* 2005, **27**:189-200.
88. Arroyo HA, De Rosa S, Ruggieri V, de Davila MT, Fejerman N: **Epilepsy, occipital calcifications, and oligosymptomatic celiac disease in childhood.** *J Child Neurol* 2002, **17**:800-806.
89. Calvani M Jr, Parisi P, Guaitolini C, Parisi G, Paolone G: **Latent coeliac disease in a child with epilepsy, cerebral calcifications, drug-induced systemic lupus erythematosus and intestinal folic acid malabsorption associated with impairment of folic acid transport across the blood-brain barrier.** *Eur J Pediatr* 2001, **160**:288-292.
90. Johnson AM, Dale RC, Wienholt L, Hadjivassiliou M, Aeschlimann D, Lawson JA: **Coeliac disease, epilepsy, and cerebral calcifications: association with TG6 autoantibodies.** *Dev Med Child Neurol* 2013, **55**:90-93.
91. Sobrido MJ, Coppola G, Oliveira J, Hopfer S, Geschwind DH: *Primary Familial Brain Calcification.* 2013.
92. Wang C, Li Y, Shi L, Ren J, Patti M, Wang T, de Oliveira JR, Sobrido MJ, Quintans B, Baquero M et al.: **Mutations in SLC20A2 link familial idiopathic basal ganglia calcification with phosphate homeostasis.** *Nat Genet* 2012, **44**:254-256.
93. Schottlaender L, Mencacci N, Koepf M, Hanna M, Hardy J, Lees A, Houlden H: *Interesting Clinical Features Associated with Mutations in the SLC20A2 Gene.* 2012.
94. Hsu SC, Sears RL, Lemos RR, Quintans B, Huang A, Spiteri E, Nevarez L, Mamah C, Zatz M, Pierce KD et al.: **Mutations in SLC20A2 are a major cause of familial idiopathic basal ganglia calcification.** *Neurogenetics* 2013, **14**:11-22.
95. Lemos RR, Oliveira MF, Oliveira JRM: **Reporting a new mutation at the SLC20A2 gene in familial idiopathic basal ganglia calcification.** *Eur J Neurol* 2013, **20**:e43-e44.
96. Zhang Y, Guo X, Wu A: **Association between a novel mutation in SLC20A2 and familial idiopathic basal ganglia calcification.** *PLoS One* 2013, **8**:e57060.
97. Jensen N, Schröder HD, Hejbøl EK, Füchtbauer E-M, de Oliveira JRM, Pedersen L: **Loss of function of Slc20a2 associated with familial idiopathic basal ganglia calcification in humans causes brain calcifications in mice.** *J Mol Neurosci* 2013, **51**:994-999.
98. Nicolas G, Pottier C, Maltete D, Coutant S, Rovelet-Lecruc A, Legallic S, Rousseau S, Vaschalde Y, Guyant-Marechal L, Augustin J et al.: **Mutation of the PDGFRB gene as a cause of idiopathic basal ganglia calcification.** *Neurology* 2013, **80**:181-187.
99. Kakita A, Suzuki A, Nishiwaki K, Ono Y, Kotake M, Ariyoshi Y, Miura Y, Oiso Y: **Stimulation of Na-dependent phosphate transport by platelet-derived growth factor in rat aortic smooth muscle cells.** *Atherosclerosis* 2004, **174**:17-24.
100. Villa-Bellosta R, Levi M, Sorribas V: **Vascular smooth muscle cell calcification and SLC20 inorganic phosphate transporters: effects of PDGF, TNF- $\alpha$ , and Pi.** *Pflug Arch Eur J Phy* 2009, **458**:1151-1161.
101. Tsang HG, Rashdan NA, Whitelaw CBA, Corcoran BM, Summers KM1, MacRae VE: **Large animal models of cardiovascular disease.** *Cell Biochem Funct* 2016 <http://dx.doi.org/10.1002/cbf.3173>.
- This paper provides a current indepth review of large animal models relevant to cardiovascular disease.
102. Hsu PD, Lander ES, Zhang F: **Development and applications of CRISPR-Cas9 for genome engineering.** *Cell* 2014, **157**:1262-1278.
- This review describes the development and applications of Cas9 for a variety of research and translational applications.