## **Short Communication**

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# Type-1 Cannabinoid Receptor Activity During Alzheimer's Disease Progression

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**Abstract**. The activity of  $CB_1$  cannabinoid receptors was studied in postmortem brain samples of Alzheimer's disease (AD) patients during clinical deterioration.  $CB_1$  activity was higher at earlier AD stages in limited hippocampal areas and internal layers of frontal cortex, but a decrease was observed at the advanced stages. The pattern of modification appears to indicate initial hyperactivity of the endocannabinoid system in brain areas that lack classical histopathological markers at earlier stages of AD, indicating an attempt to compensate for the initial synaptic impairment, which is then surpassed by disease progression. These results suggest that initial  $CB_1$  stimulation might have therapeutic relevance.

Keywords: Alzheimer's disease, cannabinoid receptors, functional autoradiography, G-protein, ligand binding

### INTRODUCTION

The decline in synaptic function appears at early stages of Alzheimer's disease (AD), which correlates with cognitive dysfunction in AD patients [1, 2] at areas innervated by the cholinergic cells of the basal forebrain [3, 4]. The neuropathological markers, used for the classification of AD patients in different stages, have been found in subjects without dementia that may represent a preclinical stage of the illness [5].

The study of the endocannabinoid synapse is particularly interesting with regard to AD because cannabinoid receptor expression and other components of the endocannabinoid system have been found to be modified [6]. Reduction of CB<sub>1</sub> receptor density has been described in the hippocampus and caudateputamen [7]. The localization of the cannabinoid receptors in the brain suggests its involvement in the modulation of learning and memory [8]. Some cannabinoid compounds are able to induce amnesia and memory deficits in mice [9] and regulate fear-conditioned memory [10]. In addition, increase in cannabinoid tone appears to induce neuronal survival [11]. The genetic deletion of CB<sub>1</sub> receptors induces neuronal loss in the hippocampus, accompanied by

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a decline in cognitive functions [12]. Therefore, the activity mediated by CB<sub>1</sub> receptors might regulate some cognitive functions and neuroprotective actions.

In the present study, we analyze the activity and density of  $CB_1$  cannabinoid receptors in postmortem tissue from patients during AD progression.

#### MATERIALS AND METHODS

Brain tissue samples from 17 control cases and 36 AD patients were obtained from the tissue bank of the Hospital of Bellvitge, Barcelona. The AD patients were divided into three groups according to Braak's stages [13, 14], and were matched for age, postmortem delay, and freezing storage time.

Tissue sections were incubated with  $0.04\,\mathrm{nM}$  guanosine [ $^{35}\mathrm{S}$ ]5'-O-[gamma-thio]triphosphate ([ $^{35}\mathrm{S}$ ]GTP $\gamma$ S). Agonist-stimulated binding was measured in the presence of the specific cannabinoid receptor agonist WIN55,212-2 ( $10^{-4}\mathrm{M}$ ), and consecutive slices were incubated with 3 nM [ $^{3}\mathrm{H}$ ]CP55,940 (more details in Supplementary Material).

Differences between groups of patients were analyzed using the Kruskal-Wallis non-parametric test followed by Dunn's *post-hoc* test. Correlations were applied to compare the [<sup>35</sup>S]GTPγS with the [<sup>3</sup>H]CP55,940 binding sites (Pearson's or Spearman's test).

#### **RESULTS**

In the present study, we used functional  $[^{35}S]GTP\gamma S$  autoradiography to analyze the frontal cortex (Brodmann area 8), amygdala, basal forebrain (nucleus basalis of Meynert, nbM), striatum, hippocampus, and entorhinal cortex for  $CB_1$ -mediated activation of  $G_{i/o}$  proteins in the presence of the cannabinoid agonist WIN55,212-2. We also employed  $[^{3}H]CP55,940$  autoradiography to calculate the density of  $CB_1$  receptors.

Analysis of the [ $^{35}$ S]GTP $\gamma$ S binding stimulated by WIN55,212-2 (Table 1) showed an upward trend during AD stages I-II of the functional CB $_1$  receptors at layer VI of frontal cortex.

The activity of CB<sub>1</sub> receptors was lower during AD stages V-VI than during stages I-II in the pyramidal and radiatum layers of the hippocampal CA1. The increased activity measured in the AD I-II group compared with the control group was partially responsible for this significant effect. This increase was statistically significant in the hilus of the dentate gyrus

during stages I-II and decreased during stages V-VI. At stages III-IV, the  $CB_1$  activity was similar to the control group in most of the hippocampal areas (Supplementary Fig. 1). The increased activity at stages I-II might be delaying the deterioration of  $CB_1$ -mediated activity. In contrast, in the lateral nucleus of the amygdala,  $CB_1$ -mediated activity decreased from the initial stages of the disease. In the nbM and striatal areas,  $CB_1$  activity was not altered in AD.

We quantified the cannabinoid receptor density by measuring the specific labeling of the radioligand [<sup>3</sup>H]CP55,940 (Table 2). The CB<sub>1</sub> cannabinoid receptors were upregulated in layer VI of the frontal cortex in patients with stage III-IV AD. In the hippocampus, CB<sub>1</sub> receptor density was altered in different hippocampal subfields in AD patients depending on the AD stage. Receptor density was increased relative to control cases mainly during AD stages III-IV. The density of receptors decreased to or fell below control levels at the most advanced stages (Supplementary Fig. 1). In the amygdala, CB<sub>1</sub> density was very low, and the measured densities of cannabinoid receptors were maintained during disease progression. Cannabinoid receptor density was not altered in the nbM of AD patients. In the caudate-putamen, the CB<sub>1</sub> receptor density increased during the initial stages of AD compared with control densities and a trend to decrease to control levels was observed during the next stages.

#### DISCUSSION

The aim of the present study was to analyze the status of the endocannabinoid system during AD progression. Patients were divided in three groups according to Braak's neuropathological stages (stages I-II, III-IV, and V-VI) [13, 15]. We observed regulation of the activation of the signaling cascade by  $G_{i/o}$  proteins mediated through cannabinoid receptors during AD progression. The data obtained from the [<sup>3</sup>H]CP55,940 autoradiography did not correlate with the WIN55,212-2-stimulated binding. Therefore, receptor density and receptor efficiency are modulated separately. The different lipid composition of the neuronal membranes can modulate the CB<sub>1</sub> activity [16].

The frontal cortex tissue sections from AD patients exhibit an apparent increase of the CB<sub>1</sub> activity in the AD I-II stages compared to AD III-IV. Previous studies that analyzed CB<sub>1</sub> receptor density and expression in cortical areas did not describe changes in AD [7, 17], but other binding and PET studies showed that CB<sub>1</sub> densities were reduced in frontal cortex [6,

Table 1 Autoradiographic densities for the specific binding of  $[^{35}S]GTP\gamma S$  in human brain (nCi/g t.e.) stimulated by WIN55,212-2 in frontal cortex, hippocampus, and entorhinal cortex of control and AD patients

Brain area	Control	AD I-II	AD III-IV	AD V-VI
Frontal cortex	(n = 4)	(n = 5 - 8)	(n = 6)	
Layer I-III	$45 \pm 33$	$64 \pm 23$	$53 \pm 23$	_
Layer IV	$20 \pm 15$	$71 \pm 26$	$53 \pm 33$	_
Layer V	$59 \pm 55$	$130 \pm 49$	$82 \pm 47$	_
Layer VI	$78 \pm 68$	$212 \pm 430$	$125 \pm 60$	_
Hippocampus	(n = 4-7)	(n = 3-5)	(n = 3-4)	(n = 4 - 8)
CA1				
Lacunosum moleculare	$101 \pm 34$	$238 \pm 76$	$29 \pm 37$	$129 \pm 17$
Oriens	$134 \pm 58$	$197 \pm 43$	_	$100 \pm 56$
Pyramidal	$296 \pm 82$	$512 \pm 109$	_	$194 \pm 25^{*,d}$
Radiatum	$166 \pm 31$	$321 \pm 70$	_	$154 \pm 33^{*,d}$
CA3				
Lacunosum moleculare	$176 \pm 113$	$484 \pm 120$	$323 \pm 152$	$140 \pm 19$
Oriens	$147 \pm 39$	$227 \pm 65$	_	$91 \pm 34$
Pyramidal	$252 \pm 88$	$436 \pm 156$	_	$162 \pm 31$
Radiatum	$218 \pm 53$	$451 \pm 122$	_	$93 \pm 37$
Dentate gyrus				
Granular	$80 \pm 23$	$201 \pm 75$	$27 \pm 33$	$0 \pm 6^{*,d}$
Hilus	$87 \pm 22$	$236 \pm 83^{*,a}$	$15 \pm 29$	$11 \pm 6^{*,d}$
Molecular	$258 \pm 61$	$523 \pm 142$	$171 \pm 62$	$80 \pm 32^{*,d}$
Subiculum				
Lacunosum moleculare	$110 \pm 42$	$208 \pm 72$	$42 \pm 40$	$90 \pm 36$
Oriens	$101 \pm 54$	$219 \pm 32$	$64 \pm 28$	$73 \pm 25$
Pyramidal	$301 \pm 44$	$525 \pm 123$	$205 \pm 69$	$152 \pm 36^{*,c,d}$
Radiatum	$236 \pm 89$	$324 \pm 162$	$111 \pm 56$	$138 \pm 49$
Entorhinal cortex	(n = 4-7)	(n = 3-5)	(n = 3-4)	(n = 4 - 8)
Layer I	$165 \pm 35$	$417 \pm 239$	$116 \pm 109$	$108 \pm 56$
Layer II-III	$218 \pm 55$	$364 \pm 226$	$175 \pm 136$	$105 \pm 29$
Layer IV-VI	$169 \pm 45$	$407 \pm 203$	$173 \pm 107$	$88 \pm 35$
Amygdala	(n=4)	(n = 8-12)	(n = 7-9)	
Lateral nucleus	$321 \pm 159$	$-68 \pm 51^{*,a}$	$58 \pm 88^{*,b}$	_
Basal nucleus (magnocellular)	$44 \pm 46$	$90 \pm 32$	$55 \pm 15$	_
Basal forebrain	(n=7)	(n = 12)	(n = 9)	(n=5)
Nucleus basalis (Meynert)	$138 \pm 38$	$173 \pm 66$	$97 \pm 55$	$113 \pm 64$
Striatum	(n=6)	(n = 13)	(n = 11)	(n = 9)
Caudate-putamen	$277 \pm 97$	$197 \pm 48$	$170 \pm 45$	$169 \pm 50$

Data are mean  $\pm$  SEM values. (n): number of cases used. (–): less than three samples available. The p values were calculated by the Kruskal-Wallis non-parametric test followed by Dunn's test. <sup>a</sup>AD I-II versus control, <sup>b</sup>AD III-IV versus control, <sup>c</sup>AD V-VI versus control, <sup>d</sup>AD I-II versus AD V-VI. \*p<0.05.

18, 19]. The increase of CB<sub>1</sub> receptor activity that we observed during the initial stages of AD might indicate a neuroprotective action mediated by endocannabinoids in response to initial neural damage, which has been extensively reviewed [20]. Analysis of the [³H]CP55,940 binding sites during disease progression revealed a significant increase of CB<sub>1</sub> receptor density at layer VI of the frontal cortex in the AD III-IV patient group. The results indicate that the regulation of CB<sub>1</sub>-mediated activity precedes the increase in receptor density. The increased efficiency could be less metabolically costly to the cell than the increased availability of new receptors.

Recent studies have reported a reduction in the enzyme responsible for the synthesis of anandamide

in the cortex of AD patients [21]. The regulation of the CB<sub>1</sub> density and activity that we describe at cortex might be a compensatory mechanism to balance the anandamide signaling.

In a previous study CB<sub>1</sub> receptor density has been found decreased at the hippocampus in AD patients [7]. But when we analyzed it in detail during the progression of the disease, the CB<sub>1</sub> activation was greater during AD stages I-II, decreased to levels similar to the control group during AD stages III-IV, and continued to fall below control group levels during AD stages V-VI. These effects were significant at the pyramidal layers and the dentate gyrus, areas in which the cannabinoid receptors are more densely located at synaptic terminals. Therefore, the increase of activity

Table 2
Autoradiographic densities for the specific binding of [<sup>3</sup>H]CP55,940 in the human brain samples from control and AD patients (fmol/mg)

Brain area	Controls	AD I-II	AD III-IV	AD V-VI
Frontal cortex	(n=9)	(n = 9-10)	(n = 6)	
Layer I-III	$57.8 \pm 13.9$	$56.2 \pm 7.8$	$67.1 \pm 12.8$	_
Layer IV	$59.7 \pm 16.4$	$51.1 \pm 5.4$	$69.4 \pm 10.9$	_
Layer V	$54.6 \pm 13.1$	$60.4 \pm 7.8$	$64.9 \pm 8.6$	_
Layer VI	$50.4 \pm 9.8$	$68.5 \pm 6.1$	$91.2 \pm 8.6^{*,b}$	_
Hippocampus				
CA1	(n = 4-6)	(n = 4-7)	(n = 5)	(n = 4-5)
Lacunosum moleculare	$40.6 \pm 5.1$	$48.4 \pm 6.5$	$51.4 \pm 8.4$	$42.8 \pm 3.2$
Oriens	$40.6 \pm 8$	$42.2 \pm 3.1$	$39.3 \pm 4.6$	$43.9 \pm 5.4$
Pyramidal	$64.6 \pm 9.6$	$84.1 \pm 10$	$87.5 \pm 15.9$	$48.1 \pm 2.1^{*,d}$
Radiatum	$42.5 \pm 5.9$	$46.8 \pm 5.7$	$55.7 \pm 9.6$	$44.4 \pm 4.9$
CA3	(n = 6-7)	(n = 7 - 8)	(n = 6)	(n=7)
Lacunosum moleculare	$44.2 \pm 6.7$	$53.9 \pm 5.2$	$65.8 \pm 5.5$	$40.5 \pm 3.3^{*,d}$
Oriens	$42.6 \pm 5.1$	$42.2 \pm 3.7$	$51.9 \pm 7.6$	$44.5 \pm 4.7$
Pyramidal	$68.3 \pm 7.3$	$88.4 \pm 6.2$	$98.3 \pm 9.6$	$58.1 \pm 3.5^{*,c,d}$
Radiatum	$41.6 \pm 5.0$	$47.0 \pm 8.9$	$59.6 \pm 6.2^{*,b}$	$44.9 \pm 4.5$
Dentate gyrus	(n = 4-6)	(n = 6)	(n = 7)	(n = 4-5)
Granular	$80.2 \pm 10.1$	$95.8 \pm 3.5$	$99.5 \pm 8.5$	$70.8 \pm 8.5^{*,c}$
Hilus	$64.3 \pm 9.9$	$67.7 \pm 6.3$	$74.6 \pm 6.6$	$65.2 \pm 5.6$
Subiculum	(n = 5-7)	(n = 6-7)	(n = 5-6)	(n = 6)
Lacunosum moleculare	$44.6 \pm 6.5$	$46.4 \pm 7.5$	$64.8 \pm 7.1$	$36.7 \pm 1.2^{*,d}$
Oriens	$45.5 \pm 6.3$	$38.3 \pm 4.6$	$55.9 \pm 6.3$	$42.4 \pm 2.7$
Pyramidal	$59.4 \pm 6.5$	$101.1 \pm 2.7^{*,a}$	$107.3 \pm 13.7$	$49.1 \pm 4.2^{*,d}$
Radiatum	$44.8 \pm 6.2$	$59.3 \pm 8.6$	$72.4 \pm 5.6$	$36.3 \pm 1.1^{*,d}$
Entorhinal cortex	(n=4)	(n = 3)	(n = 3)	(n=3)
Layer I	$71.3 \pm 17$	$55.1 \pm 15.8$	$60.9 \pm 0.6$	$58.7 \pm 14.9$
Layer II-III	$53.0 \pm 10.8$	$73.2 \pm 5.9$	$94.5 \pm 3.9^{*,b}$	$67.8 \pm 22.1^{*,d}$
Layer IV-VI	$48.5 \pm 9.1$	$63.5 \pm 9.5$	$70.0 \pm 3.2$	$60.5 \pm 14^{*,d}$
Amygdala	(n = 3-4)	(n = 6-9)	(n = 4-5)	
Lateral nucleus	$7.1 \pm 1.9$	$5.3 \pm 0.9$	$4.3 \pm 0.9$	_
Basal nucleus (magnocellular)	$3.9 \pm 0.5$	$4.5 \pm 0.4$	$3.5 \pm 0.8$	_
Basal forebrain	(n=4)	(n = 6)	(n=4)	
Nucleus basalis (Meynert)	$47.1 \pm 4.7$	$40.1 \pm 0.9$	$41.5 \pm 8.0$	_
Striatum	(n=5)	(n = 12)	(n=8)	(n=5)
Caudate-putamen	$62.7 \pm 8.6$	$94.7 \pm 4.8^{*a}$	$75.4 \pm 6.9$	$66.4 \pm 4.5^{*,c}$

Data are mean  $\pm$  SEM values. (n): number of cases used. (–): less than 3 samples available. The p values were calculated by the Kruskal-Wallis non-parametric test followed by Dunn's test. <sup>a</sup>AD I-II versus control, <sup>b</sup>AD III-IV versus control, <sup>c</sup>AD I-II versus AD V-VI, <sup>d</sup>AD III-IV versus AD V-VI. \*p < 0.05.

during AD stages I-II might be an initial response to neural impairment.

Moreover, an increase of monoacylglycerol lipase activity has been described in AD stages V-VI that might contribute to the accumulation of 2-arachidonoyl glycerol, resulting in synaptic failure with the consequent downregulation of CB<sub>1</sub> receptors [22]. However, immunohistochemical assays have described no changes in CB<sub>1</sub> receptors [23].

We observed that the regulation on  $CB_1$  densities was delayed during the advance of AD compared with the modulation of  $CB_1$  activity. The [ $^3H$ ]CP55,940 binding sites were decreased at later stages in the pyramidal layers of different hippocampal areas and the inner layers of the entorhinal cortex.

The nbM cholinergic cells innervate the amygdala, where the functional CB<sub>1</sub> receptors decreased in the

lateral nucleus of the amygdala in AD patients. In contrast, the number and expression of cannabinoid receptors in the human amygdala are low [24, 7]. We also observed low CB<sub>1</sub> receptor density in the amygdala, which was conserved in AD patients. This finding suggests that the down-regulation of cannabinoid activity is not indicating modulation in receptor density. On the contrary, CB<sub>1</sub> activity in the striatal area was conserved in AD patients, but CB<sub>1</sub> density was high and was increased at the initial AD stages, continuing to a reduction with the progression of the disease, which might be caused by the loss of afferents from areas such as the globus pallidus and the substantia nigra, where a reduction of CB<sub>1</sub> of receptors had been reported [7]. Although the incidence of co-morbidity with parkinsonism is frequent in AD patients, we were unaware if this was the case in any

of the cases included in the present study. In addition, maintained levels of CB<sub>1</sub> cannabinoid receptors have been described in Parkinson's disease [25].

In summary,  $CB_1$  receptors were more efficient in the patients at the earlier AD stages, specifically in hippocampal areas. These regulations on  $CB_1$  signaling might precede to the accumulation of the neuropathological markers of the AD in specific brain areas. On the contrary, at the most advanced stages of AD,  $CB_1$  efficacy diminished in both the hippocampus and the frontal cortex. The modulation of  $CB_1$  density follows the same pattern, but occurs later in the course of AD.

The initial hyperactivity of the endocannabinoid system accounts for the possible compensation of synaptic impairment; however, the intimate and unknown cause of AD continues with neurodegeneration and determines a loss of  $CB_1$  synapses.  $CB_1$  stimulation might have therapeutic relevance during the initial and moderate stages of AD.

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#### SUPPLEMENTARY MATERIAL

Supplementary tables are available in the electronic version of this article: http://dx.doi.org/10.3233/JAD-140492.

#### REFERENCES

- Perry EK, Tomlinson BE, Blessed G, Bergmann K, Gibson PH, Perry RH (1978) Correlation of cholinergic abnormalities with senile plaques and mental test scores in senile dementia. Br Med J 6150, 1457-1459.
- [2] DeKosky ST, Harbaugh RE, Schmitt FA, Bakay RA, Chui HC, Knopman DS, Reeder TM, Shetter AG, Senter HJ, Markesbery WR (1992) Cortical biopsy in Alzheimer's disease: Diagnostic accuracy and neurochemical, neuropathological, and cognitive correlations. Intraventricular Bethanecol Study Group, Ann Neurol 32, 625-632.
- [3] Davies CA, Mann DM, Sumpter PQ, Yates PO (1987) A quantitative morphometric analysis of the neuronal and synaptic content of the frontal and temporal cortex in patients with Alzheimer's disease. *J Neurol Sci* **78**, 151-164.
- [4] Scheff SW, Price DA, Schmitt FA, DeKosky ST, Mufson EJ (2007) Synaptic alterations in CA1 in mild Alzheimer disease and mild cognitive impairment. *Neurology* 68, 1501-1508.
- [5] Price JL, McKeel DW Jr, Buckles VD, Roe CM, Xiong C, Grundman M, Hansen LA, Petersen RC, Parisi JE, Dickson

- DW, Smith CD, Davis DG, Schmitt FA, Markesbery WR, Kaye J, Kurlan R, Hulette C, Kurland BF, Higdon R, Kukull W, Morris JC (2009) Neuropathology of nondemented aging: Presumptive evidence for preclinical Alzheimer disease. *Neurobiol Aging* **30**, 1026-1036.
- [6] Ramírez BG, Blázquez C, Gómez del Pulgar T, Guzmán M, de Ceballos ML (2005) Prevention of Alzheimer's disease pathology by cannabinoids: Neuroprotection mediated by blockade of microglial activation. *J Neurosci* 25, 1904-1913.
- [7] Westlake TM, Howlett AC, Bonner TI, Matsuda LA, Herkenham M (1994) Cannabinoid receptor binding and messenger RNA expression in human brain: An in vitro receptor autoradiography and in situ hybridization histochemistry study of normal aged and Alzheimer's brains. Neuroscience 63, 637-652.
- [8] Riedel G, Davies SN (2005) Cannabinoid function in learning, memory and plasticity. Handb Exp Pharmacol 168, 445-477.
- [9] Puighermanal E, Marsicano G, Busquets-Garcia A, Lutz B, Maldonado R, Ozaita A (2009) Cannabinoid modulation of hippocampal long-term memory is mediated by mTOR signaling. *Nat Neurosci* 12, 1152-1158.
- [10] Maćkowiak M, Chocyk A, Dudys D, Wedzony K (2009) Activation of CB1 cannabinoid receptors impairs memory consolidation and hippocampal polysialylated neural cell adhesion molecule expression in contextual fear conditioning. *Neuroscience* 158, 1708-1716.
- [11] Marsicano G, Goodenough S, Monory K, Hermann H, Eder M, Cannich A, Azad SC, Cascio MG, Gutiérrez SO, van der Stelt M, López-Rodriguez ML, Casanova E, Schütz G, Zieglgänsberger W, Di Marzo V, Behl C, Lutz B (2003) CB1 cannabinoid receptors and on-demand defense against excitotoxicity. Science 302, 84-88.
- [12] Braak H, Braak E (1991) Neuropathological stageing of Alzheimer-related changes. Acta Neuropathol 82, 239-259.
- [13] Bilkei-Gorzo A, Racz I, Valverde O, Otto M, Michel K, Sastre M, Zimmer A (2005) Early age-related cognitive impairment in mice lacking cannabinoid CB1 receptors. *Proc Natl Acad Sci U S A* 102, 15670-15675.
- [14] Khachaturian ZS (1985) Diagnosis of Alzheimer's disease. Arch Neurol 42, 1097-1105.
- [15] Braak H, Alafuzoff I, Arzberger T, Kretzschmar H, Del Tredici K (2006) Staging of Alzheimer disease-associated neurofibrillary pathology using paraffin sections and immunocytochemistry. Acta Neuropathol 112, 389-404.
- [16] Oddi S, Dainese E, Fezza F, Lanuti M, Barcaroli D, De Laurenzi V, Centonze D, Maccarrone M (2011) Functional characterization of putative cholesterol binding sequence (CRAC) in human type-1 cannabinoid receptor. *J Neurochem* 116, 858-865.
- [17] Lee JH, Agacinski G, Williams JH, Wilcock GK, Esiri MM, Francis PT, Wong PT, Chen CP, Lai MK (2010) Intact cannabinoid CB1 receptors in the Alzheimer's disease cortex. *Neurochem Int* 57, 985-989.
- [18] Solas M, Francis PT, Franco R, Ramirez MJ (2013) CB2 receptor and amyloid pathology in frontal cortex of Alzheimer's disease patients. *Neurobiol Aging* 34, 805-808.
- [19] Ahmad R, Goffin K, Van den Stock J, De Winter FL, Cleeren E, Bormans G, Tournoy J, Persoons P, Van Laere K, Vandenbulcke M (2013) *In vivo* type 1 cannabinoid receptor availability in Alzheimer's disease. *Eur Neuropsychopharmacol* 24, 242-250.
- [20] Campbell VA, Gowran A (2007) Alzheimer's disease; taking the edge off with cannabinoids? Br J Pharmacol 152, 655-662.

- [21] Jung KM, Astarita G, Yasar S, Vasilevko V, Cribbs DH, Head E, Cotman CW, Piomelli D (2012) An amyloid β(42)dependent deficit in anandamide mobilization is associated with cognitive dysfunction in Alzheimer's disease. *Neurobiol Aging* 33, 1522-1532.
- [22] Mulder J, Zilberter M, Pasquaré SJ, Alpár A, Schulte G, Ferreira SG, Köfalvi A, Martín-Moreno AM, Keimpema E, Tanila H, Watanabe M, Mackie K, Hortobágyi T, de Ceballos ML, Harkany T (2011) Molecular reorganization of endocannabinoid signaling in Alzheimer's disease. *Brain* 134, 1041-1060.
- [23] Benito C, Núñez E, Pazos MR, Tolón RM, Romero J (2007) The endocannabinoid system and Alzheimer's disease. *Mol Neurobiol* 36, 75-81.
- [24] Herkenham M, Lynn AB, Little MD, Johnson MR, Melvin LS, de Costa BR, Rice KC (1990) Cannabinoid receptor localization in brain. *Proc Natl Acad Sci U S A* 87, 1932-1936.
- [25] Farkas S, Nagy K, Jia Z, Harkany T, Palkovits M, Donohou SR, Pike VW, Halldin C, Máthé D, Csiba L, Gulyás B (2012) The decrease of dopamine D<sub>2</sub>/D<sub>3</sub> receptor densities in the putamen and nucleus caudatus goes parallel with maintained levels of CB<sub>1</sub> cannabinoid receptors in Parkinson's disease: A preliminary autoradiographic study with the selective dopamine D<sub>2</sub>/D<sub>3</sub> antagonist [³H]raclopride and the novel CB<sub>1</sub> inverse agonist [¹<sup>25</sup>I]SD7015. *Brain Res Bull* 87, 504-510.