

PDF hosted at the Radboud Repository of the Radboud University Nijmegen

The following full text is a publisher's version.

For additional information about this publication click this link.

<http://hdl.handle.net/2066/152882>

Please be advised that this information was generated on 2017-12-05 and may be subject to change.

Transmission and Progression to Disease of *Mycobacterium tuberculosis* Phylogenetic Lineages in The Netherlands

Hanna Nebenzahl-Guimaraes,^{a,b,c} Lilly M. Verhagen,^{d,e} Martien W. Borgdorff,^{f,g} Dick van Soolingen^{a,h}

National Institute for Public Health and the Environment (RIVM), Bilthoven, The Netherlands^a; Life and Health Sciences Research Institute (ICVS), School of Health Sciences, University of Minho, Braga, Portugal^b; ICVS/3Bs, PT Government Associate Laboratory, Braga/Guimarães, Portugal^c; Wilhelmina Children's Hospital Utrecht, Utrecht, The Netherlands^d; Laboratory of Pediatric Infectious Diseases, Radboud University Medical Centre, Nijmegen, The Netherlands^e; Public Health Service, Amsterdam, The Netherlands^f; Department of Clinical Epidemiology, Academic Medical Centre, University of Amsterdam, Amsterdam, The Netherlands^g; Department of Medical Microbiology, Radboud University Nijmegen Medical Centre, Nijmegen, The Netherlands^h

The aim of this study was to determine if mycobacterial lineages affect infection risk, clustering, and disease progression among *Mycobacterium tuberculosis* cases in The Netherlands. Multivariate negative binomial regression models adjusted for patient-related factors and stratified by patient ethnicity were used to determine the association between phylogenetic lineages and infectivity (mean number of positive contacts around each patient) and clustering (as defined by number of secondary cases within 2 years after diagnosis of an index case sharing the same fingerprint) indices. An estimate of progression to disease by each risk factor was calculated as a bootstrapped risk ratio of the clustering index by the infectivity index. Compared to the Euro-American reference, *Mycobacterium africanum* showed significantly lower infectivity and clustering indices in the foreign-born population, while *Mycobacterium bovis* showed significantly lower infectivity and clustering indices in the native population. Significantly lower infectivity was also observed for the East African Indian lineage in the foreign-born population. Smear positivity was a significant risk factor for increased infectivity and increased clustering. Estimates of progression to disease were significantly associated with age, sputum-smear status, and behavioral risk factors, such as alcohol and intravenous drug abuse, but not with phylogenetic lineages. In conclusion, we found evidence of a bacteriological factor influencing indicators of a strain's transmissibility, namely, a decreased ability to infect and a lower clustering index in ancient phylogenetic lineages compared to their modern counterparts. Confirmation of these findings via follow-up studies using tuberculin skin test conversion data should have important implications on *M. tuberculosis* control efforts.

Curbing tuberculosis (TB) transmission is a challenge in high-burden countries. However, even in low-prevalence settings, controlling TB is an important requirement due to human migration from higher-incidence areas to Western countries (1). In Western countries, studies on transmission are more feasible, as all cases undergo extended diagnostic algorithms and all clinical and demographic data are recorded. Current molecular typing methods, such as variable number of tandem repeat (VNTR) typing and restriction fragment length polymorphism (RFLP) typing, allow identification of clusters of *Mycobacterium tuberculosis* isolates with identical genotypes that, in population-based studies, reveal recent transmission (2, 3). Spoligotyping and VNTR typing can identify the genotype family of the isolate, revealing bacterial variation via the identification of phylogenetic lineages (4, 5).

While many studies have elucidated the variation in the disease's spread and outcome attributable to host and environmental factors, there is also evidence that bacterial factors may affect the spread of tuberculosis (6). In The Netherlands, for example, one study showed that the number of positive contacts around a case increases with growing cluster size (7). In a subsequent study in the same setting, cluster size growth was not different between phylogenetic lineages after controlling for host risk factors (8). However, this study could not distinguish between transmission rates and progression to disease. There are, however, indications that progression to disease is partly dependent on bacterial variation. It has, e.g., been postulated that some *Mycobacterium africanum* strains might transmit equally well as other *M. tuberculosis* complex strains but might be less associated with progression to disease (9). We will refer to these two properties that affect the

degree of clustering as infectivity (the bacterium's ability to establish an initial infection in the human host) and progression to disease (the bacterium's capacity to produce disease) (10).

In the low-incidence context of The Netherlands, with a globally representative cohort of patients, we aim to determine differences in indices of infectivity, clustering, and estimated progression to disease of different mycobacterial lineages using fingerprinting data and contact investigation. This will provide insights into the role of bacteriological factors in TB transmission, which itself may affect future TB control measures.

MATERIALS AND METHODS

Data collection and DNA fingerprinting. The National Institute for Public Health and the Environment (RIVM) is a reference laboratory for secondary laboratory diagnosis of all TB cases in The Netherlands, offer-

Received 20 May 2015 Returned for modification 19 June 2015
Accepted 23 July 2015

Accepted manuscript posted online 29 July 2015

Citation Nebenzahl-Guimaraes H, Verhagen LM, Borgdorff MW, van Soolingen D. 2015. Transmission and progression to disease of *Mycobacterium tuberculosis* phylogenetic lineages in The Netherlands. *J Clin Microbiol* 53:3264–3271. doi:10.1128/JCM.01370-15.

Editor: G. A. Land

Address correspondence to Hanna N. Guimaraes, hanna.guimaraes@gmail.com.

Copyright © 2015, American Society for Microbiology. All Rights Reserved.

doi:10.1128/JCM.01370-15

ing identification, drug susceptibility testing, and molecular typing for each TB case. DNA fingerprints of all nationwide *M. tuberculosis* complex isolates and their cluster statuses have been stored in an RFLP/VNTR database since 1993. The registration committee of The Netherlands Tuberculosis Register (NTR) approved this retrospective study and provided anonymized demographic and clinical information for patients. Because these data are deidentified by name, DNA fingerprinting results were matched by sex, date of birth, year of diagnosis, and postal code. All notified culture-positive cases of *M. tuberculosis* between 1993 and 2011 were included in the study. For patients with multiple isolates sharing identical fingerprints, only the isolate with the earliest diagnosis date was included. Contaminating isolates were excluded.

Isolates recovered from patients between 1993 and 2009 underwent IS6110 typing and polymorphic GC-rich sequence (PGRS) RFLP typing ($n = 15,073$), and those from 2004 onward were subjected to VNTR typing ($n = 5,870$) (11, 12). In the period of 2004 to 2008, both RFLP and 24-locus VNTR typing were performed to obtain a smooth transition in typing methods and to evaluate VNTR typing performance (3). In addition, 4,433 randomly selected isolates were spoligotyped ($n = 4,433$). We defined a cluster as a group of patients who shared *M. tuberculosis* isolates with identical RFLP or VNTR patterns or, if strains had fewer than five IS6110 copies, identical PGRS RFLP patterns.

Conventional contact investigation. Systematic contact investigation by TB Public Health Services in The Netherlands is conducted per the stone-in-the-pond principle, in which the decision to extend conventional contact investigation to the next ring of contacts is based on the prevalence of infection in the investigated ring (13). Contacts are defined by the frequency and intimacy of their contacts with the TB index case. The tuberculin skin test (TST) is used to investigate presumably exposed contacts. If the number of TST-positive contacts in the first ring suggests a high spread of tuberculosis, a larger ring of contacts is investigated. We have defined positive contacts as contacts with a TST induration ≥ 10 mm and/or contacts who received a diagnosis of TB disease. If contact investigations become very large, identified TB infections and secondary cases are less likely to be related to the index case. To minimize the probability that positive contacts in our research were unrelated to the defined index case, we only included contacts in the first ring around the index patient. First-ring contacts are defined as contacts that are physically close to the index patient, considering environmental factors, such as room size, ventilation, air purification, and air circulation. In addition, the patient and the contact must be able to indicate where they met and must have a long-standing relationship to qualify as a first-ring contact. Examples of first-ring contacts are household members, close work colleagues, and close friends.

Classification into phylogenetic lineages. The phylogenetic lineages of isolates were determined using a combination of spoligotyping, the MIRU (mycobacterial interspersed repetitive unit) best match analysis offered by the MIRU-VNTRplus online tool, and RFLP similarity, as described in a previous study using the same data set (8, 14). Three species (*M. africanum*, *Mycobacterium bovis*, and *M. tuberculosis*) and four major phylogenetic lineages of *M. tuberculosis* were identified: the Euro-American, Central Asian strain (CAS), East African Indian (EAI), and Beijing genotypes. Strains not assigned a phylogenetic lineage or assigned to multiple major phylogenetic families per cluster were not analyzed. Strains classified as either T or U (unknown) also were excluded due to the ambiguity of these classifications.

Definitions. For our infectivity index, we took the mean number of positive contacts around each patient who underwent contact investigation. We excluded patients with missing data on contact investigation or those who had zero contacts investigated, as well as those for whom we lacked ethnicity information. Because TB transmission almost exclusively results from patients with pulmonary TB, we also excluded patients with extrapulmonary TB, leaving us with a total of 2,809 cases (Fig. 1).

For our clustering index, we used the number of secondary cases occurring within 2 years of the index case diagnosis. The 2-year cutoff has

been shown to best reflect recent transmission as opposed to disease reactivation (1, 15). We defined index cases as patients who had strains with RFLP or VNTR patterns not seen in other patients in the previous 2 years. We searched for index cases based on RFLP-typing data from 1995 to 2007 and for index cases based on VNTR typing from 2007 to 2009. We excluded RFLP-defined index cases from 1993 and 1994 and VNTR-defined index cases from 2005 and 2006 ($n = 2,684$), because we could not determine whether the strains of these index cases were uncovered in the previous 2 years. Similarly, we excluded RFLP-defined index cases occurring after 2007 and VNTR-defined index cases occurring after 2009 ($n = 950$), because we could not follow these index cases for a full 2 years. Secondary cases from these index cases (included in the counts) were also excluded. Finally, we excluded cases between 1995 and 2007 occurring < 2 years after a previous patient with the same RFLP fingerprint yet diagnosed > 2 years after a cluster's start ($n = 722$) and cases occurring between 2007 and 2009 that occurred < 2 years after a previous patient with the same VNTR fingerprint yet > 2 years after a cluster's start ($n = 40$). After excluding extrapulmonary cases, 4,432 patients remained: 2,881 nonclustered index patients, 607 index patients who were the first patient of a cluster, and 944 secondary cases within 2 years of a cluster's start (Fig. 1).

Finally, estimates of progression to disease were calculated as risk ratios (RR) of the population risk of disease given exposure to a risk factor by the population risk of infection given exposure to the same risk factor (dividing the clustering odds ratios [ORs] by the infectivity ORs).

Statistical analysis. We used a multivariate negative binomial regression model to determine the association between phylogenetic lineages and the infectivity and clustering indices. Since TST is poorly specific among *Mycobacterium bovis* BCG contacts and positive TSTs may represent old infections, we divided our data sets into native and foreign-born (FB) cohorts in order to address important differences between the two: FB patients are often BCG vaccinated (in contrast to native Dutch patients, who are not), while the prevalence of infection is higher among FB patients. Second-generation patients (born to FB patients) were included in the native cohorts, given that, like native patients, they are not BCG vaccinated and they have already been born in a setting of lower prevalence of infection. Studies carried out in The Netherlands have also previously demonstrated that contact investigation practices vary by demographic characteristics of the index patient (16). As such, in both analyses, we adjusted for index patient-related factors, including demographic, behavioral, and sputum smear status. In addition, the logarithm of the number of investigated contacts around a source case was used as an offset in the multivariate model assessing the association between phylogenetic lineages and the spread index, since the greater number of contacts around a source are investigated, the likelier it is to detect TST positive contacts. Variables with P values of ≤ 0.20 were entered into the multivariate model. Crude and adjusted ORs are presented with 95% confidence intervals (CIs). Estimates of TB progression were calculated for any risk factor that was significant in either multivariate regression model. To calculate the variance for the estimate of TB progression, we performed a bootstrapping procedure, running our multivariate negative binomial regression models 10,000 times on bootstrapped data sets. The median of the resulting 10,000 RRs was used as the estimate of TB progression, while the 2.5th and 97.5th percentiles were used as the 95% cutoffs for the estimate CI. All analyses were conducted using SAS (Windows version 9.3), SPSS program for Windows version 20.0 (SPSS Inc., Chicago, IL, USA) and R (version 3.1.2 for Windows).

RESULTS

Between January 1993 and December 2011, 18,294 isolates were collected from the same number of notified TB cases in The Netherlands, and their clustering statuses were ascertained, of which 15,601 (85%) were successfully matched with the NTR data. Of these, 15,224 (98%) were noncontaminated *M. tuberculosis* cul-

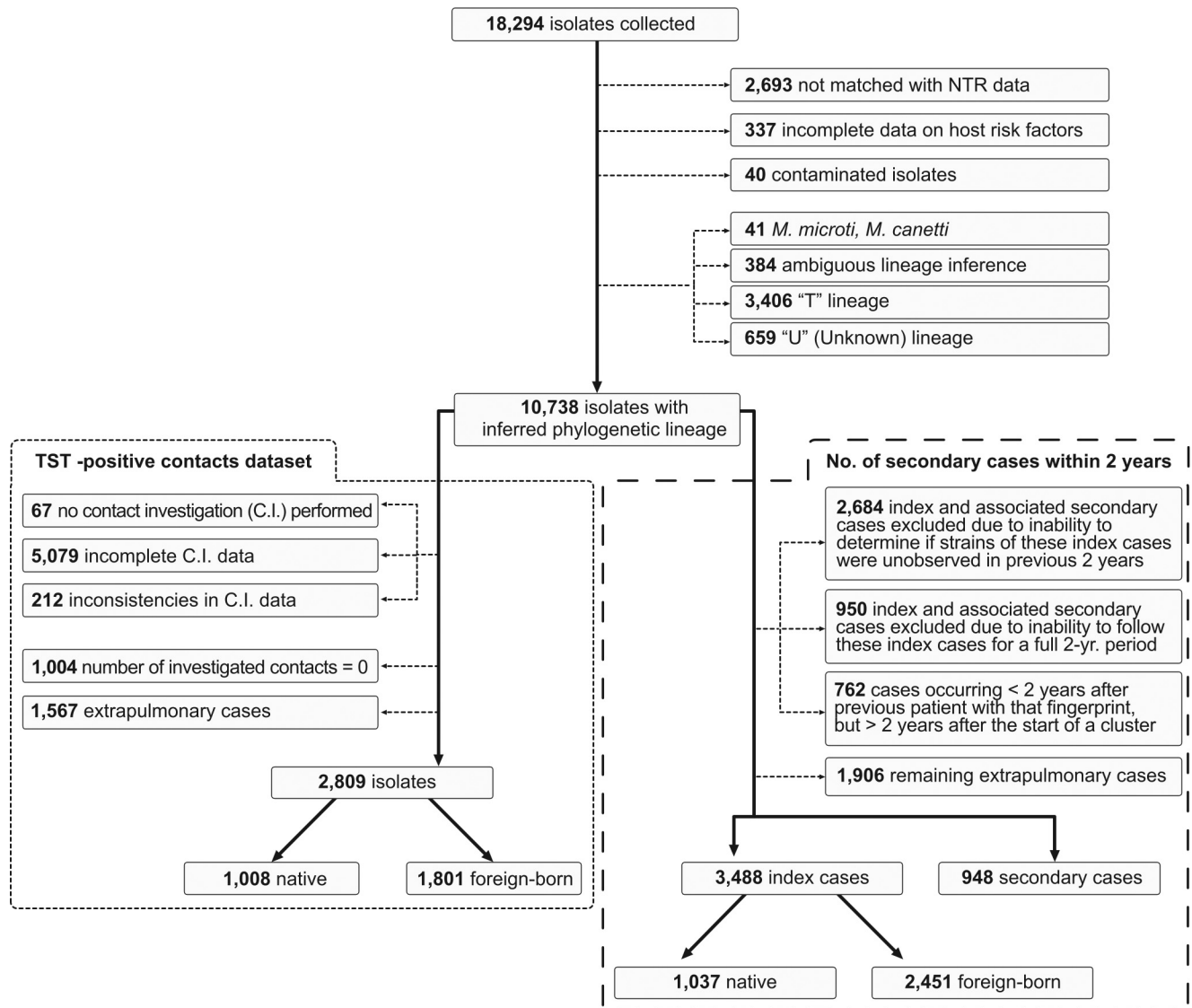


FIG 1 Flow-diagram of exclusion criteria applied to data set.

tures with completely ascertained information on host risk factors. After phylogenetic lineage assignment, there were 10,738 isolates that were *M. bovis*, *M. africanum*, or *M. tuberculosis* of the Euro-American, Beijing, CAS, or EAI lineages (Fig. 1). The mean age of the patients carrying these strains was 41 years (standard deviation, 20 years); 6,394 (60%) were male; and 7,762 (72%) were foreign born.

Mycobacterial genotypes. The Euro-American lineage was predominant in both the infectivity (78% in native cohort; 56% in FB cohort) and clustering (79% in native cohort; 64% in FB cohort) data sets. In contrast, both *M. africanum* and *M. bovis* represented less than 1% of all cases in the infectivity data set. In the clustering data set, both *M. africanum* and *M. bovis* represented only 2% of all cases (Tables 1 and 2).

Infectivity by mycobacterial lineage. The proportion of cases in which a contact investigation was performed in The Netherlands was approximately equal between lineages, though slightly lower in the FB cohort for Beijing and EAI compared to the native

counterpart (Fig. 2). The average number of TST-positive contacts declined significantly in the >65 years age category in the native cohort and in the <20 years age category in the FB cohort. Smear positivity was associated with an increased average number of TST-positive contacts in both native and FB cohorts. There were no significant differences in infectivity by gender, homelessness, and alcohol use in the two cohorts, although use of intravenous drugs in the native populations and rural residence in the FB population were associated with a decreased average number of TST-positive contacts. The mean number of TST-positive contacts around an index case was significantly lower for *M. bovis* than for the Euro-American reference lineage in the native population in multivariable analysis. In the FB population, *M. africanum* and EAI presented a significantly lower number of TST-positive contacts (Table 1).

Clustering by mycobacterial lineage. The number of secondary cases declined significantly with increasing age (>65 years) in both the native and FB cohorts. Smear positivity was also associ-

TABLE 1 Risk factors among native and foreign-born index cases for infectivity (number of TST-positive contacts per index case)

Characteristic	Native cohort				Foreign-born cohort					
	No. of index cases	Mean no. of TST-positive contacts/index case	Univariate analysis Relative no. (95% CI)	P	Multivariate analysis, relative no. (95% CI)	No. of index cases	Mean no. of TST-positive contacts/index case	Univariate analysis Relative no. (95% CI)	P	Multivariate analysis, relative no. (95% CI)
Age, yr				0.00					0.033	
0–19	99	1.02	1.0 (0.68–1.6)		0.96 (0.65–1.4)	200	0.83	1.11 (0.83–1.5)		0.69 (0.53–0.89)
20–39	363	0.99	1 (Ref) ^a		1 (Ref)	1024	0.75	1 (Ref)		1 (Ref)
40–64	339	0.82	0.83 (0.63–1.1)		0.91 (0.69–1.2)	459	0.60	0.81 (0.65–1.0)		0.96 (0.79–1.2)
≥65	207	0.51	0.51 (0.37–0.72)		0.50 (0.36–0.70)	118	0.49	0.66 (0.45–0.97)		0.80 (0.57–1.1)
Sex				0.15					0.32	
Male	635	0.78	1 (Ref)		1 (Ref)	1,125	0.73	1 (Ref)		
Female	373	0.93	1.2 (0.93–1.5)		1.0 (0.82–1.3)	676	0.66	0.73 (0.65–0.82)		
Smear positivity				<0.001					<0.001	
Negative	395	0.52	0.50 (0.39–0.65)		0.48 (0.38–0.62)	754	0.47	0.54 (0.45–0.65)		0.55 (0.46–0.65)
Positive	613	1.04	1 (Ref)		1 (Ref)	1,047	0.87	1 (Ref)		1 (Ref)
Lineage				0.00					0.006	
Euro-American	786	0.91	1 (Ref)		1 (Ref)	1,157	0.75	1 (Ref)		1 (Ref)
Beijing	112	0.67	0.73 (0.50–1.1)		1.1 (0.77–1.6)	182	0.55	0.74 (0.54–1.0)		0.94 (0.71–1.2)
CAS	30	0.23	0.26 (0.11–0.61)		0.64 (0.27–1.5)	198	0.86	1.2 (0.86–1.5)		1.0 (0.79–1.3)
EAI	62	0.56	0.62 (0.37–1.04)		0.78 (0.47–1.3)	237	0.53	0.71 (0.54–0.94)		0.64 (0.49–0.83)
<i>M. africanum</i>	5	0.40	0.44 (0.067–2.9)		0.44 (0.055–3.5)	15	0.27	0.36 (0.11–1.1)		0.30 (0.10–0.89)
<i>M. bovis</i>	13	0.23	0.25 (0.068–0.94)		0.23 (0.059–0.94)	12	0.17	0.22 (0.052–0.96)		0.51 (0.11–2.4)
Residency				0.07					0.002	
Urban	690	0.90	1 (Ref)		1 (Ref)	1,067	0.79	1 (Ref)		1 (Ref)
Rural	318	0.70	0.78 (0.60–1.0)		0.78 (0.60–1.0)	734	0.58	0.74 (0.62–0.89)		0.71 (0.60–0.84)
Alcohol abuse				0.19					0.69	
No	969	0.85	1 (Ref)		1 (Ref)	1,779	0.70	1 (Ref)		
Yes	39	0.54	0.64 (0.33–1.2)		0.59 (0.30–1.2)	22	0.59	0.84 (0.36–1.9)		
Drug abuse				0.01					0.97	
No	953	0.86	1 (Ref)		1 (Ref)	1,722	0.70	1 (Ref)		
Yes	55	0.40	0.47 (0.26–0.83)		0.43 (0.24–0.78)	79	0.71	1.0 (0.65–1.6)		
Traveler to country of endemicity				0.17					0.17	
No	973	0.85	1 (Ref)		1 (Ref)	1,759	0.69	1 (Ref)		1 (Ref)
Yes	35	0.51	0.61 (0.30–1.2)		0.53 (0.25–1.1)	42	1.02	1.5 (0.83–2.6)		1.5 (0.88–2.4)
Homeless				0.72					0.94	
No	978	0.84	1 (Ref)			1,753	0.70	1 (Ref)		
Yes	30	0.73	0.88 (0.43–1.8)			48	0.69	0.98 (0.56–1.7)		
Site of disease				0.21					0.099	
Pulmonary	891	0.86	1 (Ref)		1 (Ref)	1,430	0.73	1 (Ref)		1 (Ref)
Pulmonary + extrapulmonary	117	0.67	0.78 (0.53–1.1)		1.0 (0.70–1.5)	371	0.60	0.83 (0.66–1.0)		0.88 (0.71–1.1)

^a Ref, reference.

ated with an increased number of secondary cases in both cohorts, and female gender was associated with an increased number of secondary cases only among the FB. Rural residence was associated with a decreased number of secondary cases only in the FB cohort. Relative to the Euro-American reference in the multivariable analysis, the number of secondary cases was significantly lower for *M. bovis* in the native-born population and for *M. africanum* in the FB population (Table 2).

Estimates of progression to disease by mycobacterial lineage.

Estimates of progression to disease were significantly lower in the >65 years age category in both ethnic cohorts and significantly higher in the 0- to 19-years age category in the FB cohort. Additionally, in the FB-born population, estimates of progression to disease were significantly lower in smear-negative patients. Both alcohol and drug abuse were significantly associated with higher estimates in the native population. No

TABLE 2 Risk factors among native and foreign-born index cases for clustering (number of secondary cases within 2 years of an index case)

Characteristic	Native cohort					Foreign-born cohort				
	No. of index cases	Mean no. of second cases per index case	Univariate analysis		Multivariate analysis, relative no. (95% CI)	No. of index cases	Mean no. of second cases per index case	Univariate analysis		Multivariate analysis, relative no. (95% CI)
			Relative no. (95% CI)	P				Relative no. (95% CI)	P	
Age, yr										
0–19	59	0.46	1.02 (0.53–1.90)	0.001	1.04 (0.56–1.93)	276	0.36	1.32 (0.92–1.92)	0.716	1.42 (0.99–2.03)
20–39	216	0.45	1 (Ref ^a)		1 (Ref)	1,411	0.27	1 (Ref)		1 (Ref)
40–64	267	0.34	0.65 (0.41–0.98)		0.68 (0.46–1.02)	569	0.31	1.14 (0.86–1.51)		1.06 (0.80–1.42)
≥65	495	0.1	0.22 (0.12–0.33)		0.21 (0.13–0.32)	195	0.08	0.30 (0.16–0.55)		0.30 (0.16–0.55)
Sex										
Female	387	0.23	0.87 (0.61–1.23)	0.4255		975	0.23	0.51 (0.29–0.90)	0.019	1.29 (1.01–1.66)
Male	650	0.27	1 (Ref)			1,476	0.31	1 (Ref)		1 (Ref)
Smeared positivity										
Negative	393	0.18	0.58 (0.40–0.83)	0.0027	0.62 (0.44–0.88)	1,076	0.17	0.48 (0.38–0.61)	<.0001	0.50 (0.39–0.65)
Positive	644	0.3	1 (Ref)		1 (Ref)	1,375	0.36	1 (Ref)		1 (Ref)
Lineage										
Euro-American	756	0.25	1 (Ref)	0.1971	1 (Ref)	1,385	0.3	1 (Ref)	0.081	1 (Ref)
Beijing	73	0.36	1.44 (0.79–2.62)		1.20 (0.68–2.11)	354	0.28	0.93 (0.66–1.32)		1.07 (0.76–1.52)
CAS	28	0.29	1.15 (0.41–3.09)		0.79 (0.30–2.10)	290	0.28	0.93 (0.64–1.36)		0.87 (0.59–1.27)
EAI	118	0.33	1.34 (0.82–2.19)		0.90 (0.54–1.50)	335	0.22	0.74 (0.51–1.07)		0.80 (0.57–1.17)
<i>M. africanum</i>	8	0.13	0.50 (0.04–5.22)		0.26 (0.03–2.53)	65	0.15	0.52 (0.22–1.21)		0.47 (0.31–0.94)
<i>M. bovis</i>	54	0.06	0.22 (0.06–0.78)		0.11 (0.006–0.56)	22	0.09	0.30 (0.06–1.67)		0.31 (0.06–1.67)
Residency										
Urban	785	0.35	1 (Ref)	0.02	1 (Ref)	1,547	0.34	1 (Ref)	0.004	1 (Ref)
Rural	252	0.23	0.65 (0.45–0.94)		0.99 (0.69–1.43)	904	0.24	0.70 (0.55–0.89)		0.77 (0.61–0.99)
Alcohol abuse										
No	1,009	0.24	1 (Ref)	0.02	1 (Ref)	2,430	0.28	1 (Ref)	0.964	
Yes	28	0.68	2.8 (1.22–6.42)		1.88 (0.89–3.99)	21	0.29	1.93 (0.28–3.74)		
Drug abuse										
No	1,017	0.25	1 (Ref)	0.05	1 (Ref)	2,398	0.28	1 (Ref)	0.905	
Yes	20	0.65	2.64 (0.98–7.03)		1.42 (0.58–3.49)	53	0.26	1.53 (0.42–2.18)		
Traveler to country of endemicity										
No	988	0.26	1 (Ref)	0.07	1 (Ref)	2,391	0.28	1 (Ref)	0.167	1 (Ref)
Yes	49	0.10	0.39 (0.14–1.09)		0.26 (0.09–0.70)	60	0.15	0.53 (0.22–1.30)		0.47 (0.20–1.14)
Homeless										
No	1,028	0.25	1 (Ref)	0.75		2,401	0.27	1 (Ref)	0.233	1 (Ref)
Yes	9	0.33	1.31 (0.25–6.99)			50	0.44	1.61 (0.74–3.49)		1.19 (0.5–2.56)
Site of disease										
Pulmonary	883	0.26	1 (Ref)	0.22	1 (Ref)	1,949	0.29	1 (Ref)	0.129	1 (Ref)
Pulmonary + extrapulmonary	154	0.19			0.94 (0.58–1.53)	502	0.23	0.79 (0.58–1.07)		0.92 (0.67–0.99)

^a Ref, reference.

significant differences were found across phylogenetic lineages (Table 3).

DISCUSSION

In this study, we observed variations between the infectivity and clustering indices of different phylogenetic subgroups of *M. tuberculosis*, *M. bovis*, and *M. africanum* after controlling for clinical and demographic index host factors. *M. africanum* and *M. bovis*

showed both significantly lower infectivity and clustering indices in the FB and native populations, respectively. A significantly lower infectivity was also observed for the EAI lineage in the larger FB population.

Our findings around *M. africanum* are consistent with previous experiments characterizing its reduced ESAT-6 (early secretory antigenic target-6) immunogenicity and candidate genes behind its attenuated phenotype (17). However, they are only

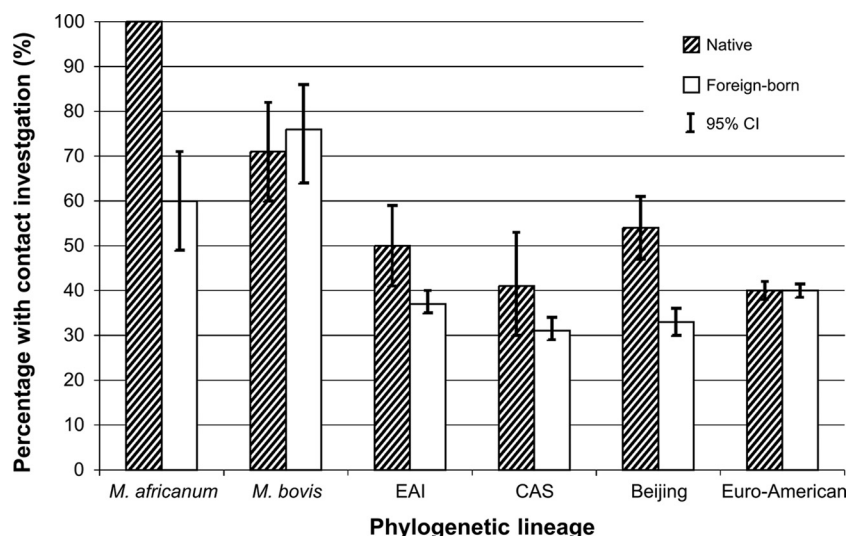


FIG 2 Proportion of cases in which contact investigation was performed by phylogenetic lineage.

TABLE 3 Estimates of progression to disease by risk factor

Characteristic	Median of bootstrapped progression-to-disease RR (95% CI)	
	Native cohort	Foreign-born cohort
Age, yr		
0–19	1.09 (0.54–2.39)	2.05 (1.25–3.74)
20–39	1 (Ref) ^a	1 (Ref)
40–64	0.83 (0.56–1.25)	1.15 (0.79–1.74)
≥65	0.76 (0.58–0.94)	0.60 (0.40–0.87)
Smear positivity		
Negative	0.62 (0.27–1.52)	0.26 (0.17–0.38)
Positive	1 (Ref)	1 (Ref)
Lineage		
Euro-American	1 (Ref)	1 (Ref)
Beijing	1.33 (0.60–3.48)	1.32 (0.64–2.35)
CAS	2.47 (0.73–11.26)	0.86 (0.54–1.43)
EAI	1.30 (0.57–3.30)	1.16 (0.82–1.68)
<i>M. africanum</i>	2.07 (0.96–2.11)	1.14 (0.70–2.11)
<i>M. bovis</i>	0.89 (0.66–1.31)	0.92 (0.45–2.00)
Residency		
Urban	1 (Ref)	1 (Ref)
Rural	1.67 (1.01–3.01)	1.65 (0.91–2.42)
Alcohol abuse		
No	1 (Ref)	
Yes	9.78 (1.52–159.85)	
Drug abuse		
No	1 (Ref)	
Yes	3.79 (1.20–22.68)	
Traveler to country with endemicity		
No	1 (Ref)	1 (Ref)
Yes	0.77 (0.41–1.30)	0.39 (0.13–0.95)

^a Ref, reference.

partially consistent with those from a study conducted in the Gambia, where *M. africanum* was shown to transmit equally well to household contacts but less likely than *M. tuberculosis* to progress to disease (9). While numbers in our native population were too low to detect any associations in both indices, in the larger FB cohort, our findings suggest that lower infectivity might also be a component of the overall lower transmissibility of *M. africanum*. Perhaps because of this lower infectivity, we did not observe the previously reported lower estimate of progression to disease in *M. africanum*. Possible explanations for this disparity may lie in the slightly different definition of infectivity used in the Gambia, where they used the incidence of TST conversion (using a follow-up period of 3 months) specifically within households as the outcome. In addition, we may not be comparing exactly the same genotype; in our FB cohort, only 3 of 183 (1.7%) *M. africanum* strains with a known birth country came from the Gambia.

In a cohort of native and FB TB cases in Montreal, the EAI lineage was also significantly associated with lower number of TST-positive contacts around index cases and with less clustering (lower proportion of patients clustering, as defined by identical RFLP or spoligotypes) in multivariable analysis (18). It is interesting to observe this trend in our study, which includes only pulmonary cases of EAI, given the association this lineage has with the extrapulmonary site of disease (16). In a secondary cohort of only FB cases in the Montreal study, the EAI lineage was significantly associated with less TST positivity but not with less clustering (18). This again agrees with our study, where we observed a significant association of EAI with lower infectivity but not with lower clustering. These findings on the EAI genotype are hard to explain using the molecular epidemiological data from Vietnam, where approximately 40% of cases are caused by EAI strains and another 40%, by the Beijing genotype strains (19). If EAI strains are less successful at infecting, one would expect them to disappear in a few generations and be replaced by other, more fit, strains. This shift is perhaps occurring at the very moment, as Beijing genotype isolates have been associated with a lower age of patients and, hence, with active transmission.

Although *M. bovis* was spread significantly less in the native

population and although the estimates of average number of secondary cases were lower than other lineages, the fact that there were three documented secondary cases (from three different index cases with unique fingerprints) does not rule out the possible occurrence of transmission of *M. bovis* in The Netherlands, where pasteurization practices have been in place for decades. Ingestion of unpasteurized dairy products has been suggested as the likely route of infection in extrapulmonary cases in second-generation immigrants in The Netherlands who may have traveled back to their country of origin (20). Yet, all three *M. bovis* index cases with secondary cases in our clustering cohort also had pulmonary manifestations; two of these index cases were FB but had no indication of recent travel to a country of *M. bovis* endemicity. Indeed, instances of human-to-human transmission of *M. bovis* have been documented in other settings (21, 22). Together these observations suggest that, from a public health perspective, contact investigation and treatment of pulmonary *M. bovis* patients should not altogether differ from those of *M. tuberculosis* patients.

Unlike studies conducted in other populations, where the Beijing strain was associated with greater virulence and transmissibility, we did not find that the Beijing strain had higher indices of infectivity, clustering, or progression to disease in The Netherlands (23, 24). This is concordant with other recent studies conducted in similarly low-incidence, immigrant-receiving settings, such as the United States and Canada, which concluded that Beijing strains are no more of a public health threat than non-Beijing strains (25, 26). The observed higher success rate of Beijing strains may therefore result from circumstances characteristic of high-prevalence settings, such as mass use of BCG vaccination, development of resistance, crowding of the human population, and other unknown factors.

Other clinical and demographic factors positively associated with either infectivity or clustering indices, such as smear positivity, a lower age, and residing in an urban area, have been similarly described in previous studies (27–29). The significantly lower estimate of progression to disease given an elderly source likely reflects a lower dose of infection (due to a less close contact) and propensity for older patients to have older contacts themselves, as well as the higher proportion of long, latent infections (possibly associated with lower virulence) in this age category (30). Likewise, the significant association between alcohol and drug abuse with higher estimates of progression to disease can be linked to the direct effects of both substances on immunity, the indirect effects of substance-related disorders (i.e., malnutrition), and other potential confounding factors, such as homelessness (31, 32). There are two possible reasons behind the less-expected association between use of intravenous drugs and the lower average number of TST-positive contacts in the native cohort. Contacts of drug abusers are often intravenous drug users themselves, a scenario in which the accurate definition of a first-ring contact is prone to misclassification (contacts could be misclassified as first-ring contacts while they actually do not have much contact with an index case and, therefore, do not become TST positive). It has also been described that drug use can comprise cellular immunity (even in the absence of HIV infection) so that TST sensitivity in drug users is lower (33, 34).

The low prevalence setting of this study means that the investigation of the role of the *M. tuberculosis* genotype on transmission is less likely to be confounded by a high background infection pressure, where a TST result is more likely to fail at distinguishing

recent from past infection. Furthermore, in The Netherlands there is no routine BCG vaccination program that could affect the interpretation of TST results, making TST a suitable tool for the detection of recent *M. tuberculosis* infection in contact investigations. This advantage applies solely to the native cohort, however, as patients in the FB cohort are far more likely to have been BCG vaccinated than native patients (40% versus 8%, respectively) and have had higher exposure to TB in their country of origin; both of these factors might lead to an overestimation of infectivity. It is encouraging, however, to observe the same trend of lower infectivity in EAI result in another study which did adjust for the probability of previous latent TB (18). On the other hand, the facts that FB patients often have FB contacts and that contact tracing in this group is less efficient imply that we might have also underestimated infectivity (and, by implication, biased the progression to disease index upward) in this group. The same reasoning applies to cases of addiction to alcohol and drugs, where an increased likelihood of homelessness means infected contacts are less likely to be found.

It is important to remember the potential shortcomings from the molecular epidemiology data underpinning these findings. A lack of clinical follow-up data of infected contacts meant that we were unable to link infected contacts to secondary cases and, thus, to estimate the proportion of secondary cases infected by a specific index case. In this low-burden country, however, there is likely a large overlap in the number of infected contacts around an index case and the number of secondary cases occurring within 2 years of that index case. It nevertheless meant that we could not control for risk factors across the transmission chain, such as rates of latent TB treatment and existing medical risk factors in secondary cases, which could influence the likelihood of progression to disease or the susceptibility to infection of the host, respectively. Studies using a prospective cohort approach (i.e., with access to household contacts and TST conversion data) that can bypass some of these issues are warranted to confirm these findings.

In sum, the lower infectivity or overall transmissibility observed in this study for *M. bovis*, *M. africanum*, and EAI—all, ancient lineages—matches the hypothesis that modern strains, as a consequence of their access to rapidly increasing numbers of susceptible hosts, have been selected for more rapid disease progression and transmission (35). Validation of this scenario via future experimental studies could have important implications on how TB control efforts may be determined not only by index case host characteristics, but also by a bacterial signature, such as phylogenetic lineage.

ACKNOWLEDGMENTS

This study was supported by the Portuguese Foundation for Science and Technology (FCT) (reference SFRH/BD/33902/2009 to H.N.-G).

We thank Rogier Donders, Megan Murray, and Maha Farhat for their statistical support and discussion of the methodology used. We thank the staff of the RIVM mycobacteriological laboratory for their work on the RFLP and VNTR typing of *M. tuberculosis* isolates, and we thank the Municipal Health Services for their voluntary collaboration in the nationwide tuberculosis surveillance.

REFERENCES

1. Borgdorff MW, van den Hof S, Kremer K, Verhagen L, Kalisvaart N, Erkens C, van Soolingen D. 2010. Progress towards tuberculosis elimination: secular trend, immigration and transmission. *Eur Respir J* 36:339–347. <http://dx.doi.org/10.1183/09031936.00155409>.

2. Goguet de la Salmoniere YO, Li HM, Torrea G, Bunschoten A, van Embden J, Gicquel B. 1997. Evaluation of spoligotyping in a study of the transmission of *Mycobacterium tuberculosis*. *J Clin Microbiol* 35:2210–2214.
3. de Beer JL, van Ingen J, de Vries G, Erkens C, Sebek M, Mulder A, Sloot R, van den Brandt AM, Enaimi M, Kremer K, Supply P, van Soolingen D. 2013. Comparative study of IS6110 restriction fragment length polymorphism and variable-number tandem-repeat typing of *Mycobacterium tuberculosis* isolates in The Netherlands, based on a 5-year nationwide survey. *J Clin Microbiol* 51:1193–1198. <http://dx.doi.org/10.1128/JCM.03061-12>.
4. Kato-Maeda M, Gagneux S, Flores LL, Kim EY, Small PM, Desmond EP, Hopewell PC. 2011. Strain classification of *Mycobacterium tuberculosis*: congruence between large sequence polymorphisms and spoligotypes. *Int J Tuberc Lung Dis* 15:131–133.
5. Allix-Béguec C, Harmsen D, Weniger T, Supply P, Niemann S. 2008. Evaluation and strategy for use of MIRU-VNTR_{plus}, a multifunctional database for online analysis of genotyping data and phylogenetic identification of *Mycobacterium tuberculosis* complex isolates. *J Clin Microbiol* 46:2692–2699. <http://dx.doi.org/10.1128/JCM.00540-08>.
6. Kik SV, Verver S, van Soolingen D, de Haas PE, Cobelens FG, Kremer K, van Deutekom H, Borgdorff MW. 2008. Tuberculosis outbreaks predicted by characteristics of first patients in a DNA fingerprint cluster. *Am J Respir Crit Care Med* 178:96–104. <http://dx.doi.org/10.1164/rccm.200708-1256OC>.
7. Verhagen LM, van den Hof S, van Deutekom H, Hermans PW, Kremer K, Borgdorff MW, van Soolingen D. 2011. Mycobacterial factors relevant for transmission of tuberculosis. *J Infect Dis* 203:1249–1255. <http://dx.doi.org/10.1093/infdis/jir013>.
8. Nebenzahl-Guimaraes H, Borgdorff MW, Murray MB, van Soolingen D. 2014. A novel approach: the propensity to propagate (PTP) method for controlling for host factors in studying the transmission of *Mycobacterium tuberculosis*. *PLoS One* 9:e97816. <http://dx.doi.org/10.1371/journal.pone.0097816>.
9. de Jong B, Hill PC, Aiken A, Awine T, Antonio M, Adetifa IM, Jackson-Sillah DJ, Fox A, Deriemer K, Gagneux S, Borgdorff MW, McAdam KP, Corrah T, Small PM, Adegbola RA. 2008. Progression to active tuberculosis, but not transmission, varies by *M. tuberculosis* lineage in The Gambia. *J Infect Dis* 198:1037–1043. <http://dx.doi.org/10.1086/591504>.
10. Rhee JT, Piatek AS, Small PM, Harris LM, Chaparro SV, Kramer FR, Alland D. 1999. Molecular epidemiologic evaluation of transmissibility and virulence of *Mycobacterium tuberculosis*. *J Clin Microbiol* 37:1764–1770.
11. van Embden JD, Cave MD, Crawford JT, Dale JW, Eisenach KD, Gicquel B, Hermans P, Martin C, McAdam R, Shinnick TM. 1993. Strain identification of *Mycobacterium tuberculosis* by DNA fingerprinting: recommendations for a standardized methodology. *J Clin Microbiol* 31:406–409.
12. Supply P, Allix C, Lesejean S, Cardoso-Oelemann M, Rüsch-Gerdes S, Willery E, Savine E, de Haas P, van Deutekom H, Roring S, Bifani P, Kurepina N, Kreiswirth B, Sola C, Rastogi N, Vatin V, Gutierrez MC, Fauville M, Niemann S, Skuce R, Kremer K, Loch C, van Soolingen D. 2006. Proposal for standardization of optimized mycobacterial interspersed repetitive unit-variable-number tandem repeat typing of *Mycobacterium tuberculosis*. *J Clin Microbiol* 44:4498–4510. <http://dx.doi.org/10.1128/JCM.01392-06>.
13. van Soolingen D, Borgdorff MW, de Haas PEW, Sebek MM, Veen J, Dessens M, Kremer K, van Embden JD. 1999. Molecular epidemiology of tuberculosis in The Netherlands: a nationwide study from 1993 through 1997. *J Infect Dis* 180:726–736. <http://dx.doi.org/10.1086/314930>.
14. Weniger T, Krawczyk J, Supply P, Niemann S, Harmsen D. 2010. MIRU-VNTR_{plus}: a web tool for polyphasic genotyping of *Mycobacterium tuberculosis* complex bacteria. *Nucleic Acids Res* 38:W326–W331. <http://dx.doi.org/10.1093/nar/gkq351>.
15. Jasmer RM, Hahn JA, Small PM, Daley CL, Behr MA, Moss AR, Creasman JM, Schechter GF, Paz EA, Hopewell PC. 1999. A molecular epidemiologic analysis of tuberculosis trends in San Francisco, 1991 to 1997. *Ann Intern Med* 130:971–978. <http://dx.doi.org/10.7326/0003-4819-130-12-199906150-00004>.
16. Mulder C, van Deutekom H, Huisman EM, Meijer-Veldman W, Erkens CG, van Rest J, Borgdorff MW, van Leth F. 2011. Coverage and yield of tuberculosis contact investigations in The Netherlands. *Int J Tuberc Lung Dis* 15:1630–1637. <http://dx.doi.org/10.5588/ijtld.11.0027>.
17. Gehre F, Otu J, DeRiemer K, de Sessions PF, Hibberd ML, Mulders W, Corrah T, de Jong BC, Antonio M. 2014. Deciphering the growth behavior of *Mycobacterium africanum*. *PLoS Negl Trop Dis* 7:e2220. <http://dx.doi.org/10.1371/journal.pntd.0002220>.
18. Albanna AS, Reed MB, Kotar KV, Fallow A, McIntosh FA, Behr MA, Menzies D. 2011. Reduced transmissibility of East African Indian strains of *Mycobacterium tuberculosis*. *PLoS One* 6:e25075. <http://dx.doi.org/10.1371/journal.pone.0025075>.
19. Buu TN, Huyen MN, Lan NT, Quy HT, Hen NV, Zignol M, Borgdorff MW, Cobelens FG, van Soolingen D. 2009. The Beijing genotype is associated with young age and multidrug-resistant tuberculosis in rural Vietnam. *Int J Tuberc Lung Dis* 13:900–906.
20. Majoor CJ, Magis-Escorra C, van Ingen J, Boeree MJ, van Soolingen D. 2011. Epidemiology of *Mycobacterium bovis* disease in humans, The Netherlands, 1993 to 2007. *Emerg Infect Dis* 17:457–453. <http://dx.doi.org/10.3201/eid1703.101111>.
21. Sunder S, Lanotte P, Godreuil S, Martin C, Boschiroli ML, Besnier JM. 2009. Human-to-human transmission of tuberculosis caused by *Mycobacterium bovis* in immunocompetent patients. *J Clin Microbiol* 47:1249–1251. <http://dx.doi.org/10.1128/JCM.02042-08>.
22. Etchehoury I, Valencia GE, Morcillo N, Sequeira MD, Imperiale B, López M, Caimi K, Zumarraga MJ, Catalá A, Romano MI. 2010. Molecular typing of *Mycobacterium bovis* isolates in Argentina: first description of a person to person transmission case. *Zoonoses Public Health* 57:375–381. <http://dx.doi.org/10.1111/j.1863-2378.2009.01233.x>.
23. Yang C, Luo T, Sun G, Qiao K, Sun G, DeRiemer K, Mei J, Gao Q. 2012. *Mycobacterium tuberculosis* Beijing strains favor transmission but not drug resistance in China. *Clin Infect Dis* 55:1179–1187. <http://dx.doi.org/10.1093/cid/cis670>.
24. Tounousova OS, Mariandyshv A, Bjune G, Sandven P, Caugant DA. 2003. Molecular epidemiology and drug resistance of *Mycobacterium tuberculosis* isolates in the Archangel prison in Russia: predominance of the W-Beijing clone family. *Clin Infect Dis* 37:665–672. <http://dx.doi.org/10.1086/377205>.
25. Langlois-Klassen D, Senthilselvan A, Chui L, Kunimoto D, Saunders LD, Menzies D, Long R. 2013. Transmission of *Mycobacterium tuberculosis* Beijing strains, Alberta, Canada, 1991 to 2007. *Emerg Infect Dis* 19:701–711. <http://dx.doi.org/10.3201/eid1905.121578>.
26. Gagneux S, Small PM. 2007. Global phylogeography of *Mycobacterium tuberculosis* and implications for tuberculosis product development. *Lancet Infect Dis* 7:328–337. [http://dx.doi.org/10.1016/S1473-3099\(07\)70108-1](http://dx.doi.org/10.1016/S1473-3099(07)70108-1).
27. Lohmann EM, Koster BF, le Cessie S, Kamst-van Agterveld MP, van Soolingen D, Arend SM. 2012. Grading of a positive sputum smear and the risk of *Mycobacterium tuberculosis* transmission. *Int J Tuberc Lung Dis* 16:1477–1484. <http://dx.doi.org/10.5588/ijtld.12.0129>.
28. Borgdorff MW, Nagelkerke NJD, de Haas PEW, van Soolingen D. 2001. Transmission of *Mycobacterium tuberculosis* depending on the age and sex of source cases. *Am J Epidemiol* 154:934–943. <http://dx.doi.org/10.1093/aje/154.10.934>.
29. Marais BJ, Gie RP, Schaaf HS, Hesselning AC, Obihara CC, Starke JJ, Enarson DA, Donald PR, Beyers N. 2004. The natural history of childhood intrathoracic tuberculosis: a critical review of literature from the prechemotherapy era. *Int J Tuberc Lung Dis* 8:392–402.
30. Wallinga J, Teunis P, Kretzschmar M. 2006. Using data on social contacts to estimate age-specific transmission parameters for respiratory-spread infectious agents. *Am J Epidemiol* 164:936–944. <http://dx.doi.org/10.1093/aje/kwj317>.
31. Lönnroth K, Williams BG, Stadlin S, Jaramillo E, Dye C. 2008. Alcohol use as a risk factor for tuberculosis: a systematic review. *BMC Public Health* 8:2458–2458. <http://dx.doi.org/10.1186/1471-2458-8-289>.
32. Szabo G. 1997. Alcohol's contribution to compromised immunity. 1997. *Alcohol Health Res World* 21:30–38.
33. Carballo-Diéguez A, Sahs J, Goetz R, el Sadr W, Sorell S, Gorman J. 1994. The effect of methadone on immunological parameters among HIV-positive and HIV-negative drug users. *Am J Drug Alcohol Abuse* 20:317–329. <http://dx.doi.org/10.3109/00952999409106017>.
34. Alonzo NC, Bayer BM. 2002. Opioids, immunology, and host defenses of intravenous drug abusers. *Infect Dis Clin North Am* 16:553–569. [http://dx.doi.org/10.1016/S0891-5520\(02\)00018-1](http://dx.doi.org/10.1016/S0891-5520(02)00018-1).
35. Portevin D, Gagneux S, Comas I, Young D. 2011. Human Macrophage Responses to Clinical Isolates from the *Mycobacterium tuberculosis* Complex Discriminate between Ancient and Modern Lineages. *PLoS Pathog* 7:e1001307. <http://dx.doi.org/10.1371/journal.ppat.1001307>.