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## Fungal Rhino Sinusitis in Tehran, Iran

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### Abstract

**Background:** Fungal rhino sinusitis (FRS) is an important infection of para nasal sinuses, which encompasses two main categories; invasive and noninvasive forms according to histopathological findings. *Aspergillus* spp are the most common species isolated from noninvasive form, while Mucorales are more frequently isolates from acute infections.

**Methods:** Four hundred fifty patients suspected to fungal rhino sinusitis were investigated in a cross-sectional prospective study from June 2009 to Sep 2013. All patients under went endoscopic sinus surgery of the middle meatus. Tissue biopsies were investigated for culture, histopathology and molecular examination.

**Results:** Totally, 87 patients were diagnosed with fungal rhinosinusitis. *A. flavus* was the most common etiological agent of chronic invasive form (CIFRS), allergic fungal rhino sinusitis (AFRS) and fungus ball (FB), while *Rhizopus oryzae* (26.7%) was the most common cause of infection in acute invasive fungal rhino sinusitis (AIFR). However, a few rare species such as *Shyzoophyllum commune* and *Fusarium proliferatum* were also isolated.

**Conclusion:** Diabetes is the most important predisposing factor for patients with acute invasive form of sinusitis and the most involved sinuses were unilateral multiple sinuses and maxillary sinus.

**Keywords:** Fungal rhinosinusitis, AFRS, AIFR, Fungus ball

### Introduction

Fungal rhino sinusitis (FRS) is a group of infections affecting in paranasal including invasive and noninvasive forms. The invasive infections encompasses acute invasive or fulminant (AIFRS), Chronic granulomatos invasive (CGFRS) and chronic invasive (CIFRS). The noninvasive diseases include saprophytic fungal infestation (SFI), fungus ball (FB) and Allergic fungal rhino sinusitis (AFRS) (1, 2). These forms have been classified according to histopathology finding such as fungal invasion to vessels, bone erosion and infection period (1, 3). Acute invasive commonly happen in patients with immune suppressed status in a pe-

riod less than 4 weeks (4). However, in all of invasive forms hyphal invasion to vessels and bone erosion are observed (2,5). In other hand noninvasive are an extra mucosal infection and most of the patient are immune competent (6,7).

*Aspergillus* sp. is the most common species reported as a major cause of fungal sinusitis (5,8-11) with the exception of acute invasive form that the most frequent fungal isolated belong to the Zygomycete order (8). However, etiologic agents of these infections may vary according to type of sinusitis and geographical epidemiology (4).

Up until now, the majorities of studies performed in Iran were limited to a few case reports or limited to particular form of fungal rhino sinusitis in a special patient (12, 13). In addition, the identified fungal agents were most fungal species identified by conventional methods (13,14).

The link between clinical features and etiological agent is an important factor to apply proper administration of antifungal agent (15). The application of complete identification of fungal agent seems quite necessary. This may apply a combination of culture, morphology, and molecular base assays.

In the present study we used both microscopic morphology and sequence-based identification assays using ITS and B-tubulin rDNA gene in order to identify causing agent own to species level. We also aimed to link the fungal sinusitis sub types to the etiological agent and the associated predisposing factors in patients with suspected rhino sinusitis.

## Materials and Methods

In a prospective cross-sectional study, 450 patients with suspected fungal rhino sinusitis referred to Amir-Alam Hospital, Tehran were examined. The patients had headache, rhinorrhea, obstruction and nasal discharge; inflammation or polyps in endoscopic examinations with sinus involvement in CT scan findings after obtaining informed consent were included in this study. All patients underwent endoscopic sinus surgery of the middle meatus. Tissue biopsies were taken to evaluate histopathological and mycological characteristics of these infections. Some portions of tissue samples were inoculated on two multi points of Sabouraud Dextrose Agar plate (belief, Italy) supplemented with chloramphenicol (0.5mg/ml) and incubated for 10 days at 30 and 37°C (16). All samples with positive culture identified by macroscopic features and morphological criteria were confirmed by PCR. From the 10 days colony on 4% sabouraud dextrose agar fungal DNA was extracted by using a conical grinder according to the previously described method (17).

Three pairs of forward and reverse primers were designed based on the sequence of translation elongation factor-1, ITS rDNA and tubulin genes for cases of *Fusarium* sp., *Aspergillus* sp. and *micoral* sp. Demateaceous and indistinguishable species) were identified respectively by macroscopic features and morphological criteria (18-20). PCR was carried out according to PCR programs, selected for each primer (16, 19, 20). The PCR products were subjected to DNA sequencing with the same primers.

The obtained sequences were analyzed in Gen Bank database and revealed high identities recorded and then the results were compared with macroscopic features and microscopic structures.

## Results

In total, 87 patients were diagnosed with fungal rhino sinusitis over the period of this study. Fungal elements were identified in both direct examination and pathological finding. Thirty-three (30.7%) out of 87 patients had negative culture. Overall the mean patient age was (10-86) with a female to male ratio of 1.16/1 and based on clinical histological criteria patients were classified (Table 1). Overall, 81% of patients lived in urban areas and most prevalent of disease were house keeper women and worker men.

Depending on the type of sinusitis there were differences between fungal species. *A. flavus* was the most common etiological agent in CIFRS, AFRS and FB while *Rhizopusoryze* (26.7%) was the most common cause of infection in AIFR although some rare species such *Syzyphyllum commune* and *F. proliferatum* also report (Table2).

All patients with acute invasive form of sinusitis had underlying disease such as diabetes (71.4%) and malignancy (28.6%) whereas diabetes and malignancy in patients with non- invasive fungal sinusitis was 12.2% and 6.1% respectively. Headache (46.2%), nasal discharge (34.6%) and facial pain (33.3%) were most symptoms but these were different according to sinusitis form. The most involved sinuses were unilateral multiple sinuses and maxillary sinus.

**Table 1:** Distribution of patients with fungal rhino sinusitis based on age, Tehran, 2012-13

Sinusitis type	NO	Mean	Range	F:M
AIFRS	28	41	(10-86)	1.15/1
FB	33	44	(20-69)	1.5/1
AFS	16	36	(11-67)	1:1/3
CIFRS	1	55	55	-
Total	78	41	(10-86)	1.16/1

**Table 2:** Distribution of isolated fungal species based on rhino sinusitis type, Tehran, 2012-2013

Fungal species	Rhino sinusitis types				Total
	AIFRS No-PER (%)	FB No-PER(%)	AFRS No-PER (%)	CIFRS No-PER (%)	
<i>Aspergillus flavus</i>	2-4.4	14-31.1	9-20	0	25-55.5
<i>Aspergillus fumigatus</i>	0	1-2.2	2-4.4	0	3-6.6
<i>Rhizopus oryza</i>	12-26.7	0	0	0	12-26.7
<i>Fusarium proliferatum</i>	0	1-2.2	0	0	1-2.2
<i>Shizophyllum commune</i>	0	1-2.2	0	0	1-2.2
<i>Nattrassia mangiferae</i>	0	0	1-2.2	0	1-2.2
<i>Candida albicans</i>	0	0	2-4.4	0	2-4.4
Total	14-31.1	17-37.7	14-31	0	45- 100

## Discussion

Fungal rhino sinusitis has been considered as an uncommon disorder and its frequency are increasing in recent years. (21). This study is the first large study from Iran, and other studies had been conducted of limited (12-14, 22, 23). Thus many of fungal isolates particularly Zygomycetes are identified in genus level that show different in vitro susceptibility to antifungal agents (24). Isolation and identification of fungal agent are tremendously important for proper management of infections caused by the less common fungi (25, 26). Therefore, combination of PCR sequencing with conventional methods able us to identify fungal isolates precisely specially in some rare species such as *S. commune* which despite the morphological study, due to the lack of diagnostic structure the only successful method for identification was PCR sequencing.

In this study majority of patient had non-invasive disease. This is similar to previous studies but different in sub type disorder frequency (5, 8, 27).

In contrast with our study, Challa et al. in South India reported a low frequency of non-invasive fungal sinusitis (25%) vs. invasive form (75%) (9). FB was the most common form that was similar to previous studies (5, 7). The only one case (1.3%) of CIFRS reported in a diabetic man was similar to Granville et al. , Monotone et al. and Das et al. study who reported 2.1% , 1.2% and 1% of patients with CIFRS in their patient groups respectively (4, 28) although the incidence CIFRS was higher (10%) in the Micheal et al. study (8).

Incidence AFRS was lower than other studies (28 ,29) but Panda et al. and Chakrabarti, et al. reported much lower incidence of this type of sinusitis (5,11). This difference could be due to variation in diagnostic criteria for disease defining because the existence of eosinophilic mucin (EM) with fungal elements in histopathological examination or the existence EM without fungal elements in histopathological examination (30). So it should not be ignored which could be due to contamination especially in some study samples collection by nasal irrigation fluid

through nasal passage and nasal brush or swab from nasal septums (31-34).

*Aspergillus* species are the most common fungal agents of the paranasal sinuses while according to geographical conditions there is a difference between *Aspergillus* species (5, 7-9, 27). *A. fumigatus* has the highest frequency in some reports (4,7) but in our study *A. flavus* had the highest frequency similar to other studies (5,8,27,35).

*Rhizopus* sp., *A. fumigatus* and *A. flavus* are as the main causes of the acute invasive sinusitis in around the world (5, 8, 9, 27). In our study *Rhizopus oryza* was the most common fungal isolated in acute invasive sinusitis and all patients with this form showed an immune deficiency such as malignancy or uncontrolled diabetes.

Most variation in fungal species reported in AFRS. Although the *Aspergillus* sp. and dematiaceous species are most isolated but fungal species vary in disparate geographic area in different studies (4). However in our study dematiaceous had a very small role in AFRS and like some other study *A. flavus* was most common fungal species isolated in patients with AFRS (5, 9, 35).

Although the over growth of the *Aspergillus* sp. compared to the dematiaceous species in routine culture media should not be ignored (4), but hyaline septate hyphae, which branch at a 45 ° angle observed in direct examination was able to rollout infection due to dematiaceous fungal species (36).

33.3% of all samples submitted to laboratory with normal saline container had negative culture that was similar to some other studies (2, 36, 37). This could be due to the damaged hyphae during processing or when limited specimen is available and fungal agent lost their vitality. So histopathological examination provide only a presumptive diagnosis (38).

## Conclusion

This study showed the potential assistance by using the PCR sequencing method in combination with a conventional assay for precise identification of fungal species in fungal

rhino sinusitis. However, it seems in cases where fungal elements envisage and where either culture result is negative or culture is not done, fungal DNA detection from tissue specimens for improvement of detection is necessary. In this situation we have more complete information about fungal rhino sinusitis epidemiology in Iran that help to better management and control of this infection by ENT specialists.

## Ethical considerations

Ethical issues (Including plagiarism, informed consent, misconduct, data fabrication and/or falsification, double publication and/or submission, redundancy, etc.) have been completely observed by the authors.

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## References

1. Chakrabarti A, Denning DW, Ferguson BJ, Ponikau J, Buzina W, Kita H, et al. (2009). Fungal rhinosinusitis. *Laryngoscope*, 119(9): 1809-18.
2. Pagella F, Matti E, Bernardi FD, Semino L, Cavanna C, Marone P, et al. (2007). Paranasal sinus fungus ball: diagnosis and management. *Mycoses*,50(6):451-6.
3. Deshazo RD (1998). Fungal sinusitis. *Am J Med Sci*,316(1):39-45.
4. Montone KT, Livolsi VA, Feldman MD, Palmer J, Chiu AG, Lanza DC, et al. (2012). Fungal rhinosinusitis: a retrospective microbiologic and pathologic review of 400 patients at a single university medical center. *Int J Otolaryngol*, Article ID 684835, pages 9.
5. Panda NK, Sharma SC, Chakrabarti A, Mann S (1998). Paranasal sinus mycoses in north India. *Mycoses*,41(7-8):281-6.
6. Ferguson BJ (2000). Fungus balls of the paranasal sinuses. *Otolaryngol Clin North Am*,33(2):389-98.

7. Dufour X, Kauffmann-Lacroix C, Ferrie J, Goujon J, Rodier M, Klossek J (2006). Paranasal sinus fungus ball: epidemiology, clinical features and diagnosis. A retrospective analysis of 173 cases from a single medical center in France, 1989-2002. *Med Mycol*, 44(1):61-7.
8. Michael RC, Michael JS, Ashbee RH, Mathews MS (2008). Mycological profile of fungal sinusitis: An audit of specimens over a 7-year period in a tertiary care hospital in Tamil Nadu. *Ind J Pathol Microbiol*, 51(4):493.
9. Challa S, Uppin SG, Hanumanthu S, Panigrahi MK, Purohit AK, Sattaluri S, et al. (2010). Fungal rhinosinusitis: a clinicopathological study from South India. *Eur Arch Otorhinolaryngol*, 267(8):1239-45.
10. Jain S, Das S, Gupta N, Malik JN (2013). Frequency of fungal isolation and antifungal susceptibility pattern of the fungal isolates from nasal polyps of chronic rhinosinusitis patients at a tertiary care centre in north India. *Med Mycol*, 51(2):164-169.
11. Chakrabarti A, Sharma S, Chander J (1992). Epidemiology and pathogenesis of paranasal sinus mycoses. *Otolaryngol Head Neck Surg*, 107(6 Pt 1):745-750.
12. Kordbacheh P, Badiie P, Alborzi A, Zaini F, Mirhendi H, Mahmoudi M, et al. (2008). Acute Fulminant Fungal Sinusitis in Patients with Acute Leukemia. *Iran J Public Health*, 37(4): 46-51.
13. Eslamifar A, Razzaghi-Abyaneh M, Vazir-Nezami M, Moghadasi H, Ramezani A, Shams-Ghahfarokhi M, et al. (2008). Frequency and Identification of Fungal Strains in Patients with Chronic Rhinosinusitis. *Iran J Pathol*, 3(3):135-9.
14. Okhovvat AR, Karim M, Hashemi SM, Hashemi SM, Berjis N, Amiridavan M, et al. (2010). Fungal Sinusitis and Treatment. *J Isfahan Med Sci*, 27(103):866-73.
15. Taneja T, Saxena S, Aggarwal P, Reddy V (2011). Fungal infections involving maxillary sinus—a difficult diagnostic task. *J Clin Exp Dental*, 3(2):172-6.
16. Nazeri M, Mohammadi Ardehali M, Moazeni M, Hashemi S J, Ehteram H, Rezaie S (2012). A case of Fungus ball type pansinusitis caused by *Schizophyllum commune*. *Med Mycol Case Reports*, 1(1): 115-118.
17. Rezaei-Matehkolaei A, Makimura K, de Hoog S, Shidfar MR, Zaini F, Eshraghian M, et al. (2013). Molecular epidemiology of dermatophytosis in Tehran, Iran, a clinical and microbial survey. *Med Mycol*, 51(2):203-7.
18. Arunmozhi Balajee S, Sigler L, Brandt ME (2007). DNA and the classical way: identification of medically important molds in the 21st century. *Med Mycol*, 45(6):475-90.
19. Geiser DM, del Mar Jiménez-Gasco M, Kang S, Makalowska I, Veeraraghavan N, Ward TJ, et al. (2004). Fusarium-ID v. 10: A DNA sequence database for identifying Fusarium. *Molecular Diversity and PCR-detection of Toxigenic Fusarium Species and Ochratoxigenic Fungi*: Springer. p. 473-9.
20. May GS, Gambino J, Weatherbee JA, Morris NR (1985). Identification and functional analysis of beta-tubulin genes by site specific integrative transformation in *Aspergillus nidulans*. *J Cell Biol*, 101(3):712-9.
21. Chakrabarti A, Das A, Panda NK (2004). Overview of fungal rhinosinusitis. *Indian J Otolaryngol Head Neck Surg*, 56(4):251-8.
22. Hashemian F, Bakhshaei M (2012). The role of fungi in chronic rhinosinusitis in the high altitude region of Iran. *Iran J Otorhinolaryngol*. 24(66):29-33.
23. Naghibzadeh B, Razmpa E, Alavi S, Emami M, Shidfar M, NAGHIBZADEH G, et al. (2011). Prevalence of fungal infection among Iranian patients with chronic sinusitis. *Acta Otorhinolaryngol Ital*, 31(1):35- 38.
24. Dannaoui E, Meis JF, Loeberberg D, Verweij PE (2003). Activity of posaconazole in treatment of experimental disseminated zygomycosis. *Antimicrob Agents Chemother*, 47(11):3647-50.
25. Pfaller M, Diekema D (2004). Rare and emerging opportunistic fungal pathogens: concern for resistance beyond *Candida albicans* and *Aspergillus fumigatus*. *J Clin Microbiol*, 42(10): 4419-31.
26. Walsh T, Groll A, Hiemenz J, Fleming R, Roilides E, Anaissie E (2004). Infections due to emerging and uncommon medically important fungal pathogens. *Clin Microbiol Infect*, 10(s1):48-66.
27. Das A, Bal A, Chakrabarti A, Panda N, Joshi K (2009). Spectrum of fungal rhinosinusitis;

- histopathologist's perspective. *Histopathology*, 54(7):854-9.
28. Granville L, Chirala M, Cernoch P, Ostrowski M, Truong LD (2004). Fungal sinusitis: histologic spectrum and correlation with culture. *Hum Pathol*,35(4):474-81.
  29. Taxy JB (2006). Paranasal fungal sinusitis: contributions of histopathology to diagnosis: a report of 60 cases and literature review *Am J Surg Pathol*,30(6):713-20.
  30. deShazo RD, Swain RE (1995). Diagnostic criteria for allergic fungal sinusitis. *J Allergy Clin Immunol*,96(1):24-35.
  31. Polzehl D, Weschta M, Podbielski A, Riechelmann H, Rimek D (2005). Fungus culture and PCR in nasal lavage samples of patients with chronic rhinosinusitis. *J Med Microbiol*,54(1):31-7.
  32. Catten MD, Murr AH, Goldstein JA, Mhatre AN, Lalwani AK (2001). Detection of fungi in the nasal mucosa using polymerase chain reaction. *Laryngoscope*, 111(3):399-403.
  33. Braun H, Buzina W, Freudenschuss K, Beham A, Stammberger H (2009). 'Eosinophilic Fungal Rhinosinusitis': A Common Disorder in Europe? *Laryngoscope*.113(2):264-9.
  34. Ponikau JU, Sherris DA, Kern EB, Homburger HA, Frigas E, Gaffey TA, et al.(1999). editors. The diagnosis and incidence of allergic fungal sinusitis. *Mayo Clin Proc*, 74(9): 877-884.
  35. Al-Dousary SH. Allergic fungal sinusitis: Radiological and microbiological features of 59 cases(2008). *AnnSaudi Med*, 28(1)17-21.
  36. Grosjean P, Weber R (2007). Fungus balls of the paranasal sinuses: a review.*Eur Arch Otorhinolaryngol*,264(5):461-70.
  37. Klossek JM, Serrano E, Péloquin L, Percodani J, Fontanel JP, Pessey JJ (1997). Functional endoscopic sinus surgery and 109 mycetomas of paranasal sinuses. *Laryngoscope*, 107(1):112-7.
  38. Lau A, Chen S, Sorrell T, Carter D, Malik R, Martin P, et al. (2007). Development and clinical application of a panfungal PCR assay to detect and identify fungal DNA in tissue specimens. *J Clin Microbiol*,45(2):380-5.