



Phosphorylation of unique domains of Src family kinases

Irene Amata[†], Mariano Maffei and Miquel Pons*

Biomolecular NMR Laboratory, Department of Organic Chemistry, University of Barcelona, Barcelona, Spain

Edited by:

Allegra Via, Sapienza University, Italy
Andreas Zanzoni, Technological
Advances for Genomics and Clinics,
Inserm UMR1090, France

Reviewed by:

Philipp Selenko, Leibniz Society,
Germany
Julie D. Forman-Kay, Hospital for Sick
Children, Canada

*Correspondence:

Miquel Pons, Biomolecular NMR
Laboratory, Department of Organic
Chemistry, University of Barcelona,
Baldri Reixac, 10-12, 08028
Barcelona, Spain
e-mail: mpons@ub.edu

[†]Present address:

Irene Amata, Laboratoire de
Cristallographie et RMN Biologiques –
Université Paris Descartes, Paris,
France

Members of the Src family of kinases (SFks) are non-receptor tyrosine kinases involved in numerous signal transduction pathways. The catalytic, SH3 and SH2 domains are attached to the membrane-anchoring SH4 domain through the intrinsically disordered “Unique” domains, which exhibit strong sequence divergence among SFk members. In the last decade, structural and biochemical studies have begun to uncover the crucial role of the Unique domain in the regulation of SFk activity. This mini-review discusses what is known about the phosphorylation events taking place on the SFk Unique domains, and their biological relevance. The modulation by phosphorylation of biologically relevant inter- and intra- molecular interactions of Src, as well as the existence of complex phosphorylation/dephosphorylation patterns observed for the Unique domain of Src, reinforces the important functional role of the Unique domain in the regulation mechanisms of the Src kinases and, in a wider context, of intrinsically disordered regions in cellular processes.

Keywords: IDPs, IDRs, phosphorylation, unique domain, SFks, Src

INTRODUCTION

The Src kinase family is composed of 10 proteins: Src, Frk, Lck, Lyn, Blk, Hck, Fyn, Yrk, Fgr, and Yes. Src family kinases (SFks) are membrane-associated, non-receptor tyrosine kinases that act as important signaling intermediaries regulating a variety of outputs, such as cell proliferation, differentiation, apoptosis, migration, and metabolism. All SFks share the same domain arrangement: a large catalytic C-terminal domain is preceded by regulatory Src 2 and 3 homology domains (SH2 and SH3, respectively) and the membrane-anchoring SH4 N-terminal region, which contain myristoylation and palmitoylation sites as well as positively charged residues. The SH3 and SH4 domains are linked by an intrinsically disordered segment of 50–90 residues, called the Unique domain (UD) because of the lack of sequence similarity among the different SFks. However, the UD of each individual SFk member is well conserved between different organisms suggesting a more specific role than that of a simple spacer. **Figure 1** shows the sequences of the UDs of the 10 SFks, highlighting known phosphorylation sites.

Efforts to find specific UD functions during the last two decades have confirmed an active role of the UD in the regulation of SFk members. In the case of Lck, the UD mediates association with the cytoplasmic tail of CD4 and CD8 α through a zinc clasp structure (Kim et al., 2003). For most of the other SFk members the detailed mechanisms remain obscure. The Unique domains of Fyn and Lyn were observed to be cleaved during induction of apoptosis in intact hematopoietic cells, revealing a novel mechanism for the specific regulation of different SFks, with important consequences

for their cellular localization and activity (Luciano et al., 2001). Several SFks contain residues in the N-terminal region that are phosphorylated and dephosphorylated in cellular processes (Joung et al., 1995; Hansen et al., 1997; Johnson et al., 2000). Moreover, swapping the Unique domains of Src and Yes interchanges the functional specificity of the two SFks (Hoey et al., 2000; Summy et al., 2003; Werdich and Penn, 2005). The versatility and relevance of the active role of the Unique domain in Src function was confirmed by the discovery of binding by the Unique domain to different targets, such as acidic lipids, the SH3 domain, and calmodulin (Pérez et al., 2013). The amino-acid sequence of the UD region responsible for some of these interactions is conserved in Src, Fyn, and Yes. The three proteins are co-expressed in many tissues and are partially redundant, in the sense that deficiency in one of them can be compensated by the action of the other two.

The involvement of SFk signaling in growth, motility and cell survival makes them important oncology targets. Activation of SFks is tightly regulated in healthy cells (Boggon and Eck, 2004), while the kinase activation is often deregulated in cancer, giving rise to altered cellular shape, function, and growth (Vlahovic and Crawford, 2003). Among the SFks, Src is the most studied and the most commonly discussed in the context of cancer (Yeatman, 2004). However, there has been growing interest in the other SFks in both normal physiological and pathological states.

The UD is an intrinsically disordered region (IDR). Intrinsically disordered proteins (IDPs) are relatively more prevalent among signaling and cancer-related proteins (Iakoucheva et al., 2002). The connection between disorder and diseases such as

proteins, thereby promoting DNA synthesis complex assembly and cell proliferation. Both p56^{Lyn} Tyr32 and MCM7 Tyr600 phosphorylation are enhanced in proliferating cells and correlated with poor survival of breast cancer patients (Huang et al., 2013).

All Unique domains contain aromatic residues, a rare feature in intrinsically disordered domains. A tyrosine residue is present between positions 25 and 34 in seven SFKs, and is known to be phosphorylated in five of them: Lyn, Hck, Lck (Hornbeck et al., 2012), Yes (Ariki et al., 1997), and Frg (Oppermann et al., 2009). Phosphorylation of Tyr34 of Frg has been observed by mass spectrometry mainly in samples from leukemia patients.

Erk is the only SFK that is not myristoylated. Although it contains three tyrosine residues in its Unique domain, phosphorylation has only been reported for Tyr46, at the interface between the Unique and SH3 domains. The modification was observed by mass spectrometry mainly in samples from lung, liver, and gastric cancer (Stokes et al., 2012).

PHOSPHORYLATION OF THE UNIQUE DOMAIN OF Fyn

Fyn is ubiquitously expressed together with Src and Yes, whereas other members of the Src family are expressed only in specific cell types. Fyn is primarily localized to the cytoplasmic leaflet of the plasma membrane, where it phosphorylates tyrosine residues on key targets involved in a variety of different signaling pathways. Fyn characterization has been mainly focused on its immune and neurological function. However, Fyn has also been recognized as an important mediator of mitogenic signaling and regulator of cell-cycle entry, growth, and proliferation, integrin-mediated interactions as well as cell adhesion and migration (Kawakami et al., 1988; Chen et al., 2001; Li et al., 2003; Kim et al., 2010). In particular, Fyn is over-expressed in several cancers, such as glioblastoma multiforme and melanoma. The role of Fyn over-expression in these systems, however, has not been well defined as yet.

Fyn presents multiple phosphorylation sites that can affect its kinase activity. Activation of Platelet-Derived Growth Factor (PDGF) β -receptor by binding to PDGF leads to activation of a member of the SFK (Alonso et al., 1995). The Unique domain of Fyn was found to be phosphorylated at Tyr28 (Hansen et al., 1997). The functional role of this phosphorylation was confirmed by the observation of significantly reduced activation following PDGF stimulation of a Fyn mutant in which Tyr28 was replaced by phenylalanine (Hansen et al., 1997). It was also proposed that the autophosphorylation of the N-terminal tyrosine residues plays a key role in the activation of Fyn, by complementing the PDGF receptor-induced phosphorylation of Tyr28.

Serine 21 within the UD of Fyn is part of a RxxS motif targeted by protein kinase A (PKA). Mutation of Ser21 to Alanine (Ser21Ala-Fyn) blocks PKA phosphorylation of Fyn and alters its tyrosine kinase activity (Yeo et al., 2011). In the same work, the authors showed that the over-expression of Ser21Ala-Fyn mutant in cells lacking Src/Yes/Fyn kinases (SYF cells) led to decreased tyrosine phosphorylation of focal adhesion kinase, resulting in reduced focal adhesion targeting, and slow lamellipodia dynamics and cell migration. These important changes in cell motility demonstrate a key role of UD phosphorylation at Ser21 in the Fyn

kinase activity that controls assembly and disassembly of focal adhesions in response to signals arising from cell-extracellular matrix interactions.

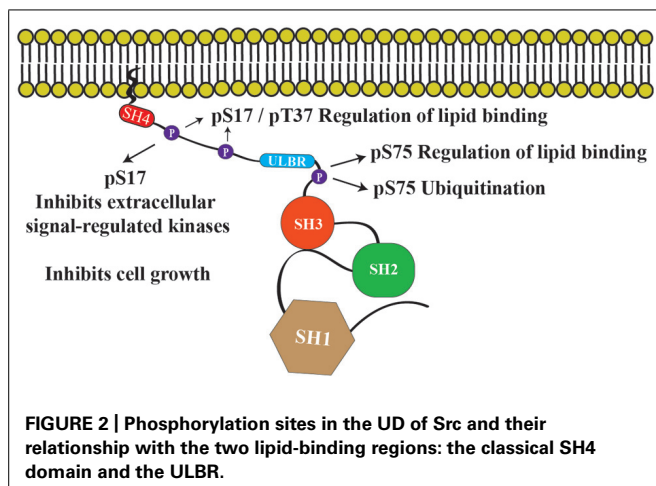
Protein kinase A is a crucial component of integrin-mediated signaling pathways. Interaction of cells with their substrate or adhesion to other cells leads to activation of PKA (Whittard and Akiyama, 2001), and induce the phosphorylation of a variety of protein substrates (Walsh and Van Patten, 1994). Four SFKs (Fyn, Src, Lck, and Fgr) contain a conserved RxxS motif suggesting they can be regulated by PKA.

PHOSPHORYLATION OF THE UNIQUE DOMAIN OF Src

Src is a non-receptor protein tyrosine kinase with a key role in regulating cell-to-matrix adhesion, migration, and junctional stability (Frame, 2004). Thus, precise regulation of Src activity is critical for normal cell growth. The inactive state of Src is obtained by phosphorylated tyrosine near the C-terminus of Src (Tyr530 in mammalian Src; Tyr527 in chicken Src), which is recognized by its SH2 domain, while the SH3 domain interacts with a polyproline motif located in the linker region between the SH2 and kinase domains; these intramolecular interactions restrict access to the kinase domain (Xu et al., 1997). Dephosphorylation of Tyr530 is followed by autophosphorylation at Tyr419, leading to full activation of the kinase. Active Src may be deactivated by rephosphorylation of Tyr530 by C-terminal Src kinase (Csk; Piwnicka-Worms et al., 1987; Nada et al., 1991; Brown, 1996).

Although the phosphorylation of Src at Ser17 by PKA (cAMP-dependent protein kinase) is a well-characterized process, its biological significance remains unclear (Obara et al., 2004). Potential roles in protein-protein interactions or cellular localization have been postulated for this phosphorylation site. For instance, it has been observed that the treatment of 3T3 fibroblasts with PDGF results in the translocation of Src from the plasma membrane to the cytosol, concomitant with an increase in phosphorylation of Ser17 by PKA (Walker et al., 1993). This observation suggests that this phosphorylation could interfere with the electrostatic interactions that act to anchor Src to the lipid bilayer. PKA phosphorylation of Src at Ser17 is also required in cAMP activation of Rap1, inhibition of extracellular signal-regulated kinases, and inhibition of cell growth, although the mechanism by which this phosphorylation mediates these processes is not known (Obara et al., 2004).

Interaction of Src with lipids is not restricted to the SH4 domain (Figure 2). The unique lipid binding region (ULBR) was discovered following NMR observations that revealed a partially structured region within the UD of Src (Pérez et al., 2009). While phosphorylation of Ser17 by PKA disturbed the interaction of the SH4 domain with lipids, phosphorylation of Thr37 and Ser75 by p25-Cdk5 decreased lipid binding by the ULBR (Pérez et al., 2009). Cross-effects were observed, suggesting a cooperative interaction of the two lipid binding regions with membranes. Conformational effects of phosphorylation at Ser17, Thr37, and Ser75 in the isolated UD are strictly local, indicating that electrostatic repulsion of the phosphorylated residues with the negatively charged lipids is the main mechanism by which lipid binding is disrupted.



The ULBR major regulatory role of the UD was shown experimentally in *Xenopus laevis* oocytes. Mutations abolishing lipid binding by the ULBR but not affecting the SH4 domain resulted in a conditional lethal phenotype after progesterone induced maturation (Pérez et al., 2013).

Previous results had identified a functional role for Thr37 and Ser75 (Thr34 and Ser72 in chicken Src) as well as Thr46 in chicken Src (with no correspondence in humans). These residues are phosphorylated by cyclin-dependent kinase 1 (Cdk1/Cdc2) during mitosis. These phosphorylations were found to activate Src by disrupting the interaction between the SH2 domain and Tyr527/Tyr530 (chicken/human) and facilitating the dephosphorylation by protein tyrosine phosphatases (Shenoy et al., 1992; Stover et al., 1994).

In addition (or possibly related to) its effect on lipid binding, phosphorylation of Ser75 is important in other aspects of Src regulation. Mitosis-independent phosphorylation of this site was observed in neurons and in certain cancer cell lines; this phosphorylation was shown to be due to Cdk5, a widely distributed proline-directed kinase with a substrate specificity similar to Cdk1 (Kato and Maeda, 1999). Active Src is reported to be irreversibly destroyed by Cullin-5-dependent ubiquitination and proteosomal degradation (Hakak and Martin, 1999; Laszlo and Cooper, 2009). More recently, it was shown that phosphorylation at Ser75 promotes the ubiquitin-dependent degradation of Src (Pan et al., 2011). Ser75-Src phosphorylation in epithelial cells was found to depend on the activation state of Src: only active Src was phosphorylated and eventually marked for ubiquitination. Thus, Cdk5-dependent phosphorylation of Ser75 within the UD of Src represents a mechanism to restrict the availability of active Src (Pan et al., 2011). Sequence alignment suggests that serine residues are present in homologous positions of Yes and possibly Frk.

NMR and mass-spectrometry studies of human Src UD added to *X. laevis* egg extracts showed the phosphorylation of Ser75 and Ser69, as well as Ser 17 (Pérez et al., 2013). Phosphorylation of Ser75 and Ser69 seem to be mutually interfering *in vitro*. Interestingly, *X. laevis* Src has a glycine residue at position 69 and phosphorylation of Ser69 in human Src had only been

previously detected by mass spectrometry in extracts of cancer lines HCT116 and MDA-MB-435S (Oppermann et al., 2009). It is tempting to speculate that pathological phosphorylation of Ser 69 in human cells could result in decreased phosphorylation of Ser75 and reduced degradation of active Src, leading to an oncogenic phenotype caused by Src overactivation.

The interplay between various phosphorylation sites within the UD emphasizes its role as a signaling integration hub. Further input signals, in addition to phosphorylation, include calcium-dependent interaction with calmodulin and the allosteric interaction with Src SH3 domain (Pérez et al., 2013; Figure 2). A further level of cross-talk between various phosphorylation events was observed when the time-dependent phosphorylation of the UD was studied by real-time NMR in cell extracts (Amata et al., 2013). In these experiments it was observed that the activity of a PKA-like kinase, which phosphorylates Ser17, also repressed the phosphatase(s) that catalyzed the dephosphorylation of Ser75. Thus, inhibition of PKA activity resulted in dephosphorylation of both sites. Similarly, inhibition of Cdk activity resulted in a reduction in the steady-state phosphorylation of both sites. On the other hand, addition of PKA caused a robust phosphorylation of Ser75 by preventing the action of the phosphatases that reverse the effect of Cdk. These results show that the phosphorylation state of the UD of Src represents a sensor of the kinase-phosphatase network active at any given moment in the cell. Experimental access to this information is possible by using real-time NMR techniques (Selenko et al., 2008; Theillet et al., 2012; Thongwichian and Selenko, 2012; Amata et al., 2013). In contrast to folded domains, intrinsically disordered domains, like the UD of Src, provide *in vivo* NMR resolution comparable to that obtainable *in vitro* (Kosol et al., 2013).

CONCLUDING REMARKS

The study of the UDs of SFKs, particularly that of Src, provides an example of multilevel regulation through phosphorylation of a membrane-bound intrinsically disordered domain. The occurrence of membrane tethering among IDRs is very common, although its relevance is not always recognized. The results arising from studies of UDs should stimulate further research to uncover similar mechanisms beyond SFKs. In particular, switchable internal lipid binding sites (the ULBR in the case of Src) in disordered domains anchored to membranes by a second, more stable, binding site close to the protein termini (the SH4 domain in the case of SFKs) can modulate the position of the active domains (the kinase or regulatory domains in the case of SFKs) with respect to the membrane surface. Lipid binding by the internal site can switch off the access of the active domains to target sites of membrane-anchored substrates/partners located beyond a particular distance from the membrane surface or, conversely, facilitate the interaction with sites near the membrane surface. Phosphorylation or other modifications preventing the interaction of the internal site would result in a release of the active domains, which can then reach sites located further apart from the membrane surface or disfavor the interaction with membrane-proximal sites. This represents a new compartmentalization mechanism based on the relative position of interacting sites with respect to the membrane

surface. This mechanism enables modulating the functional interactions between two proteins anchored next to each other in the membrane.

ACKNOWLEDGMENTS

This work was supported in part by funds from Fundació Marató TV3, the Spanish Government (BIO2010-15683), the Catalan Government (2009SGR1352), and the European Union 7FP (BioNMR contract 261863, and Marie Curie Action - COFUND to Irene Amata).

REFERENCES

- Alonso, G., Koegl, M., Mazurenko, N., and Courtneidge, S. A. (1995). Sequence requirements for binding of src family tyrosine kinases to activated growth factor receptors. *J. Biol. Chem.* 270, 9840–9848. doi: 10.1074/jbc.270.17.9840
- Amata, I., Maffei, M., Igea, A., Gay, M., Vilaseca, M., Nebreda, A. R., et al. (2013). Multi-phosphorylation of the intrinsically disordered unique domain of c-Src studied by in-cell and real-time NMR spectroscopy. *Chembiochemistry* 14, 1820–1827. doi: 10.1002/cbic.201300139
- Ariki, M., Tanabe, O., Usui, H., Hayashi, H., Inoue, R., Nishito, Y., et al. (1997). Identification of autophosphorylation sites in c-Yes purified from rat liver plasma membranes. *J. Biochem.* 121, 104–111.
- Boggon, T. J., and Eck, M. J. (2004). Structure and regulation of Src family kinases. *Oncogene* 23, 7918–7927. doi: 10.1038/sj.onc.1208081
- Brown, E. (1996). A signaling pathway to translational control. *Cell* 86, 517–520. doi: 10.1016/S0092-8674(00)80125-7
- Chen, J. W., Romero, P., Uversky, V. N., and Dunker, A. K. (2006). Conservation of intrinsic disorder in protein domains and families: I. A database of conserved predicted disordered regions. *J. Proteome Res.* 5, 879–887. doi: 10.1021/pr060048x
- Chen, R., Kim, O., Yang, J., Sato, K., Eisenmann, K. M., McCarthy, J., et al. (2001). Regulation of Akt/PKB activation by tyrosine phosphorylation. *J. Biol. Chem.* 276, 31858–31862. doi: 10.1074/jbc.C100271200
- Frame, M. C. (2004). Newest findings on the oldest oncogene; how activated src does it. *J. Cell Sci.* 117(Pt 7), 989–998. doi: 10.1242/jcs.01111
- Hakak, Y., and Martin, S. G. (1999). Ubiquitin-dependent degradation of active Src. *Curr. Biol.* 9, 1039–1042. doi: 10.1016/S0960-9822(99)80453-9
- Hansen, K., Alonso, G., Courtneidge, S. A., Rönnstrand, L., and Heldin, C.-H. H. (1997). PDGF-induced phosphorylation of Tyr28 in the N-terminus of Fyn affects Fyn activation. *Biochem. Biophys. Res. Commun.* 241, 355–362. doi: 10.1006/bbrc.1997.7743
- Hoey, J. G., Summy, J., and Flynn, D. C. (2000). Chimeric constructs containing the SH4/Unique domains of cYes can restrict the ability of Src527F to upregulate heme oxygenase-1 expression efficiently. *Cell. Signal.* 12, 691–701. doi: 10.1016/S0898-6568(00)00116-9
- Hornbeck, P. V., Kornhauser, J. M., Tkachev, S., Zhang, B., Skrzypek, E., Murray, B., et al. (2012). PhosphoSitePlus: a comprehensive resource for investigating the structure and function of experimentally determined post-translational modifications in man and mouse. *Nucleic Acids Res.* 40, D261–D270. doi: 10.1093/nar/gkr1122
- Huang, T. H., Huo, L., Wang, Y. N., Xia, W., Wei, Y., Chang, S. S., et al. (2013). Epidermal growth factor receptor potentiates MCM7-mediated DNA replication through tyrosine phosphorylation of Lyn kinase in human cancers. *Cancer Cell* 23, 796–810. doi: 10.1016/j.ccr.2013.04.027
- Iakoucheva, L. M., Brown, C. J., Lawson, D. J., Obradović, Z., and Dunker, K. A. (2002). Intrinsic disorder in cell-signaling and cancer-associated proteins. *J. Mol. Biol.* 323, 573–584. doi: 10.1016/S0022-2836(02)00969-5
- Johnson, T. M., Williamson, N. A., Scholz, G., Jaworowski, A., Wettenhall, R. E., Dunn, A. R., et al. (2000). Modulation of the catalytic activity of the Src family tyrosine kinase Hck by autophosphorylation at a novel site in the unique domain. *J. Biol. Chem.* 275, 33353–33364. doi: 10.1074/jbc.M002022200
- Joung, I., Kim, T., Stolz, L. A., Payne, G., Winkler, D. G., Walsh, C. T., et al. (1995). Modification of Ser59 in the unique N-terminal region of tyrosine kinase p56lck regulates specificity of its Src homology 2 domain. *Proc. Natl. Acad. Sci. U.S.A.* 92, 5778–5782. doi: 10.1073/pnas.92.13.5778
- Kato, G., and Maeda, S. (1999). Neuron-specific phosphorylation Cdk5 kinase is responsible of c-Src at Ser75 in human for mitosis-independent Y79 retinoblastoma. *J. Biochem.* 126, 957–961. doi: 10.1093/oxfordjournals.jbchem.a022540
- Kawakami, T., Kawakami, Y., Aaronson, S. A., and Robbins, K. C. (1988). Acquisition of transforming properties by FYN, a normal SRC-related human gene. *Proc. Natl. Acad. Sci. U.S.A.* 85, 3870–3874. doi: 10.1073/pnas.85.11.3870
- Kesavan, K. P., Isaacson, C. C., Ashendel, C. L., Geahlen, R. L., and Harrison, M. L. (2002). Characterization of the in vivo sites of serine phosphorylation on Lck identifying serine 59 as a site of mitotic phosphorylation. *J. Biol. Chem.* 277, 14666–14673. doi: 10.1074/jbc.M111911200
- Kim, H.-J., Warren, J. T., Kim, S.-Y., Chappel, J. C., DeSelm, C. J., Ross, F. P., et al. (2010). Fyn promotes proliferation, differentiation, survival and function of osteoclast lineage cells. *J. Cell. Biochem.* 111, 1107–1113. doi: 10.1002/jcb.22841
- Kim, P. W., Sun, Z.-Y. J., Blacklow, S. C., Wagner, G., and Eck, M. J. (2003). A zinc clasp structure tethers Lck to T cell coreceptors CD4 and CD8. *Science* 301, 1725–1728. doi: 10.1126/science.1085643
- Kosol, S., Contreras-Martos, S., Cedeño, C., and Tompa, P. (2013). Structural characterization of intrinsically disordered proteins by NMR spectroscopy. *Molecules* 18, 10802–10828. doi: 10.3390/molecules180910802
- Laszlo, G. S., and Cooper, J. A. (2009). Restriction of Src activity by Cullin-5. *Curr. Biol.* 19, 157–162. doi: 10.1016/j.cub.2008.12.007
- Li, S., Iakoucheva, L. M., Mooney, S. D., and Radivojac, P. (2010). Loss of post-translational modification sites in disease. *Pac. Symp. Biocomput.* 337–347. doi: 10.1142/9789814295291_0036
- Li, X., Yang, Y., Hu, Y., Dang, D., Regezi, J., Schmidt, B. L., et al. (2003). Alphabeta6-Fyn signaling promotes oral cancer progression. *J. Biol. Chem.* 278, 41646–41653. doi: 10.1074/jbc.M306274200
- Luciano, E., Ricci, J.-E., and Auberger, P. (2001). Cleavage of Fyn and Lyn in their N-terminal unique regions during induction of apoptosis: a new mechanism for Src kinase regulation. *Oncogene* 20, 4935–4941. doi: 10.1038/sj.onc.1204661
- Nada, S., Okada, M., MacAuley, A., Cooper, J. A., and Nakagawa, H. (1991). Cloning of a complementary DNA for a protein-tyrosine kinase that specifically phosphorylates a negative regulatory site of p60c-src. *Nature* 351, 69–72. doi: 10.1038/351069a0
- Obara, Y., Labudda, K., Dillon, T. J., and Stork, P. J. S. (2004). PKA phosphorylation of Src mediates Rap1 activation in NGF and cAMP signaling in PC12 cells. *J. Cell Sci.* 117, 6085–6094. doi: 10.1242/jcs.01527
- Oppermann, F. S., Gnad, F., Olsen, J. V., Hornberger, R., Greff, Z., Kéri, G., et al. (2009). Large-scale proteomics analysis of the human kinome. *Mol. Cell. Proteomics* 8, 1751–1764. doi: 10.1074/mcp.M800588-MCP200
- Pan, Q., Qiao, F., Gao, C., Norman, B., Optican, L., and Zelenka, P. S. (2011). Cdk5 targets active Src for ubiquitin-dependent degradation by phosphorylating Src(S75). *Cell Mol. Life. Sci* 68, 3425–3436. doi: 10.1007/s00018-011-0638-1
- Pérez, Y., Gairí, M., Pons, M., and Bernadó, P. (2009). Structural characterization of the natively unfolded N-terminal domain of human c-Src kinase: insights into the role of phosphorylation of the unique domain. *J. Mol. Biol.* 391, 136–148. doi: 10.1016/j.jmb.2009.06.018
- Pérez, Y., Maffei, M., Igea, A., Amata, I., Gairí, M., Nebreda, A. R., et al. (2013). Lipid binding by the Unique and SH3 domains of c-Src suggests a new regulatory mechanism. *Sci. Rep.* 3, 1295. doi: 10.1038/srep01295
- Piwonka-Worms, H., Saunders, K. B., Roberts, T. M., Smith, A. E., and Cheng, S. H. (1987). Tyrosine phosphorylation regulates the biochemical and biological properties of pp60c-src. *Cell* 49, 75–82. doi: 10.1016/0092-8674(87)90757-4
- Selenko, P., Frueh, D. P., Elsaesser, S. J., Haas, W., Gygi, S. P., and Wagner, G. (2008). In situ observation of protein phosphorylation by high-resolution NMR spectroscopy. *Nat. Struct. Mol. Biol.* 15, 321–329. doi: 10.1038/nsmb.1395
- Shenoy, S., Chackalamparampil, I., Bagrodia, S., Lin, P.-H., and Shalloway, D. (1992). Role of p34cdc2-mediated phosphorylations in two-step activation of pp60c-src during mitosis. *Proc. Natl. Acad. Sci. U.S.A.* 89, 7237–7241. doi: 10.1073/pnas.89.15.7237
- Stokes, M. P., Farnsworth, C. L., Moritz, A., Silva, J. C., Jia, X., Lee, K. A., et al. (2012). PTMScan direct: identification and quantification of peptides from critical signaling proteins by immunoaffinity enrichment coupled with LC-MS/MS. *Mol. Cell. Proteomics* 11, 187–201. doi: 10.1074/mcp.M111.015883
- Stover, R. D., Liebetanz, J., and Lydon, B. N. (1994). Cdc2-mediated Modulation of pp60c-src activity. *J. Biol. Chem.* 269, 26885–26889.
- Summy, J. M., Qian, Y., Jiang, B.-H., Guappone-Koay, A., Gatesman, A., Shi, X., et al. (2003). The SH4-Unique-SH3-SH2 domains dictate specificity in signaling

- that differentiate c-Yes from c-Src. *J. Cell Sci.* 116(Pt. 12), 2585–2598. doi: 10.1242/jcs.00466
- Theillet, F.-X., Smet-Nocca, C., Liokatis, S., Thongwichian, R., Kosten, J., Yoon, M.-K., et al. (2012). Cell signaling, post-translational protein modifications and NMR spectroscopy. *J. Biomol. NMR* 54, 217–236. doi: 10.1007/s10858-012-9674-x
- Thongwichian, R., and Selenko, P. (2012). In-cell NMR in *Xenopus laevis* oocytes. *Methods Mol. Biol.* 895, 33–41. doi: 10.1007/978-1-61779-927-3_3
- Tintori, C., Laurenzana, I., La Rocca, F., Falchi, F., Carraro, F., Ruiz, A., et al. (2013). Identification of Hck inhibitors as hits for the development of antileukemia and anti-HIV agents. *Chem. Med. Chem.* 8, 1353–1360. doi: 10.1002/cmdc.201300204
- Uros, M., Christopher, O. J., Keith, D. A., Zoran, O., and Uversky, V. N. (2009). Unfoldomics of human genetic diseases: illustrative examples of ordered and intrinsically disordered members of the human diseasome. *Protein Pept. Lett.* 16, 1533–1547. doi: 10.2174/092986609789839377
- Uversky, V. N., Oldfield, C. J., and Dunker, K. A. (2008). Intrinsically disordered proteins in human diseases: introducing the D2 concept. *Annu. Rev. Biophys.* 37, 215–246. doi: 10.1146/annurev.biophys.37.032807.125924
- Vázquez, M. L. (2007). *Biological consequences of the Phosphorylation of Serine 59 on the Tyrosine Kinase Lck*. PhD Thesis, Purdue University, West Lafayette, IN. Available at: <http://search.proquest.com/docview/304827306?accountid=28979>
- Vlahovic, G., and Crawford, J. (2003). Activation of tyrosine kinases in cancer. *Oncologist* 8, 531–538. doi: 10.1634/theoncologist.8-6-531
- Walker, F., DeBlaquiere, J., and Burgess, A. W. (1993). Translocation of pp60^{c-src} from the plasma membrane to the cytosol after stimulation by platelet-derived growth factor. *J. Biol. Chem.* 268, 19552–19558.
- Walsh, A. D., and Van Patten, M. S. (1994). Multiple pathway signal transduction by the cAMP-dependent protein kinase. *FASEB J.* 8, 1227–1236.
- Watts, J. D., Sanghera, J. S., Pelech, S. L., and Aebersold, R. (1993). Phosphorylation of serine 59 of p56lck in activated T cells. *J. Biol. Chem.* 268, 23275–23282.
- Werdich, X. Q., and Penn, J. S. (2005). Src, Fyn and Yes play differential roles in VEGF-mediated endothelial cell events. *Angiogenesis* 8, 315–326. doi: 10.1007/s10456-005-9021-x
- Whittard, J. D., and Akiyama, S. K. (2001). Positive regulation of cell-cell and cell-substrate adhesion by protein kinase A. *J. Cell Sci.* 114, 3265–3272.
- Winkler, D. G., Park, I., Kim, T., Payne, N. S., Walsht, C. T., Strominger, J. L., et al. (1993). Phosphorylation Ser-42 and Ser-59 in the N-terminal region of the tyrosine kinase p56lck. *Proc. Natl. Acad. Sci. U.S.A.* 90, 5176–5180. doi: 10.1073/pnas.90.11.5176
- Xu, W., Harrison, C. S., and Eck, J. M. (1997). Three-dimensional structure of the tyrosine kinase c-Src. *Nature* 385, 595–602. doi: 10.1038/385595a0
- Yeatman, T. J. (2004). A renaissance for SRC. *Nat. Rev. Cancer* 4, 470–480. doi: 10.1038/nrc1366
- Yeo, M. G., Oh, H. J., Cho, H.-S., Chun, J. S., Marcantonio, E. E., and Song, W. K. (2011). Phosphorylation of Ser 21 in Fyn regulates its kinase activity, focal adhesion targeting, and is required for cell migration. *J. Cell Physiol.* 226, 236–247. doi: 10.1002/jcp.22335

Conflict of Interest Statement: The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Received: 04 March 2014; accepted: 29 May 2014; published online: 30 June 2014.

Citation: Amata I, Maffei M and Pons M (2014) Phosphorylation of unique domains of Src family kinases. *Front. Genet.* 5:181. doi: 10.3389/fgene.2014.00181

This article was submitted to *Systems Biology*, a section of the journal *Frontiers in Genetics*.

Copyright © 2014 Amata, Maffei and Pons. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) or licensor are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.