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REVIEW

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ALG8-CDG: novel patients and review of the literature

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Abstract

Background: Since 1980, about 100 types of congenital disorders of glycosylation (CDG) have been reported representing an expanding group of inherited disorders. ALG8-CDG (= CDG-Ih) is one of the less frequently reported types of CDG, maybe due to its severe multi-organ involvement with coagulation disturbances, edema, massive gastrointestinal protein losing enteropathy, cataracts, and often early death. We report three additional patients, provide an update on two previously reported, and summarize features of ten patients reported in literature.

Results: Of 15 ALG8-CDG patients, three were homozygous and 12 compound heterozygous. There were multiple prenatal abnormalities in 6/12 patients. In 13/15, there were symptoms at birth, 9/15 died within 12 months. Birth weight was appropriate in 11/12, only one was small for gestational age. Prematurity was reported in 7/12. Hydrops fetalis was noticed in 3, edemas in 11/13; gastrointestinal symptoms in 9/14; structural brain pathology, psychomotoric retardation, seizures, ataxia in 12/13, muscle hypotonia in 13/14. Common dysmorphic signs were: low set ears, macroglossia, hypertelorism, pes equinovarus, campto- and brachydactyly (13/15). In 10/11, there was coagulopathy, in 8/11 elevated transaminases; thrombocytopenia was present in 9/9. Eye involvement was reported in 9/14. CDG typical skin involvement was reported in 8/13.

Conclusion: In ALG8-CDG, isoelectric focusing of transferrin in serum or plasma shows an abnormal sialotransferrin pattern. The diagnosis is confirmed by mutation analysis in *ALG8*; all patients reported so far had point mutations or small deletions. The prognosis is generally poor. Thus, a timely and correct diagnosis is important for counselling.

Keywords: Congenital disorders of glycosylation, Isoelectric focusing of transferrin, Hydrops fetalis, Cataract, Coagulopathy, Gastrointestinal symptoms

Background

Congenital disorders of glycosylation (CDG) involve a rapidly growing group of metabolic diseases that are caused by genetic defects in the synthesis of glycoproteins [1]. The first two patients were diagnosed in 1980 by the Belgian pediatrician Jaekel Jaeken [2]. To date, some 100 CDG have been identified [1, 3] (with over 700 patients reported worldwide), and about 50 of these can be identified via isoelectric focusing of transferrin [1, 3–5]. CDG have been divided in four groups: (1) disorders of protein N-glycosylation, (2) disorders of protein O-glycosylation, (3) disorders of lipid and GPI

(glycosylphosphatidylinositol) anchor glycosylation, and (4) disorders of multiple glycosylation pathways [5, 6]. Defects in protein N-glycosylation are divided into two subgroups: CDG-I disorders comprise defects in the glycan synthesis (cytosol and endoplasmic reticulum (ER)), and CDG-II disorders defects in glycan processing (Golgi apparatus) [1]. In 2008, Jaeken *et al.* proposed a nomenclature that refers to the gene name under the umbrella name of CDG, e.g. PMM2-CDG, ALG8-CDG, etc. [7]. CDG affect multiple organ systems, the severity of symptoms is highly variable and it shows a broad clinical spectrum with considerable overlap [1, 3].

ALG8-CDG (OMIM #608104) is due to inactivating mutations in the *ALG8* gene encoding dolichyl-phosphate-glucose 1-mannose 9-N-acetylglucosamine (Dol-P-Glc:Glc

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(2)Man (9)GlcNAc(2)-PP-Dol-alpha-2) glucosyltransferase (= glucosyltransferase 2) in the ER, an enzyme that attaches the second glucose residue to dolichol-PP-glycans in the ER [8]. ALG8-CDG is a CDG form with dysmorphism (retrognathia, low-set ears, pes equinovarus), muscular hypotonia, hepatomegaly, coagulopathy (thrombocytopenia), edema and ascites (including fetal hydrops), cardiorespiratory problems, and protein-losing enteropathy (watery diarrhea) [8]. Cataracts (ocular findings) have only been reported in ALG8-CDG, ALG12-CDG (CDG-Ig), other subtypes in early LLO (lipid-linked oligosaccharides) synthesis (like SRD5A3-CDG) and some unclassified CDG [9, 10].

Here, we describe three severely affected neonates from two families with ALG8-CDG with congenital cataracts, and provide an update on two previously reported children with congenital cataracts [11], where ALG8-CDG was confirmed recently. In addition, we summarize the clinical and biochemical characteristics of these five patients, further ten published patient reports and present a review of the literature.

Clinical reports - patients

Patient 1

The first child of healthy, non-consanguineous, Caucasian parents, was born in the 34th gestational week after oligohydramnios in pregnancy. 15 min after birth, the boy died of severe respiratory failure due to pulmonary hypoplasia with alveolar dysgenesis. Clinical examination showed low-set ears, macroglossia, bilateral pes equinovarus, hypoplastic thorax, generalized edema, and a pale gray mottled skin. Bilateral cataracts were noted. Autopsy showed renal tubular dysgenesis, cerebral edema and ascites.

Patient 2

The second boy, brother of Patient 1, was born at 35 weeks of gestation by Caesarean section because of a pathological cardiotocogram (CTG). Pregnancy was complicated by intrauterine hydrops fetalis with pericardial effusion and ascites as well as oligohydramnios. Birth weight was 2700 g (50th centile), head circumference 34.5 cm (75th centile) and length 48 cm (75th centile). The Apgar scores were 3/7/8 and umbilical cord pH was 7.36. Because of respiratory insufficiency, the boy was mechanically ventilated for four days. Dysmorphism (wide-open fontanel, incomplete bilateral cataract, low-set ears, hypertelorism, macroglossia, retrognathia), generalized edema, a pale mottled skin as well as pronounced muscular hypotonia were present. Laboratory investigation showed thrombocytopenia (lowest value 19,000/ μ L, decreased prothrombin time (PT)(40 %; nl 74–108) and increased partial thromboplastin time (PTT) (>200 s.; nl 28–49)).

After initial stabilization the patient's general condition worsened in the third week of life. He developed watery diarrhoea and vomiting, and subsequently acute renal failure with anuria and ascites. His condition deteriorated and at 34 days of life he died due to cardiocirculatory failure.

Patient 3

In 1996, our group reported the first, albeit at that time unclassified, ALG8-CDG patient (Table 1) [11]. The girl was the second child of healthy, unrelated, Caucasian parents and died at the age of 30 months. She presented with severe psychomotor disability, microphthalmia, cataract, brachydactyly, cortical atrophy, slight elevation of serum transaminases and blood coagulation abnormalities.

Patient 4

Six years later, a brother of Patient 3 (Fig. 1a,b) [11] was born after an uneventful pregnancy by Caesarian section in the 36th gestational week because of breech presentation. Birth weight was 2590 g, length 48 cm and head circumference 33 cm (all 25th centile). The boy presented as a floppy infant, with bilateral congenital cataracts and multiple dysmorphic features such as a prominent forehead, a wide fontanelle, macroglossia, dysplastic ears, a high-arched palate, inverted nipples, peculiar fat pads of the buttocks, and short fingers and toes. Cyanotic apnoeas were repeatedly present from the 4th day of life. Laboratory tests showed slight elevation of transaminases and transient hypoalbuminaemia, as well as pathological coagulation (with prolonged PTT (partial thromboplastin time), low antithrombin III, and several reduced coagulation factors). He suffered from vitreous bleeding during cataract operation and died at the age of four years from cardiac dysrhythmia.

Patient 5

This girl (Fig. 2a), sister of a healthy four year old girl, was born in the 37th gestational week after an uneventful pregnancy. Labour was induced because of a pathological CTG. Birth weight was 3080 g (50th centile), length 47 cm (10th centile) and head circumference 33 cm (25th centile), the Apgar scores were 8/9/10 and arterial umbilical cord pH was 7.36. She was hospitalized due to fluctuations of saturation, tachypnea and increased inflammation parameters as well as thrombocytopenia (lowest platelet count was 49 G/l; normal range: 223–510 G/l), and was treated with antibiotics. During the first days, she vomited repeatedly. She showed mild microcephaly, facial dysmorphism (bilateral cataract, low-set ears, relative macroglossia), fat pads on the arms (Fig. 2b) and thighs, hypotonia with dystonic posturing, hepatosplenomegaly, and persisting primary vitreous artery (Fig. 3). Routine laboratory parameters (including coagulation

Table 1 Findings in 15 patients with ALG8 deficiency

| Sex | Skladal [11] | | Charlwood [14] | Chantret [8] | Schollen [15] | | Eklund [9] | Vesela [16] | Stölting [17] | | Sorte [18] | Funke [19] Kouwenberg [20] | Patients 1, 2, 5 | | |
|---------------------------|--------------|-----------|----------------|--------------|---------------|---------|------------|---------------|---------------|------------|------------|-------------------------------|------------------|------------|------------|
| | F | M | M | F | F | M | M | F | M | F | F | M | M | M | F |
| Prenatal Data | | | | | | | | | | | | | | | |
| IUGR | / | / | Yes | / | Yes | No | No | No | No | No | Yes | No | No | No | No |
| Oligohydramnios | / | / | Yes | / | Yes | No | No | Yes | No | No | No | No | Yes | Yes | No |
| Hydrops fetalis | / | / | ND | / | / | / | / | Yes | No | No | / | No | Yes | Yes | No |
| Neonatal data | | | | | | | | | | | | | | | |
| Weeks of gestation | / | 36 w (CS) | 35 w (CS) | / | / | 35 w | 37 w | 29 w (CS) | At term | At term | 39 w (CS) | 35 w | 34 w (CS) | 35 w (CS) | 37 w (CS) |
| Birth weight | / | 2,590 g | 2,280 g | / | / | 2,920 g | 3,070 g | 1,420 g | 3,980 g | 3,680 g | 2,210 g | 2,920 g | 2,570 g | 2,700 g | 3,080 g |
| Clinical symptoms | | | | | | | | | | | | | | | |
| Ascites/edemas | / | | Yes | Yes | Yes | Yes | Yes | Yes | No | No | Yes | Yes | Yes | Yes | Yes |
| Onset of first symptoms | From birth | | From birth | 4 months | From birth | 8 weeks | From birth | From birth | From birth | From birth | From birth | From birth | From birth | From birth | From birth |
| Dysmorphism | Yes | | Yes | No | Yes | Yes | Yes | No | Yes | Yes | Yes | Yes | Yes | Yes | Yes |
| Gastrointestinal symptoms | / | | Yes | Yes | No | Yes | Yes | Yes | No | No | Yes | Yes | No | Yes | Yes |
| Brain involvement | Yes | | Yes | No | / | Yes | Yes | Yes | Yes | Yes | Yes | Yes | / | Yes | Yes |
| Hypotonia | / | | Yes | No | Yes | Yes | Yes | Yes | Yes | Yes | Yes | Yes | Yes | Yes | Yes |
| Skin involvement | / | | Yes | No | / | Yes | No | No | Yes | Yes | Yes | Yes | No | No | Yes |
| Eye symptoms | Cataract | | No | Retinopathy | / | No | Cataract | Optic atrophy | No | No | Nystagmus | No | Cataract | Cataract | Cataract |
| Laboratory data | | | | | | | | | | | | | | | |
| Thrombocytopenia | / | | Yes | / | Yes | Yes | Yes | Yes | / | / | Yes | Yes | / | Yes | Yes |
| Coagulopathy | Yes | | Yes | Yes | Yes | ND | Yes | Yes | / | / | Yes | Yes | / | Yes | No |
| Hyponatremia | / | | Yes | / | Yes | Yes | Yes | / | / | / | / | Yes | Yes | Yes | Yes |
| Increased transaminases | Yes | | Yes | No | / | Yes | Yes | No | / | / | Yes | Yes | / | Yes | No |

Table 1 Findings in 15 patients with ALG8 deficiency (Continued)

| Survival | 30 months | 4 years | 3 months | >3 years | 3 days | 3 months | 16 months | 2 months | >6 years | >7 years | 8,5 months | 3 months | 15 min | 34 days | 39 days |
|-------------------------|-----------|------------|-----------------|--------------|-----------------|-----------------|------------|------------|-------------|-------------|------------|-----------------|--------|------------|------------------|
| ALG8 mutations | / | Homozygous | Comp. het. | Comp. het. | Comp. het. | Comp. het. | Homozygous | Comp. het. | Comp. het. | Comp. het. | Comp. het. | Comp. het. | / | Comp. het. | Comp. het. |
| Mutation 1 (genomic) | | c.139A>C | c.139A>C | c.413delC | c.139A>C | c.673+4A>G | c.139A>C | c.139A>C | c.845C>T | c.845C>T | c.799T>C | c.673+4A>G | | c.139A>C | c.139A>C |
| Mutation 1 (functional) | | p.T47P | p.T47P | p.T138Kfs*19 | p.T47P | Splice mutation | p.T47P | p.T47P | p.A282V | p.A282V | p.S267P | Splice mutation | | p.T47P | p.T47P |
| Mutation 2 (genomic) | | c.139A>C | c.96-2A>G | c.396insA | c.96-2A>G | c.824G>A | c.139A>C | c.1090C>T | c.1436delC | c.1436delC | c.808T>C | c.824G>A | | c.1090C>T | c.1219_1220delCT |
| Mutation 2 (functional) | | p.T47P | Splice mutation | p.V133Sfs*3 | Splice mutation | p.G275D | p.T47P | p.R364* | p.P479Lfs*6 | p.P479Lfs*6 | p.F270L | p.G275D | | p.A364* | p.L407Dfs*23 |

CS Cesarean section; ND not described or no information available; dysmorphism such as low-set ears, macroglossia, pes equinovarus, campto- and brachydactyly; gastrointestinal symptoms include diarrhea, vomiting, protein-losing enteropathy; CNS defects include structural brain abnormalities, mental or psychomotor retardation, seizures; hypotonia presenting as floppy infant; skin involvement is abnormal fat distribution, wrinkly skin, cutis laxa, inverted nipples; electrolyte disturbances refers to hyponatremia; exitus, death, when written as > years it means age at last follow-up, no further information on outcome, comp. het., compound heterozygous

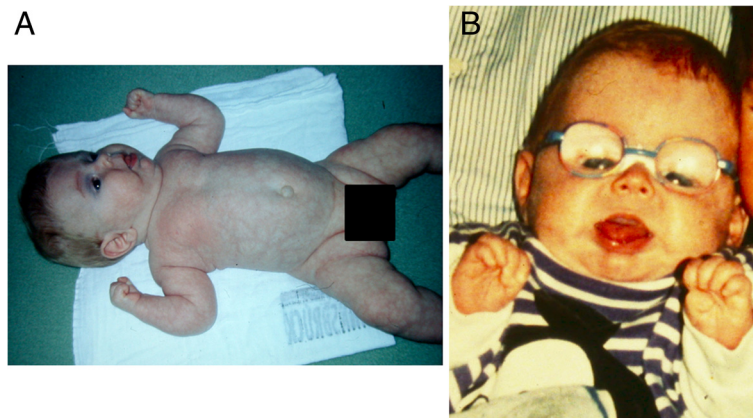


Fig. 1 a,b Patient 4 at the age of 10 months - floppy infant, with multiple dysmorphic features such as a prominent forehead, wide fontanelle, macroglossia, dysplastic ears, high palate, inverted nipples, short fingers and toes and a pale mottled skin, "fat pads" on arms and thighs [11]

parameters and thyroid function tests) were normal. Over the next weeks, the girl deteriorated progressively with recurrent episodes of apneas often combined with bradycardia. Electroencephalogram (EEG) showed a pathological brain wave pattern with decreased brain activity. On day 38 of life, a pale skin and apneas were noted, with ventricular tachycardia. The girl died from cardiovascular arrest.

Methods

Serum transferrin isoelectrofocusing was performed as reported [12]. Mutation analysis of the *PMM2* and *ALG8* genes was performed according to standard methods [1, 13].

In CDG, enzyme analyses are not available for all types of defects in the pathway. When enzymatic analysis lacks, lipid-linked oligosaccharides (LLO) in fibroblasts

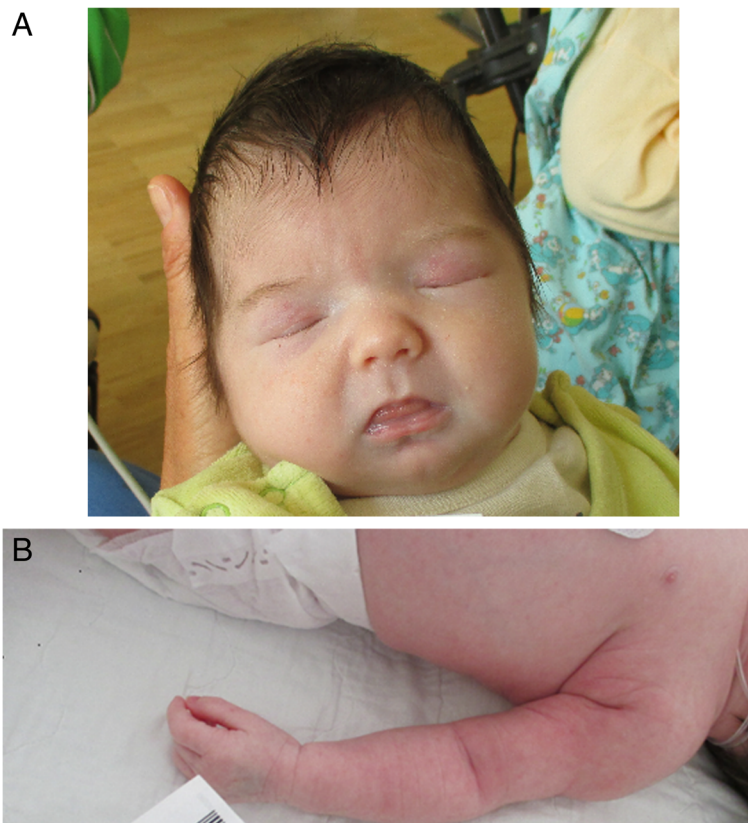


Fig. 2 a,b Patient 5 at the age of 20 days - floppy infant, with intermittent dystonic posturing, and multiple dysmorphic features such as an abnormal fat distribution on arms and thighs, macroglossia, low-set ears, additionally, cataracts and persisting primary vitreous artery were present

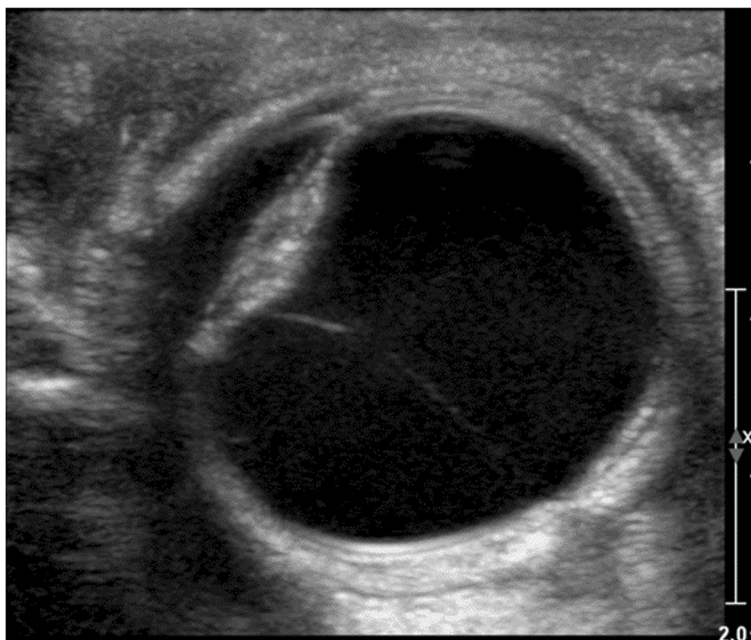


Fig. 3 Ocular sonography of Patient 5 at the age of 10 days - Persistent hyperplastic primary vitreous artery. A thin y-shaped membrane extends from the optic disc to the posterior aspect of the opacified lens representing a remnant of the hyaloid vascular system

can show characteristic profiles [13], narrowing down the diagnosis of CDG. For confirmation of the diagnosis, molecular genetic testing is necessary. Standard Sanger sequencing was performed in all five patients. In the future, with a better availability, exome or whole genome sequencing methods might be helpful as well.

Results

In all our 5 patients, the plasma isoelectric focusing of transferrin showed a type 1 pattern (CDG-I) and confirmed the clinical suspicion of an underlying CDG.

In Patient 4, sialotransferrins showed a type 1 pattern (CDG-Ix). However, PMM activity in fibroblasts was normal. Through enrollment in the Euroglycanet network, ALG8-CDG was diagnosed in 2011 (homozygous c.139A>C p.Thr47Pro). The parents are heterozygous.

In Patient 2 with prenatal hydrops, edema and cataract, the serum transferrin type 1 pattern prompted us to investigate *ALG8* gene as a candidate. A founder mutation was suspected as the family originated from the same Tyrolean region where we previously had diagnosed another pair of siblings (Patient 3 and 4) with type 1 pattern and cataract [11]. Two heterozygous mutations in the *ALG8* gene were found in DNA extracted from fibroblasts of Patient 2 (c.139A>C in exon 2, p.Thr47Pro, and c.1090C>T in exon 10, p.Arg364Ter), confirming the diagnosis of ALG8-CDG.

The clinical similarity and plasma transferring type I pattern prompted analysis of *ALG8* gene in Patient 5. This showed compound heterozygosity for the known

ALG8 mutation in exon 2, c.139A>C, p.Thr47Pro and a newly described one in exon 11, c.1219_1220delCT, p.Leu407Aspfs*23.

Both the c.139A>C and c.1090C>T mutations are known disease causing mutations described previously, c.1219_1220delCT is a frame shift mutation with subsequent premature stop codon formation.

Of the 15 patients summarized here (3 newly described, 12 reported in literature [8, 9, 14–20] – including 2 published by our group [11]), three are homozygous (including one pair of siblings) and 12 compound heterozygous (including three pairs of siblings). Of the 26 alleles detected, 9 (35 %) are c.139A>C (Table 1). If we add the analyses of the deceased siblings (Patient 1 and Patient 3), there would be 30 alleles in total, and 12 (40 %) c.139A>C.

In summary, Table 1 shows features of 15 ALG8-CDG patients in total. There are multiple prenatal abnormalities in 6/12 patients: 3 had intrauterine growth retardation, 5 oligohydramnios and 3 hydrops fetalis. In 13/15 patients symptoms were recorded at birth and 9/15 died within the first year of life, showing the severity of ALG8 deficiency. Birth weight was appropriate for gestational age in 11/12 patients and only one patient was small for gestational age (birth weight <10th centile). Prematurity (<37 weeks) was reported in 7/12 patients. Hydrops fetalis was noticed only in three patients, whereas edema was present in 11/13. Common dysmorphic signs were: low set ears, macroglossia, hypertelorism, pes equinovarus, campto- and brachydactyly (13/15). Gastrointestinal symptoms included diarrhea, vomiting, feeding problems

with failure to thrive, protein-losing enteropathy (9/14 patients). Brain involvement included psychomotor disability, seizures, ataxia, and structural abnormalities (dilatation of ventricles, corpus callosum hypoplasia, leukoencephalopathy/leukodystrophy, cortical/cerebral atrophy) in 12/13 patients. Hypotonia/'floppy infant' was present in 13/14 patients. Thrombocytopenia was present in 9/9 patients. Most patients had liver pathology: elevated serum transaminases in 8/11 and abnormal clotting factors in 8/11. Eye involvement (especially cataracts) was reported in 9/14 patients and skin involvement like fat pads, wrinkly skin, cutis laxa and inverted nipples was reported in 8/13 patients.

Discussion

ALG8-CDG is one of the less frequently reported CDG syndromes. It affects an early step of protein glycosylation and in general has a poor prognosis. It was first reported by Chantret *et al.* in 2003 [8]. There are twelve patients described to date (Skladal *et al.* [11] (n = 2), Charlwood *et al.* [14] (n = 1), Chantret *et al.* [8] (n = 1), Schollen *et al.* [15] (n = 2), Eklund *et al.* [9] (n = 1), Vesela *et al.* [16] (n = 1), Stölting *et al.* [17] (n = 2), Sorte *et al.* [18] (n = 1), and one patient reported by both Funke *et al.* [19] and Kouwenberg *et al.* [20] (Table 1). In the majority of patients (13/15), the disease had a neonatal onset and rapid progression. In 9/15 patients, the outcome was fatal within the first year of life (Table 1). So far, no adult patients (>18 years of age) affected with this subtype have been reported [21].

We reviewed the most common clinical, laboratory and genetic findings in the twelve previously reported patients diagnosed with ALG8-CDG (Table 1), add here the data of three additional patients with ALG8-CDG (Patient 1,2 and 5), and provide an update on two previously reported patients (Patient 3 and 4). For the two patients reported by Charlwood *et al.* [14] and the two reported by Skladal *et al.* (Patient 3 and 4) [11], the diagnosis of ALG8-CDG was provided years later.

In the series of 15 patients summarized in this paper, there are four pairs of siblings (Schollen [15] plus the patient reported by Charlwood *et al.* [14], Stölting [17], Patient 1/2, and Patient 3/4 in this paper) (Table 1), reflecting an ascertainment bias for firstborns deceased without diagnosis being established before a second affected child was born.

Hydrops fetalis has also been described in PMM2-CDG and ALG1-CDG [1]. In the patients described here, it was present in the two brothers (Patient 1 and 2) and in 1/12 additional patient reported in the literature (Table 1). Hyponatremia reported in eight patients is probably secondary to ascites/edema (11 patients). Unlike other CDG types, cataracts (ocular findings) have only been reported in ALG8-CDG, ALG12-CDG, other

subtypes in early LLO synthesis (like SRD5A3-CDG) and some unclassified CDG [9, 10].

CDG are a clinically and genetically heterogeneous group of disorders. Patients with ALG8-CDG show very similar clinical features with neonatal onset and fatal outcome within the first year of life. We believe that this CDG disorder might be underdiagnosed. Early discriminative symptoms in ALG8-CDG patients are multisystem involvement with dysmorphism, neurological symptoms and cataracts in about 50 % of cases. Therefore, we suggest screening for CDG sialotransferrins in serum or plasma in all infants with early multi-organ involvement (such as cataracts, non-immune hydrops, coagulopathy). Confirmation of diagnosis requires molecular genetic testing.

The diagnosis of CDG is frequently delayed due to the highly variable phenotype, with some cases showing single organ involvement and others mimicking syndromes, like skeletal dysplasia, cutis laxa syndrome, or congenital muscle dystrophy.

In MPI-CDG (=CDG-Ib) (phosphomannose isomerase deficiency), the protein-losing enteropathy and hypoglycaemia can be treated with an oral uptake of mannose [22], in SLC35C1-CDG (=CDG-IIc), the leukocyte adhesion deficiency can be treated with oral fucose [23], and in PGM1 (Phosphoglucomutase 1 deficiency), treatment with galactose leads to improved indexes of glycosylation [24]. For ALG8-CDG, there is currently no curative treatment available. However, establishing a molecular diagnosis is important for family counselling and prenatal diagnosis [25].

Informed consent

All procedures were in accordance with ethical standards of the Helsinki Declaration. Written informed consent for the case reports and images was obtained from all parents of the five patients reported.

Abbreviations

ALG8-CDG: Alpha-1,3-Glucosyltransferase CDG; CDG: Congenital disorder of glycosylation; IUGR: Intrauterine growth restriction; AGA/SGA: Appropriate for gestational age/small for gestational age.

Competing interests

The authors declare that they have no competing interests.

Authors' contributions

MH, KW, ER, UK, KP, DK treated Patient 1, 2, and 5 on the neonatal intensive care unit and the pediatric ward. DK, SSB provided expertise for metabolic diagnostic work-up; together with MH drafted the manuscript, collected data on the patients, and reviewed the literature for data on other known ALG8-CDG patients. CF and ES provided the dysmorphological description of patients. CF, ES, DL, GM, LK, JZ provided information and details on biochemical and molecular genetic analyses. KM performed the eye ultrasound, as well as other radiological examinations, and provided Fig. 3. All authors read and approved the final manuscript.

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References

- Hennet T, Cabalzar J. Congenital disorders of glycosylation: a concise chart of glycolyx dysfunction. *Trends Biochem Sci*. 2015. doi:10.1016/j.tibs.2015.03.002 (Epub ahead of print).
- Jaeken J, Vanderschueren-Lodeweyckx M, Casaer P, Snoeck L, Corbeel L, Eggemont E, et al. Familial psychomotor retardation with markedly fluctuating serum prolactin, FSH and GH levels, partial TBG deficiency, increased serum arylsulfatase A and increased CSF protein: a new syndrome? *Pediatr Res*. 1980;14:179.
- Jaeken J. Congenital disorders of glycosylation. *Ann NY Acad Sci*. 2010;1214:190–8.
- Freeze HH. Understanding human glycosylation disorders: biochemistry leads the charge. *J Biol Chem*. 2013;288:6936–45.
- Lefeber DJ, Morava E, Jaeken J. How to find and diagnose a CDG due to defective N-glycosylation. *J Inher Metab Dis*. 2011;34:849–52.
- Jaeken J. CDG: it's (nearly) all in it! *J Inher Metab Dis*. 2011;34:853–8.
- Jaeken J, Hennet T, Freeze HH, Matthijs G. On the nomenclature of congenital disorders of glycosylation. *J Inher Metab Dis*. 2008;31:669–72.
- Chantret I, Dancourt J, Dupré T, Delenda C, Bucher S, Vuillaumier-Barrot S, et al. A deficiency in dolichyl-P-glucose: Glc1Man9GlcNAc2-PP-dolichyl α 3-glucosyltransferase defines a new subtype of congenital disorders of glycosylation. *J Biol Chem*. 2003;278:9962–71.
- Eklund EA, Sun L, Westphal V, Nothrop JL, Freeze HH, Scaglia F. Congenital disorder of glycosylation (CDG)-Ih patient with a severe hepato-intestinal phenotype and evolving central nervous system pathology. *J Pediatr*. 2005;147:47–50.
- Morava E, Wosik HN, Sykut-Cegielska J, Adamowicz M, Guillard M, Wevers RA, et al. Ophthalmological abnormalities in children with congenital disorders of glycosylation type I. *Br J Ophthalmol*. 2009;93:350–4.
- Skladal D, Sperl W, Henry H, Bachmann C. Congenital cataract and familiar brachydactyly in carbohydrate-deficient glycoprotein syndrome. *J Inher Metab Dis*. 1996;19:251–2.
- Hackler R, Arndt T, Peters V, Assmann B, Hoffmann GF, Steinmetz A. Automated isoelectric focussing of serum isotransferrins for laboratory diagnosis of carbohydrate-deficient glycoprotein syndrome using the PhastSystemTM. *Scand J Clin Lab Invest*. 1998;58:66–7.
- Imbach T, Grünwald S, Schenk B, Burda P, Schollen E, Wevers RA, et al. Multi-allelic origin of congenital disorder of glycosylation (CDG)-Ic. *Hum Genet*. 2000;106:538–45.
- Charlwood J, Clayton P, Johnson A, Keir G, Mian N, Winchester B. A case of the carbohydrate-deficient glycoprotein syndrome type 1 (CDGS type 1) with normal phosphomannomutase activity. *J Inher Metab Dis*. 1997;8:17–27.
- Schollen E, Frank CG, Keldermans L, Reyntjens R, Grubenmann CE, Clayton PT, et al. Clinical and molecular features of three patients with congenital disorders of glycosylation type Ih (CDG-Ih) (ALG8 deficiency). *J Med Genet*. 2004;41:550–6.
- Vesela K, Honzik T, Hansikova H, Haeuptle MA, Semberova J, Stranak Z, et al. A new case of ALG8 deficiency (CDG Ih). *J Inher Metab Dis*. 2009;32:259–64.
- Stölting T, Omran H, Erlekotte A, Denecke J, Reunert J, Marquardt T. Novel ALG8 mutations expand the clinical spectrum of congenital disorder of glycosylation type Ih. *Mol Genet Metab*. 2009;98:305–9.
- Sorte H, Mørkrid L, Rødningen O, Kulseth MA, Stray-Petersen A, Matthijs G, et al. Severe ALG8-CDG (CDG-Ih) associated with homozygosity for two novel missense mutations detected by exome sequencing of candidate genes. *Eur J Med Genet*. 2012;55:196–202.
- Funke S, Gardeitchik T, Kouwenberg D, Mohamed M, Wortmann SB, Korsch E, et al. Early infantile symptoms in congenital disorders of glycosylation. *Am J Med Genet Part A*. 2012;161A:578–84.
- Kouwenberg D, Gardeitchik T, Mohamed M, Lefeber D, Morava E. Wrinkled skin and fat pads in patients with ALG8-CDG: Revisiting skin manifestations in congenital disorders of glycosylation. *Pediatr Dermatol*. 2014;31:1–5.
- Wolthuis DFGJ, Janssen MC, Cassiman D, Lefeber DJ, Morava-Kozicz E. Defining the phenotype and diagnostic considerations in adults with congenital disorders of N-linked glycosylation. *Expert Rev Mol Diagn*. 2014;14:217–24.
- de Lonlay P, Seta N. The clinical spectrum of phosphomannose isomerase deficiency, with an evaluation of mannose treatment for CDG-Ib. *Biochim Biophys Acta*. 2009;1792:841–3.
- Gazit Y, Mory A, Etzioni A, Frydman M, Scheuerman O, Gershoni-Baruch R, et al. Leukocyte adhesion deficiency type II: long-term follow-up and review of the literature. *J Clin Immunol*. 2010;30:308–13.
- Tegtmeyer LC, Rust S, van Scherpenzeel M, Ng BC, Losfeld ME, Timal S, et al. Multiple phenotypes in phosphoglucomutase 1 deficiency. *New Engl J Med*. 2014;370:533–42.
- Edwards M, McKenzie F, O'Callaghan S, Somerset D, Woodford P, Spilsbury J, Fietz M, et al. Prenatal diagnosis of congenital disorder of glycosylation type Ia (CDG-Ia) by cordocentesis and transferrin isoelectric focussing of serum of a 27-week fetus with non-immune hydrops. *Prenat Diagn*. 2006;26:985–8.

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