

## Leukotriene B<sub>4</sub> stimulation of macrophage cyclooxygenase metabolite synthesis

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### Introduction

The essential poly-unsaturated fatty acid, arachidonic acid (AA), is metabolised by macrophages via the cyclooxygenase (CO) and lipoxygenase (LO) pathways. In general CO metabolites, in particular prostaglandin (PG) E<sub>2</sub>, are considered to inhibit macrophage functions while LO products, in particular leukotrienes (LTs), stimulate them. Thus, the CO:LO metabolite ratio is important in regulating macrophage activities. For example, LTC<sub>4</sub> stimulates the secretion of the lysosomal enzyme  $\beta$ -glucuronidase ( $\beta$ -GLU) by macrophages. This response is enhanced by indomethacin, a CO inhibitor, and inhibited by PGE<sub>2</sub>. LTC<sub>4</sub> also stimulates macrophage PGE<sub>2</sub> synthesis indicating that LTC<sub>4</sub>-stimulated  $\beta$ -GLU secretion is self-limiting [1]. We have recently shown that calcium-ionophore (A23187)-stimulated macrophage LTB<sub>4</sub> synthesis is enhanced by indomethacin and inhibited by PGE<sub>2</sub> [2]. It is possible, therefore, that LTB<sub>4</sub>-stimulation of macrophage functions is also modified by an associated increase in endogenous PGE<sub>2</sub> formation. We now report the effect of LTB<sub>4</sub> on the release of macrophage CO metabolites [PGE<sub>2</sub>, thromboxane (TX) A<sub>2</sub> (assayed as TXB<sub>2</sub>) and prostacyclin (assayed as 6-ketoPGF<sub>1 $\alpha$</sub> )] and  $\beta$ -GLU.

### Materials and methods

The materials and methods used have been published previously [1]. Briefly, 4 day carrageenin-

elicited rat peritoneal macrophages were isolated by density gradient centrifugation over Ficoll-Isoopaque and suspended at  $2 \times 10^6$  cells ml<sup>-1</sup> in Dulbecco's modified Eagle's minimum essential medium (DMEM). Aliquots of the cell suspension (1 ml) were incubated for 1 h or 3 h at 37°C in polypropylene reaction vials. The tubes were then centrifuged and the supernatant fractions stored at -70°C for later analysis of PGE<sub>2</sub>, 6-ketoPGF<sub>1 $\alpha$</sub>  and TXB<sub>2</sub> by radioimmunoassay and for  $\beta$ -GLU activity.

### Results

#### *The effect of LTB<sub>4</sub> on macrophage PGE<sub>2</sub>, 6-ketoPGF<sub>1 $\alpha$</sub> and TXB<sub>2</sub> synthesis*

LTB<sub>4</sub> ( $10^{-12}$  to  $10^{-7}$  M) had a biphasic, stimulatory effect on the synthesis and release of PGE<sub>2</sub> and TXB<sub>2</sub> during a 1 h incubation period. A maximum stimulation of both CO metabolites was observed at  $10^{-9}$  M LTB<sub>4</sub>. The basal release of all three CO metabolites was greater during the 3 h incubation period. However, LTB<sub>4</sub> now had no effect on PGE<sub>2</sub> formation whereas TXB<sub>2</sub> synthesis was stimulated, again with a maximum effect at  $10^{-9}$  M LTB<sub>4</sub>. No effect on 6-ketoPGF<sub>1 $\alpha$</sub>  production was observed during either the 1 h or 3 h incubation periods (Table 1). LTB<sub>4</sub> ( $10^{-9}$  M and  $10^{-8}$  M) and a slight, but not significant, stimulatory effect on  $\beta$ -GLU secretion during the 3 h incubation (control = 100%,  $10^{-9}$  M = 129%,  $10^{-8}$  M = 135%).

**Table 1**

The effect of leukotriene (LT) B<sub>4</sub> on macrophage prostaglandin (PG) E<sub>2</sub>, thromboxane (TX) B<sub>2</sub> and 6-keto prostaglandin F<sub>1α</sub> (6-keto) release.

	LTB <sub>4</sub> (M)					
	0	10 <sup>-12</sup>	10 <sup>-10</sup>	10 <sup>-9</sup>	10 <sup>-8</sup>	10 <sup>-7</sup>
1 h incubation						
PGE <sub>2</sub>	0.35	0.48*	0.65*	1.32*	0.71*	0.52*
TXB <sub>2</sub>	2.33	2.78	2.97	3.27*	3.21	2.61
6-keto	1.48	1.63	1.62	1.75	1.79	1.49
3 h incubation						
PGE <sub>2</sub>	1.18	1.08	1.05	1.13	1.22	1.13
TXB <sub>2</sub>	5.75	7.74*	11.28*	12.54*	11.50*	8.60*
6-keto	3.40	3.07	3.20	3.49	3.46	3.18

Results (ng/2 × 10<sup>6</sup> cells) are mean values, n = 12. \*p < 0.05 vs control (Mann-Whitney U-test).

## Discussion

LTB<sub>4</sub> had a differential effect on the synthesis of PGE<sub>2</sub>, TXB<sub>2</sub> and 6-ketoPGF<sub>1α</sub> by carrageenin-elicited peritoneal macrophages. Whereas TXB<sub>2</sub> release was stimulated by LTB<sub>4</sub> during 1 h and 3 h incubation, PGE<sub>2</sub> formation was only stimulated during the 1 h incubation period. PGE<sub>2</sub> synthesis appeared to reach a maximum (1 h incubation with LTB<sub>4</sub> or a 3 h incubation without LTB<sub>4</sub>) after which its synthesis stopped. Release of 6-keto PGF<sub>1α</sub> was not modified by LTB<sub>4</sub> (1 h and 3 h incubations) even though its production was not maximal. LTB<sub>4</sub> had little effect on macrophage lysosomal enzyme release, under the conditions used an it has not proved possible to relate changes in CO metabolite synthesis with this marker of macrophage activation. It is possible that CO inhibitors, such as indomethacin, could enhance LTB<sub>4</sub>-stimulated enzyme release as reported for LTC<sub>4</sub>-induced lysosomal enzyme secretion [1].

LTB<sub>4</sub> has been reported to stimulate macrophage phosphatidylinositol turnover and to act as a calcium ionophore, which could result in stimulation of PLA<sub>2</sub> [3]. Both activities could stimulate the release of AA from phospholipid pools resulting in an increase in CO metabolite formation. We have previously shown that synthesis of 6-ketoPGF<sub>1α</sub> by granuloma macrophages is saturated at lower AA concentrations than the formation of PGE<sub>2</sub> or TXB<sub>2</sub> [4]. Thus it could be that the basal release of AA is sufficient to maximally stimulate 6-keto PGF<sub>1α</sub> formation. PGE<sub>2</sub> inhibits A23187-stimulated macrophage TXB<sub>2</sub> formation and LTC<sub>4</sub>-stimulated macrophage PGE<sub>2</sub> synthesis is associated with a reduction in TXB<sub>2</sub> formation [1, 2]. Thus, it could be that the apparent increase in LTB<sub>4</sub>-stimulated TXB<sub>2</sub> production observed during the 3 h incubation, compared to the 1 h data, was due to the relative reduction in PGE<sub>2</sub> synthesis. Further experiments are necessary before we can suggest a reason for the saturation of PGE<sub>2</sub> synthesis.

## References

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