

# Genome-Wide Association Study Reveals Multiple Loci Associated with Primary Tooth Development during Infancy

Demetris Pillas<sup>1,2,3,9</sup>, Clive J. Hoggart<sup>1,9</sup>, David M. Evans<sup>4,9</sup>, Paul F. O'Reilly<sup>1,9</sup>, Kirsi Sipilä<sup>5,6</sup>, Raija Lähdesmäki<sup>6,7</sup>, Iona Y. Millwood<sup>1,8</sup>, Marika Kaakinen<sup>9,10</sup>, Gopalakrishnan Netuveli<sup>2,11</sup>, David Blane<sup>2,11</sup>, Pimphen Charoen<sup>1,12</sup>, Ulla Sovio<sup>1</sup>, Anneli Pouta<sup>13</sup>, Nelson Freimer<sup>14,15,16</sup>, Anna-Liisa Hartikainen<sup>17</sup>, Jaana Laitinen<sup>18</sup>, Sarianna Vaara<sup>13</sup>, Beate Glaser<sup>4</sup>, Peter Crawford<sup>19</sup>, Nicholas J. Timpson<sup>4</sup>, Susan M. Ring<sup>20</sup>, Guohong Deng<sup>21</sup>, Weihua Zhang<sup>1</sup>, Mark I. McCarthy<sup>22,23</sup>, Panos Deloukas<sup>24</sup>, Leena Peltonen<sup>24,25,26,27</sup>, Paul Elliott<sup>1,28,9</sup>, Lachlan J. M. Coin<sup>1,9</sup>, George Davey Smith<sup>4,9</sup>, Marjo-Riitta Jarvelin<sup>1,9,10,13,9\*</sup>

**1** Department of Epidemiology and Public Health, Imperial College London, London, United Kingdom, **2** Economic and Social Research Council International Centre for Life Course Studies in Society and Health, London, United Kingdom, **3** Medical Research Council Centre of Epidemiology for Child Health, University College London Institute of Child Health, London, United Kingdom, **4** Medical Research Council Centre for Causal Analyses in Translational Epidemiology, Department of Social Medicine, University of Bristol, Bristol, United Kingdom, **5** Department of Prosthetic Dentistry and Stomatognathic Physiology, Institute of Dentistry, University of Oulu, Oulu, Finland, **6** Oral and Maxillofacial Department, Oulu University Hospital, Oulu, Finland, **7** Department of Oral Development and Orthodontics, Institute of Dentistry, University of Oulu, Oulu, Finland, **8** Clinical Trial Service Unit and Epidemiological Studies Unit (CTSU), University of Oxford, Oxford, United Kingdom, **9** Institute of Health Sciences, University of Oulu, Oulu, Finland, **10** Biocenter Oulu, University of Oulu, Oulu, Finland, **11** Department of Primary Care and Social Medicine, Faculty of Medicine, Imperial College London, United Kingdom, **12** Department of Tropical Hygiene, Faculty of Tropical Medicine, Mahidol University, Bangkok, Thailand, **13** Department of Lifecourse and Services, National Institute of Health and Welfare, Oulu, Finland, **14** Center for Neurobehavioral Genetics, University of California Los Angeles, Los Angeles, California, United States of America, **15** The Jane and Terry Semel Institute for Neuroscience and Human Behavior, Los Angeles, California, United States of America, **16** Department of Psychiatry, University of California Los Angeles, Los Angeles, California, United States of America, **17** Department of Clinical Sciences/Obstetrics and Gynecology, University of Oulu, Oulu, Finland, **18** Finnish Institute of Occupational Health, Oulu, Finland, **19** Department of Oral and Dental Science, University of Bristol, Bristol, United Kingdom, **20** Avon Longitudinal Study of Parents and Children (ALSPAC), University of Bristol, Bristol, United Kingdom, **21** Department of Gastroenterology and Hepatology, Imperial College London, London, United Kingdom, **22** Oxford Centre for Diabetes, Endocrinology and Metabolism (OCDEM), University of Oxford, Oxford, United Kingdom, **23** Wellcome Trust Centre for Human Genetics, University of Oxford, Roosevelt Drive, Headington, Oxford, United Kingdom, **24** Wellcome Trust Sanger Institute, Wellcome Trust Genome Campus, Hinxton, Cambridge, United Kingdom, **25** Institute of Molecular Medicine Finland FIMM, Nordic EMBL Partnership for Molecular Medicine, Helsinki, Finland, **26** Department of Medical Genetics, University of Helsinki, Helsinki, Finland, **27** National Institute for Health and Welfare, Public Health Genomics Unit, Helsinki, Finland, **28** Medical Research Council–Health Protection Agency Centre in Environment and Health, London, United Kingdom

## Abstract

Tooth development is a highly heritable process which relates to other growth and developmental processes, and which interacts with the development of the entire craniofacial complex. Abnormalities of tooth development are common, with tooth agenesis being the most common developmental anomaly in humans. We performed a genome-wide association study of time to first tooth eruption and number of teeth at one year in 4,564 individuals from the 1966 Northern Finland Birth Cohort (NFBC1966) and 1,518 individuals from the Avon Longitudinal Study of Parents and Children (ALSPAC). We identified 5 loci at  $P < 5 \times 10^{-8}$ , and 5 with suggestive association ( $P < 5 \times 10^{-6}$ ). The loci included several genes with links to tooth and other organ development (*KCNJ2*, *EDA*, *HOXB2*, *RAD51L1*, *IGF2BP1*, *HMG2*, *MSRB3*). Genes at four of the identified loci are implicated in the development of cancer. A variant within the *HOXB* gene cluster associated with occlusion defects requiring orthodontic treatment by age 31 years.

**Citation:** Pillas D, Hoggart CJ, Evans DM, O'Reilly PF, Sipilä K, et al. (2010) Genome-Wide Association Study Reveals Multiple Loci Associated with Primary Tooth Development during Infancy. *PLoS Genet* 6(2): e1000856. doi:10.1371/journal.pgen.1000856

**Editor:** Greg Gibson, Georgia Institute of Technology, United States of America

**Received:** November 20, 2009; **Accepted:** January 24, 2010; **Published:** February 26, 2010

**Copyright:** © 2010 Pillas et al. This is an open-access article distributed under the terms of the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

**Funding:** The NFBC1966 received financial support from the Academy of Finland (project grants 104781, 120315, 132797, and Center of Excellence in Complex Disease Genetics); University Hospital Oulu, Biocenter, University of Oulu, Finland; the European Community's Fifth/Seventh Framework Programme (EURO-BLCS, QLGI-CT-2000-01643, FP7/2007-2013); NHLBI grant 5R01HL087679-02 through the STAMPEED program (1RL1MH083268-01); ENGAGE project (HEALTH-F4-2007-201413); the Medical Research Council (studentship grant G0500539, centre grant G0600705); the Wellcome Trust (project grant GR069224), UK; the Research Council UK fellowship; the National Institute of Health Research (NIHR) Biomedical Research Centre Programme at Imperial College; and the Division of Epidemiology, Public Health and Primary Care (studentship grant DFHM G24038). The DNA extractions, sample quality controls, biobank up-keeping, and aliquotting were performed in the National Public Health Institute, Biomedicum Helsinki, Finland, and supported financially by the Academy of Finland and Biocentrum Helsinki. The UK Medical Research Council, the Wellcome Trust, and the University of Bristol provide core support for ALSPAC. CJH is funded by a European Union grant HEALTH-2007-201550 HyperGenes. DME is supported by a Medical Research Council New Investigator Award (MRC G0800582). The ICLS (International Centre for Life Course Studies in Society and Health) is funded by an ESRC award: RES-596-28-0001. The funders had no role in study design, data collection and analysis, decision to publish, or preparation of the manuscript.

**Competing Interests:** The authors have declared that no competing interests exist.

\* E-mail: m.jarvelin@imperial.ac.uk

These authors contributed equally to this work.

## Author Summary

Genome-wide association studies have been used to identify genetic variants conferring susceptibility to diseases, intermediate phenotypes, and physiological traits such as height, hair color, and age at menarche. Here we analyze the NFBC1966 and ALSPAC birth cohorts to investigate the genetic determinants of a key developmental process: primary tooth development. The prospective nature of our studies allows us to exploit accurate measurements of age at first tooth eruption and number of teeth at one year, and also provides the opportunity to assess whether genetic variants affecting these traits are associated with dental problems later in the life course. Of the genes that we find to be associated with primary tooth development, several have established roles in tooth development and growth, and almost half have proposed links with the development of cancer. We find that one of the variants is also associated with occlusion defects requiring orthodontic treatment later in life. Our findings should provide a strong foundation for the study of the genetic architecture of tooth development, which as well as its relevance to medicine and dentistry, may have implications in evolutionary biology since teeth represent important markers of evolution.

## Introduction

Heritability of primary tooth emergence is estimated to be over 70% [1]. Abnormalities in tooth development are common with tooth agenesis alone affecting up to 10% of the population, ranking it as the most common developmental anomaly in humans [2]. Such abnormalities contribute to a variety of challenging and expensive orthodontic, prosthetic and surgical treatments and account for approximately 6% of all dental health care attendances [3]. Many genes implicated in primary dentition have regulatory functions important to several developmental processes in the embryo [4], and the developing tooth is a useful model for the study of organogenesis [5]. However, despite substantial research into tooth development in mice and human malformation syndromes [5], the genetic determinants of the normal variation in human tooth development have not been established.

To identify genetic loci regulating primary dentition we performed a general population based genome-wide association (GWA) study of tooth development in infancy among individuals from the 1966 Northern Finland Birth Cohort (NFBC1966) and the Avon Longitudinal Study of Parents and Children (ALSPAC). Specifically, we tested for associations with time to first tooth eruption and number of teeth by one year of age. These phenotypes are relevant to later tooth development because teeth largely acquire their final form at a very early age [6]. The availability of longitudinal birth cohort data allowed us to investigate life-course associations with dental occlusion defects.

## Results

We tested 300,766 SNPs common to both studies (each used the Illumina platform). The analyses were adjusted for sex, gestational age and population structure (Materials and Methods). Results for the two cohorts were combined using fixed effects inverse variance meta-analysis. Five genetic loci were identified at genome-wide significance ( $P < 5 \times 10^{-8}$ ). Table 1 shows the top-ranking SNPs at each locus (see also Figure 1 and Figure 2, Figures S1, S2, S3). For all SNPs the allele associated with a delay in tooth eruption was

associated with fewer teeth at the end of infancy. Table S1 shows details of the functions of genes linked to the identified loci.

The strongest association with both phenotypes was for SNP rs8079702, located 15 kb downstream of *KCNJ2* (inward rectifier potassium channel 2) ( $P = 3.77 \times 10^{-22}$  for time of first tooth,  $P = 1.24 \times 10^{-14}$  for number of teeth; Table 1). There are no SNPs in *KCNJ2* in our data, but rs8079702 had highest correlation with SNP rs4328485 which was the closest available SNP to *KCNJ2* ( $r^2 = 0.17$ ; 1 kb away). *KCNJ2* has been implicated in Pierre Robin sequence [7] and Andersen-Tawil syndrome [8], which show abnormalities in tooth development (missing teeth, delays in eruption) and are characterized by craniofacial anomalies such as narrowing of the jaw and cleft palate [8]. The second strongest association was for SNP rs5936487, located within the *EDA* (ectodermal dysplasia protein) gene ( $P = 6.18 \times 10^{-11}$  for time of first tooth,  $P = 3.36 \times 10^{-10}$  for number of teeth). *EDA* was fundamental in forming the first teeth in organisms [9], and mutations cause hypohidrotic ectodermal dysplasia (HED) and non-syndromic disorders of tooth agenesis [10].

The three remaining loci at genome-wide significance ( $P < 5 \times 10^{-8}$ ) have SNPs located within the genes *RAD51LI* (RAD51-like1), *IGF2BP1* (insulin-like growth factor 2 mRNA binding protein 1) and *MSRB3* (methionine sulfoxide reductase B3). *RAD51LI* is involved in DNA repair and a variant in the gene has been found to confer susceptibility to breast cancer [11]. It is responsible for protein kinase activity, and the injection of activators of protein kinase C (PKC) in rats causes delays in tooth eruption [12]. *IGF2BP1* regulates the growth factor *IGF2*, and knockouts of the gene in mice suggest a role in organ development [13], while its expression is associated with ovarian cancer [14]. A microarray study in the developing mouse molar tooth found *MSRB3* to be in the top 100 most expressed genes of 34,000 examined [15].

Each of the associated SNPs explain a small fraction of the residual phenotypic variation in time to first tooth (0.2%–1.6%, NFBC1966; 0.4%–1.5%, ALSPAC) and number of teeth by one year (0.2%–1.2%, NFBC1966; 0.5%–1.6%, ALSPAC), after controlling for sex and gestational age. Selecting the SNP with the most extreme signal for either phenotype to represent each locus (“top SNPs”), and analysing them together, the additive effects of these five top SNPs explain 2.9% of the variance of both tooth eruption time and number of teeth in the NFBC1966, and 4.2% and 4.0% of the variance in tooth eruption and number of teeth in ALSPAC. Without a suitable external replication cohort these estimates were derived in the two discovery cohorts and therefore may overestimate the true values due to the “winner’s curse”. GWA studies have thus far explained only a small proportion of heritability [16], and our estimates are comparable with the variance explained in human height by a GWA study [17]. In order to identify variants with lower effect sizes or rarer variants larger sample sizes would be required.

We also summarized the predictive power of the five top SNPs by defining a ‘delayed tooth eruption’ measure as the number of alleles across the SNPs that delay tooth eruption. Figure 3 shows the number of delayed tooth eruption alleles against the mean of both time to first tooth eruption and number of teeth by one year in NFBC1966. Individuals with 8 or more delayed eruption alleles (10% of NFBC1966) have an average of 1.5 fewer teeth at 12 months, and later tooth eruption by 1.1 months, compared to individuals with 3 or fewer such alleles (11% of NFBC1966). Figure S4 shows the same plot for time to first tooth in ALSPAC.

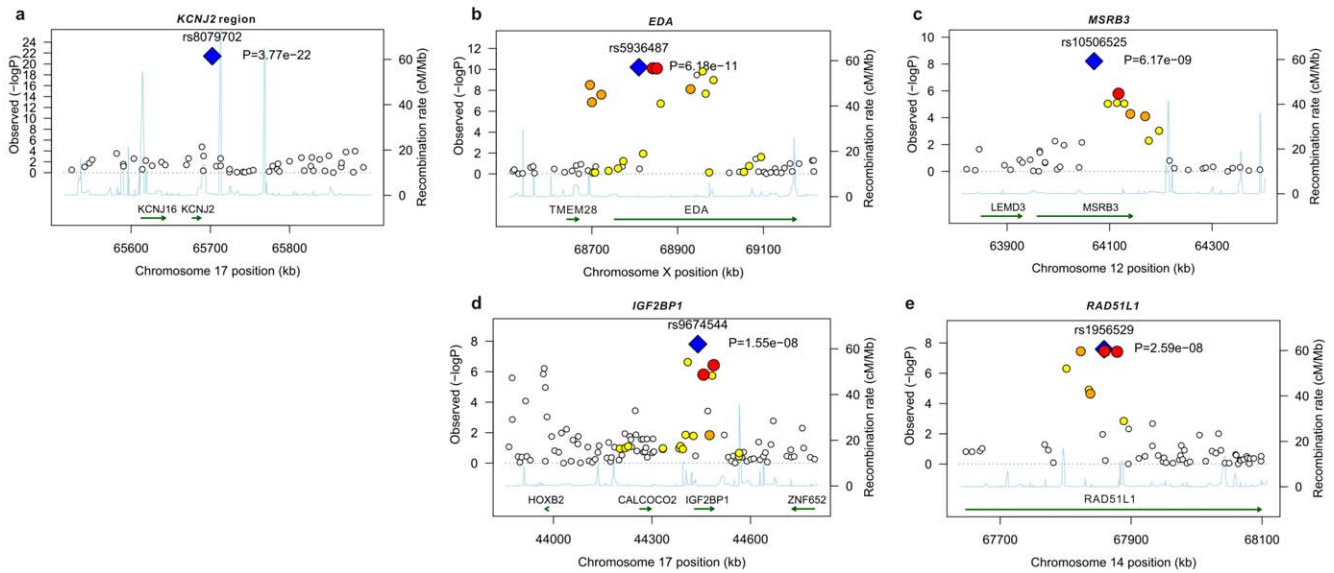
In addition to the five loci attaining genome wide significance, there were 5 loci with SNPs that had  $P$ -values between  $5 \times 10^{-6}$  and  $5 \times 10^{-8}$  (Table 1). We investigated the biological

**Table 1.** The top GWA signals at each locus from a meta-analysis of NFBC1966 and ALSPAC.

In (closest) gene/locus	SNP	Chromosome (Position)	Effect/ Other allele	Frequency of effect allele		Time to first tooth eruption				Number of teeth			
				NFBC	ALSPAC	P value NFBC	P value ALSPAC	% var NFBC/ALSPAC	Overall P	P value NFBC	P value ALSPAC	% var NFBC/ALSPAC	Overall P
<b>SNPs at genome-wide significance in meta-analysis (<math>P &lt; 5 \times 10^{-8}</math>)</b>													
(KCNJ2)	rs8079702	17 (65,702,421)	G/A	0.391	0.424	$1.89 \times 10^{-17}$	$1.62 \times 10^{-6}$	1.62/1.48	<b><math>3.77 \times 10^{-22}</math></b>	$7.78 \times 10^{-12}$	$3.22 \times 10^{-4}$	1.15/1.05	<b><math>1.24 \times 10^{-14}</math></b>
EDA	rs4844096	X (68,722,043)	G/A	0.423	0.466	$6.21 \times 10^{-6}$	$3.56 \times 10^{-4}$	0.42/0.77	<b><math>2.61 \times 10^{-8}</math></b>	$3.79 \times 10^{-9}$	$2.65 \times 10^{-3}$	0.73/0.52	<b><math>4.57 \times 10^{-11}</math></b>
"	rs5936487	X (68,809,641)	G/A	0.390	0.462	$6.65 \times 10^{-8}$	$6.90 \times 10^{-5}$	0.50/0.99	<b><math>6.18 \times 10^{-11}</math></b>	$3.89 \times 10^{-8}$	$2.10 \times 10^{-3}$	0.52/0.57	<b><math>3.36 \times 10^{-10}</math></b>
MSRB3	rs10506525	12 (64,069,645)	C/T	0.266	0.375	$1.29 \times 10^{-6}$	$5.80 \times 10^{-4}$	0.46/0.71	<b><math>6.17 \times 10^{-9}</math></b>	$2.56 \times 10^{-4}$	$5.60 \times 10^{-4}$	0.18/0.74	$8.67 \times 10^{-7}$
IGF2BP1	rs9674544	17 (44,439,710)	G/A	0.462	0.484	$2.97 \times 10^{-5}$	$5.44 \times 10^{-3}$	0.25/0.40	$8.33 \times 10^{-7}$	$2.29 \times 10^{-4}$	$9.86 \times 10^{-7}$	0.27/1.61	<b><math>1.55 \times 10^{-8}</math></b>
RAD51L1	rs1956529	14 (67,858,677)	T/C	0.383	0.364	$1.07 \times 10^{-3}$	$5.07 \times 10^{-4}$	0.16/0.73	$1.32 \times 10^{-5}$	$4.95 \times 10^{-6}$	$1.23 \times 10^{-3}$	0.51/0.60	<b><math>2.59 \times 10^{-8}</math></b>
<b>SNPs with suggestive evidence (<math>5 \times 10^{-6} &gt; P &gt; 5 \times 10^{-8}</math>) in meta-analysis</b>													
2q35	rs6435957	2 (217,586,454)	T/C	0.371	0.309	0.021	0.250	0.21/0.00	$9.99 \times 10^{-3}$	$6.85 \times 10^{-6}$	0.017	0.37/1.10	$3.64 \times 10^{-7}$
6q21	rs9386463	6 (106,200,750)	G/A	0.446	0.483	$4.55 \times 10^{-6}$	0.047	0.38/0.14	$5.99 \times 10^{-7}$	$8.29 \times 10^{-3}$	0.586	0.08/0.00	0.011
(HOXB1, HOXB2)	rs6504340	17 (43,972,018)	G/A	0.224	0.209	$3.00 \times 10^{-3}$	0.136	0.12/0.01	$9.40 \times 10^{-4}$	$2.86 \times 10^{-5}$	$6.00 \times 10^{-3}$	0.44/0.34	$6.06 \times 10^{-7}$
6q22	rs2817937	6 (121,140,156)	C/T	0.124	0.097	0.110	0.400	0.10/0.00	0.070	$6.23 \times 10^{-6}$	0.152	0.25/0.01	$3.00 \times 10^{-6}$
(HMGGA2)	rs12424086	12 (64,650,776)	C/T	0.228	0.304	$7.78 \times 10^{-5}$	0.027	0.29/0.22	$7.57 \times 10^{-6}$	$9.77 \times 10^{-5}$	0.011	0.22/0.41	$3.64 \times 10^{-6}$

SNPs at genome wide significance  $P < 5 \times 10^{-8}$  and SNPs with suggestive evidence at  $5 \times 10^{-6} > P > 5 \times 10^{-8}$ . The P value of each cohort is corrected for sex, gestational age and population structure using principal components and genomic control. The combined P value is calculated using a fixed effects inverse variance meta-analysis. When no gene is within 50 kb of the SNP the chromosome band is given. Positions of SNPs are reported in NCBI build 36 coordinates. The alleles all refer to the forward strand. The effect allele is defined as the allele associated with later tooth eruption and a smaller number of teeth. % var is the percentage of variance explained by each SNP. P values attaining overall GWA significance are in bold.

doi:10.1371/journal.pgen.1000856.t001

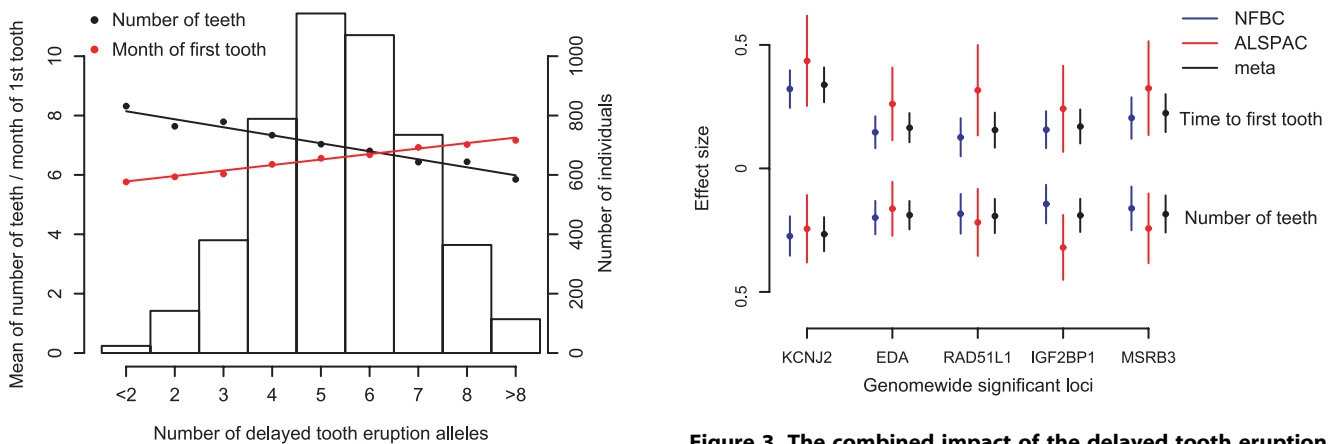


**Figure 1. Linkage disequilibrium and association at loci reaching genome-wide significance for primary tooth development in meta-analysis of NFBC1966 and ALSPAC.** (A) *KCN2* gene region for time to first tooth eruption. (B) *EDA* gene region for time to first tooth. (C) *MSRB3* gene region for time to first tooth. (D) *IGF2BP1* gene region for number of teeth at 12 months (SNP with high  $P$  at 44000 kb is that near *HOXB2*, rs6504340). Note: This is a gene-rich region, so most genes are omitted to simplify the plot. (E) *RAD51L1* gene region for number of teeth at 12 months.  $-\log_{10}$   $p$ -value is plotted against genomic position (NCBI build 36). Most significant SNP in each region is plotted in blue,  $r^2$  with top SNP is colour coded red (0.8 – 1.0), orange (0.5 – 0.8), yellow (0.2 – 0.5), and white <0.2. Gene annotations are based on Genome Browser (RefSeq Genes) and arrows represent direction of transcription. Recombination rate is estimated by LDhat using HapMap CEU sample. All  $r^2$  values are calculated in NFBC1966.

doi:10.1371/journal.pgen.1000856.g001

functions of nearby genes to see if any of these loci were related to tooth development. These signals included SNP rs6504340, which is located between the developmental regulatory genes *HOXB1* (homeobox B1) and *HOXB2* (homeobox B2). Although previous studies have indicated that tooth development is independent of a Hox patterning program [18], Homeobox genes have recently been shown to be expressed in the dental mesenchyme in the pharyngeal teeth of bony fishes [9]. SNP rs6504340 lies 500 kb upstream of rs9674544 in *IGF2BP1*, but

the two SNPs show almost no linkage disequilibrium with each other ( $r^2 = 0.002$  in NFBC1966 and  $r^2 = 0.006$  in ALSPAC). Furthermore, a test for association of rs6504340 conditional on rs9674544 was significant ( $P = 6.3 \times 10^{-5}$  in NFBC1966 and  $P = 0.01$  in ALSPAC; Materials and Methods), indicating that these represent two independent signals (Figure 1). We also identified three SNPs at 2q35, the most significant of which had  $r^2 = 0.48$  with a variant associated with breast cancer [11,19], and SNP rs12424086 located close to the *HMGA2* gene and



**Figure 2. Meta-analysis for primary tooth development by genotype for the five SNPs attaining genome-wide significance.** Estimates and 95% confidence intervals for regression coefficients are given for the effect of delayed teething allele in Gaussian regression on time to first tooth and an ordinal regression on number of teeth.

doi:10.1371/journal.pgen.1000856.g002

**Figure 3. The combined impact of the delayed tooth eruption alleles in 5 identified loci at  $P < 5 \times 10^{-8}$  in the NFBC1966.**

Subjects are classified by the number of delayed tooth eruption alleles. SNPs are chosen so that they had the strongest signal for number of teeth at each locus. Mean time of first tooth eruption is plotted in red and number of teeth by the age of one year in black. The bars represent the number of individuals for each count of 'delayed tooth eruption' alleles. The line through points is a linear regression fit.

doi:10.1371/journal.pgen.1000856.g003

6 kb away from rs1042725, the SNP identified by a GWA study for adult and childhood height [20].

Given the influence of tooth development on dental occlusion, we hypothesized that genetic determinants of early tooth eruption may associate with dental occlusion later in life. We tested for associations between the SNP with the most extreme signal for either phenotype at each of the 10 identified loci and defects in occlusion requiring orthodontic treatment by the age of 31 years in the NFBC1966 (data not available in ALSPAC). A total of 611 individuals (13.5%) reported a defect in occlusion that had required orthodontic treatment. Of the 10 SNPs tested, SNP rs6504340 (*HOXB* gene cluster) gave a significant association, where each G allele (associated with delayed tooth eruption and lower number of teeth in infancy, Table 1), increased the odds of having an occlusal defect requiring orthodontic treatment by 35%, after adjusting for sex (odds ratio (OR) = 1.35, 95% CI = 1.16–1.57;  $P = 1.13 \times 10^{-4}$ ; further adjustment for gestational age did not change the result). A smaller number of teeth at 1 year also predicted higher risk of orthodontic treatment (OR = 1.05, 95% CI = 1.01–1.09;  $P = 0.009$ ). However, when number of teeth or time to first tooth were included in the model with dental occlusion as outcome, the associations with the G allele remained ( $P = 0.001$ ,  $P = 1.71 \times 10^{-4}$ ), suggesting an independent association between rs6504340 and dental occlusion.

## Discussion

Teeth and several other organs have common growth and developmental pathways during early life [21]. The genes at the loci identified in our study have roles in organogenesis, growth and developmental processes, and cancer. Mutations in three of the genes lead to altered organogenesis and development; *KCNJ2* (teeth, jaws, palates, ears, fingers, toes), *EDA* (teeth, hair, sweat glands, salivary glands) and *IGF2BP1* (intestines) [8,13,22]. Of the loci at suggestive levels of significance, the *HOXB* gene cluster is an established regulator of development, and the *HMG2* gene has previously been associated with adult height [20]. Normal development and cancer both involve shifts between cell proliferation and differentiation [23] and genes regulating organ-specific growth are known to be involved in oncogenesis [24]. A previous study identified a common genetic link between an abnormal tooth development and cancer [25]. From our identified loci, *IGF2BP1* and *RAD51L1* have been implicated in cancer [11,14] as have *HOXB2*, 2q35, and *HMG2* [19,26,27].

We provide the first detailed insight into the genetic architecture of primary dentition and our findings could have implications for the study of other developmental and organogenic processes. Exploiting the availability of longitudinal cohort data [28] we found an association between a variant within the *HOXB* gene cluster and the requirement for orthodontic treatment due to defective occlusion by the age of 31 years. Further GWA studies of developmental processes during infancy may establish whether the genetic determinants of infant development can contribute to the study of chronic diseases, such as cancer, that occur later in life.

## Materials and Methods

### Study population and phenotype description

The data was derived from two genome-wide scans of the geographically defined prospective birth cohorts; the NFBC1966 and ALSPAC. The NFBC1966 followed pregnancies in the two northernmost provinces of Finland with expected delivery dates in 1966. ALSPAC recruited mothers during pregnancy with expected dates of delivery between April 1991 and December 1992 from Bristol and the surrounding area in the South West of

England. A total of 4,564 samples were available from the NFBC1966 and 1,518 from ALSPAC. In both cohorts, two separate measures of primary tooth development were collected: i) date of first tooth eruption (in months), and ii) number of teeth (measured at 12 months in NFBC1966 and 15 months in ALSPAC). In the NFBC1966 date of first tooth eruption and number of teeth was gathered by public health professionals during children's monthly visits to child welfare centers (parents carried a booklet where they had recorded the developmental milestones reached). In ALSPAC, parents reported the date of first tooth eruption and number of teeth at 15 months on a questionnaire. In order to ascertain the accuracy of the parental responses, a subsample were examined and validated by a dentist. Information on date of first tooth eruption was available for 4,523 individuals in the NFBC1966 (99% of available GWA samples) and 1396 (92%) in ALSPAC and for number of teeth, 4,326 (95%) in the NFBC1966 and 1,426 (94%) in ALSPAC. All aspects of the study were reviewed and approved by the Ethics Committee of the University of Oulu and the ALSPAC Law and Ethics Committee and by the respective local research committees. Participants (in NFBC1966) and parents (in ALSPAC) gave written informed consent.

### Genotyping

The Illumina HumanCNV370-Duo DNA Analysis BeadChip was used for genotyping the NFBC1966, and Illumina Human-Hap317K BeadChip for ALSPAC. The genotyping and quality control procedures have been described elsewhere [29,30]. SNPs were excluded from the analysis if the call rate in the final sample was <95%, if there was a lack of Hardy-Weinberg Equilibrium (HWE) ( $P < 10^{-4}$  in NFBC1966,  $P < 5 \times 10^{-7}$  in ALSPAC), or if the MAF was <1%. After quality control, 329,091 SNPs in NFBC1966 and 310,611 in ALSPAC were available. We report here the results from the 300,766 genotyped SNPs common to both studies.

### Statistical analyses

Age of first tooth eruption in the NFBC1966 was recorded in months, such that the first tooth could have erupted at any time between the end of previous month and the end of the recorded month. In ALSPAC it was recorded to the nearest month and 3 individuals were recorded as having no teeth after 15 months. To account for the censoring in the two cohorts the outcome was analyzed using parametric survival analysis in the R software package 2.7.1. The Gaussian distribution gave a good fit to the data in both cohorts and was used to model the underlying event time. Number of teeth in the NFBC1966 was recorded at 12 months. In ALSPAC, measurements were taken at around 15 months but there was variability in the exact time of measurement, therefore the ALSPAC analysis was adjusted for age of measurement. Teeth typically erupt in pairs from the upper and lower jaw (75% of children had an even number of teeth in the NFBC1966), making the Poisson distribution inappropriate for modeling the number of teeth. Therefore ordinal logistic regression was used as implemented by the *polr* function in the R package. Analyses of the X chromosome treated males as homozygous females. The allele frequencies of the identified SNPs on the X chromosome did not differ significantly between the sexes. GWA analyses were adjusted for sex, gestational age and population stratification using principal components (PC). Each analysis was corrected for population stratification separately by including those of the top 10 PCs that were associated with the phenotype at  $P < 0.05$  [31]. For number of teeth, PCs 3, 6 and 9 were included in ALSPAC and none in the NFBC1966. For time

to first tooth eruption no PCs were included in ALSPAC and PC 2 was included in the NFBC1966. After correction by PCs, the estimated variance inflation factors [32] for date of first tooth eruption were 1.039 and 1.047 in ALSPAC and NFBC1966 respectively, and 1.011 and 1.039 for number of teeth. Genomic control [32] was then used to correct the residual population stratification. The variance inflation factors from the meta-analyses were 1.012 for number of teeth and 1.015 date of first tooth eruption.

Results from the two studies were combined using fixed effects inverse variance meta-analysis [33]. Analyses were performed using the statistical package R and metaMapper (a meta-analysis software developed in-house). Conditional analyses were calculated using the likelihood ratio test comparing ordinal regression models, one including rs9674544 and the other including rs9674544 and rs6504340. Variance explained by each SNP was computed as 1 minus the ratio of variance of residuals of the model with age, gestational age and SNP to variance of residuals of the model with just age and gestational age. To correct for overfitting, each individual's phenotype was estimated from a model that did not include that individual. The total variance explained by the five loci reaching genome-wide significance was calculated similarly using the most associated SNPs for each phenotype at each locus. Additional tests for association with orthodontic treatment used the SNPs most associated with number of teeth at the 10 loci at  $P < 5 \times 10^{-6}$ . Table 1 reports the top GWA signals at each of the ten loci (i.e. the SNP with the strongest association with either time to first tooth eruption or number of teeth at age 1 year).

## URLs

Jackson Laboratory website, <http://www.jax.org>; NCBI, <http://www.ncbi.nlm.nih.gov>; R project, [www.r-project.org](http://www.r-project.org); UniProt, <http://www.uniprot.org>.

## Supporting Information

**Figure S1** Manhattan plots for the 300,766 SNPs from the genome-wide association meta-analysis for (A) time to first tooth eruption, and (B) number of teeth at 12 months. The (blue) line indicates the genome-wide significance threshold ( $P < 5 \times 10^{-8}$ ).  
Found at: doi:10.1371/journal.pgen.1000856.s001 (0.17 MB PDF)

## References

- Hughes TE, Bockmann MR, Seow K, Gotjamanos T, Gully N, et al. (2007) Strong genetic control of emergence of human primary incisors. *J Dent Res* 86: 1160–1165.
- Nanci A (2008) Ten Cate's oral histology. Development, structure, and function, 7th ed. St Louis: Mosby Elsevier, pg. 96.
- Anderson R, Richmond S, Thomas DW (1999) Patient presentation at medical practices with dental problems: an analysis of the 1996 General Practice Morbidity Database for Wales. *Br Dent J* 186: 297–300.
- Thesleff I (2000) Genetic basis of tooth development and dental defects. *Acta Odontol Scand* 58: 191–194.
- Tucker A, Sharpe P (2004) The cutting-edge of mammalian development; how the embryo makes teeth. *Nat Rev Genet* 5: 499–508.
- Koussoulakou DS, Margaritis LH, Koussoulakos SL (2009) A curriculum vitae of teeth: evolution, generation, regeneration. *Int J Biol Sci* 5: 226–243.
- Benko S, Fantes JA, Amiel J, Kleijnian DJ, Thomas S, et al. (2009) Highly conserved non-coding elements on either side of SOX9 associated with Pierre Robin sequence. *Nat Genet* 41: 359–364.
- Yoon G, Oberoi S, Tristani-Firouzi M, Etheridge SP, Quitania L, et al. (2006) Andersen-Tawil syndrome: prospective cohort analysis and expansion of the phenotype. *Am J Med Genet A* 140: 312–321.
- Fraser GJ, Hulsey CD, Bloomquist RF, Uyesugi K, Manley NR, et al. (2009) An ancient gene network is co-opted for teeth on old and new jaws. *PLoS Biol* 7: e31. doi:10.1371/journal.pbio.1000031.
- Mues GI, Griggs R, Hartung AJ, Whelan G, Best LG, et al. (2009) From ectodermal dysplasia to selective tooth agenesis. *Am J Med Genet A* 149A: 2037–2041.
- Thomas G, Jacobs KB, Kraft P, Yeager M, Wacholder S, et al. (2009) A multistage genome-wide association study in breast cancer identifies two new risk alleles at 1p11.2 and 14q24.1 (RAD51L1). *Nat Genet* 41: 579–584.

**Figure S2** Manhattan plots and linkage disequilibrium (LD) diagrams for five identified loci ( $P < 5 \times 10^{-8}$ ). (A) Locus 17q24 (*KCNJ2*), (B) Locus Xq13 (*EDA*), (C) Locus 14q24 (*RAD51L1*), (D) Locus 17q21.4 (*IGF2BP1*), and (E) Locus 12q14 (*MSRB3*). The (blue) line indicates the genome-wide significance threshold ( $P < 5 \times 10^{-8}$ ).

Found at: doi:10.1371/journal.pgen.1000856.s002 (0.89 MB PDF)

**Figure S3** Quantile-quantile plots of observed  $-\log_{10} P$  values versus the expectation under the null for (A) time to first tooth eruption and (B) number of teeth at 12 months. The most associated 10,000 SNPs from the meta-analysis are shown.

Found at: doi:10.1371/journal.pgen.1000856.s003 (0.07 MB PDF)

**Figure S4** Additive effect of delayed tooth eruption alleles in identified loci in ALSPAC. Subject classified by the number of delayed tooth eruption alleles. SNPs chosen had the strongest signal for time to first tooth eruption at each locus. Mean time of first tooth eruption is plotted in black. The bars represent the number of individuals for each count of “delayed tooth eruption” alleles. Lines through points are linear regression fits.

Found at: doi:10.1371/journal.pgen.1000856.s004 (0.05 MB PDF)

**Table S1** Summary of the candidate genes located within the top loci.

Found at: doi:10.1371/journal.pgen.1000856.s005 (0.08 MB PDF)

## Acknowledgments

We are grateful to all the families who took part in this study, the midwives for their help in recruiting them, Professors Paula Rantakallio and Jean Golding, founders of these cohort studies, and the whole NFBC 1966 and ALSPAC teams. We thank the Sample Logistics and Genotyping Facilities both at the Wellcome Trust Sanger Institute and the Broad Genotyping Center for generating the ALSPAC and NFBC1966 genome wide genetic data.

This publication is the work of the authors and they will serve as guarantors for the contents of this paper.

## Author Contributions

Conceived and designed the experiments: DP MRJ. Analyzed the data: DP CJH DME PC LJMC. Wrote the paper: DP CJH DME PFO KS RL IYM MK GN DB US AP NF ALH JL SV BG PC NJT SMR GD WZ MIM PD LP PE LJMC GDS MRJ.

- Wise GE, Yao S, Liu D (2006) Injections of osteoprotegerin and PMA delay tooth eruption. *Clin Anat* 19: 19–24.
- Hansen TV, Hammer NA, Nielsen J, Madsen M, Dalbaeck C, et al. (2004) Dwarfism and impaired gut development in insulin-like growth factor II mRNA-binding protein 1-deficient mice. *Mol Cell Biol* 24: 4448–4464.
- Gu L, Shigemasa K, Ohama K (2004) Increased expression of IGF II mRNA-binding protein 1 mRNA is associated with an advanced clinical stage and poor prognosis in patients with ovarian cancer. *Int J Oncol* 24: 671–678.
- Pemberton TJ, Li F-Y, Oka S, Mendoza-Fandino GA, Hsu Y-H, et al. (2007) Identification of novel genes expressed during mouse tooth development by microarray gene expression analysis. *Dev Dyn* 236: 2245–2257.
- Manolio TA, Collins FS, Cox NJ, Goldstein DB, Hindorf LA, et al. (2009) Finding the missing heritability of complex diseases. *Nature* 461: 747–753.
- Weedon MN, Lango H, Lindgren CM, Wallace C, Evans DM, et al. (2008) Genome-wide association analysis identifies 20 loci that influence adult height. *Nat Genet* 40: 575–583.
- James CT, Ohazama A, Tucker AS, Sharpe PT (2002) Tooth development is independent of a Hox patterning programme. *Dev Dyn* 225: 332–335.
- Stacey SN, Manolescu A, Sulem P, Rafnar T, Gudmundsson J, et al. (2007) Common variants on chromosomes 2q35 and 16q2 confer susceptibility to estrogen receptor-positive breast cancer. *Nat Genet* 39: 865–869.
- Weedon MN, Lettre G, Freathy RM, Lindgren CM, Voight BF, et al. (2007) A common variant of HMG2 is associated with adult and childhood height in the general population. *Nat Genet* 39: 1245–1250.
- Thesleff I, Vaahtokari A, Partanen AM (1995) Common molecular mechanisms regulating the development of teeth and other organs. *Int J Dev Biol* 39: 35–50.

22. Mikkola ML, Theleff I (2003) Ectodysplasin signalling in development. *Cytokine Growth Factor Rev* 14: 211–224.
23. Nunes FD, de Almeida FC, Tucci R, de Sousa SC (2003) Homeobox genes: a molecular link between development and cancer. *Pesqui Odontol Bras* 17: 94–98.
24. Hallikas O, Palin K, Sinjashina N, Rautiainen R, Partanen J, et al. (2006) Genome-wide prediction of mammalian enhancers based on analysis of transcription-factor binding affinity. *Cell* 124: 47–59.
25. Lammi L, Arte S, Somer M, Jarvinen H, Lahermo P, et al. (2004) Mutations in AXIN2 cause familial tooth agenesis and predispose to colorectal cancer. *Am J Hum Genet* 74: 1043–1050.
26. Collins Y, Tan DF, Pejovic T, Mor G, Qian F, et al. (2005) Identification of differentially expressed genes in clinically distinct groups of serous ovarian carcinomas using cDNA microarray. *Int J Mol Med* 14: 43–53.
27. Grier DG, Thompson A, Kwasniewska A, McGonigle GJ, Halliday HL, et al. (2005) The pathophysiology of HOX genes and their role in cancer. *J Pathol* 205: 154–171.
28. Manolio TA (2009) Cohort studies and the genetics of complex disease. *Nat Genet* 41: 5–6.
29. Sabatti C, Service SK, Hartikainen AL, Pouta A, Ripatti S, et al. (2009) Genome-wide association analysis of metabolic traits in a birth cohort from a founder population. *Nat Genet* 41: 35–46.
30. Timpson NJ, Tobias JH, Richards JB, Soranzo N, Duncan EL, et al. (2009) Common variants in the region around Osterix are associated with bone mineral density and growth in childhood. *Hum Mol Genet* 18: 1510–1517.
31. Novembre J, Stephens M (2008) Interpreting principal component analyses of spatial population genetic variation. *Nat Genet* 40: 646–649.
32. Devlin B, Roeder K (1999) Genomic control for association studies. *Biometrics* 55: 997–1004.
33. Sutton AJ, Abrams KR, Jones DR, Sheldon TA, Song F (2000) *Methods for meta-analysis in medical research*. Chichester, UK: John Wiley & Sons, pg. 58.