

## Supplementary Tables

**Table S1. Strains used in this study.**

Strain	Genotype	Source	Figure
YL515	[BY4741/2] <i>MATa</i> ; <i>his3Δ1</i> , <i>leu2Δ0</i> , <i>ura3Δ0</i>	[1]	1A-F
MB27	[YL515] <i>gtr1Δ::HIS3</i>	[1]	1A-F, 2C
MB28	[YL515] <i>gtr2Δ::HIS3</i>	[1]	1A-F, 2C
NP52-2A	[YL515] <i>ego2Δ::kanMX</i>	This study	1A-F
NP44	[YL515] <i>ego4Δ::kanMX</i>	This study	1A-F
NP51-3C	[YL515] <i>ego2Δ::kanMX</i> , <i>gtr1Δ::kanMX</i>	This study	1A, C- D, F, 2C
NP54-4D	[YL515] <i>ego2Δ::kanMX</i> , <i>gtr2Δ::kanMX</i>	This study	1A, C- D, F, 2C
NP48-5C	[YL515] <i>ego4Δ::kanMX</i> , <i>gtr1Δ::kanMX</i>	This study	1A, C- D, F, 2C
NP60-10D	[YL515] <i>ego4Δ::kanMX</i> , <i>gtr2Δ::kanMX</i>	This study	1A, C- D, F, 2C
KT1961	<i>MATa</i> ; <i>trp1</i> , <i>leu2</i> , <i>his3</i> , <i>ura3-52</i>	[2]	5C-D
CDV213	[KT1961] <i>EGO1-GFP::TRP1</i>	[3]	2A
CDV221-1A	[KT1961] <i>ego3Δ::natMX</i> , <i>EGO1-GFP::TRP1</i>	This study	2A
KP12-1C	[KT1961] <i>ego2Δ::kanMX</i> , <i>EGO1-GFP::TRP1</i>	This study	2A, 3B
KP37-4B	[KT1961] <i>ego2Δ::kanMX</i> , <i>ego3Δ::natMX</i> <i>EGO1-GFP::TRP1</i>	This study	2A
KP22-2C	[KT1961] <i>ego4Δ::kanMX</i> , <i>EGO1-GFP::TRP1</i>	This study	2A, 3B
CDV214	[KT1961] <i>EGO3-GFP::TRP1</i>	[3]	2A
CDV219-4A	[KT1961] <i>ego1Δ::natMX</i> , <i>EGO3-GFP::TRP1</i>	This study	2A, 5B
KP13-2D	[KT1961] <i>ego2Δ::kanMX</i> , <i>EGO3-GFP::TRP1</i>	This study	2A, 3B
KP39-6D	[KT1961] <i>ego2Δ::kanMX</i> , <i>ego1Δ::natMX</i> , <i>EGO3-GFP::TRP1</i>	This study	2A
KP35-2D	[KT1961] <i>ego4Δ::kanMX</i> , <i>EGO3-GFP::TRP1</i>	This study	2A, 3B
CDV210	[KT1960] <i>ego1Δ::natMX</i>	[3]	2B, 5C-D
KP27-5A	[KT1961] <i>ego1Δ::natMX</i> , <i>ego2Δ::kanMX</i>	This study	2B, 3F, 5B
KP33-3C	[KT1961] <i>ego1Δ::natMX</i> , <i>ego4Δ::kanMX</i>	This study	2B
MP268-2B	[KT1961] <i>gtr1Δ::natMX</i> , <i>gtr2Δ::natMX</i>	This study	2D-E, 5C-D, 6A-C
KP29-3B	[KT1961] <i>gtr1Δ::natMX</i> , <i>gtr1Δ::natMX</i> , <i>ego2Δ::kanMX</i>	This study	2D-E, 3F
KP34-1A	[KT1961] <i>gtr1Δ::natMX</i> , <i>gtr1Δ::natMX</i> , <i>ego4Δ::kanMX</i>	This study	2D-E
NMY51	<i>MATa</i> ; <i>his3Δ200</i> , <i>trp1-901</i> , <i>leu2-3,112</i> , <i>ade2</i> , <i>LYS2::(lexAop)4-HIS3</i> , <i>ura3::(lexAop)8-lacZ</i> , <i>ade2::(lexAop)8-ADE2</i> , <i>GAL4</i>	Dualsystems	3A, 5A
FLJ1	[NMY51] <i>ego3Δ::kanMX</i>	[4]	3G
KP12-2D	[KT1961] <i>ego2Δ::kanMX</i>	This study	3E-F
KP02	[KT1961] <i>ego4Δ::kanMX</i>	This study	3E-F
KP28-3D	[KT1961] <i>ego2Δ::kanMX</i> , <i>ego3Δ::natMX</i>	This study	3F
MP279-18B	[KT1960] <i>gtr1Δ::natMX</i> , <i>ego1Δ::kanMX</i>	This study	5B
MP279-18A	[KT1961] <i>gtr2Δ::natMX</i> , <i>ego1Δ::kanMX</i>	This study	5B
KP41-9C	[KT1961] <i>gtr1Δ::natMX</i> , <i>gtr2Δ::natMX</i> , <i>ego1Δ::kanMX</i> , <i>ego2Δ::kanMX</i> , <i>ego3Δ::kanMX</i> ( <i>egocΔ</i> )	This study	6A-C
KP41-10C	[KT1961] <i>gtr1Δ::natMX</i> , <i>ego1Δ::kanMX</i> , <i>ego2Δ::kanMX</i> , <i>ego3Δ::kanMX</i>	This study	6D-E

**Table S2. Plasmids used in this study.**

<b>Plasmid</b>	<b>Genotype</b>	<b>Source</b>	<b>Figure</b>
pRS413	<i>CEN, HIS3</i>	[5]	1A, D-F, 2A-E, 5B-D
pRS414	<i>CEN, TRP1</i>	[5]	3E-F, 5B-D
pRS415	<i>CEN, LEU2</i>	[5]	1A-C, 2A-B, D-E, 3B, E-F, 5B-D
pRS416	<i>CEN, URA3</i>	[5]	1A-F, 2A, C, 3B, E-F, 5B-D
pJU1064	[pRS413] <i>SCH9<sup>T570A</sup>-HA<sub>5</sub></i>	[6]	1B-C, 5D
pJU1058	[pRS415] <i>SCH9<sup>T570A</sup>-HA<sub>5</sub></i>	[6]	1D-E
YCplac111	<i>CEN, LEU2</i>	[7]	
pNP2529	[YCplac111] <i>ADH1p-EGO2</i>	This study	1C
pNP2530	[YCplac111] <i>ADH1p-EGO4</i>	This study	1C
YCplac33	<i>CEN, URA3</i>	[7]	
pMB1394	[YCplac33] <i>Tet<sub>ON</sub>-GTR1<sup>Q65L</sup></i>	[1]	1D-E
pMB1395	[YCplac33] <i>Tet<sub>ON</sub>-GTR1<sup>S20L</sup></i>	[1]	1D-E
pPM1619	[YCplac33] <i>Tet<sub>ON</sub>-GTR2<sup>Q66L</sup></i>	This study	1D-E
pPM1620	[YCplac33] <i>Tet<sub>ON</sub>-GTR2<sup>S23L</sup></i>	This study	1D-E
pJU650	[pRS416] <i>GTR1</i>	[4]	1F, 2D-E
pJU653	[pRS416] <i>GTR1<sup>Q65L</sup></i>	[1]	1F
pJU652	[pRS416] <i>GTR1<sup>S20L</sup></i>	[1]	1F
pJU651	[pRS416] <i>GTR2</i>	[1]	1F
pJU655	[pRS416] <i>GTR2<sup>Q66L</sup></i>	[1]	1F
pJU654	[pRS416] <i>GTR2<sup>S23L</sup></i>	[1]	1F
pFLJ1973	[YCplac33] <i>EGO1-GST</i>	This study	2B
pNP2572	[pRS416] <i>ADH1p-EGO1-TAP</i>	This study	2B
pMP1639	[pRS415] <i>GTR1-GFP</i>	[1]	2C, 5B
pMP1642	[pRS415] <i>GTR2-GFP</i>	[1]	2C, 5B
pMP1640	[pRS415] <i>GTR1<sup>Q65L</sup>-GFP</i>	This study	2C
pMP1644	[pRS415] <i>GTR2<sup>S23L</sup>-GFP</i>	This study	2C
pNP2441	[pRS413] <i>EGO2-GFP</i>	This study	3E-F, 5B
pNP2442	[pRS413] <i>EGO4-GFP</i>	This study	3E-F, 5B
pMB1344	[YCplac33] <i>GTR1-TAP</i>	[1]	2D-E
pMB1371	[YCplac33] <i>GTR1<sup>S20L</sup>-TAP</i>	[1]	2E
pMB1372	[YCplac33] <i>GTR1<sup>Q65L</sup>-TAP</i>	[1]	2E
pMP2177	[pRS414] <i>GTR2-V5-6HIS</i>	[4]	2D-E
pAI-Alg5	2 $\mu$ <i>ADH1-HA-NUB1, TRP1</i>	Dualsystems	3A, G, 5A
pDL2-Alg5	2 $\mu$ <i>ADH1-HA-NUBG, TRP1</i>	Dualsystems	3A, G, 5A
pCabWT	<i>CEN, CYC1-CUB-LexA, LEU2</i>	Dualsystems	3A, G, 5A
pFLJ2393	[pCabWT] <i>CYC1-EGO2-CUB-LexA</i>	This study	3A, G, 5A
pFLJ2394	[pCabWT] <i>CYC1-EGO4-CUB-LexA</i>	This study	3A, G
pPR3-N	2 $\mu$ <i>CYC1-NUBG-HA, TRP1</i>	Dualsystems	3A, G
pNP1689	[pPR3-N] <i>CYC1-NUBG-HA-GTR1</i>	[1]	3A, G
pNP1690	[pPR3-N] <i>CYC1-NUBG-HA-GTR1<sup>S20L</sup></i>	[1]	3A, G
pNP1691	[pPR3-N] <i>CYC1-NUBG-HA-GTR1<sup>Q65L</sup></i>	[1]	3A, G
pNP1692	[pPR3-N] <i>CYC1-NUBG-HA-GTR2</i>	This study	3A, G
pNP1693	[pPR3-N] <i>CYC1-NUBG-HA-GTR2<sup>S23L</sup></i>	This study	3A, G
pNP1694	[pPR3-N] <i>CYC1-NUBG-HA-GTR2<sup>Q66L</sup></i>	[4]	3A, G
pNP1696	[pPR3-N] <i>CYC1-NUBG-HA-EGO1</i>	[1]	3A, G, 5A

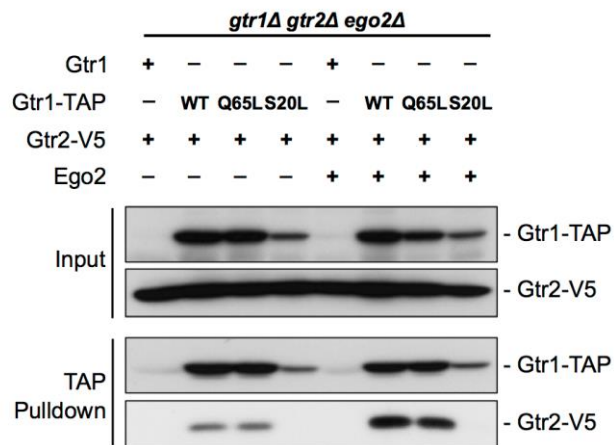
**Table S2. Plasmids used in this study - continued**

Plasmid	Genotype	Source	Figure
pMPG2221	[pPR3-N] <i>CYC1-NUBG-HA-EGO3</i>	[4]	3A, G, 5A
pKP2623	[pRS413] <i>EGO2-HA<sub>3</sub></i>	This study	3B
pKP2622	[pRS413] <i>EGO4-HA<sub>3</sub></i>	This study	3B
pET-15b	<i>PT7lac, HIS6, lacI, ApR</i>	Novagen	
pNP2564	[pET-15b] <i>HIS6-EGO1, EGO2, EGO3, EGO4</i>	This study	3C-D
pFLJ2220	[pCabWT] <i>CYC1-EGO3-CUB-LexA</i>	[4]	5A
pFLJ2734	[pPR3-N] <i>CYC1-NUBG-HA-EGO1<sup>Δ152-184</sup></i>	This study	5A
pFLJ2735	[pPR3-N] <i>CYC1-NUBG-HA-EGO1<sup>Δ169-184</sup></i>	This study	5A
pKP2736	[YCplac33] <i>EGO1-HA<sub>3</sub></i>	This study	5B-C
pKP2737	[YCplac33] <i>EGO1<sup>Δ152-184</sup>-HA<sub>3</sub></i>	This study	5B-C
pKP2738	[YCplac33] <i>EGO1<sup>Δ169-184</sup>-HA<sub>3</sub></i>	This study	5B-C
pKP2739	[YCplac33] <i>EGO1</i>	This study	5C-D
pKP2740	[YCplac33] <i>EGO1<sup>Δ152-184</sup></i>	This study	5C-D
pKP2741	[YCplac33] <i>EGO1<sup>Δ169-184</sup></i>	This study	5C-D
pKP2801	[YCplac33] <i>EGO1<sup>NT</sup>-vhhGFP4</i>	This study	6A-D
pNSIm-vhhGFP4	<i>pcDNA3-NSImb-vhhGFP4</i>	[8]	
pRH2776	[pPRS413] <i>VAC8-vhhGFP4</i>	This study	6E

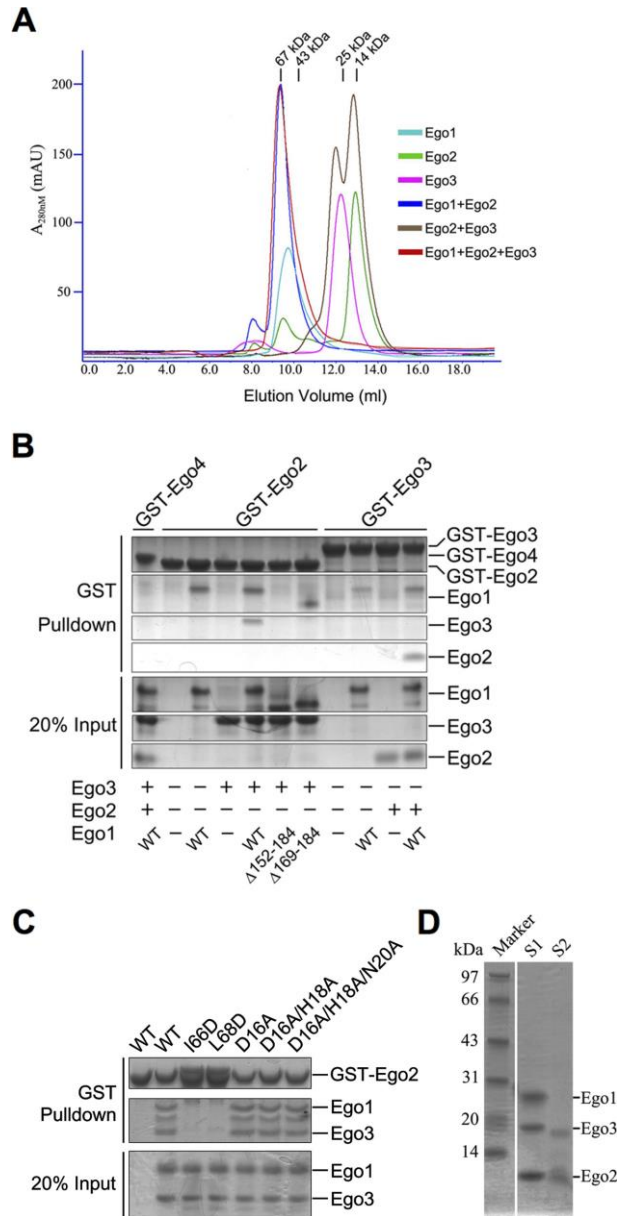
## References

- 1 Binda M, P éi-Gulli MP, Bonfils G *et al.* The Vam6 GEF controls TORC1 by activating the EGO complex. *Mol Cell* 2009; **35**:563-573.
- 2 Pedruzzi I, Dubouloz F, Cameroni E *et al.* TOR and PKA signaling pathways converge on the protein kinase Rim15 to control entry into G<sub>0</sub>. *Mol Cell* 2003; **12**:1607-1613.
- 3 Dubouloz F, Deloche O, Wanke V, Cameroni E, De Virgilio C. The TOR and EGO protein complexes orchestrate microautophagy in yeast. *Mol Cell* 2005; **19**:15-26.
- 4 Zhang T, P éi-Gulli MP, Yang H, De Virgilio C, Ding J. Ego3 functions as a homodimer to mediate the interaction between Gtr1-Gtr2 and Ego1 in the EGO complex to activate TORC1. *Structure* 2012; **20**:2151-2160.
- 5 Brachmann CB, Davies A, Cost GJ *et al.* Designer deletion strains derived from *Saccharomyces cerevisiae* S288C: a useful set of strains and plasmids for PCR-mediated gene disruption and other applications. *Yeast* 1998; **14**:115-132.
- 6 Urban J, Soulard A, Huber A *et al.* Sch9 is a major target of TORC1 in *Saccharomyces cerevisiae*. *Mol Cell* 2007; **26**:663-674.
- 7 Gietz RD, Sugino A. New yeast-*Escherichia coli* shuttle vectors constructed with *in vitro* mutagenized yeast genes lacking six-base pair restriction sites. *Gene* 1988; **74**:527-534.
- 8 Caussin E, Kanca O, Affolter M. Fluorescent fusion protein knockout mediated by anti-GFP nanobody. *Nat Struct Mol Biol* 2012; **19**:117-121.

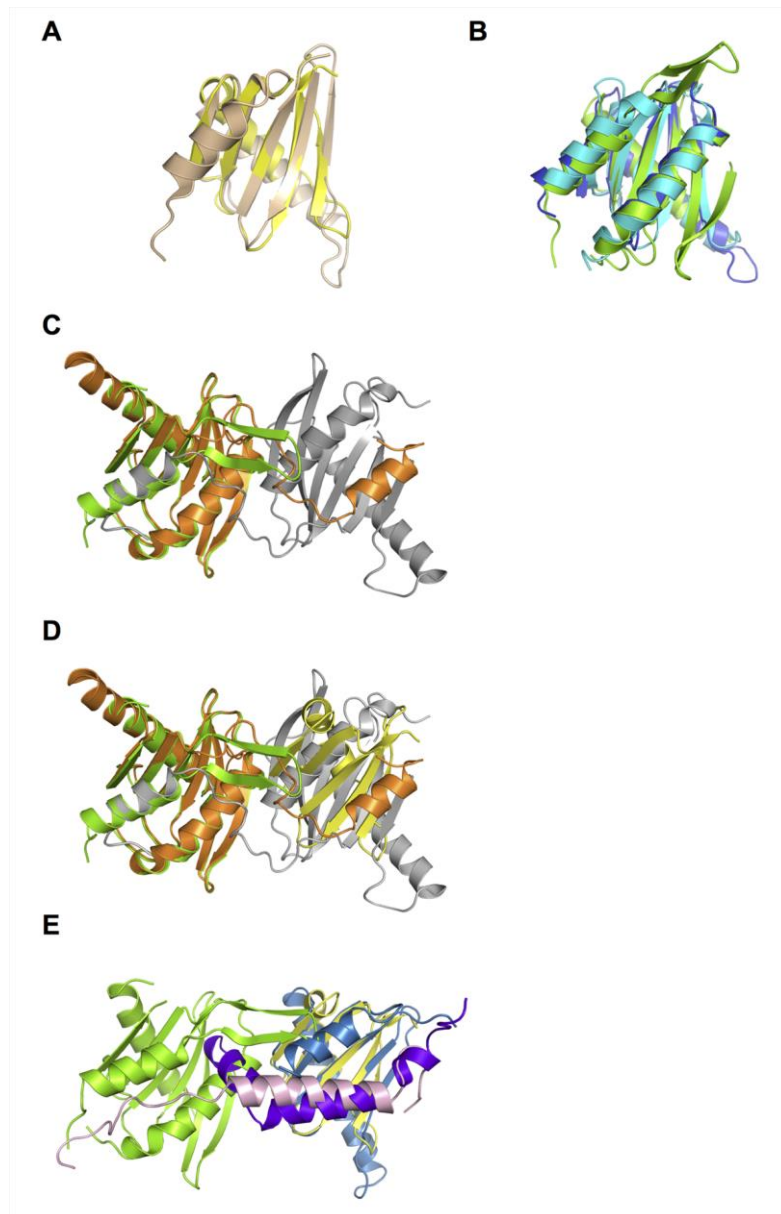
## Supplementary Figures



**Figure S1.** TAP pull-down experiments showing that the reduced interaction between Gtr1-TAP and Gtr2-V5 in an *ego2Δ* strain can be rescued by re-introduction of *EGO2* expressed from a plasmid. Lysates from cells expressing the indicated fusion proteins (input) and TAP pull-down fractions were analyzed by immunoblotting with anti-TAP and anti-V5 antibodies. Cells expressing untagged Gtr1 were used as a control.



**Figure S2.** *In vitro* functional analyses of the Ego1-Ego2-Ego3 ternary complex (EGO-TC). **(A)** Analysis of the oligomeric state of the Ego proteins in solution by size-exclusion chromatography. The Ego1-Ego2 complex was purified in a similar way as the Ego1-Ego2-Ego3 complex as described in the experimental procedures. The Ego1, Ego2, and Ego3 proteins were fused with a C-terminal His<sub>6</sub>-tag. Absorbance at 280 nm is plotted against the elution volume. Positions of the molecular weight standards are indicated. **(B)** *In vitro* GST-pull down binding assays of GST-fused Ego2, Ego3, or Ego4 with full-length or truncated Ego1 and/or Ego2 and Ego3. The results were visualized by Coomassie blue staining on SDS-PAGE. **(C)** *In vitro* GST-pull down binding assays of GST-fused wild-type or mutant Ego2 with Ego1 and Ego3. The results were visualized by Coomassie blue staining on SDS-PAGE. **(D)** SDS-PAGE of the purified EGO-TC used in the crystallization (S1) and the protein sample obtained from the crystallization solution (S2). The N-terminal region of Ego1 was degraded in the crystallization solution. The loaded proteins were visualized by Coomassie blue staining. Molecular weight markers are indicated on the left.



**Figure S3.** Structural comparisons of the Ego proteins and several representative Roadblock domain-containing proteins. **(A)** Superposition of Ego2 (colored in yellow) and LAMTOR5 (colored in wheat). **(B)** Superposition of the Ego3 monomer in the EGO-TC (colored in green), LAMTOR2 (PDB code 3CPT, colored in blue) and LAMTOR3 (PDB code 3CPT, colored in cyan). **(C)** Superposition of the Ego3 monomer in the EGO-TC (colored in green) and the Ego3 homodimer (PDB code 4FTX, one monomer colored in orange and the other in gray). **(D)** Superposition of Ego2 (colored in yellow) and Ego3 (colored in green) in the EGO-TC with the Ego3 homodimer showing that the position for Ego2 binding was occupied by the other monomer in the Ego3 homodimer. **(E)** Superposition of the EGO-TC with *Drosophila melanogaster* dynein intermediate chain and light chain complex (PDB code 3L9K). Ego1, Ego2 and Ego3 of the EGO-TC are colored in pink, yellow and green, respectively, and the dynein intermediate chain and light chain are colored in purple and skyblue, respectively.