

ICAM⁻ MELANOMA CELLS ARE RELATIVELY RESISTANT TO CD3-MEDIATED T-CELL LYSIS

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The primary activation pathway of T cells is via the T-cell receptor (TCR)/CD3 complex, which is functionally interrelated with various accessory molecules. We examined the contribution of the lymphocyte-function-associated antigen-1/intercellular adhesion molecule 1 (LFA-1/ICAM-1) interaction to CD3/TCR-mediated lysis by cytotoxic T lymphocytes (CTL). We used ICAM-1^{-or+} tumor cell lines as target cells and anti-CD3- or anti-LFA-1 containing hetero-cross-linked monoclonal antibody (MAB) to bridge CTL and target cells and simultaneously to activate CTL. The ICAM-1⁻ melanoma-derived cell line IgR39 was relatively resistant to CD3-mediated lysis by both TCR $\alpha\beta$ ⁺ and TCR $\gamma\delta$ ⁺ CTL, when compared with ICAM-1⁺ cell lines. Induction of ICAM-1 on the membrane of IgR39 cells by tumor necrosis factor (TNF) rendered these cells more susceptible to CD3-mediated lysis. Anti-ICAM-1 MAB inhibited this TNF-enhanced susceptibility to lysis, directly demonstrating that the induction of ICAM-1 was critical in the TNF-induced increase in susceptibility to lysis of IgR39 cells. CTL formed less efficient conjugates with the ICAM-1⁻ cells as compared to ICAM-1⁺ cells. Both spontaneous and CD3-induced conjugate formation as well as CD3-mediated lysis of ICAM-1⁻ tumor cells by CTL were enhanced by the addition of anti-LFA-1 containing hetero-cross-linked MAB, thereby mimicking the LFA-1/ICAM-1 interaction between CTL and target cells. Soluble anti-CD18 MAB inhibited CD3-mediated lysis of ICAM-1⁻ target cells by CTL without affecting their conjugate formation. Anti-LFA-1 MAB added after conjugate formation still inhibited lysis of both ICAM-1⁺ or⁻ tumor cells. Taken together, these findings suggest that the LFA-1/ICAM-1 interaction co-activates CD3/TCR-mediated lysis by CTL through both an enhanced CTL-target cell binding and the delivery of post-conjugate costimulatory signals.

T cells can be activated by the antigen-specific T-cell receptor (TCR) interacting with non-self major histocompatibility complex (MHC) proteins or foreign antigen associated with self MHC proteins (Ashwell and Schwartz, 1986; Martz, 1977). Lymphocytes may also be activated via non-polymorphic cell-surface receptors such as CD2, CD3 or CD16 interacting with their ligands or monoclonal antibody (MAB) (Meuer *et al.*, 1984; Bolhuis *et al.*, 1986; Van de Griend *et al.*, 1987). Recent evidence suggests that these activation sites are functionally interrelated, for instance, activation via CD2 can be regulated via CD3 and vice versa (Van de Griend *et al.*, 1987; Goedegebuure *et al.*, 1989).

The interaction between cytotoxic T lymphocytes (CTL) and the relevant target cells results in target-cell lysis by a multi-step process (Martz, 1977; Mentzer *et al.*, 1987). The initial adhesion step is the rapid formation of an intercellular bond (conjugate formation) which is antigen-independent. Two major pathways of adhesion have been identified. First, lymphocyte-function-associated antigen-1 (LFA-1) on CTL interacts with intercellular adhesion molecule-1 (ICAM-1) on the target cell (Shaw *et al.*, 1986; Makgoba *et al.*, 1988a). Second, the effector cell CD2 interacts with LFA-3 on the target cell (Selvaraj *et al.*, 1987; Dustin *et al.*, 1987; Takai *et al.*, 1987; Hunig *et al.*, 1987). Subsequently, antigen recognition via the TCR results in triggering of the CTL lytic machinery and target-cell lysis (Martz, 1977).

LFA-1 is a trans-membrane heterodimer composed of a 180-

kDa α chain (CD11a) and a 95-kDa β chain (CD18) which is expressed by most leukocytes (Springer *et al.*, 1987; Martz, 1987; Krensky *et al.*, 1983). In addition to its adhesion function, LFA-1 is also thought to be involved in T-cell activation, possibly as a signal transduction molecule, as reported for CD2 (Hunig *et al.*, 1987). Binding of anti-LFA-1 α or anti-LFA-1 β MABs to T cells has been shown to up- or down-regulate, respectively, anti-CD3 MAB-triggered T-cell proliferation in the absence of intercellular adhesion (Van Noesel *et al.*, 1988). In mice, anti-LFA-1 α MAB enhances proliferative responses of pre-activated T and B lymphocytes (Pircher *et al.*, 1986; Mishra *et al.*, 1986). Finally, a unique anti-LFA-1 α MAB induces homotypic cell-cell interactions (Keizer *et al.*, 1988).

ICAM-1 is a monomeric 90–114-kDa glycoprotein which can be expressed on cells of many lineages, especially at specific differentiation stages or after exposure of cells to certain inflammatory mediators *in vitro* or *in vivo* (Dustin *et al.*, 1986). Biochemical and functional studies have conclusively demonstrated that ICAM-1 is one of the ligands for LFA-1 (Makgoba *et al.*, 1988a,b; Marlin and Springer, 1987; Simmons *et al.*, 1988; Rothlein *et al.*, 1986). Target-cell ICAM-1 is involved in adhesion to and lysis of target cells; however, demonstration of this role is restricted to target cells which express ICAM-1 as the major LFA-1 ligand (Makgoba *et al.*, 1988b). Furthermore, ICAM-1 is a major contributor to a variety of other T-cell responses dependent on cell-cell interaction (Boyd *et al.*, 1988; Dougherty *et al.*, 1988; Altmann *et al.*, 1989; Schulz *et al.*, 1988). A previously suggested second ligand for LFA-1, *i.e.*, ICAM-2, has been identified (Staunton *et al.*, 1989). However, at this point it is unclear whether binding of the LFA-1 to the ICAM-1 ligand molecule not only facilitates the adhesion of the effector cell to the target cell, but also mediates signals that regulate CTL activation.

Lysis-resistant Burkitt's lymphoma cells express low levels of LFA-1, LFA-3 and ICAM-1 (Gregory *et al.*, 1988a; Billaud *et al.*, 1987). Epstein-Barr virus (EBV)-positive biopsy cells with a low expression of these molecules are relatively resistant to lysis by specific CTL. Prolonged *in vitro* culture of these cells induces an increased expression of the molecules which is associated with a more efficient interaction with specific CTL. These findings prompted speculation that the lack of expression of ICAM-1 and other adhesion molecules on tumor cells contributes to their escape from immunosurveillance (Gregory *et al.*, 1988a,b; Clayberger *et al.*, 1987).

The importance of the LFA-1/ICAM-1 interaction led us to further study its involvement in CTL/target-cell conjugate formation and triggering of lysis. To this end, we used ICAM-1-positive or -negative tumor-cell lines as target cells and bi-specific Ab-heteroconjugates, containing an anti-CD3 MAB

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which is covalently coupled to an anti-DNP antibody. Such bispecific reagents efficiently bridge effector cells and trinitrobenzene sulfonate (TNP)-modified target cells and simultaneously trigger the lytic machinery of the effector cell (Goedegebuure *et al.*, 1989; Karpovsky *et al.*, 1984). Our data demonstrate that induction of ICAM-1 expression on target cells enhances their susceptibility to CD3-triggered lysis. Associated findings suggest that this co-stimulatory effect results from enhanced conjugate formation as well as enhanced signal transduction by LFA-1/ICAM-1 interactions.

MATERIAL AND METHODS

Cytotoxic clones

CTL clones were obtained by limiting dilution cloning under polyclonal activation conditions as we described earlier (Van de Griend *et al.*, 1987). In short, CTL clones were expanded in 96-well round-bottomed microtiter plates (Greiner Labor Technik, Phidelsheim, FRG). One to 3×10^3 cloned CTL per well were stimulated with a mixture of irradiated (25 Gy) allogeneic PBL (2×10^4) and 1×10^4 Epstein-Barr virus (EBV)-transformed lymphoblastoid B-cell lines (BSM and APD) in a final volume of 200 μ l. The culture medium was RPMI 1640 (Flow Laboratories, Rockville, MD) buffered with bicarbonate (2 g/l) and HEPES (4.8 g/l) and supplemented with 10% non-heat-inactivated (pooled) human serum, 25 U/ml recombinant IL-2 (EuroCetus, Amsterdam, The Netherlands), 4 mM glutamine, 100 IU/ml penicillin and streptomycin, 1 μ g/ml indomethacin and 1 μ g/ml leukoagglutinin (HA15, Pharmacia, Uppsala, Sweden). Clones were harvested and replated with new feeder cells and fresh medium at weekly intervals.

Antibodies

Bispecific hetero-cross-linked antibodies (Abs) were prepared with the use of the cross-linking reagent N-succinimidyl-3-(2-pyridylthiol propionate) (SPDP) as described in detail elsewhere (Karpovsky *et al.*, 1984). Hetero-cross-linked Abs used in this study were: anti-CD3 (OKT3) \times anti-DNP; anti-CD18 (MHM23) \times anti-DNP and anti-HLA Class I (W6/32) \times anti-DNP MAb. Anti-DNP Abs were purified polyclonal rabbit Abs. MAbs used for the inhibition of lymphocyte functions and for immunofluorescence staining were: CLB54 (anti-CD18) (Miedema *et al.*, 1984), specific for the LFA-1 β chain, a generous gift from Dr. R.A.W. van Lier (CLB, Amsterdam, The Netherlands); MAb 84H10 (anti-ICAM-1) (Makgoba *et al.*, 1988a) and W6/32 (anti-HLA Class-I common epitope) (Barnstable *et al.*, 1978).

Cytotoxicity assay

Cytotoxic activity was measured in a standard 3-hr ^{51}Cr release assay (Van de Griend *et al.*, 1987; Goedegebuure *et al.*, 1989). Briefly, varying numbers of effector cells were added in triplicate in 75 μ l/well in 96-well round-bottomed microtiter plates. The hetero-cross-linked Ab (75 μ l) was added to the microtiter well at the indicated concentrations 30 min before addition of ^{51}Cr -labelled target cells (100 μ l). Target cells were incubated with 0.3 mM trinitrobenzene sulfonate (TNP) for 15 min at 37°C and then washed. ICAM-1 expression on IgR39 tumor cells was enhanced by incubation with 1000 U/ml recombinant human tumor necrosis factor (recTNF) (Cetus Corp., Emeryville, CA) for 24 h. Tumor cell lines used as target cells in this study include IgR39 and IgR37, melanoma-derived cell lines: Licr-Lon, a leukemic plasma cell line; Molt-4, a T-cell line, APD, an EBV-transformed B-cell line; K562, an erythromyeloid cell line and U937, a monocytic cell line (Van de Griend *et al.*, 1987).

Hetero-cross-linked Ab-induced cytotoxicity was blocked with MAb by the simultaneous addition of 50 μ l blocking MAb and 50 μ l heteroconjugated Ab to 50 μ l effector cells 30 min

prior to the addition of 100 μ l target cells, unless indicated otherwise. The standard error of the mean percentage lysis never exceeded 5% and was omitted for clarity of presentation.

Conjugate formation assay

Antigen-independent and hetero-cross-linked Ab-induced conjugate formation of CTL and target cells was assessed by FACS analysis (Becton Dickinson, Mountain View, CA) (Makgoba *et al.*, 1988a; Perez *et al.*, 1985; Leo *et al.*, 1987). CTL (5×10^6) were labelled with 100 μ g/ml hydroethidine in a volume of 2 ml at 37°C for 25 min. Target cells (5×10^6) were first modified with TNP (see above) and then labelled with 0.05 μ g/ml carboxy fluorescein diacetate (CFDA) in a volume of 2 ml at 37°C for 25 min. CTL and target cells were washed before use. Twenty microliters of labelled effector cells (10^7 /ml) were mixed with an equal amount of labelled target cells (10^7 /ml) with and without Ab-heteroconjugates and/or blocking MAb. Ab-heteroconjugates and MAb were preincubated with the CTL for 15 min at 37°C before the addition of tumor target cells. The final volume was adjusted to 100 μ l. Conjugate assay medium was RPMI 1640 supplemented with 10% FCS. CTL and target-cell mixtures were centrifuged for 5 sec in a serofuge and conjugates were allowed to form at 21°C for 10 min. Conjugate formation was stopped by the addition of 1 ml ice-cold PBS + 1% BSA. Cells were immediately resuspended and analyzed on a FACScan. Ten thousand total fluorescent events were collected and the number of events simultaneously emitting red and green fluorescence (conjugates) was divided by the total number of events emitting red fluorescence (total effector cells) yielding the percentage of CTL that had formed conjugates. No conjugates were observed when CTL and target cells were mixed and analyzed immediately.

Immunofluorescence assay

Five hundred thousand cells were pelleted and incubated with 10–20 μ l MAb at 4°C for 30 min. After washing, the cells were incubated with 10 μ l 1:40 diluted goat anti-mouse, conjugated with FITC (Nordic, Tilburg, The Netherlands). The cells were washed twice with PBS + 1% BSA + 0.01% sodium azide and analyzed on the FACScan.

RESULTS

Melanoma IgR39 is relatively resistant to CD3-mediated lysis

A panel of histologically distinct tumor cell lines were compared in their susceptibility to lysis by TCR $\alpha\beta^+$ /CD3 $^+$ and TCR $\gamma\delta^+$ /CD3 $^+$ CTL clones triggered via CD3, using hetero-cross-linked anti-CD3 \times anti-DNP. Most tumor cell lines were susceptible to CD3-mediated lysis (Fig. 1). The melanoma-derived cell line IgR39, however, showed a relative resistance to CD3-mediated lysis. This was found for both TCR $\alpha\beta^+$ and TCR $\gamma\delta^+$ CTL and hence is independent of the sublineage of the CTL.

The level of CD3-mediated lysis depended on the amount of anti-CD3 added. At higher anti-CD3 concentrations (15 ng/ml or more) the tumor-cell line IgR39 was effectively lysed (Fig. 1). Thus, the relative resistance to CD3-mediated lysis of IgR39 melanoma cells by cloned TCR $\alpha\beta^+$ and TCR $\gamma\delta^+$ CTL can be overcome at higher effective densities of anti-CD3 MAb on the target cells.

Expression of cellular adhesion molecules on tumor cells

Because the relative resistance of IgR39 melanoma cells to CD3-mediated lysis was independent of the type of CTL clone used, we hypothesized that the composition of surface molecules on the target cells, particularly the known ligands for CTL/target-cell interaction ICAM-1 and LFA-3, determined its relative resistance to CD3-mediated lysis by CTL. The TNP

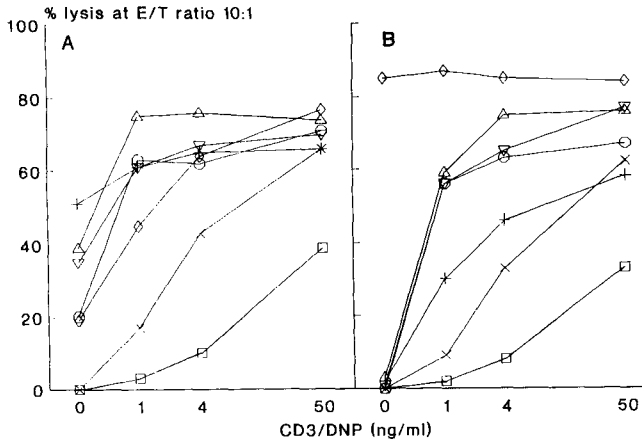


FIGURE 1 - IgR39 melanoma cell line is relatively resistant to CD3 mediated lysis by TCR $\gamma\delta^+$ /CD3 $^+$ clone WiK (panel a) or TCR $\alpha\beta^+$ /CD3 $^+$ clone D11 (panel b). TNP-modified tumor cell lines used as target cells were: \square , IgR39; $+$, IgR37; \times , APD; Δ , Molt-4; \diamond , Licr-Lon; ∇ , K562 and \circ , U937. Target-cell lysis was induced by addition of the indicated concentration hetero-cross-linked anti-CD3 MAb. Target-cell line Licr-Lon expressed the HLA Class-I antigen (Cw3), specifically recognized by TCR $\alpha\beta^+$ /CD3 $^+$ clone D11.

modification of the IgR39 cells was not the cause of the relative resistance because IgR39 target cells reportedly showed levels of sensitivity to CD2-mediated lysis comparable to those of other tumor cell lines (Goedegebuure *et al.*, 1989). The expression of LFA-3 and ICAM-1 on the tumor cell lines was then determined. Contrary to the panel of other cultured tumor cells tested in Figure 1, the IgR39 cells lacked surface expression of ICAM-1 (Fig. 2). All human tumor cell lines tested here expressed LFA-3. TNP modification of the tumor cell lines did not affect expression of ICAM-1 and LFA-3 (data not shown).

The "resistance" of IgR39 target cells to CD3-mediated lysis is due to lack of ICAM-1 expression

To assess the functional correlation between lack of ICAM-1 expression and "resistance" to CD3-mediated lysis, we used

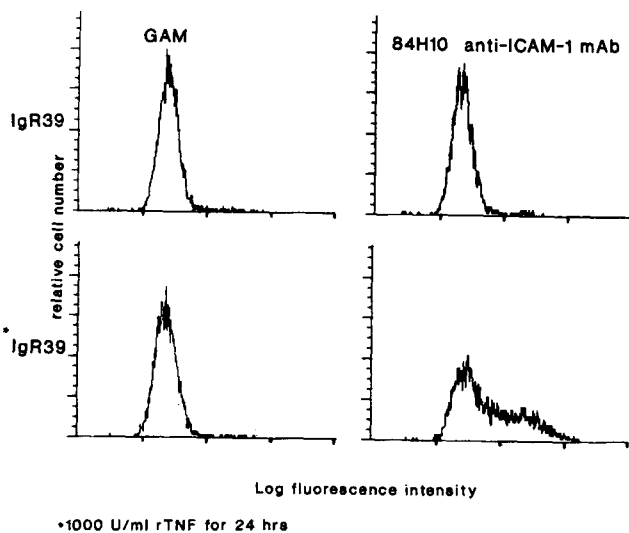


FIGURE 2 - TNF induces ICAM-1 expression on the ICAM-1 $^-$ melanoma cell line IgR39. IgR39 was cultured in the presence of 1000 U/ml recTNF for 24 hr.

TNF which is known to induce ICAM-1 expression on many cell types (Dustin *et al.*, 1986). Incubation of IgR39 cells with 1000 U/ml recTNF for 24 hr induced ICAM-1 expression as determined by FACS-analysis (Fig. 2). This expression of ICAM-1 was accompanied by a simultaneous increase in susceptibility to CD3-mediated lysis (Fig. 3). Further evidence that ICAM-1 is the critical factor determining the susceptibility to CD3-mediated lysis of IgR39 target cells comes from anti-ICAM-1 MAb inhibition studies. ICAM-1 specific MAb again abolished the increased levels of CD3-mediated lysis of IgR39 target cells obtained by TNF treatment (Fig. 3). Indeed, the level of lysis of TNF-untreated ICAM-1 $^-$ IgR39 cells was, as expected, not affected by anti-ICAM-1 MAb. Thus, TNF-induced expression of ICAM-1 can account for the TNF-induced increase in susceptibility of IgR39 target cells to CD3-mediated CTL lysis.

Signalling properties of LFA-1 in CTL activation

To examine the signalling abilities of LFA-1 in the activation of the lytic machinery of CTL, we tested the effect of anti-CD18 (LFA-1 β) MAb on spontaneous and anti-CD3 MAb induced conjugate formation and cytotoxicity. As shown in Table I, anti-CD18 MAb, as expected, inhibited spontaneous conjugate formation of CTL with ICAM-1 $^+$ Licr-Lon cells but minimally with ICAM-1 $^-$ IgR39 cells. The latter also suggests that ICAM-1 $^-$ IgR39 cells lack sufficient expression of alternative ligands for LFA-1 (*e.g.* ICAM-2) (Staunton *et al.*, 1989). Anti-CD3 MAb containing heteroconjugated Ab enhanced the percentage conjugates formed between CTL and tumor cells. CD18-specific MAb had virtually no inhibitory effect on CD3-mediated conjugate formation of CTL to ICAM-1 $^{+or-}$ tumor cells (Table I), but did inhibit CD3-mediated lysis of either ICAM-1 $^{+or-}$ target cells (Fig. 4). Interestingly, this significant inhibitory effect of CD18-specific MAb on lysis contrasts with the marginal inhibitory effects on conjugate formation. These findings point to a regulatory role for LFA-1 in CD3-mediated CTL activation post CTL/target-cell conjugate formation.

Anti-CD18 \times anti-DNP hetero-cross-linked Ab coactivates CD3-mediated lysis of ICAM-1 $^-$ target cells

Experiments were designed to mimic LFA-1/ICAM-1 inter-

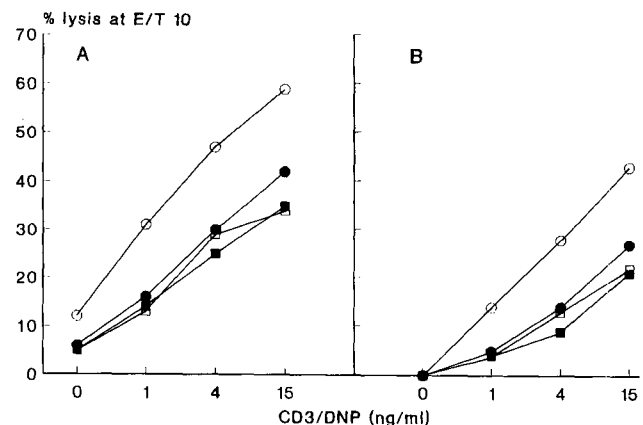


FIGURE 3 - Anti-ICAM-1 MAb abolishes the increased susceptibility of TNF-treated IgR39 cells to CD3-mediated lysis by TCR $\gamma\delta^+$ /CD3 $^+$ clone WiK (panel a) and TCR $\alpha\beta^+$ /CD3 $^+$ clone D11 (panel b). IgR39 cells were cultured with or without 1,000 U/ml recTNF for 24 hr and TNP-modified before use as target cells. Hetero-cross-linked anti-CD3 MAb and anti-ICAM-1 MAb (84 H10, dilution 1/1,000 of ascites) were added simultaneously to the CTL effectors prior to the addition of target cells \square , IgR39; \blacksquare , IgR39 + anti-ICAM-1 MAb; \circ , TNF-treated IgR39 and \bullet , TNF-treated IgR39 + anti-ICAM-1 MAb.

TABLE I – INHIBITION OF CONJUGATE FORMATION BY ANTI-CD18 (LFA-1 β) MAb

Addition of heteroconjugate ²	Anti-CD18 MAb ³	Percentage conjugates ⁴ between TCR $\gamma\delta^+$ CTL ¹ and TNP-target cells	
		ICAM-1 ⁺ (Licr-Lon)	ICAM-1 ⁻ (Igr39)
—	—	34	23
—	+	17	19
Anti-CD3 × anti-DNP	—	52	35
Anti-CD3 × anti-DNP	+	48	31

¹TCR $\gamma\delta^+$ effector cell: WiK. ²Heteroconjugate concentration 1 μ g/ml. ³Anti-CD18 (CLB54 ascites, 1/1000). ⁴For calculation of percentage conjugates see "Material and Methods".

TABLE II – ANTI-CD18 × ANTI-DNP Ab CO-ACTIVATES LYSIS OF ICAM-1⁻ TARGET CELL BY TCR $\alpha\beta$ CTL

Addition of Ab heteroconjugates	Percentage lysis of ICAM-1 ⁻ Igr39 by TCR $\alpha\beta^+$ clones	
	Clone D11	Clone 75
—	1	0
aCD18 × aDNP	0	0
aCD3 × aDNP	37	37
aCD3 × aDNP +	70	68
aCD18 × aDNP		

Effector to target cell ratio of 7 (clone D11), 14 (clone 75). Heteroconjugate concentration aCD3 × aDNP 25 μ g/ml; aCD18 × aDNP 25 μ g/ml.

DISCUSSION

Initiation of CD3/TCR complex-mediated T-cell functions is believed to be interrelated with various accessory molecules. The transmembrane molecule LFA-1 is one of the most important accessory molecules. Here, we have studied the contribution of the LFA-1/ICAM-1 interaction to CD3/TCR complex-mediated lysis by TCR $\alpha\beta^+$ and TCR $\gamma\delta^+$ CTL at different stages of the lytic cycle. The use of hetero-cross-linked anti-CD3 × anti-DNP Ab makes it possible to compare the susceptibilities to lysis of different tumor cell lines by an individual CTL clone. An ICAM-1⁻ melanoma tumor cell line Igr39 shows a relative resistance to CD3/TCR-mediated cytolysis by TCR $\alpha\beta^+$ as well as by TCR $\gamma\delta^+$ cloned lymphocytes. The low susceptibility of Igr39 tumor cells to CD3/TCR-mediated cytolysis was directly related to the lack of ICAM-1 expression because TNF treatment of ICAM-1⁻ Igr39 cells induced ICAM-1 expression and indeed simultaneously increased susceptibility to CD3/TCR-mediated cytolysis. The Igr39 cell line is not sensitive to TNF lysis (data not shown). Moreover, anti-ICAM-1 MAb down-regulated this TNF-induced, increased susceptibility to lysis, demonstrating that it was functionally due to up-regulation of ICAM-1 and not other unknown ligands.

The relative resistance of Igr39 cells to CD3-mediated cytolysis can be overcome by higher concentrations of anti-CD3 MAb. Thus, an efficient LFA-1/ICAM-1 interaction may lower the requirement for either high antigen density on target cells or, alternatively, may compensate for a low-affinity TCR on CTL by establishing a more effective CTL-target-cell interaction.

The interaction between CTL and target cells, resulting in target cell lysis, is a multi-step process (Martz, 1977; Mentzer *et al.*, 1987). Thus the relative resistance of the ICAM-1⁻ Igr39 target cells to CD3/TCR-mediated lysis may reflect either a less efficient conjugate formation, or a decreased level of activation of the lytic machinery. This study shows that CTL formed conjugates less efficiently with ICAM-1⁻ than with ICAM-1⁺ tumor cells. Hetero-cross-linked anti-CD3 MAb enhanced conjugate formation with both the ICAM-1-negative and -positive target cells. Addition of anti-CD18 × anti-DNP Ab increased spontaneous and CD3-mediated conjugate formation with the ICAM-1⁻ cell line and consequently enhanced CD3/TCR-mediated lysis of this target cell. Hetero-cross-linked anti-CD18 MAb failed to enhance CD3/TCR-mediated lysis of ICAM-1⁺ tumor cells, because it could not further enhance the relatively efficient conjugate formation with this target. It was remarkable that hetero-cross-linked anti-HLA Class I did not enhance CD3-mediated conjugate formation with the ICAM-1⁻ target cells (data not shown) and therefore did not affect CD3-mediated lysis.

Our data suggest that the relative resistance to CD3-mediated lysis of the ICAM-1⁻ tumor cells is not exclusively associated with decreased conjugate formation. The LFA-1/ICAM-1 interaction also appears to synergize with CD3/

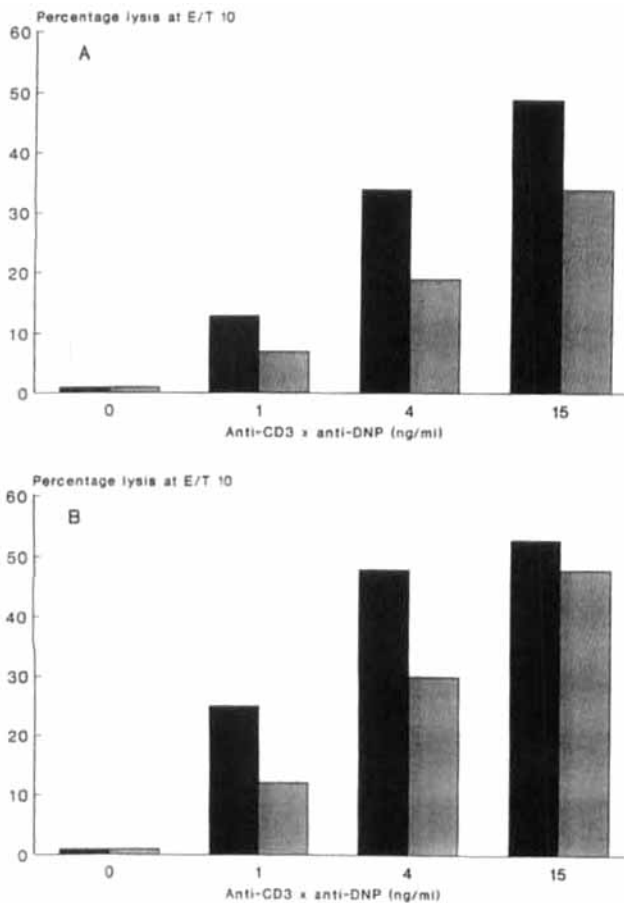


FIGURE 4 – Anti-CD18 MAb inhibits CD3-mediated lysis of ICAM-1⁻ Igr39 cells (panel a) or ICAM-1⁺ Licr-Lon cells (panel b) by TCR $\gamma\delta^+$ /CD3⁺ clone WiK. Target-cell lysis is induced by the indicated concentration of anti-CD3 MAb in the absence (solid bars) or presence (hatched bars) of anti-CD18 MAb (CLB54, 1/1000 of ascites).

actions, using anti-CD18 × anti-DNP hetero-cross-linked Ab. Addition of anti-CD18 × anti-DNP Ab-heteroconjugate increased the susceptibility of ICAM-1⁻ Igr39 target cells to CD3-mediated lysis by TCR $\alpha\beta^+$ CTL (Table II). Similar results were obtained using TCR $\gamma\delta^+$ CTL (Table III). The susceptibility to lysis of ICAM-1⁺ target cells was much less affected (Table III). Remarkably, anti-CD18 × anti-DNP Ab alone also considerably enhanced the MHC-unrestricted lysis of ICAM-1⁻ but not ICAM-1⁺ target cells by TCR $\gamma\delta^+$ CTL. Control, anti-HLA Class I × anti-DNP Ab had no effect on MHC-unrestricted or CD3-mediated lysis.

TABLE III - ANTI-CD18 × ANTI-DNP Ab CO-ACTIVATES LYSIS OF THE ICAM-1⁻ TARGET CELL BY TCR $\gamma\delta$ CTL

Ab heteroconjugates ¹	ICAM-1 ⁻ (Igr39)				ICAM-1 ⁺ (Licr-Lon)			
	Exp. 1	Exp. 2	Exp. 3	Exp. 4	Exp. 1	Exp. 2	Exp. 3	Exp. 4
—	15 ²	2	13	3	3	3	4	11
—	aCD18 × aDNP	40	6	37	9	7	3	5
—	aClassI × aDNP	nt	2	16	3	nt	1	4
aCD3 × aDNP	—	24	11	44	29	49	37	59
aCD3 × aDNP	aCD18 × aDNP	42	31	70	56	49	46	69
aCD3 × aDNP	aClassI × aDNP	nt	12	40	23	nt	33	63

Effector TCR $\gamma\delta$ ⁺/CD3⁺ clone: WiK. ¹Heteroconjugate concentration aCD3 × aDNP = 0.1 μ g/ml; aCD18 × aDNP = 0.1 μ g/ml; aClassI × aDNP = 0.1 μ g/ml. ²% specific lysis at an E/T ratio of 24.

TCR-mediated lysis at a post-conjugate stage. LFA-1-specific MAbs reportedly block lysis by CTL and NK cells through inhibition of conjugate formation (Makgoba *et al.*, 1988a; Leo *et al.*, 1987). Here we show that, although anti-CD18 MAb inhibited CD-3 mediated cytotoxicity by CTL of ICAM-1⁻ tumor cells, it did not affect their conjugate formation. These findings suggest that LFA-1 can transduce down-regulatory signals after conjugate formation, although the possibility that the inhibitory effect of anti-CD18 MAb is solely due to decreased recycling of CTL is to be formally excluded. A further argument in favor of a signalling function for LFA-1 comes from our previously reported observation that ICAM-1⁻ Igr39 tumor cells show a susceptibility to CD2-mediated lysis comparable to that of ICAM-1⁺ tumor cells (Goedegebuure *et al.*, 1989). The relative resistance of ICAM-1⁻ target cells to CD3- but not CD2-mediated lysis demonstrates that it is not merely due to a decreased conjugate-forming ability but suggests a signalling function for the LFA-1/ICAM-1 interaction. Hetero-cross-linked anti-CD18 MAb enhanced MHC-unrestricted lysis of the ICAM-1⁻ target cell by TCR $\gamma\delta$ ⁺ CTL but not by TCR $\alpha\beta$ ⁺ CTL in 2 out of 4 experiments. Still, LFA-1 cross-linking to the target cell *per se* is not sufficient to trigger the cytolytic machinery of TCR $\alpha\beta$ ⁺ and TCR $\gamma\delta$ ⁺ CTL because anti-LFA-1 MAb-expressing hybridomas, cross-linking LFA-1, were not lysed by the CTL (Hoffman *et al.*, 1985; unpublished observation). Hence, LFA-1 transduces a co-activation signal rather than a primary signal. Whether the functional interrelation between LFA-1 and the CD3/TCR complex de-

pends on co-clustering of these molecules, as has been demonstrated for CD3/TCR and the accessory molecule CD4 (Saizawa *et al.*, 1987), remains to be established.

In summary, our results demonstrate the central role of LFA-1 in T-cell activation. LFA-1 appears to be involved in CD3/TCR-mediated lysis at 3 distinct stages of the lytic cycle. Firstly, as known, it promotes binding of CTL to target cells; secondly, it delivers co-stimulatory signals to CD3/TCR in triggering cell-mediated lysis. Thirdly, it can transduce down-regulatory signals after conjugate formation, *i.e.* anti-CD18 MAb inhibits CD3-mediated bispecific antibody-triggered lysis. Consequently, tumor cells lacking ICAM-1 expression may escape lysis by CTL. The relative resistance of ICAM-1⁻ tumor cells to lysis might be overcome by the induction of ICAM-1 on the tumor-cell surface, *e.g.* with TNF or IFN- γ (Dustin *et al.*, 1986). Also, the use of anti-CD2 × anti-target Abs which activate CTL independently of the additional LFA-1/ICAM-1 interaction or the combined use of bispecific anti-CD3 × anti-target and anti-LFA-1 × anti-target Ab may induce efficient lysis of ICAM-1⁻ target cells.

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