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**Cite this article:** Warnock RCM, Parham JF, Joyce WG, Lyson TR, Donoghue PCJ. 2015 Calibration uncertainty in molecular dating analyses: there is no substitute for the prior evaluation of time priors. *Proc. R. Soc. B* **282**: 20141013.

http://dx.doi.org/10.1098/rspb.2014.1013

Received: 26 April 2014 Accepted: 23 October 2014

#### **Subject Areas:**

palaeontology, evolution, bioinformatics

#### **Keywords:**

molecular clock, calibration, cross-validation, fossil record, Bayesian, priors

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Electronic supplementary material is available at http://dx.doi.org/10.1098/rspb.2014.1013 or via http://rspb.royalsocietypublishing.org.

## THE ROYAL SOCIETY PUBLISHING

# Calibration uncertainty in molecular dating analyses: there is no substitute for the prior evaluation of time priors

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Calibration is the rate-determining step in every molecular clock analysis and, hence, considerable effort has been expended in the development of approaches to distinguish good from bad calibrations. These can be categorized into a priori evaluation of the intrinsic fossil evidence, and a posteriori evaluation of congruence through cross-validation. We contrasted these competing approaches and explored the impact of different interpretations of the fossil evidence upon Bayesian divergence time estimation. The results demonstrate that a posteriori approaches can lead to the selection of erroneous calibrations. Bayesian posterior estimates are also shown to be extremely sensitive to the probabilistic interpretation of temporal constraints. Furthermore, the effective time priors implemented within an analysis differ for individual calibrations when employed alone and in differing combination with others. This compromises the implicit assumption of all calibration consistency methods, that the impact of an individual calibration is the same when used alone or in unison with others. Thus, the most effective means of establishing the quality of fossil-based calibrations is through a priori evaluation of the intrinsic palaeontological, stratigraphic, geochronological and phylogenetic data. However, effort expended in establishing calibrations will not be rewarded unless they are implemented faithfully in divergence time analyses.

#### 1. Introduction

The molecular clock uniquely combines evidence from both molecular sequences and palaeontological and geological temporal constraints on sequence divergence, to establish evolutionary timescales. However, the precision of divergence time estimates is often so broad that they do not provide for effective tests of evolutionary hypotheses. It has been demonstrated that there is a modest limit on the gains in precision that can be obtained with increasing sequence data, beyond which increased precision can be obtained only by increasing the precision of fossil calibrations [1–3]. Unfortunately, identifying calibrations that are merely accurate is difficult enough. Two principal (but not necessarily mutually exclusive) approaches have emerged: (i) the *a priori* assessment of the empirical fossil anatomical, phylogenetic, stratigraphic and geochronological evidence, versus (ii) the *a posteriori* evaluation of the consistency of calibrations within a set.

A priori best practice requires that fossil calibrations comprise a conservative minimum constraint on a clade's age, minimizing phylogenetic uncertainty. In converting this into a calibration, the approach most widely adopted is to assign

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a non-uniform probability density (e.g. lognormal, exponential), fixed on the minimum constraint, that expresses a generalized view of the degree to which minima approximate divergence dates [3-12], invariably established without justification [13]. Alternatively, qualitatively justified 'soft maxima' have been established on palaeontological and geological grounds, based on the absence of evidence for a lineage antedating its oldest fossil record, qualified by the presence of taphonomic controls provided by sister lineages [6] (cf. [14-16]), and known gaps and facies biases in the rock record [15,17]. Effectively, the a priori establishment of minimum and maximum constraints based on fossil evidence removes them from equivocation. However, in practice, this approach is necessarily conservative in the identification of fossil taxa suitable for use in calibration, and in interpreting their age, such that calibrations established in this way are often a poor, or imprecise, approximation of divergence dates.

Alternatively, *a posteriori* methods have been developed to assess the relative quality of calibrations through the consistency with which each calibration, within a set, estimates the others when used in isolation. The underlying assumption is that calibrations should be consistent and inconsistent calibrations should be rejected. This approach has also been used to consider competing phylogenetic positions for critical fossils [18–23]. In attempting to address phylogenetic and stratigraphic uncertainty, Marshall [24] established, and Dornburg *et al.* [25] extended, a method for selecting the fossil calibration(s) among a set that provide the best approximation of the antiquity of the respective lineage(s). *A posteriori* methods keep fossil data at arm's length, assessing internal consistency or its veracity measured with respect to branch length.

We set out to evaluate the performance a posteriori versus a priori approaches for assessing calibration quality. Our analyses are based on the now classic empirical dataset that encompasses extant turtle phylogeny used to first demonstrate a number of a posteriori methods of evaluating calibration quality (e.g. [24-26]) and in debate about the importance of establishing calibration quality a priori [27-29]. We also employ a completely revised set of calibrations for this phylogeny, constructed following the principles of best practice [30]. These exemplify the impact of the a priori evaluation of fossil calibrations and, as such, they can be considered accurate, if not precise. Since debate over calibration quality has not considered seriously the impact of different approaches for establishing maximum constraints, we first explore the impact of different approaches to constraining node ages. We simulated the approach of assigning a non-uniform probability density to fossil-based minimum constraints, and contrast these results to those of analyses in which a uniform density is employed. We employed the cross-validation method of Near et al. [26,31] to measure consistency among calibrations based on minimum constraints, and adopted a novel crossvalidation approach considering the entire timespan between minimum and maximum constraints [32].

Crucially, our results demonstrate that: (i) *a posteriori* methods have led to the recurrent selection of erroneous constraints, and (ii) the effective time priors implemented in an analysis differ for individual calibrations when employed alone or in variable combinations with additional constraints—this means that estimates of calibration quality based on consistency do not provide a faithful indication of how a given calibration will impact the analysis in combination with

others. *A posteriori* approaches to assessing calibration quality cannot therefore substitute for the *a priori* evaluation of fossil evidence in establishing accurate constraints. However, the accuracy of any calibration may be compromised by the way in which the calibrations are effectively implemented in the Bayesian estimation of divergence times.

#### 2. Material and methods

## (a) Modelling non-uniform and uniform priors using fossils

Bayesian molecular clock analyses were performed using the approximate likelihood approach implemented in MCMCTREE [2,3,33], because it is computationally efficient [34] and uses a more predictable procedure in the construction of the joint time prior, in comparison to BEAST [3,35]. However, we reproduced our analyses in BEAST 1.6.1 [8,36] using uniform priors, to explore differences in the construction of the joint time prior. Fossil-based minimum and maximum constraints were established for this dataset following best practice [27,30].

Non-uniform priors express approximations of divergence timing relative to a minimum constraint, however, such calibrations are rarely evidence-based [13]. Although there are objective approaches to informing non-uniform prior densities (e.g. [37]), the turtle fossil record has not yet been documented in a manner that would allow time priors to be established in this way. By contrast, uniform priors allow the user to accommodate a view that nothing is known about the time of divergence relative to the constraints. We present this as a null hypothesis that given the absence of evidence to the contrary, there is an equal prior probability of the timing of the divergence event, per unit time, spanning the minimum and maximum bounds; this is not an uninformative prior. We implemented hard minima, such that the probability that a divergence time postdates the minimum constraint approximates zero. Where applicable, we implemented soft maxima, allowing 2.5% of the probability to exceed maximum constraints [3].

We explored the use of non-uniform calibration priors, permuting the truncated Cauchy distribution, to reflect variable non-uniform probabilities of divergence timing relative to the minimum constraints [12]. A maximum bound must be specified at the root of the tree and so we retained a uniform distribution at the root, corresponding to the fossil-based calibration available for the age of crown turtles [27]. All molecular clock analyses were performed without sequence data to examine the effective priors, compared to the specified priors.

#### (b) A posteriori evaluation of calibration quality

We implemented the original cross-validation method described in [26] to compare the consistency between our calibrations. Consistency was assessed: (i) relative to minimum constraints only, and (ii) relative to minimum and maximum constraints [32]. For each individual calibration, during each round of cross-validation, the tree was calibrated using a single uniform calibration prior, with a hard minimum and soft maximum constraint based on fossil evidence [27]. A soft maximum age constraint was applied at the root using the fossil-based maximum for the age of this node.

Finally, we compared three *a posteriori* approaches to evaluating calibrations [24–26] to the *a priori* evaluation of fossil evidence. To assess the quality of calibrations selected using *a posteriori* methods, we contrasted the selection of calibrations based on assessments of calibration quality among the Testudines dataset used in the seminal application of the cross-validation method (and its derivatives) [24–26]. This was compared to the *a priori* assessment of calibration

quality based on the intrinsic palaeontological evidence used to establish these constraints, which formed the basis of an independent study [27]. Further details of all materials and methods are provided in the electronic supplementary material.

#### 3. Results

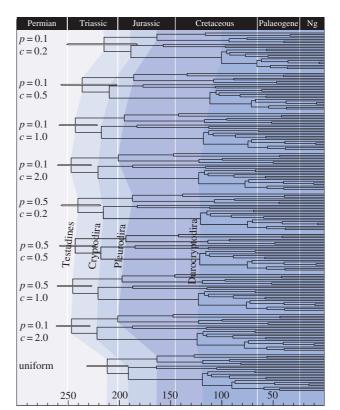
## (a) The impact of non-uniform and uniform calibration priors

Increasing the uncertainty in the timing of divergence relative to fossil minima, based on a non-uniform prior, led to an increase in both prior and posterior age estimates across all nodes (figure 1; electronic supplementary material, table S1). Increasing uncertainty also generated more diffuse credibility intervals, except at the root (node 1: Testudines). The mean root age increased from 215 to 246 Ma, but the 95% intervals were attenuated with increasing uncertainty at the internal nodes. The results appear to be influenced strongly by the limit on the root: the posterior estimates appear to become only as ancient as the soft maximum at the base of the tree will allow (figure 1); note that MCMCTREE requires the user to specify a soft maximum constraint at the root of the tree. In BEAST, if the upper (soft) constraint is not specified, then this limit will be specified indirectly by other parameters. The results obtained using uniform priors are different to those obtained using non-uniform priors, including the proposed temporal sequence of non-hierarchically dependent divergence events (figure 1; electronic supplementary material, table S1). For example, the posterior confidence intervals obtained using uniform priors suggest that of the two major groups of turtles, Cryptodira (node 8) originated before Pleurodira (node 2). By contrast, there is substantial overlap between the estimates obtained using nonuniform priors for the age of these nodes and, consequently, a coincident time of origin cannot be rejected.

#### (b) A posteriori evaluation of calibration quality

The results of the cross-validation analysis showed that the most consistent calibrations based on fossil minima are the most inconsistent calibrations based on minima and maxima; these are the nodes that tend to produce the youngest average estimates (figure 2a-d). Conversely, the most inconsistent calibrations based on fossil minima tend to produce the oldest average estimates; these nodes are the most frequently underestimated and tend to overestimate the age at other nodes. The results of the cross-validation analysis, which considered the minimum constraints only, are presented in figure 2a,c. The  $\bar{D}_x$  and the SS values for each calibration reflect the average differences between the mean molecular estimates and the minimum constraints of all other nodes. All  $D_x$  values are positive and range from 14.29 to 46.24 Myr, indicating that most posterior estimates of divergence times do not postdate fossil minima.

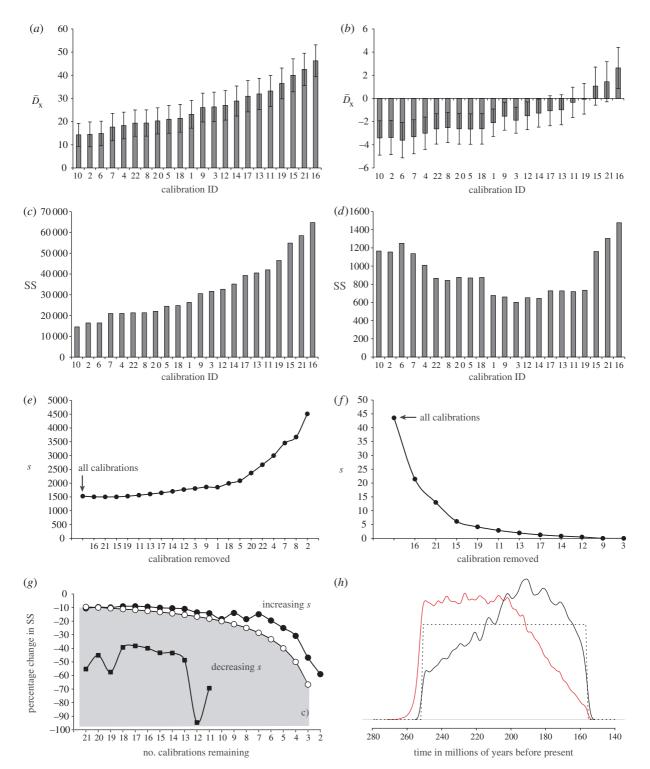
The results of the cross-validation analysis, in which both minimum and maximum age constraints are considered, are presented in figure  $2b_rd$ . The  $\bar{D}_x$  and the sum of squared differences (SS) values for each calibration reflect the average differences between the mean estimates and the minimum or maximum constraints of all other nodes. Most  $\bar{D}_x$  values are slightly negative and range from 3.60 to 2.63 Myr. SS values are two orders of magnitude smaller than those based on minimum constraints alone (figure  $2c_rd$ ). This reflects the use of conservative maximum



**Figure 1.** The posterior mean estimates obtained when the truncated Cauchy distribution was used to approximate the time of divergence relative to fossil-based minima in MCMCTREE. Results are shown for two values of the location parameter p (0.1, 0.5) and four values of the scale parameter c (0.1, 0.5, 1, 2) and results are contrasted to those obtained when a uniform distribution is used to constrain node ages between the fossil-based minima and maxima. The branching order (and corresponding node labels) is the same as those shown in figure 3. The branch lengths represent the posterior means of the node ages. Four nodes are connected across the analyses to facilitate comparison. The 95% higher posterior density is indicated at the root of the tree. Ages are presented in millions of years before present. (Online version in colour.)

age constraints and the informative maximum limit placed at the root (251.4 Ma), which precludes estimates from becoming unjustifiably ancient. Few molecular estimates are likely to exceed their respective maxima. Regardless of the direction or magnitude of inconsistency, cross-validation analyses demonstrate that independent calibrations produce appreciably different divergence estimates. However, the removal of any calibrations did not significantly reduce the variance among calibrations and molecular estimates (figure 2e-g).

A comparison between three a posteriori approaches to assessing calibration quality shows that different numerical methods of defining calibration quality identify variable suites of constraints as either accurate or inaccurate (table 1). The calibrations selected for rejection using alternative a posteriori methods are neither supported by each other, nor by the available fossil evidence on which the constraints are based. The a priori evaluation of palaeontological evidence led to a ubiquitous, and in some cases substantial (up to 89%), revision of the minimum (and maximum) age constraints for all nodes. Although a posteriori approaches correctly discriminate some of the constraints that were necessarily revised based on fossil evidence, they also eliminated a number of accurate constraints, while retaining a number of inaccurate constraints (table 1). This means that a posteriori methods can lead to the selection of calibrations that are not supported by available fossil evidence.



**Figure 2.** (a-d) Histograms showing the average difference  $(\bar{D}_x)$  (a,b) and the sum of squared differences (SS) (c,d) between molecular- and fossil-based estimates obtained during each round of cross-validation. The values obtained when fossil-based minima are used to calculate  $\bar{D}_x$  (a) and SS (c), and the values obtained when both minima and maxima are considered in the estimation of  $\bar{D}_x$  (b) and SS (d). The error bars shown in (a,b) reflect the standard error of the mean. (e-f) Plots showing the impact of sequentially removing calibrations on the average squared deviation(s) between molecular—and fossil—estimates considering minima only (e) and both minima and maxima (f). Sequential removal of the most inconsistent calibrations revealed a consistent, albeit statistically non-significant increase in s when only the fossil-based minima are used in the estimation of SS (f). By contrast, there is a steady albeit statistically non-significant decline in s when minimum and maximum temporal constraints are used to calculate SS (e). The value of s will increase if the rate of change of the denominator, n(n-1) in the equation used to calculate s, exceeds the rate of change of the numerator s SS, where s represents the total number of calibrations [38]. (s) Illustrates the percentage change in s SS with the sequential removal of calibrations, estimated using minima (black circles) or both minima and maxima (black squares). This is contrasted to the rate of change of s SS when fossil minima are used in the calculation, hence we observe an increase in s. The converse is true when both minimum and maximum constraints are incorporated into the estimation of SS. Changes in s fluctuate markedly with the removal of calibrations as the value of s approaches zero; SS s 0 with the inclusion of the final 10 calibrations. (s) The effect of truncation in the establishment of the joint time prior at the root (node 1, figure 3) in BEAST (black) and MCMCTREE (red).

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250	and general	10331						(manual)		
					Near <i>et al.</i> [26]	Marshall [24]	Dornburg <i>et al.</i> [25]	Joyce <i>et al.</i> [27]		
1 (1)	Testudines	Proterochersis robusta	210 <sub>F/F/M</sub>	1	consistent	1	omitted <i>a priori</i>	phyl. misplaced	155.6	251.4
2 (3)	Pleurodira	Araripemys barretoi	110 <sub>F</sub> <sub>W/F/M</sub>	134.4	consistent	l	consistent	accurate minima	111	165.2
3 (4)	Pelomedusoides	Cearachelys placidoi	110 <sub>F-M/F/M</sub>	134.4	consistent	1	consistent	accurate minima	92.8	149.5
4 (14)	Pelomedusidae	Pelusios rusingae	18ғ.ғ.м	22.0	consistent	1	inconsistent	inaccurately dated	5.3	149.5
5 (11)	Chelidae	Yaminuechelys gasparinii	71ғ.ғ.м	86.7	inconsistent		consistent	accurate minima	65.2	149.5
6 (15)	Chelodininae	Chelodina sp. and Elseya sp.	15ғ.ғ.м	18.3	inconsistent	1	inconsistent	accurate minima	11.6	149.5
7 (16)	Chelus – Phrynops	Chelus sp.	11.6 <sub>F/F/М</sub>	14.2	inconsistent	1	inconsistent	accurate minima	13.4	149.5
8 (2)	Cryptodira	Sandownia harrisi	110 <sub>F-M/F/M</sub>	1	consistent	l	omitted <i>a priori</i>	phyl. misplaced	124	200.2
10 (6)	Trionychidae	Aspideretes maortuensis	100 <sub>F-M/F/M</sub>	122.1	consistent	selected for	consistent	phyl. untested	17.3	149.5
						calibration				
13 (5)	Chelonioidea	Santanachelys gaffneyi	110 <sub>F/F/M</sub>	134.3	inconsistent	inconsistent	consistent/ inconsistent	phyl. misplaced	48.4	149.5
15 (7)	Kinosternoidea	Hoplochelys sp.	65 <sub>F/F/M</sub>	79.4	consistent	l	consistent	accurate minima, alternative	70	149.5
								fossils now available		
16 (10)	Kinosternidae	Baltemys sp.	50 <sub>F/F/M</sub>	61.1	consistent	1	consistent	accurate minima	52.8	149.5
19 (12)	Emydidae	Chrysemys antiqua	34 <sub>FF.M</sub>	41.5	consistent	l	consistent	accurate minima	32	100.5
20 (17)	Graptemys – Trachemys	Trachemys inflata	5 <sub>F/F/M</sub>	6.1	inconsistent	1	inconsistent	phyl. untested	3	34
21 (8)	Testuguria	Hadrianus majusculus	52 <sub>F/F/M</sub>	63.5	consistent	l	consistent	accurate minima	50.3	100.5
22 (9)	Heosemys – Mauremys	"Ocadia" crassa	50 <sub>F/F/M</sub>	61.1	inconsistent	1	consistent	phyl. untested	5.3	65.8
— (13)	I	Lindholmemydidae	90 <sub>F/F/M</sub>	109.9	consistent	l	consistent	phyl. untested	ı	ı
9 (18)	Trionychia	1	l	1	1	l	l	I	124	177.6
11 (19)	Durocryptodira	l	1	1	l	l	l	I	9.88	149.5
12 (20)	Americhelydia	1	I	1	l	l	l	I	70	149.5
14 (21)	Chelydroidea	1	l	1	1	1	l	I	70	149.5
17 (—)	Testudinoidea	I	I	1	1	1	I	I	50.3	149.5
18 (—)	Emydidae – <i>Platysternon</i>	1	1	1	1	1	l	ı	32	100.5

<sup>\*</sup>Node ID corresponds to those shown in figure 3. Numbers in brackets refer to the corresponding ID in Near et al. [26].

<sup>&</sup>lt;sup>b</sup>Fossil specimens used to assess calibration quality *a posteriori*.

Minimum (M) or fixed (F) age constraint used to assess calibration quality a posteriori in Near et al. [26], Marshall [24] and Dornburg et al. [25].

<sup>&</sup>lt;sup>d</sup>Soft maximum age constraint used to assess calibration quality a posteriori in Dornburg et al. [25]. Maxima established using Marshall [24]; eqn. (11).

<sup>&</sup>lt;sup>e</sup>A priori assessment of minimum fossil constraints only. Assessments indicate whether fossil specimens have been phylogenetically (phy.) misplaced or untested, inaccurately dated or provide appropriate minimum constraints.

#### (c) Effective versus user-specified calibration priors

Analysis without sequence data demonstrates that the specified calibration priors are not faithfully implemented in the joint estimation of divergence times. This occurs in association with both non-uniform and uniform probability distributions. In the case of non-uniform time priors, increasing the uncertainty associated with the calibrations produced more imprecise specified time priors, but the effective marginal densities still do not match the specified time prior. This change, between the specified and the effective priors, is particularly significant at the root where the uniform specified prior is transformed into a distinctly non-uniform effective prior. This had a large impact on the prior credibility intervals for the root. The upper (maximum) 95% prior interval at the root always exceeded the specified soft maximum (251.5 Ma) by up to 5 Myr, but the lower (minimum) 95% prior interval became older (up to 55 Myr) than the specified minimum (155.6 Ma) (electronic supplementary material, table S1).

We compared the specified uniform age priors in MCMCTREE to: (i) the effective priors for each node during independent rounds of cross-validation, and (ii) the effective priors observed at each node when all calibrations are combined in a single analysis (figure 3). Even during cross-validation, when a single uniform calibration is employed, the marginal calibration densities do not always match the specified uniform densities. The largest discrepancies between the specified and effective priors are associated with nodes that have the broadest calibration spans and, hence, overlap most with the specified constraints on ancestral nodes (e.g. node 4: Pelomedusidae).

#### (d) Comparison between BEAST and MCMCTREE

BEAST and MCMCTREE derived similar prior and posterior estimates of divergence times (figure 4), though MCMCTREE produced slightly older mean estimates and wider credibility intervals. The largest difference was observed in estimates of root age. When all calibrations are combined in a single analysis, the effective prior densities obtained using BEAST and MCMCTREE are similar and exhibit the same direction of skew and modality—with the main exception of the root (node 1: Testudines; figure 2h). The effective root age prior implemented in MCMCTREE indicates that an older time of divergence is more likely. Conversely, in BEAST, the effective root age prior suggests that younger divergence times are more likely.

#### 4. Discussion

## (a) The impact of non-uniform and uniform calibration priors

In the absence of fossil-based maximum constraints, the specified uncertainty associated with constraints may be made subjectively large (or small). Estimates of divergence times are evidently sensitive to the parameters used to specify the prior density. In figure 1, we contrast the posteriors obtained using non-uniform priors, to those obtained using uniform fossil-based minima and soft maxima where each node is constrained using a uniform prior. The comparison shows that these analyses yield very different results, including

differences in the relative temporal sequence, not merely absolute timing, of speciation events.

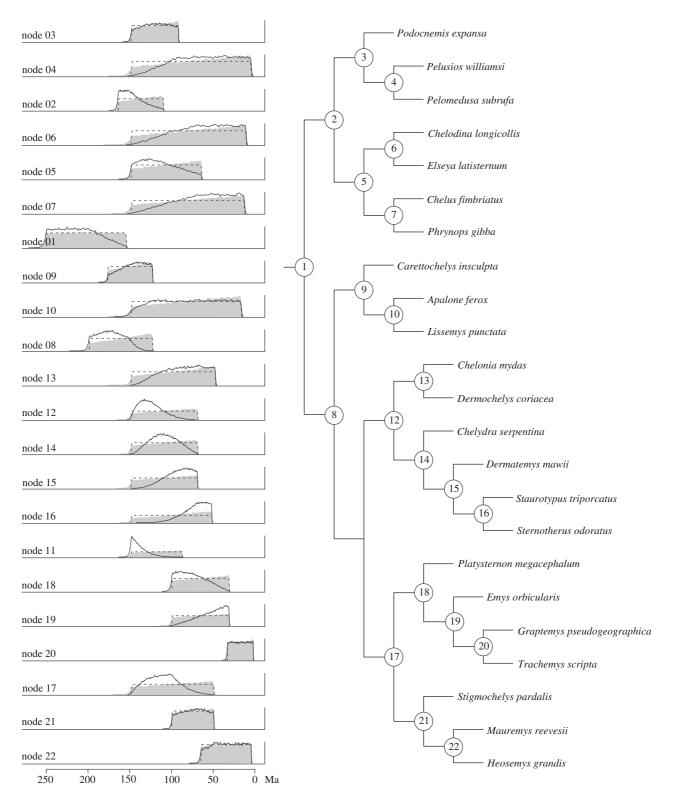
These results corroborate the findings of previous studies [12,13,32], demonstrating that subtle changes in the parameters that describe the priors have an adverse impact upon the posterior divergence time estimates. Since there is frequently no material basis for selecting among the parameters, or the distributions that they control, in the majority of studies the time priors are, quite literally, unjustified [13]. Note that if no alternate evidence exists, relying on the default settings for the calibration priors in BEAST or MCMCTREE is equivalent to accepting them and their implicit assumptions about the time of divergence relative to the constraints. For instance, if a soft maximum is not defined explicitly, this constraint will be specified indirectly by other parameters. Alternative approaches to informing calibration priors-for example, those based on stratigraphic occurrence data (e.g. [37,39]) represent an exciting area of development. However, these methods require large, well-curated fossil databases that are rarely available and, therefore, implemented.

## (b) The impact of specified and effective calibration priors

In all contemporary Bayesian molecular clock programs, the initial specified calibration priors will not be the same as the effective calibration priors actually implemented in the estimation of divergence times [12,13,30,32,35]. This is because the specified calibrations are truncated in the construction of the joint prior on times, to satisfy the expectation that ancestral nodes are older than descendent nodes [3,40]. Truncation is particularly obvious, given multiple overlapping constraints (e.g. [13]). However, even if there is no temporal or topological overlap between a pair of calibrations, their interaction with the tree prior can still result in differences in the effective priors (figure 3).

BEAST and MCMCTREE differ in the way they construct the joint prior on times. Effectively, the same palaeontological constraints are implemented as different time priors by these two principal software packages. In BEAST, the specified and effective priors can be very different, even when only a single calibration is employed. This is because BEAST uses a multiplicative construction, by multiplying the calibration densities by the tree prior, which is necessary for the estimation of topology [35]. This can sometimes cause the effective calibration priors to violate the palaeontological constraints, and it is difficult to predict the impact of including multiple constraints [35].

Differences between the models that underlie BEAST and MCMCTREE manifest themselves in both the prior and posterior estimates of divergence times. In particular, we have demonstrated that the same fossil constraints will lead to different effective time priors. The largest difference between BEAST and MCMCTREE posterior estimates was observed at the root and is probably a direct consequence of differences in modality observed in the specification of the root age prior (figure 2h). Since the posteriors are sensitive to different time priors, as evidenced by the impact of variable non-uniform and uniform priors, this has material consequences for posterior molecular clock estimates. It is clear is that the effort expended in establishing accurate palaeontological time priors will not be repaid unless they are reflected in the effective time priors. The specified priors should be

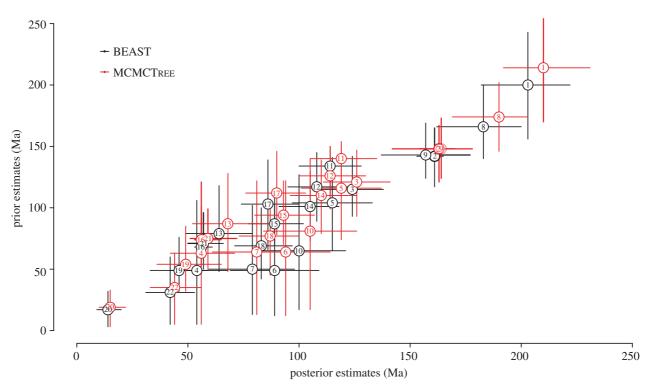


**Figure 3.** Plots contrasting the user-specified uniform calibration priors (dashed lines), firstly, with the effective marginal priors observed when each node is used for calibration during independent rounds of cross-validation (grey plots) and secondly, with the effective priors observed when all calibrations are combined in the final analysis in MCMCTREE (black lines). This diagram illustrates how the interaction between different calibrations in the joint prior can result in effective priors that deviate substantially from the initial user-specified distributions.

permuted experimentally until the primary palaeontological constraints are reflected in the effective time prior.

# (c) A posteriori versus a priori approaches to assessing calibration quality

Time priors have a substantial impact upon the outcome of divergence time analyses, and so it is necessary to discriminate between 'good' and 'bad' calibrations. Hence, there has been a great deal of effort expended in establishing criteria on which fossil calibrations should be based [5–7,13,30,32,41,42], and in developing methodological approaches to discriminating misleading fossil calibrations [18–26,31,38,43,44]. The *a posteriori* original cross-validation approach [26,31] and its subsequent developments [22,24,25,38] emphasize calibration consistency as the most desirable quality in a set of calibrations. The underlying assumptions of the cross-validation approach to assessing calibration quality have been criticized previously



**Figure 4.** Plot contrasting the prior and posterior estimates (mean and 95% credibility intervals) obtained using BEAST (black) and MCMCTREE (red). Node labels correspond to those shown in figure 3. (Online version in colour.)

[5,24]. The results of our analyses identify two additional and ultimately fatal problems with the cross-validation approach: (i) cross-validation methods demonstrably result in the selection of calibrations that are not supported by the available fossil evidence (table 1), and (ii) this approach is compromised within the Bayesian framework because the effective priors for a given calibration vary depending on the presence or the absence of other constraints (figure 3). This final point is particularly problematic since it demonstrates violation of the basic implicit assumption of cross-validation methods, that individual calibrations perform in the same manner regardless of whether they are employed individually or in combination with other calibrations.

Our comparison of *a posteriori* assessments of calibration quality to the *a priori* evaluation of calibration quality based on the intrinsic fossil evidence (table 1) demonstrates that cross-validation methods do not identify accurate calibrations consistently—that is, calibrations supported on the basis of independent (palaeontological, phylogenetic and geological) evidence. In addition, the evaluation of the available fossil evidence *a priori* using best practices [30] led to a substantial revision of the age constraints for many nodes.

The advantage of implementing the cross-validation approach within the Bayesian framework is that it can account for the expected probability that the age of a node may be considerably older than its first appearance in the fossil record (or any specified minimum age constraint). None of the revised calibrations [27] were identified as statistically inconsistent through the cross-validation methods [26,32] implemented in this study. This may reflect the fact that we considered the mean estimates relative to minima, or minimum—maximum divergence time priors. It is much easier for divergence time estimates to be compatible with broad constraints than with precise node ages. If consistency is a desirable quality in a suite of calibrations, it could be argued that the penalty for achieving this quality is a loss

of precision over the age of component nodes since the calibrations are ultimately more accurate but less precise. Though expert evaluation of palaeontological evidence may be best practice, perhaps a less conservative approach to evaluating fossil evidence might result in more precise calibration constraints. Hence, *a posteriori* approaches, including the cross-validation family of methods, may be an appealing alternative to wrestling with the complexities of deriving a temporal calibration from fossil, phylogenetic, stratigraphic and geochronological data. This appeal is demonstrated by the continued development (e.g. [18,20,25,32]) and application (e.g. [45–47]) of *a posteriori* methods.

Regardless, our analyses highlight the fact that the effective calibrations employed in divergence time estimation invariably differ from those specified by the user (figure 3). Consequently, different combinations of calibrations, in combination with the tree prior, will produce different joint time prior constructs—this occurs regardless of the (non-uniform or uniform) prior probability densities employed, or the approach used (directly or indirectly) to specify the (soft) maximum constraints. Thus, the manner in which a given calibration is implemented in the estimation of divergence times is not equivalent if it is employed alone or in combination with others. Furthermore, because different calibration priors have a material impact on the posteriors, consistency among either the effective priors or posteriors is not a reliable means of evaluating the relative accuracy of calibrations.

Cross-validation methods share the same implicit expectation that the influence of a single calibration on a molecular clock analysis is the same regardless of whether it is employed alone or in combination with a suite of other calibrations. Our results demonstrate that this expectation is not met since the effective time prior for any one node is not the same as the user-specified calibration, and the effective time prior differs depending upon its precise temporal and topological relationship to other calibrations. This observation calls into question

the entire approach of the cross-validation family of methods for evaluating calibration through consistency, regardless of whether consistency is perceived to be an appropriate quality of a set of calibrations. Thus, cross-validation approaches to assessing the quality of calibrations based on consistency cannot be considered a reliable means of establishing accuracy, not merely because they are biologically questionable [48], but because they are flawed, both logically and methodologically.

Evaluating calibrations a priori places emphasis on palaeontological accuracy. At the very least, fossil minima should postdate divergence events and fossil maxima predate divergence events. Our results show that there should be no alternative to the careful evaluation of fossil evidence, in terms of comparative anatomy, phylogenetic affinity, stratigraphic occurrence and its geochronological interpretation. There can be no justification for using calibrations that are contradicted by this independent body of evidence. However, we also show that the best efforts of field palaeontologists, comparative anatomists, phylogeneticists, biostratigraphers and geochronologists may be of moot significance if carefully researched calibrations are not implemented (rather than merely specified) in molecular clock analyses. At the very least, it should be a basic requirement of every molecular clock analysis that the effective time priors are evaluated in comparison to the specified time priors by first running the analysis without sequence data [13,35]. Ultimately, it is important only that the effective time priors reflect accurately the palaeontological constraints on divergence time estimation.

#### 5. Conclusion

Bayesian posterior estimates of divergence times are extremely sensitive to the time priors. We have demonstrated that slight changes in the specification of the prior probabilities have an adverse impact on posterior time estimates. In addition, we have shown that a posteriori approaches of assessing calibration quality can be used to explore qualitatively the relationship between minimum and maximum constraints and the putative time of divergence, but do not provide justification for the removal of any calibrations. However, we have also demonstrated that a posteriori methods which rely on crossvalidation are incoherent since they rely on the implicit assumption that the performance of each calibration is the same regardless of whether it is employed alone or in combination with others. This assumption is violated within the Bayesian framework because the effective calibrations employed in the joint estimation of divergence times are never the same as the user-specified calibrations when more than one calibration is employed. The effective time priors

always depend on the temporal and topological relationship among all calibrations included in the analysis. Although *a priori* justification generates calibrations that are based on all available evidence, which are consequently superior in terms of accuracy, they are not immune to the effects of establishing the joint time prior. Every molecular clock study should consider carefully the disparity between the specified and effective priors. This phenomenon has broad implications for any study that relies on the accurate estimation of evolutionary rates and times. Our results also underscore the need to consider simultaneously the multifaceted issues associated with calibration, such as the nature of the diverse data on which the calibrations are based and the ability of existing molecular clock methods to effectively represent these constraints.

Finally, best practice a priori protocols for establishing calibrations should not remain static. Recent methodological developments in approaches to calibration require additional types of palaeontological data, such as tip calibration using fossils as terminal taxa [49], or probabilistic approaches to constraining divergence times based on the distribution of stratigraphic occurrences [37,39]. These methods hold great promise for the development of increasingly accurate and precise evolutionary timescales for groups with a good fossil records and maybe even for entirely extinct lineages (e.g. [50]). However, for lineages with little or no fossil record-those groups for which the molecular clock was established—these novel calibration methods cannot be applied. Consequently, node-based calibrations will continue to play an important role in molecular dating. As we have demonstrated, establishing accurate constraints should not rely on a posteriori methods, and so node-based calibrations established using a priori methods will remain especially significant for groups for whom the molecular clock is the only means of establishing a reliable timescale.

Data accessibility. Data available from the Dryad Digital Repository: http://doi.org/10.5061/dryad.j8776.

Acknowledgements. We thank Jun Inoue, Mario dos Reis and Ziheng Yang for their advice on molecular clock methodology. We are also grateful to Callum Wright for assisting with the use of the Advanced Computing Research Centre facilities, University of Bristol, using which the majority of the work presented in this study was conducted.

Funding statement. This study was supported by NERC studentship NE/I528250/1 to R.C.M.W. and P.C.J.D., NERC grant no. NE/G009600/1 to P.C.J.D. and BBSRC grant nos. BB/G006660/1 and BB/J00538X/1 to Ziheng Yang and P.C.J.D. This work also benefitted from a working group on fossil calibrations (Synthesizing and databasing fossil calibrations: divergence dating and beyond) at the National Evolutionary Synthesis Center (NESCent) (National Science Foundation no. EF-0905606), and an NESCent studentship to R.C.M.W.

#### References

- dos Reis M, Yang ZH. 2013 The unbearable uncertainty of Bayesian divergence time estimation. *J. Syst. Evol.* 51, 30–43. (doi:10.1111/J.1759-6831.2012.00236.X)
- Rannala B, Yang ZH. 2007 Inferring speciation times under an episodic molecular clock. *Syst. Biol.* 56, 453 – 466. (doi:10.1080/10635150701420643)
- Yang ZH, Rannala B. 2006 Bayesian estimation of species divergence times under a molecular clock using multiple fossil calibrations with soft bounds.
- Mol. Biol. Evol. **23**, 212–226. (doi:10.1093/Molbev/Msj024)
- Barnett R, Barnes I, Phillips MJ, Martin LD, Harington CR, Leonard JA, Cooper A. 2005 Evolution of the extinct Sabretooths and the American cheetah-like cat. *Curr. Biol.* 15, R589 – R590. (doi:10.1016/J.Cub.2005.07.052)
- Benton MJ, Donoghue PCJ, Asher RJ. 2009 Calibrating and constraining molecular clocks. In *The timetree of*
- *life* (eds SB Hedges, S Kumar), pp. 35–86. Oxford, UK: Oxford University Press.
- Benton MJ, Donoghue PCJ. 2007 Paleontological evidence to date the tree of life. *Mol. Biol. Evol.* 24, 26–53. (doi:10.1093/Molbev/Msl150)
- Donoghue PCJ, Benton MJ. 2007 Rocks and clocks: calibrating the Tree of Life using fossils and molecules. *Trends Ecol. Evol.* 22, 424–431. (doi:10. 1016/J.Tree.2007.05.005)

- Drummond AJ, Ho SYW, Phillips MJ, Rambaut A. 2006 Relaxed phylogenetics and dating with confidence. *Plos Biol.* 4, 699–710. (doi:10.1371/ Journal.Pbio.0040088)
- Hedges SB, Kumar S. 2004 Precision of molecular time estimates. *Trends Genet.* 20, 242–247. (doi:10.1016/J.Tig.2004. 03.004)
- Ho SYW. 2007 Calibrating molecular estimates of substitution rates and divergence times in birds.
   J. Avian Biol. 38, 409–414. (doi:10.1111/J.2007. 0908-8857.04168.X)
- Ho SYW, Phillips MJ. 2009 Accounting for calibration uncertainty in phylogenetic estimation of evolutionary divergence times. *Syst. Biol.* 58, 367–380. (doi:10.1093/Sysbio/Syp035)
- Inoue J, Donoghue PCJ, Yang ZH. 2010 The impact of the representation of fossil calibrations on Bayesian estimation of species divergence times. Syst. Biol. 59, 74–89. (doi:10.1093/Sysbio/ Syp096)
- Warnock RCM, Yang ZH, Donoghue PCJ. 2012 Exploring uncertainty in the calibration of the molecular clock. *Biol. Lett.* 8, 156–159. (doi:10. 1098/Rsbl.2011.0710)
- Behrensmeyer AK, Kidwell SM, Gastaldo RA. 2000 Taphonomy and paleobiology. *Paleobiology* 26, 103 – 147. (doi:10.1666/0094-8373(2000)26[103:Tap]2.0.Co;2)
- 15. Holland SM. 1995 The stratigraphic distribution of fossils. *Paleobiology* **21**, 92–109.
- Holland SM. 2000 The quality of the fossil record: a sequence stratigraphic perspective. *Paleobiology* 26, 148–168. (doi:10.1666/0094-8373(2000)26 [148:Tqotfr]2.0.Co;2)
- Smith AB, McGowan AJ. 2007 The shape of the Phanerozoic marine palaeodiversity curve: how much can be predicted from the sedimentary rock record of Western Europe? *Palaeontology* 50, 765-774. (doi:10.1111/J.1475-4983.2007.00693.X)
- Andújar C, Soria-Carrasco V, Serrano J, Gómez-Zurita J. 2014 Congruence test of molecular clock calibration hypotheses based on Bayes factor comparisons. *Methods Ecol. Evol.* 5, 226–242. (doi:10.1111/2041-210X.12151)
- Lee MSY, Oliver PM, Hutchinson MN. 2009
   Phylogenetic uncertainty and molecular clock calibrations: a case study of legless lizards (Pygopodidae, Gekkota). *Mol. Phylogenet. Evol.* 
   50, 661–666. (doi:10.1016/J.Ympev.2008.11.024)
- Lee MSY, Skinner A. 2011 Testing fossil calibrations for vertebrate molecular trees. *Zool. Scr.* 40, 538-543. (doi:10.1111/J.1463-6409.2011. 00488.X)
- 21. Pyron RA. 2010 A likelihood method for assessing molecular divergence time estimates and the placement of fossil calibrations. *Syst. Biol.* **59**, 185–194. (doi:10.1093/Sysbio/Syp090)
- Rutschmann F, Eriksson T, Abu Salim K, Conti E.
   2007 Assessing calibration uncertainty in molecular dating: the assignment of fossils to alternative

- calibration points. *Syst. Biol.* **56**, 591 608. (doi:10. 1080/10635150701491156)
- Sanders KL, Lee MSY. 2007 Evaluating molecular clock calibrations using Bayesian analyses with soft and hard bounds. *Biol. Lett.* 3, 275–279. (doi:10. 1098/Rsbl.2007.0063)
- Marshall CR. 2008 A simple method for bracketing absolute divergence times on molecular phylogenies using multiple fossil calibration points. Am. Nat. 171, 726 – 742. (doi:10.1086/587523)
- Dornburg A, Beaulieu JM, Oliver JC, Near TJ. 2011 Integrating fossil preservation biases in the selection of calibrations for molecular divergence time estimation. Syst. Biol. 60, 519-527. (doi:10.1093/ Sysbio/Syr019)
- Near TJ, Meylan PA, Shaffer HB. 2005 Assessing concordance of fossil calibration points in molecular clock studies: an example using turtles. *Am. Nat.* 165, 137 146. (doi:10.1086/427734)
- Joyce WG, Parham JF, Lyson TR, Warnock RCM, Donoghue PCJ. 2013 A divergence dating analysis of turtles using fossil calibrations: an example of best practices. J. Paleontol. 87, 612–634. (doi:10.1666/ 12-149)
- Near TJ, Meylan PA, Shaffer HB. 2008 Caveats on the use of fossil calibrations for molecular dating: a reply to Parham and Irmis. *Am. Nat.* 171, 137 – 140. (doi:10.1086/524201)
- Parham JF, Irmis RB. 2008 Caveats on the use of fossil calibrations for molecular dating: a comment on Near et al. Am. Nat. 171, 132 – 136. (doi:10. 1086/524198)
- Parham JF et al. 2012 Best practices for justifying fossil calibrations. Syst. Biol. 61, 346–359. (doi:10. 1093/Sysbio/Syr107)
- 31. Near TJ, Bolnick DI, Wainwright PC. 2005 Fossil calibrations and molecular divergence time estimates in centrarchid fishes (Teleostei: Centrarchidae). *Evolution* **59**, 1768–1782. (doi:10. 1111/j.0014-3820.2005.tb01825.x)
- Clarke JT, Warnock RCM, Donoghue PCJ. 2011
   Establishing a time-scale for plant evolution. New Phytol. 192, 266–301. (doi:10.1111/J.1469-8137. 2011.03794.X).
- 33. Yang ZH. 2007 PAML 4: phylogenetic analysis by maximum likelihood. *Mol. Biol. Evol.* **24**, 1586 1591. (doi:10.1093/Molbev/Msm088)
- dos Reis M, Yang ZH. 2011 Approximate likelihood calculation on a phylogeny for Bayesian estimation of divergence times. *Mol. Biol. Evol.* 28, 2161–2172. (doi:10.1093/Molbev/Msr045)
- Heled J, Drummond AJ. 2012 Calibrated tree priors for relaxed phylogenetics and divergence time estimation. Syst. Biol. 61, 138–149. (doi:10.1093/ Sysbio/Syr087)
- Drummond AJ, Rambaut A. 2007 BEAST:
   Bayesian evolutionary analysis by sampling trees. BMC Evol. Biol. 7, 214. (doi:10.1186/1471-2148-7-214)
- 37. Wilkinson RD, Steiper ME, Soligo C, Martin RD, Yang ZH, Tavare S. 2011 Dating primate divergences

- through an integrated analysis of palaeontological and molecular data. *Syst. Biol.* **60**, 16–31. (doi:10. 1093/Sysbio/Syq054)
- Noonan BP, Chippindale PT. 2006 Vicariant origin of Malagasy reptiles supports Late Cretaceous Antarctic land bridge. Am. Nat. 168, 730—741. (doi:10.1086/ 509052)
- Heath TA, Huelsenbeck JP, Stadler T. 2014 The fossilized birth-death process for coherent calibration of divergence-time estimates. *Proc. Natl Acad. Sci. USA* 111, E2957 – E2966. (doi:10.1073/pnas.1319091111)
- Drummond AJ, Nicholls GK, Rodrigo AG, Solomon W.
   2002 Estimating mutation parameters, population history and genealogy simultaneously from temporally spaced sequence data. *Genetics* 161, 1307 – 1320.
- 41. Müller J, Reisz RR. 2005 Four well-constrained calibration points from the vertebrate fossil record for molecular clock estimates. *Bioessays* **27**, 1069 1075. (doi:10.1002/Bies.20286)
- Reisz RR, Müller J. 2004 Molecular timescales and the fossil record: a paleontological perspective. *Trends Genet*.
   20, 237 241. (doi:10.1016/J.Tiq.2004.03.007)
- Burbrink FT, Lawson R. 2007 How and when did Old World ratsnakes disperse into the New World? *Mol. Phylogenet. Evol.* 43, 173 – 189. (doi:10.1016/J. Ympev.2006.09.009)
- Near TJ, Sanderson MJ. 2004 Assessing the quality of molecular divergence time estimates by fossil calibrations and fossil-based model selection. *Phil. Trans. R. Soc. Lond. B* 359, 1477 – 1483. (doi:10. 1098/Rstb.2004.1523)
- 45. Harrington RC, Benavides E, Near TJ. 2013
  Phylogenetic inference of nuptial trait evolution in the context of asymmetrical introgression in North American darters (Teleostei). *Evolution* **67**, 388–402. (doi:10.1111/J.1558-5646.2012.01797.X)
- Pozzi L, Hodgson JA, Burrell AS, Sterner KN, Raaum RL, Disotell TR. 2014 Primate phylogenetic relationships and divergence dates inferred from complete mitochondrial genomes. *Mol. Phylogenet. Evol.* 75, 165 – 183. (doi:10.1016/j.ympev.2014.02.023)
- Yang JS, Lu B, Chen DF, Yu YQ, Yang F, Nagasawa H, Tsuchida S, Fujiwara Y, Yang WJ. 2013 When did decapods invade hydrothermal vents? Clues from the Western Pacific and Indian Oceans. *Mol. Biol. Evol.* 30, 305–309. (doi:10.1093/Molbev/Mss224)
- 48. Hugall AF, Foster R, Lee MSY. 2007 Calibration choice, rate smoothing, and the pattern of tetrapod diversification according to the long nuclear gene RAG-1. *Syst. Biol.* **56**, 543 563. (doi:10.1080/10635150701477825)
- Ronquist F, Klopfstein S, Vilhelmsen L, Schulmeister S, Murray DL, Rasnitsyn AP. 2012 A total-evidence approach to dating with fossils, applied to the early radiation of the Hymenoptera. Syst. Biol. 61, 973 – 999. (doi:10.1093/sysbio/sys058)
- Lee MSY, Cau A, Naish D, Dyke GJ. 2014
   Morphological clocks in paleontology, and a mid-Cretaceous origin of crown Aves. Syst. Biol. 63, 442 – 449. (doi:10.1093/Sysbio/Syt110)