

Original Article

Accuracy and the Influence of Marrow Fat on Quantitative CT and Dual-Energy X-ray Absorptiometry Measurements of the Femoral Neck In Vitro

J. W. Kuiper¹, C. van Kuijk¹, J. L. Grashuis¹, A. G. H. Ederveen² and H. E. Schütte¹

¹Department of Experimental Radiology, Erasmus University Medical School and University Hospital 'Dijkzigt', Rotterdam; and

²Department of Endocrinology, Organon Scientific Development Group, Organon International, Oss, The Netherlands

Abstract. Bone mineral measurements with quantitative computed tomography (QCT) and dual-energy X-ray absorptiometry (DXA) were compared with chemical analysis (ChA) to determine (1) the accuracy and (2) the influence of bone marrow fat. Total bone mass of 19 human femoral necks in vitro was determined with QCT and DXA before and after defatting. ChA consisted of defatting and decalcification of the femoral neck samples for determination of bone mineral mass (BmM) and amount of fat. The mean BmM was 4.49 g. Mean fat percentage was 37.2% (23.3%–48.5%). QCT, DXA and ChA before and after defatting were all highly correlated ($r > 0.96$, $p < 0.0001$). Before defatting the QCT values were on average 0.35 g less than BmM and the DXA values were on average 0.65 g less than BmM. After defatting, all bone mass values increased; QCT values were on average 0.30 g more than BmM and DXA values were 0.29 g less than BmM. It is concluded that bone mineral measurements of the femoral neck with QCT and DXA are highly correlated with the chemically determined bone mineral mass and that both techniques are influenced by the femoral fat content.

Keywords: Absorptiometry; Accuracy; Bone; Comparative study; Dual-energy X-ray; Femur; In vitro; Marrow fat, Quantitative CT

Introduction

Single energy quantitative computed tomography (QCT) and dual energy X-ray absorptiometry (DXA) are commonly used non-invasive methods for measuring bone mineral content (BMC) and/or density (BMD). A reported disadvantage of QCT is the influence of the variable marrow fat content of bone on the accuracy of bone mineral measurements. Estimates of the accuracy-error of QCT measurements in various body sites range from 2% to 30% [1,2]. Dual-energy QCT methods were suggested as a solution to the fat-error problem and render higher accuracy, but at the cost of precision [1,3]. DXA and the preceding technique of dual photon absorptiometry (DPA) utilize a dual-energy method which separates soft tissue from bone but does not correct for marrow fat [4]. Various influences of marrow and soft tissue fat of DXA measurements on the spine have been reported [5–8]. Magnetic resonance studies show that the amount of marrow fat in the skeleton varies with age and location [9,10] and is known to be influenced by state of health and medication [11].

The proximal femur, like the spine, is an area where many osteoporotic fractures occur. In particular the femoral neck has, unlike the spine, a complex geometry and a non-uniform distribution of bone mineral and marrow fat, which is likely to influence bone mineral measurements. Measurement of bone mineral in the proximal part of the femur with the use of DXA has become more or less standardized and generally applicable. Bone mineral measurements with QCT in the

femur are still experimental. Various authors have presented QCT studies on the proximal part [12–14], the shaft [3] and on the distal part (e.g. condyles) [15,16], but standardization of a QCT method for measuring bone mineral in the proximal femur has not been established.

The aim of this study was to evaluate the measurement of bone mass with QCT and DXA in the femoral neck and to compare the results with chemical analysis (ChA) in order to determine the accuracy of the bone mineral measurements. In addition, the influence of marrow fat on these measurements were investigated.

Materials and Methods

Femoral Neck Specimen

Nineteen femurs from embalmed human cadavers (10 males and 9 females, mean age 83.2 ± 4.5 years) were used. The proximal femurs were cleaned from all surrounding bones and soft tissues and mounted on a Perspex plate in a position comparable to the anatomical position of a recumbent patient. Long-distance radiographs of the mounted proximal femur were used to position the sawing lines for the femoral neck. Positioning of the sawing lines was done with the use of a template which had positioning lines perpendicular to the longitudinal axis of the femoral neck. These lines were drawn on the Perspex mounting plate. The femur was then frozen (-80 °C) to prevent loss of bone marrow during sawing, and a 20-mm thick slice of the femoral neck was taken out with the use of a high-speed band saw. This 20-mm thick slice was used for bone mass measurements using QCT, DXA and ChA.

Chemical Analysis

The individual constituent of bone (i.e. fat, bone mineral, collagen, etc.) were determined with ChA according to the following procedure. Directly after sawing the weight of bone samples was measured (WetM), and the samples were then degassed underwater at -0.95 bar for 24 h at room temperature. The bone samples were placed in the mounting device to prevent loss of bone marrow. After degassing, QCT and DXA scans (session 1; QCT 1 and DXA 1) were done. The samples were then dried under vacuum at a temperature of 45 °C until constant weight was attained; this was considered dry mass (DryM1). Next the dried bones were defatted in 100% trichloroethylene for 2 weeks and subsequently dried and weighed again (DryM2). The defatted samples were again degassed under water and QCT and DXA measurements (session 2; QCT 2 and DXA 2) were done. After the second scan session the samples were dried and decalcified in decalcification fluid (37 g/l sodium formiate and 170 ml/l formic acid in aqua dest) for 2 weeks with a fluid change after 1 week. Thereafter the remainder (OrgM) was dried again until

constant weight; the bone mineral mass (BmM) could then be calculated. The difference between DryM1 and DryM2 was considered to be the amount of fat (FatM). The amount of fat was also calculated as the percentage of WetM in a bone sample. This was considered to give the best representation of the physiological state of bone in vivo. OrgM was considered to be the organic constituents of the bone matrix. BmM was considered to be the amount of bone mineral in the bone sample.

QCT Measurements

A Perspex mounting device was developed for reproducible fixation of the femur specimen (Fig. 1, shaded part). This mounting device was placed in a circular water bath together with solid reference material. The femurs were degassed at room temperature, positioned in the mounting device and then scanned. CT scans were done using a Siemens Somatom Plus (Siemens, Erlangen, Germany) after calibration and quality checks of the CT system. Scanning parameters were: 80 kV, 125 mA, 1 s scan time. The water bath was positioned in the center of the gantry and 12 contiguous scans were done with a slice thickness of 2 mm, ensuring complete scanning of the specimen. The first and the last scan were done through the Perspex plates which enclosed the femoral neck. To ensure a reproducible slice position in the specimen, the starting position was selected from a scout scan using a thin metal ring fixed on the mounting device as reference point.

From scans containing femoral neck images (generally 10), CT values of the object and reference material were obtained, using an ISG Allegro 3-D workstation with version 5.1 software. For seed-controlled contour detection of the femur a threshold of 78 HU was selected. This threshold was used because it allowed an optimal distinction between the bone sample and the surrounding water bath. After a contour had been drawn the mean pixel value within this contour was determined. Solid reference material (Image Analysis

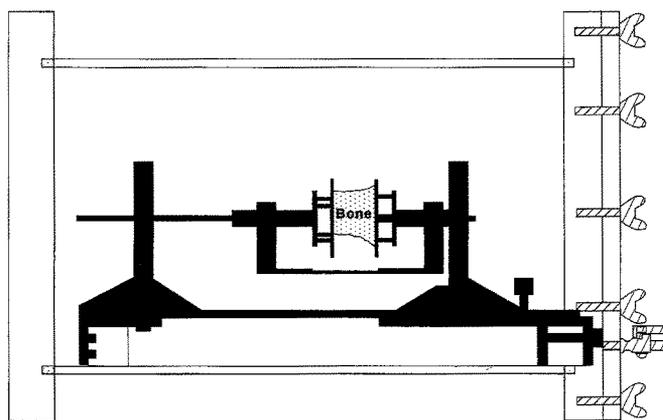


Fig. 1. Set-up of the mounting device and water bath for QCT measurements. The mounting device (shaded part) is shown with a sample of femur neck positioning in center (side view).

Inc., Columbia, Ky) was used for calibration. Using the measured volume and BMD of all 2-mm scans the total bone mass of the femoral neck was calculated for sessions 1 and 2.

DXA Measurements

The mounting device described above was also used for DXA measurements. It was placed in a square water bath with a water level of 15 cm, stimulating soft tissue. Scans were done using a Lunar DPX-L scanner with version 1.3 software. After calibration and quality assurance tests, scans were done using the Spine scan option in Medium scan mode with a sample size of 1.2×1.2 mm at 3000 μ A current. Spine scan option was used because there was no complete femur and it allowed a fixed start position and scans perpendicular to the longitudinal axis of the femoral neck. All scans were manually analyzed by the same person and the BMC (in grams) for each femoral neck specimen calculated.

Statistical Analysis

Statistical analysis was performed with the use of Statgraphics, Statistical Graphics System, version 6.0 software (STSC Inc., Rockville, MD).

Results

Results of ChA, QCT and DXA measurements are presented in Table 1; all weights are in grams. Correlations between QCT, DXA and BmM measurements

before and after defatting were all high ($r > 0.96$), p values < 0.0001).

Accuracy

The initial bone mineral measurements show that the mean QCT value (QCT 1) was 0.35 g less than mean BmM and the mean DXA value (DXA 1) was 0.65 g less than mean BmM. Compared with ChA-BmM, QCT 1 results show a consistent underestimation of BmM, ranging from 0.31 to 0.37 g (Fig. 2). The slope of the regression line, at 0.984, is close to optimal. An accuracy error for QCT can therefore be presented as a mean difference of -0.35 g between QCT-BmM and ChA-BmM.

DXA 1 results also show an underestimation of ChA-BmM, ranging from 0.36 to 0.99 g (Fig. 3). The slope of the regression line, at 0.880, is less optimal. DXA results tend to produce a larger underestimation of ChA-BmM at higher bone mass value. Thus, the DXA accuracy error is not a constant value and, therefore, can better be expressed as a relative value of 14.6% (percentage of ChA-BmM).

Fat Influence

Defatting of the femoral neck gave a mean fat percentage by weight of $37.2 \pm 7.9\%$ with a range of 23.3%–48.5%.

Compared with bone mass measurements before defatting, QCT and DXA showed an increase for both methods after defatting. The QCT 2 measurements increased 0.64 g and the values rose above the ChA-

Table 1. Results of ChA, QCT and DXA measurements (in grams)

No.	WetM	DryM1	DryM2	FatM	OrgM	BmM	QCT 1	QCT 2	DXA 1	DXA 2
1	23.78	16.94	9.15	7.80	4.06	5.09	4.85	5.41	4.23	4.56
2	27.12	21.59	8.74	12.85	3.47	5.27	4.35	5.39	4.21	4.59
3	23.56	19.23	8.07	11.16	3.48	4.59	4.46	5.01	3.85	4.37
4	18.68	13.84	4.78	9.05	2.27	2.51	1.97	2.74	1.95	2.42
5	23.55	18.28	7.44	10.84	3.31	4.14	3.67	4.62	3.25	4.03
6	25.11	19.97	10.48	9.49	4.21	6.27	5.83	6.43	5.20	5.80
7	21.26	17.01	11.66	5.35	4.34	7.32	6.92	7.22	6.66	6.67
8	15.74	10.45	6.79	3.66	2.85	3.94	3.76	4.02	3.65	3.76
9	21.63	15.60	8.98	6.62	3.94	5.04	4.38	4.87	4.18	4.61
10	25.11	18.23	8.14	10.09	3.73	4.41	4.07	4.91	3.67	4.17
11	11.25	9.01	3.75	5.27	1.65	2.10	1.76	2.16	1.74	1.95
12	22.52	17.65	9.46	8.20	3.96	5.49	5.41	6.23	4.79	5.28
13	27.09	19.83	10.23	9.60	4.59	5.64	5.29	6.16	5.00	5.28
14	17.92	13.28	7.19	6.09	3.37	3.82	3.44	3.93	3.50	3.44
15	16.13	10.29	6.22	4.07	2.68	3.54	3.31	3.62	3.17	3.27
16	18.97	14.93	7.20	7.73	3.34	3.86	3.76	4.44	3.33	3.73
17	19.45	15.03	7.33	7.70	3.23	4.10	3.94	4.60	3.45	3.88
18	22.65	17.00	8.63	8.36	3.82	4.81	4.61	5.34	4.09	4.81
19	13.98	10.47	5.98	4.50	2.67	3.31	2.88	3.78	3.04	3.19
Mean	20.82	15.72	7.91	7.81	3.42	4.49	4.14	4.78	3.84	4.20
SD	4.42	3.68	1.96	2.56	0.75	1.25	1.25	1.26	1.12	1.13

For abbreviations see text.

Fig. 2. Results of QCT versus ChA (BmM) measurements of femur neck. QCT 1, session 1 (scan results on complete bone sample); QCT 2, session 2 (scan results of defatted bone samples).

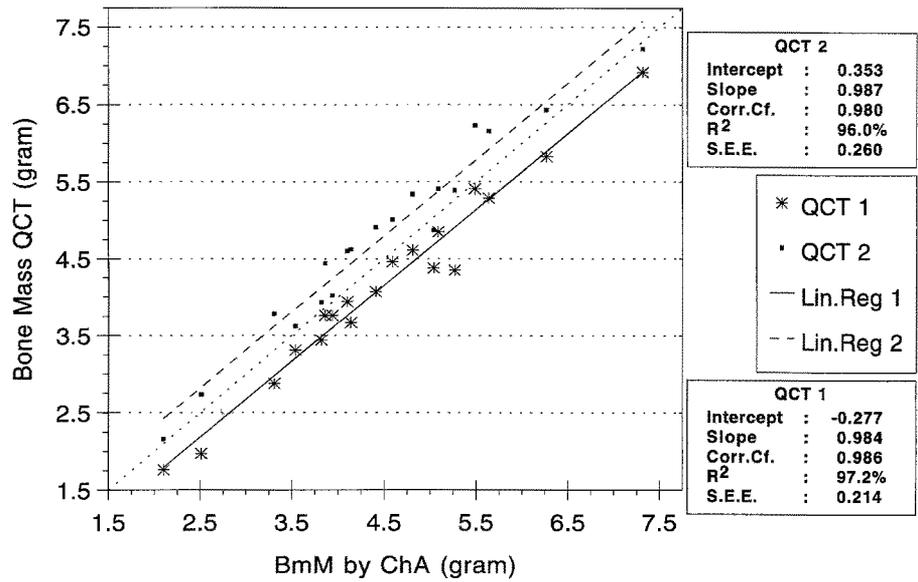


Fig. 3. Results of DXA versus ChA (BmM) measurements of femur neck. DXA 1, session 1 (scan results on complete bone sample); DXA 2, session 2 (scan results of defatted bone samples).

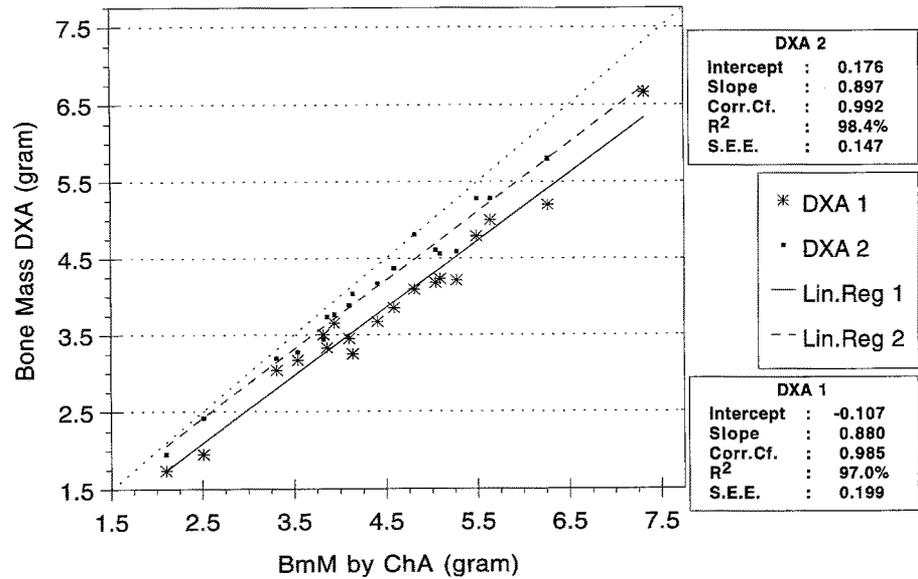
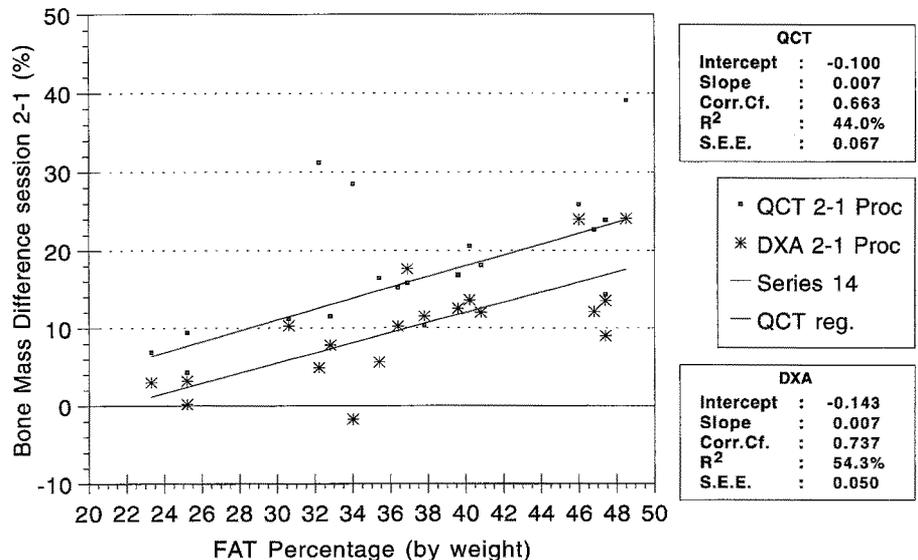


Fig. 4. Differences between sessions 1 and 2 (normal and defatted bone respectively) as a percentage of the baseline measurement for QCT and DXA measurements versus fat percentage of the femoral neck determined by ChA.



BmM values. The mean QCT 2 value was 0.30 g more than BmM (range 0.26–0.33 g, Fig. 2). The DXA measurements after defatting (DXA 2) increased by 0.36 g and still gave an underestimation of the ChA-BmM values in the range 0.04–0.58 g (Fig. 3).

Figure 4 shows the percentage differences between measurements before and after defatting versus the fat percentage by weight. The regression lines show that for QCT the measurement error due to the variable amount of fat in the femoral neck ranges from 7.2% to 25.3%. For DXA these values are between 1.2% and 17.6% respectively. In this study, the average increase in bone mass given as a percentage of the initially measured bone mass is, for both methods, about 0.7% per percentage increase in fat weight. This shows that when there is a higher fat percentage by weight in the femoral neck, the measurement errors of both QCT and DXA increase.

Discussion

In this study the QCT measurements were, as for DXA, for cortical and trabecular bone together. We consider that this is the only way to make an optimal comparison between the two radiological methods and ChA. An ashing method was not used on the bone samples. Our ChA has the advantage over ashing that the various compartments (e.g. water, fat, bone mineral and organic constituents) can be separated stepwise, thus allowing measurements at each stage.

There are two minor issues that were not considered in this study. We used material from embalmed human cadavers. Reports on the influence of the fixative on bone mineral measurements are lacking. We assume that this influence in our study is small and cannot account for differences between measurements before and after defatting.

Also, Whitehouse et al. [17] suggested that, when compared with *in vivo* measurements, temperature differences between objects and calibration material may give rise to an underestimation of accuracy errors (related to marrow fat) in QCT bone mineral measurements *in vitro*. This temperature factor will probably influence both QCT and DXA results *in vitro*. All our measurements were at room temperature and may therefore underestimate fat-related errors *in vivo*.

Accuracy

Our accuracy measurements showed that both methods underestimated the amount of bone mineral in the femoral neck. The trends show that QCT has a consistent underestimation while for DXA the measurement error increases at higher bone masses. The accuracy is influenced by a number of factors, one of which is the variable amount of marrow fat. Other, mainly technical factors are more or less the same for QCT and DXA.

For QCT, besides the variation by fat, the choice of

calibration material may affect the initial underestimation and later the overestimation [18,19]. For DXA the exact calibration procedures are not clear but Mazess et al. [20] reported that the calibration material used is considered good when the fat content is about 1.5–2 times the mass of the ash content of the specimen. This does not explain the deviation at higher BmM values and the underestimation of the ChA-BmM values in our study, as the fat content was about 1.7 times the bone content. Ho et al. [21] reported comparable results for DXA of the spine, with a tendency to underestimate ash weight and an accuracy error of 8.9%.

Other, possibly minor influences on the measurements include the partial volume effects and beam-hardening artifacts. Both QCT and DXA have the problem of partial volume effects. With QCT, using a 2 mm scan thickness, partial volume effects are inevitable in scans at the edges of the bone samples and these scans could not always be optimally analyzed. For DXA a scanline was 1.2 mm wide, and low scan profiles at the edges of the bone were excluded from measurement. Because of these partial volume effects there is small tendency for both methods to underestimate the chemically determined bone mass. These partial volume effects only influence the absolute accuracy, not the estimates of the fat error. The position of the calibration material for QCT measurements was inside the water tank and close to the (relatively small) object. Beam-hardening effects and field non-uniformity should, therefore, be minimal and/or have little effect on the QCT results. The existence of beam-hardening effects for DXA are reported by Goodsitt [7] and Blake et al. [22], but the magnitude of these errors in our study is unclear.

Fat Influence

In our bone samples we found a 25.2% variation in fat content. Both QCT and DXA are clearly influenced by the variable amount of fat and show an identical trend, with a 0.7% difference in measured bone mass for every 1% of fat weight. The only difference is the intercept of the regression lines. However, correlations and R^2 are relatively low and the values for both methods show considerable spread and overlap.

The QCT method used was a single-energy measurement. DXA is a dual-energy method, but should not be confused or compared with dual-energy QCT. Dual-energy QCT uses the two energies to estimate the fat content and the bone mineral content. DXA utilizes the dual energies for separation of the soft tissue from bone. It can roughly estimate the fat content of soft tissue, but cannot estimate the fat content of bone. According to Mazess et al. [20] a decrease of 0.05 g/cm² for each 1 g/cm² of fat can be expected with DXA measurements.

Our results show that, before and after defatting, both QCT and DXA are valid techniques for measuring bone mineral in the femoral neck. However, both

techniques are influenced by the variable amount of marrow fat. The variation in amount of fat may not only be age specific but also exists between individuals. This will cause problems in longitudinal and cross-sectional measurements. Possible changes in amount of fat, as occurs in some therapies and diseases (e.g. corticosteroids, Gaucher diseases (Cushing disease) may also influence outcome of longitudinal bone mineral measurements.

The results of this study can provide basic information for use in interpreting future *in vivo* and clinical QCT and DXA measurements on the proximal femur. Further research is aimed at *in vitro* measurements with QCT and DXA on proximal femurs with the surrounding (pelvis) bone and soft tissue and at local variations in the amount of fat throughout the femoral neck.

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