Novel Outcome Measures for Clinical Trials in Cystic Fibrosis

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Summary. Cystic fibrosis (CF) is a common inherited condition caused by mutations in the gene encoding the CF transmembrane regulator protein. With increased understanding of the molecular mechanisms underlying CF and the development of new therapies there comes the need to develop new outcome measures to assess the disease, its progression and response to treatment. As there are limitations to the current endpoints accepted for regulatory purposes, a workshop to discuss novel endpoints for clinical trials in CF was held in Anaheim, California in November 2011. The pros and cons of novel outcome measures with potential utility for evaluation of novel treatments in CF were critically evaluated. The highlights of the 2011 workshop and subsequent advances in technologies and techniques that could be used to inform the development of clinical trial endpoints are summarized in this review. **Pediatr Pulmonol. 2015**; **50:302–315.** © 2014 The Authors. *Pediatric Pulmonology* published by Wiley Periodicals, Inc.

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INTRODUCTION

Cystic fibrosis (CF) is caused by mutations in the gene encoding the CF transmembrane conductance regulator (CFTR) protein, an ion channel that transports chloride ions across epithelial cell membranes. Therapeutic progress has been realized over the last 20 years with improved health, health-related quality of life (HRQoL) and overall survival.¹ These improvements may, in part, be due to changes in therapeutic approach and patient management. There has been a recent shift in the treatment paradigm, with a reactive approach based on responding to acute declines in respiratory health giving way to a proactive approach of preventing exacerbations and loss of lung function (Fig. 1),² as well as improving the functional and emotional wellbeing of individuals.³

As life expectancy improves and decline in lung function is reduced, the traditional endpoints for studies in patients with CF, such as spirometry parameters, are becoming less appropriate for assessing drug effects.⁴ In addition, newborn screening has been widely adopted and the introduction of disease-modifying therapies aimed at correcting the function of the defective CFTR protein⁵ that could be started early in life seems imminent, particularly following the recent approval of the first CFTR potentiator.⁶ Hence there is a need for novel endpoints that allow detection of clinical benefits starting in young children and continuing into adulthood, which are also acceptable to regulatory authorities. More sensitive outcome measures may assist identification of individuals who could benefit from a novel therapy, reduce the sample size and shorten the duration of intervention studies.⁴

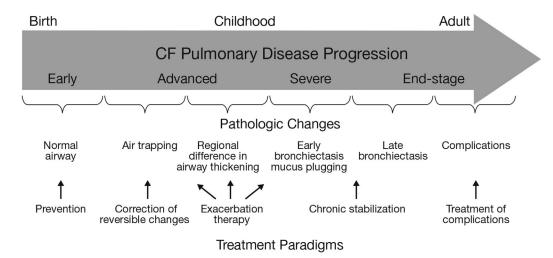


Fig. 1. Stages of disease progression and pathologic changes that occur in the airways of patients with CF as they age, along with possible treatment approaches. Reprinted with permission of the American Thoracic Society. Copyright 2014 American Thoracic Society. Ramsey BW. 2007. Use of lung imaging studies as outcome measures for development of new therapies in cystic fibrosis. Proc Am Thorac Soc;4(4):359–63. Official Journal of the American Thoracic Society.²

A workshop to discuss novel endpoints for clinical trials in CF was held in Anaheim, California, in November 2011. The pros and cons of novel outcome measures with potential to be used for evaluation of novel treatments in CF were critically assessed. The aim of this review is to summarize the highlights of this workshop, together with more recent developments of clinical trial endpoints for CF, in order to identify potential alternatives that may be useful in future studies.

Pulmonary Function Testing

Parameters derived from spirometry, such as forced expiratory volume in 1 sec (FEV₁), still play an important role as outcome measures in clinical trials. However, with improved therapeutic options and earlier interventions, FEV₁ has become less useful as a tool for differentiating between interventions, as differences in FEV₁ are reduced, so establishing statistical significance is more difficult. Recently, end expiratory flows such as FEF_{25–75} and FEF₇₅ have gained new interest as markers of early lung disease. End expiratory flows show more variability compared to FEV₁, but they are substantially impaired in early disease and have been shown to respond to therapy in clinical studies.^{7–9}

Unfortunately, standard spirometry cannot be routinely used in children below the age of 6. For these young children, preschool and infant pulmonary function tests have been developed.¹⁰

Infant Pulmonary Function Testing

The raised volume rapid thoracoabdominal compression (RVRTC) technique (Table 1) is a reproducible, repeatable and safe infant pulmonary function test (iPFT), which allows the use of functional outcome parameters for the diagnosis of airflow limitation and monitoring of CF in sedated infants.^{11,12} Furthermore, standardized guidelines for measurement and interpretation of results obtained using RVRTC have been published and commercial equipment is now available.¹²

Through the use of the RVRTC technique, investigators have shown that lung function parameters are significantly diminished in infants with CF, even in those as young as 3 months old.^{11,13,14} This finding suggests that RVRTC parameters can identify early disease and may therefore serve as useful clinical trial endpoints in infants with CF. Early identification of disease is particularly relevant in CF as therapeutic interventions aimed at preserving lung function may be most effective when administered within the first 6 months of life.¹⁵ In addition, RVRTC parameters have been shown to be sensitive to interventions in infants with CF. A study has recently revealed that forced expiratory flows and volumes improved following antibiotic therapy for a pulmonary exacerbation in a small subset of infants.¹⁶ In another recent clinical trial, RVRTC parameters were evaluated as an exploratory endpoint in a subset of patients randomized to either 7% hypertonic saline or isotonic saline for 1 year.¹⁷ When adjusted for baseline infant lung function, age, gender, height, and weight, the mean change in FEV in 0.5 sec (FEV_{0.5}) over 1 year was significantly greater (by 38 ml) in the group treated with 7% hypertonic saline.¹⁷ However, it should be noted that acceptable measurements were only obtained in 45 of 73 infants in this study, and the clinical significance of a 38 ml difference is not known. Nevertheless, given these recent findings, the commercial availability of equipment, and the promising potential of iPFTs noted in the literature to date, it seems worth pursuing the inclusion of functional parameters as an outcome measure for clinical trials in infants.

However, disadvantages of the RVRTC technique that form a hurdle for use as a primary endpoint must also be considered. Infants must be sedated, equipment is expensive, extensive training of personnel is required, performing measurements is time consuming, and normative data are limited.^{11,12,18} Despite these challenges, Davis et al. were able to conduct a prospective, longitudinal, observational study with a commercial device in 10 centers in the US.¹¹ Personnel at each site underwent rigorous training, and quality control and independent reading of all data were performed by an expert panel.¹¹ In this study, key RVRTC parameters were, on average, abnormal in infants with CF compared with healthy historical control subjects.¹¹ The authors concluded that infant RVRTC parameters were not yet ready to be adopted as primary efficacy endpoints for multicenter clinical trials, particularly at inexperienced sites and based on acceptability rates.¹¹ However, the use of iPFTs as a secondary endpoint should still be strongly considered.

Multiple Breath Washout and the Lung Clearance Index

Parameters from the non-invasive, simple multiple breath washout (MBW) tests include functional residual capacity and lung clearance index (LCI) (Table 1). LCI is determined by washout of an inert gas during tidal breathing.¹⁹ As the washout takes longer to complete in the presence of mucus retention or inflammation, LCI increases with disease severity.¹⁹ LCI measures ventilation inhomogeneity, which helps assess changes in small airways that are not apparent while using spirometry.²⁰ LCI has been shown to be superior to spirometry for the detection of early lung disease in CF,^{20–23} while MBW has been shown to be equivalent to RVRTC for detecting early disease in infants.²⁴

LCI has been demonstrated to correlate with structural lung changes seen on computed tomography (CT) scans in cross sectional studies, suggesting that these techniques have similar sensitivity for the detection of CF lung disease, and that using both methods in individual patients

| | | Disease severity (structural | | | | | Status of | |
|---|---|--|---|--------------------------|---------------------------------------|---|---|------------------------------------|
| | | or functional | Type of | MCID | Test | | standardization | Cost of |
| Assessment | Age group | changes assessed) | evidence | defined | frequency | Safety aspects | (reproducibility, validity) | equipment |
| Infant pulmonary function testing (RVRTC) ^{11–18} | <3 years of age | Detection of early functional changes | Well validated | No | Unrestricted | Oral sedation | Well defined | Moderate to high |
| MBW/LCI ^{4,19–35} | All | All | Limited | No | Unrestricted | Oral sedation (optional for young children) | Required for each trial | Moderate to high |
| CT scans ^{25,36,45–58,60–63} | For all ages | Early and advanced structural changes | Well validated | Yes/No | Restricted | Radiation | Well defined and required for each trial | Moderate |
| MR1 ^{40-42,57,66-78} | All (depending on local expertise) | All (morphology and function) | Moderately well validated | No | Unrestricted | Only when contrast media injection is used | Required for each trial | High |
| PET scan ^{43,44,79–86} | For children 26 years of age | Detection of structural changes as for CT, in addition detection of active inflammation | Few studies in CF available | No | Restricted | Radiation | In development | High |
| Mucociliary clearance ^{87–93,95} | >6 years of age | Early or advanced | Not validated | No | Restricted (4–6 scans per year) | Radiation | Standardized between few sites | Moderate |
| NPD ^{119–134} | All, specialized at young age | Not applicable | Well validated | No | Unrestricted | Minimal | Well established | Moderate, requires expertise |
| ICM ^{133–142} | All | Not applicable | Limited | No | Restricted | Requires rectal biopsy | Reasonably standardized | Moderate, requires expertise |
| Sweat chloride ^{5,120–122,143–149} | All | Not applicable | Well validated | No | Unrestricted | None | Well established | Low |
| CF, cystic fibrosis; CT, computed tomography; FDA, Food and Drug Administration; ICM, intestinal current measurement; LCI, lung clearance index; MBW, multiple breath washout; MCID minimal clinically important difference; MRI, magenetic resonance imaging; NPD, nasal potential difference; PET, positron emission tomography; RVRTC, raised volume rapid thoracoabdominal compression. | ıted tomography; FDA ifference; MRI, magen | . Food and Drug Administrat etic resonance imaging; NPD. | ion; ICM, intestinal , nasal potential diffe | current m srence; PE' | easurement; LC T, positron emis | II, lung clearance sion tomography; | Drug Administration; ICM, intestinal current measurement; LCI, lung clearance index; MBW, multiple breath washout; MCID nee imaging; NPD, nasal potential difference; PET, positron emission tomography; RVRTC, raised volume rapid thoracoabdomina | 1 washout; MCID thoracoabdomina |

TABLE 1— Techniques Under Investigation For Clinical Studies in Young Children, and the Status of Their Validation

Novel Outcome Measures in CF 305

Pediatric Pulmonology

could elicit complementary information.^{21,25} A study in pre-school children showed that an abnormal LCI at preschool age could predict lung function abnormalities at school age.⁴ In addition, a study including early school age children demonstrated that LCI correlated with quality of life, and that elevated LCI values could predict pulmonary exacerbations (defined as changes in respiratory status requiring intravenous antibiotics).²⁶ Furthermore, LCI may also be a suitable outcome measure to assess early intervention strategies in pediatric patients, as studies among children with CF using normal spirometry have shown LCI to be more sensitive than FEV_1 for detecting response to treatment with dornase alfa, hypertonic saline, and ivacaftor (in patients with a G551D-CFTR mutation and normal FEV₁).^{27–29}

One strength of LCI is that it has low variability, both within and between tests,^{30,31} indicating that LCI is a suitable endpoint for longitudinal studies.¹⁹ However, limitations of LCI include that it may require sedation in infants to increase its success rate. Furthermore, LCI is affected by irregular breathing patterns, requirement for expensive equipment such as mass spectrometers with associated software to follow the concentration of the inert gas, and the need for longer washouts as the disease progresses.¹⁹ While sulfur hexafluoride-based mass spectrometry measurements are considered to be the gold standard,³² a number of devices have become commercially available, but will need to undergo thorough validation to assure accuracy of the measurement. Sulfur hexafluoride gas mass spectrometry is not commercially available,³² favoring nitrogen-based MBW that only requires 100% oxygen which is readily available in centers. The use of nitrogen-based MBW is increasingly being favored over sulfur hexafluoride-based methods and is more affordable than methods that require mass spectroscopy. A device that determines nitrogen concentration indirectly, by measuring both carbon dioxide and oxygen concentrations, and using Dalton's law of partial pressures, has recently been validated.³³ In relation to its feasibility, the European CF Society clinical trial network (ECFS-CTN) has selected a commercially available nitrogen MBW system to be used in the network. Over the last year many centers within the network have acquired the system and are currently being trained in its use. Similar efforts are ongoing in North America and training centers have been established in both London and Toronto.

Overall, LCI has potential as a clinical and research outcome measure in young children with CF⁴ and in single-dose, as well as multiple-dose studies.²⁸ With regard to the need for sedation, studies that will further assess the suitability of LCI for clinical trials are currently ongoing.^{29,34} A recent study demonstrating the utility of LCI as an outcome measure in a multi-center trial.³⁵

Imaging as an Outcome Measure

For the past 40 years, disease progression has been evaluated through lung function tests and plain chest radiographs. The implementation of multi-detector chest CT scan technology has provided clinicians with a more sensitive method for imaging CF lung damage.³⁶ Various types of imaging techniques are now used to determine the presence and extent of lung disease in patients with CF, including CT and chest magnetic resonance imaging (MRI). In addition, scoring systems have been developed to quantify and characterize the structural abnormalities detected through CT and MRI in patients with CF at various stages of the disease.^{37–39} These systems assess structural changes such as bronchiectasis, trapped air, airway wall thickening, mucus, and opacities. More recently, chest MRI techniques have been developed allowing the assessment of functional characteristics of the lung, as well as the evaluation of lung morphology.^{40–42} Positron emission tomography (PET) imaging with [18F]fluorodeoxyglucose ([18]FDG; FDG-PET) can also be used as a non-invasive technique to quantify lung inflammation.^{43,44}

CT Scans

Chest CT scans have been shown to be more sensitive at detecting disease severity than spirometry.^{25,36,45,46} In particular, CT can be used to detect bronchiectasis and trapped air, which reflects abnormal ventilation and perfusion in infants.⁴⁷ Studies have shown that there are weak associations between the presence and extent of structural lung damage and functional parameters.^{45,47,48} It has also been shown that infection, inflammation and abnormal chest CT findings are already present in a significant proportion of infants with CF at 3 months of age^{46,49} and that these early structural changes are progressive.^{46,50} These findings suggest that chest CT could be used to detect the presence and extent of structural lung disease, particularly as the majority of infants with lung disease may be asymptomatic.⁴⁶

Evaluation of the ability of chest CT to identify lung abnormalities not detected by spirometry is an important step in validating its use in the diagnosis and monitoring of CF lung disease.^{51,52} Two studies showed that chest CT scores are predictive of the respiratory tract exacerbation rate, which is considered a key clinical efficacy outcome measure in CF clinical studies,^{51,53} while another showed that the CT scan bronchiectasis score (Brody-II system) was strongly associated with the respiratory tract exacerbation rate.⁵¹ Moreover, time to first respiratory tract exacerbation and hospitalization was significantly associated with quartiles of bronchiectasis score as recorded by CT scan.⁵¹

Data from CT scans have also been correlated with survival. In patients screened for lung transplantation, individuals with a higher volume of infection/inflammation-like changes were shown to have a higher risk of dying on the waiting list.⁵⁴ In addition, there was a

correlation between HRQoL scores and the presence of structural changes on CT scans.⁵⁵ Recent studies further indicate that CT scans, and identification of bronchiectasis in particular, can be useful in identifying children at risk for worse pulmonary outcomes and could be used to guide treatment decisions.^{36,56}

The use of chest CTs in clinical trials has been shown to be viable.⁵⁵ However, as with spirometry, use of this technique requires standardization, data transfer, and centralized reading of images. A great advantage of chest CT for clinical trials is that most CF centers have scanners. However, as CT is based on ionizing radiation, the radiation dose has to be minimized, especially in young patients and when scans need to be repeated within a relatively short study period.^{36,57} This can be achieved by optimizing low-dose scanning protocols and current CT techniques.⁵⁸ Careful consideration should be given to the balance between radiation exposure and potential benefits.³⁷ It is also necessary to define the optimal balance between image modality, image quality, and radiation exposure for each study protocol, at each study site in multicenter clinical trials. In a cohort study of children participating in the Wisconsin CF Neonatal Screening Project, quantitative chest radiography was shown to have excellent sensitivity for detecting abnormal chest CT.⁵⁹ In addition, efforts should be directed at implementing strategies that have been shown to reduce the radiation dose associated with chest CT protocols.60-65 Using these strategies allows the radiation dose of a chest CT to be reduced to within the range of a routine chest radiograph.^{64,65} Regarding its feasibility, over the last year 15 centers within the ECFS-CTN participated in a CT standardization effort. Each center was asked to characterize and optimize the CT scanners using age-specific phantoms; personnel were also trained in the use of a spirometry controlled chest CT protocol. Similar efforts were made in 2 centers in North America and in 9 centers in Australia. A training center has been established in Erasmus MC Rotterdam.

Overall, bronchiectasis and trapped air detected by chest CT are feasible and utilizable surrogate outcome measures for clinical studies of novel treatments for patients with CF (Table 1). Furthermore, due to the increased sensitivity of CT scans compared with spirometry, smaller sample sizes will be needed in clinical studies employing CT scan endpoints.⁴⁶ As CT is a sensitive method of detection, it may be useful early in life to detect signs of disease and act as a trigger for initiating therapy before lung function measures, such as FEV₁, become noticeably impaired.

MRI

Pulmonary MRI has been introduced as a research and diagnostic tool, primarily to overcome the limitations of CT scans.⁴¹ However, pulmonary MRI also has some

limitations, such as the signal diminishing deeper into the lung, respiratory and/or cardiac motion artifacts, and magnetic field distortions due to lung parenchyma.⁴¹ Over the last decade, technical advances have addressed these limitations in functional and morphologic assessment of various pulmonary diseases, including airway diseases.⁴¹ In particular, motion artifacts can be reduced by faster imaging, so that controlled breathing is no longer necessary for acceptable images.⁶⁶

In patients with CF, MRI is useful for detecting morphologic changes in airways and lung parenchyma, in particular inflammation and mucus plugging,⁶⁷ and can be used in patients of almost all ages.^{42,68} In cross-sectional studies, there is a strong correlation between MRI and CT results in patients with CF.⁴² Both MRI and CT are able to detect most large morphologic changes in the CF lung, but MRI is less sensitive for the detection of small airway disease.^{57,69–71} New sequences are in development that might further improve the resolution of MRI.

MRI has no radiation exposure, a clinically acceptable scan time (15–30 min), and is superior to CT for the assessment of functional changes such as altered pulmonary perfusion.⁵⁷ Further, a reproducible morpho-functional MR-scoring-system has been developed, allowing CF lung disease over a broad severity range to be monitored,^{40,68} although hyperperfusion is not taken into account.

MRI is also sensitive to the effects of treatments including antibiotics.⁷² The above mentioned features of MRI may make it particularly suitable for monitoring the course of functional pulmonary changes, such as pulmonary perfusion in response to investigational therapeutic interventions, and assessing other treatment effects in clinical trials. A recent study in infants and preschool children with CF demonstrated that MRI was able to detect abnormalities in lung structure and perfusion, as well as response to treatment for exacerbations.⁶⁸ These findings confirm the potential of MRI for non-invasive monitoring and as an outcome measure in interventional trials for early CF lung disease.⁶⁸

Overall, although MRI is a feasible and utilizable surrogate outcome measure for clinical studies in patients with CF (Table 1), further validation and standardization are still needed.⁷³ In addition, while there are no age limitations to the use of MRI, imaging of children below the age of 6 years remains challenging. New methods are in development, but are not yet in routine use.⁷⁴ Experimental lung imaging techniques that provide additional information on ventilation beyond conventional MRI, such as hyperpolarized noble gas (³He and¹²⁹Xe) MRI^{75–77} and Fourier decomposition MRI,⁷⁸ are currently being evaluated.

PET Scanning

The utility of the FDG-PET scan is based on its unique ability to image active infection and inflammation in the

lungs.^{44,79} In particular, neutrophil (the predominant inflammatory cell in the lungs of patients with CF) activation can be detected using FDG-PET.⁴⁴ Indeed, FDG uptake in the damaged lung has been correlated with uptake by neutrophils and may also be a biomarker of eosinophilic inflammation.^{79,80} In patients with CF, a decline in FEV₁ has been associated with the rate of FDG uptake.⁸¹ In contrast, adult patients with stable CF do not show enhanced FDG uptake compared with control patients, despite high sputum neutrophil levels.⁸²

As the combination of CT and PET scans enables the localization of inflammation to anatomical hotspots, the use of hybrid FDG-PET/CT scanning was evaluated to monitor lung inflammation and/or infection in patients with CF.⁸³ This study reported the presence of localized areas of increased uptake of FDG that may represent active focal infections or inflammatory processes in the lungs.⁸³ Moreover, in this study, the resolution of the acute infection resulted in either a disappearance or great reduction in the high-intensity areas of uptake.⁸³ These findings were recently confirmed in a FDG-PET/CT study in 20 pediatric patients treated for a pulmonary exacerbation.43 The above evidence, in conjunction with clinical observation, has led to the suggestion that FDG-PET/CT scanning may be more valuable than CT for assessing responses to antibiotic treatment in CF patients with acute lung infection.⁸³ Furthermore, FDG-PET/CT imaging could be enhanced by quantitative measurements of regional ventilation and perfusion, to increase sensitivity for detecting functional changes in the lungs, as demonstrated previously for PET scanning.^{84–86}

However, FDG-PET/CT exposes patients to relatively high doses of ionizing radiation, which limits its repeated use (Table 1). The combination of PET and MRI may overcome this limitation as it could provide valuable results with reduced radiation burden, which is of particular importance in children where radiation exposure must be minimized.

The advantages of FDG-PET and FDG-PET/CT scans warrant further research. The development of improved image analysis methods may help to validate this tool as an endpoint for clinical trials.

Mucociliary Clearance

Although impaired mucociliary clearance (MCC) is a hallmark of CF, current evidence suggests that MCC varies significantly in children with CF lung disease and normal pulmonary function, with MCC values reported both within and below the normal range in this cohort.⁸⁷ Furthermore, published studies on MCC as an outcome measure have used different methods and yielded inconsistent results.^{88–90} Robinson et al. demonstrated a global reduction in MCC in CF patients regardless of their lung function, including from the large airways.⁹⁰ In

contrast, Donaldson et al. demonstrated that the 1 hr mucus-clearance rate, which is dominated by largeairway clearance, did not differ significantly between CF patients and controls, whereas clearance in peripheral lung regions and cumulative mucus clearance at 24 hr (both reflective of the smaller airways) were significantly reduced.⁸⁸ Finally, another study showed that long-term lung clearance measured over 21 days was not slower in patients with CF than in healthy subjects.⁸⁹ In this study, greater clearance was reported in CF patients on days 1-7 and no difference in clearance was found between groups from day 7 to day 21.89 Potential interpretations of the absence of a difference in particle clearance after 7 days include the absence of a small airway MCC problem, or that clearance via mechanisms other than mucociliary transport (e.g., macrophage-mediated clearance) dominate during these very long time domains.⁸⁹

MCC has several challenges that must be addressed before use in clinical trials. Particle delivery and the resulting pattern of lung deposition are critical determinants of the observed clearance rate in MCC studies, and therefore must be carefully controlled.⁹¹ Particle deposition depends on aerosol characteristics (average particle size, distribution of size), the breathing pattern used during inhalation (flow rate, tidal volume), and anatomical features of the airways (i.e., degree of obstruction and lung size).⁹¹ Although total deposition tends to be equivalent between patients with CF and controls, scans have shown that deposition is patchy in patients with CF. Particle clearance is also likely to be heterogeneous.^{89,91} potentially complicating the characterization and comparison of clearance rates. To minimize intra- and intersubject variability in particle deposition, aerosol delivery to the bronchial airways must occur in a reproducible manner.⁹¹ Variation in aerosol delivery methods likely explains differences between prior studies, and failure to control these variables can degrade the ability to accurately characterize the response to therapeutic interventions. A further limitation of MCC as an endpoint is spontaneous cough and associated additional mucus clearance that has been reported as a main adverse event following inhaled medications.⁸⁷ Finally, MCC is not sensitive to all therapeutic agents; while sensitive to agents that effectively change airway surface liquid hydration, it was not altered by dornase alfa in previous studies.92,93

Despite these limitations, recent findings suggest that MCC has the potential to be used as an outcome measure in CF clinical trials. Results from one study using MCC to assess the efficacy of hypertonic saline suggest that there is an association between improvements in MCC and lung function tests.⁸⁸ In this study, MCC, FEV₁, and forced vital capacity significantly improved over treatment time in patients receiving hypertonic saline, but not in those who received the ineffective combination of amiloride

and hypertonic saline.⁸⁸ In the recent GOAL study, the profound effect of restoring CFTR function on MCC, using ivacaftor in patients with the G551D mutation, also demonstrated the tight link between CFTR function and MCC.⁹⁴ In this multicenter study, MCC more than doubled within 1 month of starting ivacaftor, and this effect was maintained after 3 months of treatment. These data suggest that MCC measurements may provide a useful tool for the study of novel CFTR modulators in the therapeutic pipeline.

At present, measurement of MCC is a promising, but incompletely developed, biomarker for CF clinical research (Table 1). A standard operating procedure has recently been developed, which has facilitated the conduct of multicenter studies.⁹⁵ It should also be borne in mind that MCC is not sensitive to all therapeutic agents, and therefore its application must be used selectively in future clinical trials.

Biomarkers as Outcome Measures

Sputum Biomarkers

Sputum is easily obtainable and a rich source of biomarkers of inflammation and infection in patients with CF.^{96–98} As airway inflammation plays a central role in CF lung disease, sputum biomarkers of inflammation that can be used to monitor disease activity or evaluate response to therapy would be valuable. Biomarkers may also provide further insight into the pathophysiology of CF lung disease.

Findings from small single-center studies are limited, but support an association between sputum biomarkers and disease status in CF, as determined by pulmonary function tests, chest radiograph scores, HRQoL measures, and illness severity scores (e.g., Shwachman–Kulczycki score).⁹⁷ Significant correlations between FEV₁ and sputum inflammatory measures, including neutrophil counts, interleukin (IL)-8, and neutrophil elastase, have also been demonstrated in a diverse CF population across multiple centers participating in four CF clinical trials.⁹⁹

Several studies have demonstrated good reproducibility of cell counts and inflammatory mediators in induced sputum.^{97,100–102} Emerging longitudinal analyses of sputum biomarkers provide variability estimates over time, allowing investigators to derive sample size calculations for interventional trials.^{103,104}

Reductions in sputum biomarkers following therapeutic intervention have been demonstrated in a number of clinical studies.^{105–107} Following intravenous antibiotic therapy in patients with CF, reductions in neutrophil counts, IL-8 concentration and neutrophil elastase activity were associated with improvements in FEV₁.¹⁰⁶ Importantly, sputum induction was relatively well tolerated in CF patients, even during acute pulmonary exacerbations.¹⁰⁶ In the initial US azithromycin trial, there were modest differences in sputum elastase between the placebo and treated groups at the end of treatment in favor of the azithromycin group, suggesting that azithromycin may exert an anti-inflammatory effect by preventing a worsening of protease-mediated inflammation over time.¹⁰⁷ Two other studies, one investigating the anti-inflammatory effects of ibuprofen,¹⁰⁸ the other examining the CFTR potentiator ivacaftor in G551D subjects,⁹⁴ did not show significant changes in sputum biomarkers of airway inflammation. While these studies raise concerns about the utility of sputum biomarkers, the lack of significant changes could also be attributed to the relatively short treatment periods and the need for more prolonged therapies before substantially impacting airway inflammation. Other potential sputum biomarkers that might be relevant to CF clinical trials include biomarkers of structural injury and matrix degradation;¹⁰⁹ physical properties, including wet and dry weight, surface tension and impedance; rheology (viscosity, elasticity), and microparticles;¹¹⁰ as well as biochemical constituents besides inflammatory mediators (mucins, fibrin, DNA).

Overall, sputum assessments provide a non-invasive endpoint for assessing novel therapies for CF, especially in patients over 10 years of age. Ongoing investigations into the variability and reproducibility of this technique are a practical step before widespread application in clinical trial methodology. In addition, longitudinal analyses are essential for the validation of biomarkers of inflammation, as correlates of disease severity and progression. Data are now emerging from ongoing investigations evaluating the ability of sputum biomarkers to predict key clinical events in CF, including lung function decline,^{104,111} pulmonary exacerbations,^{103,105} development of bronchiectasis⁴⁷ and even survival.¹⁰³ Taken together, these data provide strong support for the use of sputum biomarkers of airway inflammation as tools to monitor disease activity and as outcome measures in CF clinical trials.

Blood-Based Biomarkers

Serum and plasma biomarkers have the potential to provide a relatively non-invasive means of evaluating pulmonary inflammation and infection in patients with CF. Several circulating biomarkers have been investigated, including C-reactive protein, serum amyloid A, calprotectin, neutrophil elastase antiprotease complexes, plasma sCD14, a protein complex containing alfa-1 antitrypsin and CD16b (AAT:CD16b), and cytokines including IL-6 and IL-8.^{112–115} A recent systematic review summarized the results of studies that have used blood-based biomarkers to monitor response to treatment during pulmonary exacerbations.¹¹⁶ In a clinical trial in CF patients 6–18 years of age uninfected with *Pseudomonas aeruginosa*, azithromycin significantly reduced circulating neutrophil counts and systemic markers of

inflammation including C-reactive protein, serum amyloid A, and calprotectin.¹¹⁷ Reduction in these inflammatory markers correlated with improvements in lung function and weight gain, providing indirect evidence that these changes were associated with clinically meaningful outcomes. This was the first study to demonstrate the utility of a panel of systemic inflammatory markers in a CF interventional trial and these data provide evidence that systemic biomarkers have added value and should be included in future CF clinical trials.

Exhaled Breath Condensate

Exhaled breath condensate (EBC) may provide a potential source of biomarkers that could be useful in a variety of diseases, including CF.¹¹⁸ Several potential EBC biomarkers have been investigated in CF, including EBC pH, fractional exhaled nitric oxide, leukotriene-B4, 8-isoprostane, hydrogen peroxide, interferon- γ , IL-10, IL-4, tumor necrosis factor, purines and glucose.¹¹⁸ At present there is insufficient evidence to support the use of any of these biomarkers as outcome measures in clinical studies.

Assessment of CFTR Activity

Nasal Potential Difference

It is well established that CF is caused by mutations in the CFTR gene, resulting in disruption of chloride and bicarbonate transport across epithelial cell membranes.¹¹⁹ These abnormalities can be evaluated by measuring the transepithelial potential difference (PD) across the nasal epithelium. The degree of CFTR dysfunction, as measured by nasal PD, correlates with the number and severity of CFTR gene mutations.¹²⁰ Measurement of nasal PD has therefore proved useful in clinical studies of novel CFTR therapeutic agents¹²¹⁻¹²⁴ and is used in clinical practice as one of the diagnostic criteria for CF.^{125–127} However, there are several different methods for measuring nasal PD, as well as important considerations with respect to its use in clinical trials, such as the requirement for well-trained and dedicated personnel to provide accurate assay results.¹²⁸ Commercially-available equipment, which has made this measurement easier to perform, is now in use.¹²⁹ The agar nasal catheter has shown greater reliability than the perfusion nasal catheter for measurement of nasal PD,¹³⁰ and its sensitivity and specificity are generally understood.^{131,132} The tool has been particularly useful for evaluating the response, including the effect of different doses of the CFTR potentiator ivacaftor, in G551D CF patients.¹²¹

PD measurements can also be made in the lower airway via bronchoscopy,^{133,134} thus permitting direct assessment of the airways. This could be particularly important for evaluating the effect of inhaled drugs or gene therapy in which only pulmonary delivery is expected. A study

has shown that invasive bronchoscopic methods can be utilized safely and reliably in children as young as 1 year of age, albeit under anesthesia.¹³³ Therefore, this measure could be a useful functional endpoint assay for studies of either CFTR gene transfer or for future trials evaluating inhaled therapeutics.

Intestinal Current Measurement (ICM)

Intestinal current measurement (ICM) was introduced about two decades ago as an ex vivo diagnostic method for CF and has been the subject of renewed interest.^{135–140} The technique can distinguish pancreatic-sufficient individuals, indicating its utility in quantifying patient phenotype.¹⁴¹ ICM has some advantages over nasal and lower airway PD techniques, including easy access to intestinal tissue in all age groups and minimal tissue damage or remodeling triggered by bacterial or viral infections.136 Moreover, ICM allows novel CFTR therapies to be studied in native human epithelium ex vivo without increasing risk to the patient and is able to detect low levels of functionally active CFTR.¹³⁶ The potential use of ICM in clinical trials is being evaluated, and has been combined with protein detection by Western blotting.¹³⁶ In addition, standardized guidelines for this technique are now available, following collaboration between the European CF Society Diagnostic Network Working Group, the European CF Society Clinical Trials Network and the CF Foundation Therapeutics Development Network.¹⁴²

Sweat Test

Determination of sweat electrolytes has been performed in clinical laboratories for over 40 years and remains the gold standard diagnostic test for CF.¹⁴³ The measure provides a sensitive indicator of CFTR activity, and correlates well with the CF phenotype.¹²⁰ Sweat chloride measurements are feasible in multicenter clinical trials,^{5,121,122,143} and use of the macroduct collection system allows analysis in a central laboratory, facilitating standardized methodology among centers. Furthermore, the procedure does not place a significant burden on patients and is rarely associated with complications.¹⁴³ Several lines of evidence suggest that sweat chloride is a robust outcome measure in clinical trials evaluating agents directed at restoring CFTR function.^{5,121,122} As sweat chloride levels change significantly with small changes in CFTR activity, as seen in genotype-phenotype correlations,¹²⁰ their determination allows the evaluation of the effect of CFTR modulators with relatively low activity,¹²² providing a reasonable assay for doseresponse relationships. However, change in sweat chloride has not proven to be predictive of lung function response on an individual basis in studies involving a CFTR potentiator.¹⁴⁴ Recently, sweat rate and sweat conductivity have been determined in conjunction with

sweat chloride levels, to assess the secretory function of the sweat gland.^{145,146} These techniques may be particularly well suited to detecting altered glandular activity, although this needs to be demonstrated in prospective studies.

In conclusion, application of ion channel measurements in clinical trials appears feasible given its established use, favorable safety and acceptability profile and validity (Table 1). Widespread use in recent years has improved understanding of its sensitivity, specificity, and reproducibility.^{121,122,145,147,148} However, the need for well-trained personnel and use of accepted standard operating procedures should be employed when including sweat chloride as an outcome measure in clinical trials. In addition, concerns regarding whether pharmacodynamic activity in the absorptive epithelium of the sweat gland correlates with activity in the secretory epithelia of the airway and other organs still need to be addressed.¹⁴⁹ This could be achieved by characterizing CFTR, and its response to novel therapies, in different tissues. Other questions include whether tissue (i.e., skin) penetration or differences in the absorptive function of CFTR following pharmacologic rescue of F508del may impair its ability to detect the efficacy of multi-agent therapy.

CONCLUSION

Spirometry is the standard clinical trial endpoint in older children and adults with CF. However, it is now well recognized that, over time, decline in FEV_1 has become relatively small, thus making it an insufficiently sensitive marker of CF to serve as a primary endpoint in clinical studies. There is consequently an urgent need for alternative sensitive and accurate surrogate outcome measures that detect early lung disease and track disease progression through early childhood into adulthood.⁴

Ideally endpoints must be accurate, reproducible, sensitive, and reflect patient function and survival. They should also predict the efficacy of therapy,^{99,145,147} as well as being feasible for use in clinical studies with small patient numbers, non-invasive and inexpensive. The most promising and feasible new sensitive outcome measures that can be used in today's clinical trials to measure severity of CF lung disease in (young) children and adults, are the LCI and the bronchiectasis scores derived from chest CT. Tests to measure CFTR activity, such as the sweat test, are of key importance to establish the effect of CFTR modifiers on CFTR function.

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REFERENCES

- 1. Cystic Fibrosis Foundation. Cystic Fibrosis Foundation Patient Registry. 2008 annual data report. Bethesda, Matyland. Last accessed 14 August 2013. http://www.cff.org/treatments/Care-CenterNetwork/PatientRegistryReport/.
- Ramsey BW. Use of lung imaging studies as outcome measures for development of new therapies in cystic fibrosis. Proc Am Thorac Soc 2007;4:359–363.
- Yuksel H, Yilmaz O. A new model for cystic fibrosis management: control concept. Pneumologia 2011;60:150–154.
- 4. Aurora P, Stanojevic S, Wade A, Oliver C, Kozlowska W, Lum S, Bush A, Price J, Carr SB, Shankar A, et al. Lung clearance index at 4 years predicts subsequent lung function in children with cystic fibrosis. Am J Respir Crit Care Med 2011;183:752–758.
- 5. Ramsey BW, Davies J, McElvaney NG, Tullis E, Bell SC, Drevinek P, Griese M, McKone EF, Wainwright CE, Konstan MW, et al. A CFTR potentiator in patients with cystic fibrosis and the G551D mutation. N Engl J Med 2011;365:1663–1672.
- Davis PB, Yasothan U, Kirkpatrick P. Ivacaftor. Nat Rev Drug Discov 2012;11:349–350.
- Bakker EM, Borsboom GJ, van der Wiel-Kooij EC, Caudri D, Rosenfeld M, Tiddens HA. Small airway involvement in cystic fibrosis lung disease: routine spirometry as an early and sensitive marker. Pediatr Pulmonol 2013;48:1081–1088.
- Bakker M, Borsboom G, Tiddens H. The authors reply. Pediatr Pulmonol 2014;49:1256–1257.
- Bakker EM, Volpi S, Salonini E, van der Wiel-Kooij EC, Sintnicolaas CJ, Hop WC, Assael BM, Merkus PJ, Tiddens HA. Improved treatment response to dornase alfa in cystic fibrosis patients using controlled inhalation. Eur Respir J 2011;38:1328– 1335.
- 10. Rosenfeld M, Allen J, Arets BH, Aurora P, Beydon N, Calogero C, Castile RG, Davis SD, Fuchs S, Gappa M, et al. An official American Thoracic Society workshop report: optimal lung function tests for monitoring cystic fibrosis, bronchopulmonary dysplasia, and recurrent wheezing in children less than 6 years of age. Ann Am Thorac Soc 2013;10:S1–S11.
- 11. Davis SD, Rosenfeld M, Kerby GS, Brumback L, Kloster MH, Acton JD, Colin AA, Conrad CK, Hart MA, Hiatt PW, et al. Multicenter evaluation of infant lung function tests as cystic fibrosis clinical trial endpoints. Am J Respir Crit Care Med 2010;182:1387–1397.
- ATS/ERS statement: raised volume forced expirations in infants: guidelines for current practice. Am J Respir Crit Care Med 2005;172:1463–1471.
- 13. Ranganathan SC, Dezateux C, Bush A, Carr SB, Castle RA, Madge S, Price J, Stroobant J, Wade A, Wallis C, et al. Airway function in infants newly diagnosed with cystic fibrosis. Lancet 2001;358:1964–1965.
- 14. Hoo AF, Thia LP, Nguyen TT, Bush A, Chudleigh J, Lum S, Ahmed D, Balfour L, Carr I, Chavasse SB, et al. Lung function is abnormal in 3-month-old infants with cystic fibrosis diagnosed by newborn screening. Thorax 2012;67:874–881.
- 15. Linnane BM, Hall GL, Nolan G, Brennan S, Stick SM, Sly PD, Robertson CF, Robinson PJ, Franklin PJ, Turner SW, et al. Lung function in infants with cystic fibrosis diagnosed by newborn screening. Am J Respir Crit Care Med 2008;178:1238–1244.
- Pittman JE, Johnson RC, Davis SD. Improvement in pulmonary function following antibiotics in infants with cystic fibrosis. Pediatr Pulmonol 2012;47:441–446.
- 17. Rosenfeld M, Ratjen F, Brumback L, Daniel S, Rowbotham R, McNamara S, Johnson R, Kronmal R, Davis SD. Inhaled hypertonic saline in infants and children younger than 6 years with cystic fibrosis: the ISIS randomized controlled trial. JAMA 2012;307:2269–2277.

- The raised volume rapid thoracoabdominal compression technique. The Joint American Thoracic Society/European Respiratory Society Working Group on Infant Lung Function. Am J Respir Crit Care Med 2000;161:1760–1762.
- Horsley A. Lung clearance index in the assessment of airways disease. Respir Med 2009;103:793–799.
- 20. Gustafsson PM, Aurora P, Lindblad A. Evaluation of ventilation maldistribution as an early indicator of lung disease in children with cystic fibrosis. Eur Respir J 2003;22:972–979.
- 21. Gustafsson PM, De Jong PA, Tiddens HA, Lindblad A. Multiplebreath inert gas washout and spirometry versus structural lung disease in cystic fibrosis. Thorax 2008;63:129–134.
- 22. Ellemunter H, Fuchs SI, Unsinn KM, Freund MC, Waltner-Romen M, Steinkamp G, Gappa M. Sensitivity of Lung Clearance Index and chest computed tomography in early CF lung disease. Respir Med 2010;104:1834–1842.
- 23. Aurora P, Bush A, Gustafsson P, Oliver C, Wallis C, Price J, Stroobant J, Carr S, Stocks J. Multiple-breath washout as a marker of lung disease in preschool children with cystic fibrosis. Am J Respir Crit Care Med 2005;171:249–256.
- 24. Lum S, Gustafsson P, Ljungberg H, Hülskamp G, Bush A, Carr SB, Castle R, Hoo AF, Price J, Ranganathan S, et al. Early detection of cystic fibrosis lung disease: multiple-breath washout versus raised volume tests. Thorax 2007;62:341–347.
- 25. Owens CM, Aurora P, Stanojevic S, Bush A, Wade A, Oliver C, Calder A, Price J, Carr SB, Shankar A, et al. Lung Clearance Index and HRCT are complementary markers of lung abnormalities in young children with CF. Thorax 2011;66:481–488.
- 26. Vermeulen F, Proesmans M, Boon M, Havermans T, De Boeck K. Lung clearance index predicts pulmonary exacerbations in young patients with cystic fibrosis. Thorax 2014;69:39–45.
- 27. Amin R, Subbarao P, Jabar A, Balkovec S, Jensen R, Kerrigan S, Gustafsson P, Ratjen F. Hypertonic saline improves the LCI in paediatric patients with CF with normal lung function. Thorax 2010;65:379–383.
- 28. Amin R, Subbarao P, Lou W, Jabar A, Balkovec S, Jensen R, Kerrigan S, Gustafsson P, Ratjen F. The effect of dornase alfa on ventilation inhomogeneity in patients with cystic fibrosis. Eur Respir J 2011;37:806–812.
- 29. Davies J, Sheridan H, Bell N, Cunningham S, Davis SD, Elborn JS, Milla CE, Starner TD, Weiner DJ, Lee PS, Ratjen F. Assessment of clinical response to ivacaftor with lung clearance index in cystic fibrosis patients with a G551D-CFTR mutation and preserved spirometry: a randomised controlled trial. Lancet Respir Med 2013;1:630–638.
- Fuchs SI, Eder J, Ellemunter H, Gappa M. Lung clearance index: normal values, repeatability, and reproducibility in healthy children and adolescents. Pediatr Pulmonol 2009;44:1180–1185.
- Fuchs SI, Gappa M. Lung clearance index: clinical and research applications in children. Paediatr Respir Rev 2011;12: 264–270.
- 32. Fuchs SI, Buess C, Lum S, Kozlowska W, Stocks J, Gappa M. Multiple breath washout with a sidestream ultrasonic flow sensor and mass spectrometry: a comparative study. Pediatr Pulmonol 2006;41:1218–1225.
- 33. Jensen R, Stanojevic S, Gibney K, Salazar JG, Gustafsson P, Subbarao P, Ratjen F. Multiple breath nitrogen washout: a feasible alternative to mass spectrometry. PLoS ONE 2013;8:e56868.
- 34. Kent L, Reix P, Innes JA, Zielen S, Le BM Braggion, Lever S, Arets HG, Brownlee K, Bradley JM, Bayfield K, O'Neill K, et al. Lung clearance index: evidence for use in clinical trials in cystic fibrosis. J Cyst Fibros 2014;13:123–138.
- 35. Subbarao P, Stanojevic S, Brown M, Jensen R, Rosenfeld M, Davis S, Brumback L, Gustafsson P, Ratjen F. Lung clearance index as an outcome measure for clinical trials in young children with cystic

fibrosis. A pilot study using inhaled hypertonic saline. Am J Respir Crit Care Med 2013;188:456–460.

- 36. Daines C, Morgan W. The importance of imaging in cystic fibrosis. Am J Respir Crit Care Med 2011;184:751–752.
- 37. De Jong PA, Ottink MD, Robben SG, Lequin MH, Hop WC, Hendriks JJ, Pare PD, Tiddens HA. Pulmonary disease assessment in cystic fibrosis: comparison of CT scoring systems and value of bronchial and arterial dimension measurements. Radiology 2004;231:434–439.
- 38. Eichinger M, Puderbach M, Fink C, Gahr J, Ley S, Plathow C, Tuengerthal S, Zuna I, Muller FM, Kauczor HU. Contrastenhanced 3D MRI of lung perfusion in children with cystic fibrosis-initial results. Eur Radiol 2006;16:2147–2152.
- 39. Loeve M, van Hal PT, Robinson P, De Jong PA, Lequin MH, Hop WC, Williams TJ, Nossent GD, Tiddens HA. The spectrum of structural abnormalities on CT scans from patients with CF with severe advanced lung disease. Thorax 2009;64:876–882.
- 40. Eichinger M, Optazaite DE, Kopp-Schneider A, Hintze C, Biederer J, Niemann A, Mall MA, Wielputz MO, Kauczor HU, Puderbach M. Morphologic and functional scoring of cystic fibrosis lung disease using MRI. Eur J Radiol 2012;81:1321–1329.
- 41.Ohno Y, Koyama H, Yoshikawa T, Nishio M, Matsumoto S, Iwasawa T, Sugimura K. Pulmonary magnetic resonance imaging for airway diseases. J Thorac Imaging 2011;26:301–316.
- 42. Puderbach M, Eichinger M, Haeselbarth J, Ley S, Kopp-Schneider A, Tuengerthal S, Schmaehl A, Fink C, Plathow C, Wiebel M, et al. Assessment of morphological MRI for pulmonary changes in cystic fibrosis (CF) patients: comparison to thin-section CT and chest X-ray. Invest Radiol 2007;42:715–725.
- 43. Amin R, Charron M, Grinblat L, Shammas A, Grasemann H, Graniel K, Ciet P, Tiddens H, Ratjen F. Cystic fibrosis: detecting changes in airway inflammation with FDG PET/CT. Radiology 2012;264:868–875.
- 44. Chen DL, Schuster DP. Positron emission tomography with [18F] fluorodeoxyglucose to evaluate neutrophil kinetics during acute lung injury. Am J Physiol Lung Cell Mol Physiol 2004;286:L834– L840.
- 45. De Jong PA, Lindblad A, Rubin L, Hop WC, de Jongste JC, Brink M, Tiddens HA. Progression of lung disease on computed tomography and pulmonary function tests in children and adults with cystic fibrosis. Thorax 2006;61:80–85.
- 46. Sly PD, Ware RS, de Klerk N, Stick SM. Randomised controlled trials in cystic fibrosis: what, when and how?. Eur Respir J 2011;37:991–993.
- 47. Sly PD, Gangell CL, Chen L, Ware RS, Ranganathan S, Mott LS, Murray CP, Stick SM. Risk factors for bronchiectasis in children with cystic fibrosis. N Engl J Med 2013;368:1963–1970.
- 48. Hall GL, Logie KM, Parsons F, Schulzke SM, Nolan G, Murray C, Ranganathan S, Robinson P, Sly PD, Stick SM, et al. Air trapping on chest CT is associated with worse ventilation distribution in infants with cystic fibrosis diagnosed following newborn screening. PLoS ONE 2011;6:e23932.
- 49. Stick SM, Brennan S, Murray C, Douglas T, von Ungern-Sternberg BS, Garratt LW, Gangell CL, de KN, Linnane B, Ranganathan S, et al. Bronchiectasis in infants and preschool children diagnosed with cystic fibrosis after newborn screening. J Pediatr 2009;155:623–628.
- 50. Mott LS, Park J, Murray CP, Gangell CL, de Klerk NH, Robinson PJ, Robertson CF, Ranganathan SC, Sly PD, Stick SM. Progression of early structural lung disease in young children with cystic fibrosis assessed using CT. Thorax 2012;67:509–516.
- 51. Loeve M, Gerbrands K, Hop WC, Rosenfeld M, Hartmann IC, Tiddens HA. Bronchiectasis and pulmonary exacerbations in children and young adults with cystic fibrosis. Chest 2011; 140:178–185.

- 52. Loeve M, Tiddens H, Gorbunova V, Hop WC, de Bruijne M. Reversibility of trapped air on CT scans of CF patients, an automated approach. Pediatr Pulmonol 2011;46:-359. Abstract 349.
- 53.Brody AS, Sucharew H, Campbell JD, Millard SP, Molina PL, Klein JS, Quan J. Computed tomography correlates with pulmonary exacerbations in children with cystic fibrosis. Am J Respir Crit Care Med 2005;172:1128–1132.
- 54. Loeve M, Hop WC, de Bruijne M, van Hal PT, Robinson P, Aitken ML, Dodd JD, Tiddens HA. Chest computed tomography scores are predictive of survival in patients with cystic fibrosis awaiting lung transplantation. Am J Respir Crit Care Med 2012;185:1096–1103.
- 55. Tepper LA, Utens EM, Caudri D, Bos AC, Gonzalez-Graniel K, Duivenvoorden HJ, van der Wiel EC, Quittner AL, Tiddens HA. Impact of bronchiectasis and trapped air on quality of life and exacerbations in cystic fibrosis. Eur Respir J 2013;42:371–379.
- 56. Sanders DB, Li Z, Brody A, Farrell PM. Bronchiectasis on chest CT predicts future lung disease severity in children with CF. Pediatr Pulmonol 2011;46(Suppl 34):330. Abstract 327.
- 57. Eichinger M, Heussel CP, Kauczor HU, Tiddens H, Puderbach M. Computed tomography and magnetic resonance imaging in cystic fibrosis lung disease. J Magn Reson Imaging 2010;32:1370–1378.
- Brenner DJ, Hall EJ. Computed tomography–an increasing source of radiation exposure. N Engl J Med 2007;357:2277–2284.
- 59. Sanders DB, Li Z, Rock MJ, Brody AS, Farrell PM. The sensitivity of lung disease surrogates in detecting chest CT abnormalities in children with cystic fibrosis. Pediatr Pulmonol 2012;47:567–573.
- 60. Willemink MJ, De Jong PA. Pediatric chest computed tomography at a radiation dose approaching a chest radiograph. Am J Respir Crit Care Med 2013;188:626–627.
- 61. Kubo T, Lin PJ, Stiller W, Takahashi M, Kauczor HU, Ohno Y, Hatabu H. Radiation dose reduction in chest CT: a review. AJR Am J Roentgenol 2008;190:335–343.
- 62. Kalra MK, Woisetschlager M, Dahlstrom N, Singh S, Digumarthy S, Do S, Pien H, Quick P, Schmidt B, Sedlmair M, et al. Sinogramaffirmed iterative reconstruction of low-dose chest CT: effect on image quality and radiation dose. AJR Am J Roentgenol 2013;201:W235–W244.
- 63.Bang DH, Lim D, Hwang WS, Park SH, Jeong OM, Kang KW, Kang H. Lateral topography for reducing effective dose in lowdose chest CT. AJR Am J Roentgenol 2013;200:1294–1297.
- 64. Kuo W, Ciet P, Tiddens HA, Zhang W, Guillerman RP, van Straten M. Monitoring cystic fibrosis lung disease by computed tomography. Radiation risk in perspective. Am J Respir Crit Care Med 2014;189:1328–1336.
- 65. Brody AS, Guillerman RP. Don't let radiation scare trump patient care: 10 ways you can harm your patients by fear of radiationinduced cancer from diagnostic imaging. Thorax 2014;69:782– 784.
- 66. Matsuoka S, Hunsaker AR, Gill RR, Jacobson FL, Ohno Y, Patz S, Hatabu H. Functional MR imaging of the lung. Magn Reson Imaging Clin N Am 2008;16:275–289.
- 67.Puderbach M, Eichinger M, Gahr J, Ley S, Tuengerthal S, Schmahl A, Fink C, Plathow C, Wiebel M, Muller FM, et al. Proton MRI appearance of cystic fibrosis: comparison to CT. Eur Radiol 2007;17:716–724.
- 68. Wielpütz MO, Puderbach M, Kopp-Schneider A, Stahl M, Fritzsching E, Sommerburg O, Ley S, Sumkauskaite M, Biederer J, Kauczor HU, et al. Magnetic resonance imaging detects changes in structure and perfusion, and response to therapy in early cystic fibrosis lung disease. Am J Respir Crit Care Med 2014;189:956– 965.
- 69.Failo R, Wielopolski PA, Tiddens HA, Hop WC, Mucelli RP, Lequin MH. Lung morphology assessment using MRI: a robust

ultra-short TR/TE 2D steady state free precession sequence used in cystic fibrosis patients. Magn Reson Med 2009;61:299–306.

- 70. Ciet P, Serra G, Bertolo S, et al. Comparison of chest-MRI to chest-CT to monitor cystic fibrosis (CF) lung disease. 97th Annual Scientific Assembly and Annual Meeting of the Radiological Society of North America. Chicago, IL, USA, 27 November - 02 December 2011 (Abstract).
- 71.Puderbach M, Eichinger M. The role of advanced imaging techniques in cystic fibrosis follow-up: is there a place for MRI?. Pediatr Radiol 2010;40:844–849.
- 72. Redmond A, Sweeney L, MacFarland M, Mitchell M, Daggett S, Kubin R. Oral ciprofloxacin in the treatment of pseudomonas exacerbations of paediatric cystic fibrosis: clinical efficacy and safety evaluation using magnetic resonance image scanning. J Int Med Res 1998;26:304–312.
- 73. Jannin P, Fitzpatrick JM, Hawkes DJ, Pennec X, Shahidi R, Vannier MW. Validation of medical image processing in imageguided therapy. IEEE Trans Med Imaging 2002;21:1445–1449.
- 74. MacKenzie JD, Vasanawala SS. Advances in pediatric MR imaging. Magn Reson Imaging Clin N Am 2008;16:385–402.
- 75. Fain SB, Korosec FR, Holmes JH, O'Halloran R, Sorkness RL, Grist TM. Functional lung imaging using hyperpolarized gas MRI. J Magn Reson Imaging 2007;25:910–923.
- 76. Stadler A, Stiebellehner L, Jakob PM, Arnold JF, Eisenhuber E, von Katzler I, Bankier AA. Quantitative and O₂ enhanced MRI of the pathologic lung: findings in emphysema, fibrosis, and cystic fibrosis. Int J Biomed Imaging 2007;2007:23624.
- 77. Sun Y, O'Sullivan BP, Roche JP, Walvick R, Reno A, Baker D, Mansour JK, Albert MS. Using hyperpolarized 3He MRI to evaluate treatment efficacy in cystic fibrosis patients. J Magn Reson Imaging 2011;34:1206–1211.
- 78. Bauman G, Puderbach M, Deimling M, Jellus V, Chefd'hotel C, Dinkel J, Hintze C, Kauczor HU, Schad LR. Non-contrastenhanced perfusion and ventilation assessment of the human lung by means of fourier decomposition in proton MRI. Magn Reson Med 2009;62:656–664.
- Chen DL, Kinahan PE. Multimodality molecular imaging of the lung. J Magn Reson Imaging 2010;32:1409–1420.
- 80. Harris RS, Venegas JG, Wongviriyawong C, Winkler T, Kone M, Musch G, Vidal Melo, de Prost MF, Hamilos N, Afshar DL, et al. 18F-FDG uptake rate is a biomarker of eosinophilic inflammation and airway response in asthma. J Nucl Med 2011;52:1713– 1720.
- 81. Dolovich MB, Schuster DP. Positron emission tomography and computed tomography versus positron emission tomography computed tomography: tools for imaging the lung. Proc Am Thorac Soc 2007;4:328–333.
- 82. Labiris NR, Nahmias C, Freitag AP, Thompson ML, Dolovich MB. Uptake of 18fluorodeoxyglucose in the cystic fibrosis lung: a measure of lung inflammation?. Eur Respir J 2003;21:848–854.
- 83. Klein M, Cohen-Cymberknoh M, Armoni S, Shoseyov D, Chisin R, Orevi M, Freedman N, Kerem E. F-fluorodeoxyglucose-PET/ CT imaging of lungs in patients with cystic fibrosis. Chest 2009;136:1220–1228.
- Musch G, Venegas JG. Positron emission tomography imaging of regional lung function. Minerva Anestesiol 2006;72:363–367.
- 85. Schroeder T, Vidal Melo MF, Musch G, Harris RS, Winkler T, Venegas JG. PET imaging of regionalF-FDG uptake and lung function after cigarette smoke inhalation. J Nucl Med 2007; 48:413–419.
- 86. Musch G, Winkler T, Harris RS, Vidal Melo MF, Wellman TJ, de Prost N, Kradin RL, Venegas JG. Lung [F]fluorodeoxyglucose uptake and ventilation-perfusion mismatch in the early stage of experimental acute smoke inhalation. Anesthesiology 2014;120: 683–693.

- 87. Laube BL, Sharpless G, Carson KA, Kelly A, Mogayzel PJ, Jr. Acute inhalation of hypertonic saline does not improve mucociliary clearance in all children with cystic fibrosis. BMC Pulm Med 2011;11:45.
- 88. Donaldson SH, Bennett WD, Zeman KL, Knowles MR, Tarran R, Boucher RC. Mucus clearance and lung function in cystic fibrosis with hypertonic saline. N Engl J Med 2006;354:241–250.
- 89.Lindström M, Camner P, Falk R, Hjelte L, Philipson K, Svartengren M. Long-term clearance from small airways in patients with cystic fibrosis. Eur Respir J 2005;25:317–323.
- Robinson M, Eberl S, Tomlinson C, Daviskas E, Regnis JA, Bailey DL, Torzillo PJ, Menache M, Bye PT. Regional mucociliary clearance in patients with cystic fibrosis. J Aerosol Med 2000;13:73–86.
- 91. Donaldson SH, Corcoran TE, Laube BL, Bennett WD. Mucociliary clearance as an outcome measure for cystic fibrosis clinical research. Proc Am Thorac Soc 2007;4:399–405.
- 92. Laube BL, Auci RM, Shields DE, Christiansen DH, Lucas MK, Fuchs HJ, Rosenstein BJ. Effect of rhDNase on airflow obstruction and mucociliary clearance in cystic fibrosis. Am J Respir Crit Care Med 1996;153:752–760.
- 93. Robinson M, Hemming AL, Moriarty C, Eberl S, Bye PT. Effect of a short course of rhDNase on cough and mucociliary clearance in patients with cystic fibrosis. Pediatr Pulmonol 2000;30: 16–24.
- 94. Rowe SM, Heltshe SL, Gonska T, Donaldson SH, Borowitz D, Gelfond D, Sagel SD, Khan U, Mayer-Hamblett N, Van Dalfsen JM, et al. Clinical mechanism of the cystic fibrosis transmembrane conductance regulator potentiator ivacaftor in G551D-mediated cystic fibrosis. Am J Respir Crit Care Med 2014;190:175–184.
- 95.Bennett WD, Laube BL, Corcoran T, Zeman K, Sharpless G, Thomas K, Wu J, Mogayzel PJ, Jr., Pilewski J, Donaldson S. Multisite comparison of mucociliary and cough clearance measures using standardized methods. J Aerosol Med Pulm Drug Deliv 2013;26:157–164.
- 96. Rogers GB, Hoffman LR, Johnson MW, Mayer-Hamblett N, Schwarze J, Carroll MP, Bruce KD. Using bacterial biomarkers to identify early indicators of cystic fibrosis pulmonary exacerbation onset. Expert Rev Mol Diagn 2011;11:197–206.
- 97.Sagel SD, Chmiel JF, Konstan MW. Sputum biomarkers of inflammation in cystic fibrosis lung disease. Proc Am Thorac Soc 2007;4:406–417.
- Al-Saleh S, Dell SD, Grasemann H, Yau YC, Waters V, Martin S, Ratjen F. Sputum induction in routine clinical care of children with cystic fibrosis. J Pediatr 2010;157:1006–1011.
- 99. Mayer-Hamblett N, Aitken ML, Accurso FJ, Kronmal RA, Konstan MW, Burns JL, Sagel SD, Ramsey BW. Association between pulmonary function and sputum biomarkers in cystic fibrosis. Am J Respir Crit Care Med 2007;175:822–828.
- 100. Ordoñez CL, Kartashov AI, Wohl ME. Variability of markers of inflammation and infection in induced sputum in children with cystic fibrosis. J Pediatr 2004;145:689–692.
- 101. Reinhardt N, Chen CI, Loppow D, Schink T, Kleinau I, Jörres RA, Wahn U, Magnussen H, Paul KP. Cellular profiles of induced sputum in children with stable cystic fibrosis: comparison with BAL. Eur Respir J 2003;22:497–502.
- 102. Smountas AA, Lands LC, Mohammed SR, Grey V. Induced sputum in cystic fibrosis: within-week reproducibility of inflammatory markers. Clin Biochem 2004;37:1031–1036.
- 103. Liou TG, Adler FR, Keogh RH, Li Y, Jensen JL, Walsh W, Packer K, Clark T, Carveth H, Chen J, et al. Sputum biomarkers and the prediction of clinical outcomes in patients with cystic fibrosis. PLoS ONE 2012;7:e42748.
- 104. Sagel SD, Wagner BD, Anthony MM, Emmett P, Zemanick ET. Sputum biomarkers of inflammation and lung function decline in

children with cystic fibrosis. Am J Respir Crit Care Med 2012;186:857-865.

- 105. Colombo C, Faelli N, Tirelli AS, Fortunato F, Biffi A, Claut L, Cariani L, Daccò V, Prato R, Conese M. Analysis of inflammatory and immune response biomarkers in sputum and exhaled breath condensate by a multi-parametric biochip array in cystic fibrosis. Int J Immunopathol Pharmacol 2011;24:423–432.
- 106. Ordoñez CL, Henig NR, Mayer-Hamblett N, Accurso FJ, Burns JL, Chmiel JF, Daines CL, Gibson RL, McNamara S, Retsch-Bogart GZ, et al. Inflammatory and microbiologic markers in induced sputum after intravenous antibiotics in cystic fibrosis. Am J Respir Crit Care Med 2003;168:1471–1475.
- 107. Saiman L, Marshall BC, Mayer-Hamblett N, Burns JL, Quittner AL, Cibene DA, Coquillette S, Fieberg AY, Accurso FJ, Campbell PW, III. Azithromycin in patients with cystic fibrosis chronically infected with Pseudomonas aeruginosa: a randomized controlled trial. JAMA 2003;290:1749–1756.
- 108. Chmiel JF, Konstan MW, Lymp J, Mayer-Hamblett N, Hilliard KA, Accurso FJ, Ramsey B. Assessment of induced sputum as a tool to evaluate anti-inflammatory agents in CF. Pediatr Pulmonol 2014;42(S30):228. Abstract.
- 109. Laguna TA, Wagner BD, Luckey HK, Mann SA, Sagel SD, Regelmann W, Accurso FJ. Sputum desmosine during hospital admission for pulmonary exacerbation in cystic fibrosis. Chest 2009;136:1561–1568.
- 110. Porro C, Lepore S, Trotta T, Castellani S, Ratclif L, Battaglino A, Di Gioia S, Martinez MC, Conese M, Maffione AB. Isolation and characterization of microparticles in sputum from cystic fibrosis patients. Respir Res 2010;11:94.
- 111. Hector A, Kormann MS, Mack I, Latzin P, Casaulta C, Kieninger E, Zhou Z, Yildirim AÖ, Bohla A, Rieber N, et al. The chitinaselike protein YKL-40 modulates cystic fibrosis lung disease. PLoS ONE 2011;6:e24399.
- 112. Fantino E, Gangell CL, Hartl D, Sly PD. Airway, but not serum or urinary, levels of YKL-40 reflect inflammation in early cystic fibrosis lung disease. BMC Pulm Med 2014;14:28.
- 113. Reeves EP, Bergin DA, Fitzgerald S, Hayes E, Keenan J, Henry M, Meleady P, Vega-Carrascal I, Murray MA, Low TB, et al. A novel neutrophil derived inflammatory biomarker of pulmonary exacerbation in cystic fibrosis. J Cyst Fibros 2012;11:100–107.
- 114. Quon BS, Ngan DA, Wilcox PG, Man SF, Sin DD. Plasma sCD14 as a biomarker to predict pulmonary exacerbations in cystic fibrosis. PLoS ONE 2014;9:e89341.
- 115. Wojewodka G, De Sanctis JB, Bernier J, Berube J, Ahlgren HG, Gruber J, Landry J, Lands LC, Nguyen D, Rousseau S, et al. Candidate markers associated with the probability of future pulmonary exacerbations in cystic fibrosis patients. PLoS ONE 2014;9:e88567.
- 116. Shoki AH, Mayer-Hamblett N, Wilcox PG, Sin DD, Quon BS. Systematic review of blood biomarkers in cystic fibrosis pulmonary exacerbations. Chest 2013;144:1659–1670.
- 117. Ratjen F, Saiman L, Mayer-Hamblett N, Lands LC, Kloster M, Thompson V, Emmett P, Marshall B, Accurso F, Sagel S, Anstead M. Effect of azithromycin on systemic markers of inflammation in patients with cystic fibrosis uninfected with Pseudomonas aeruginosa. Chest 2012;142:1259–1266.
- 118. Grob NM, Aytekin M, Dweik RA. Biomarkers in exhaled breath condensate: a review of collection, processing and analysis. J Breath Res 2008;2:037004.
- 119. Rowe SM, Miller S, Sorscher EJ. Cystic fibrosis. N Engl J Med 2005;352:1992–2001.
- 120. Wilschanski M, Dupuis A, Ellis L, Jarvi K, Zielenski J, Tullis E, Martin S, Corey M, Tsui LC, Durie P. Mutations in the cystic fibrosis transmembrane regulator gene and in vivo transepithelial potentials. Am J Respir Crit Care Med 2006;174:787–794.

Novel Outcome Measures in CF 315

- 121. Accurso FJ, Rowe SM, Clancy JP, Boyle MP, Dunitz JM, Durie PR, Sagel SD, Hornick DB, Konstan MW, Donaldson SH, et al. Effect of VX-770 in persons with cystic fibrosis and the G551D-CFTR mutation. N Engl J Med 2010;363:1991–2003.
- 122. Clancy JP, Rowe SM, Accurso FJ, Aitken ML, Amin RS, Ashlock MA, Ballmann M, Boyle MP, Bronsveld I, Campbell PW, et al. Results of a phase IIa study of VX-809, an investigational CFTR corrector compound, in subjects with cystic fibrosis homozygous for the F508del-CFTR mutation. Thorax 2012;67:12–18.
- 123. Kerem E, Hirawat S, Armoni S, Yaakov Y, Shoseyov D, Cohen M, Nissim-Rafinia M, Blau H, Rivlin J, Aviram M, et al. Effectiveness of PTC124 treatment of cystic fibrosis caused by nonsense mutations: a prospective phase II trial. Lancet 2008;372:719–727.
- 124. Wilschanski M, Yahav Y, Yaacov Y, Blau H, Bentur L, Rivlin J, Aviram M, Bdolah-Abram T, Bebok Z, Shushi L, et al. Gentamicin-induced correction of CFTR function in patients with cystic fibrosis and CFTR stop mutations. N Engl J Med 2003;349:1433–1441.
- 125. Mayell SJ, Munck A, Craig JV, Sermet I, Brownlee KG, Schwarz MJ, Castellani C, Southern KW. A European consensus for the evaluation and management of infants with an equivocal diagnosis following newborn screening for cystic fibrosis. J Cyst Fibros 2009;8:71–78.
- 126. Rosenstein BJ, Cutting GR. The diagnosis of cystic fibrosis: a consensus statement. Cystic Fibrosis Foundation Consensus Panel. J Pediatr 1998;132:589–595.
- 127.Farrell PM, Rosenstein BJ, White TB, Accurso FJ, Castellani C, Cutting GR, Durie PR, Legrys VA, Massie J, Parad RB, et al. Guidelines for diagnosis of cystic fibrosis in newborns through older adults: Cystic Fibrosis Foundation consensus report. J Pediatr 2008;153:S4–S14.
- 128. Naehrlich L, Ballmann M, Davies J, Derichs N, Gonska T, Hjelte L, van Konigsbruggen-Rietschel S, Leal T, Melotti P, et al. Nasal potential difference measurements in diagnosis of cystic fibrosis: an international survey. J Cyst Fibros 2014;13:24–28.
- 129. Rowe SM, Clancy JP, Wilschanski M. Nasal potential difference measurements to assess CFTR ion channel activity. Methods Mol Biol 2011;741:69–86.
- 130. Solomon GM, Konstan MW, Wilschanski M, Billings J, Sermet-Gaudelus I, Accurso F, Vermeulen F, Levin E, Hathorne H, Reeves G, et al. An international randomized multicenter comparison of nasal potential difference techniques. Chest 2010;138:919–928.
- 131. Clancy JP, Rowe SM, Bebok Z, Aitken ML, Gibson R, Zeitlin P, Berclaz P, Moss R, Knowles MR, Oster RA, et al. No detectable improvements in cystic fibrosis transmembrane conductance regulator by nasal aminoglycosides in patients with cystic fibrosis with stop mutations. Am J Respir Cell Mol Biol 2007;37:57–66.
- 132. Rowe SM, Liu B, Hill A, Hathorne H, Cohen M, Beamer JR, Accurso FJ, Dong Q, Ordoñez CL, Stone AJ, et al. Optimizing nasal potential difference analysis for CFTR modulator development: assessment of ivacaftor in CF subjects with the G551D-CFTR mutation. PLoS ONE 2013;8:e66955.
- 133. Davies JC, Davies M, McShane D, Smith S, Chadwick S, Jaffe A, Farley R, Collins L, Bush A, Scallon M, et al. Potential difference measurements in the lower airway of children with and without cystic fibrosis. Am J Respir Crit Care Med 2005;171:1015–1019.
- 134. Dransfield MT, Wilhelm AM, Flanagan B, Courville C, Tidwell SL, Raju SV, Gaggar A, Steele C, Tang LP, Liu B, et al. Acquired cystic fibrosis transmembrane conductance regulator dysfunction in the lower airways in COPD. Chest 2013;144:498–506.

- 135. Cohen-Cymberknoh M, Yaakov Y, Shoseyov D, Shteyer E, Schachar E, Rivlin J, Bentur L, Picard E, Aviram M, Israeli E, et al. Evaluation of the intestinal current measurement method as a diagnostic test for cystic fibrosis. Pediatr Pulmonol 2013;48:229– 235.
- 136. De Boeck K, Derichs N, Fajac I, de Jonge HR, Bronsveld I, Sermet I, Vermeulen F, Sheppard DN, Cuppens H, Hug M, et al. New clinical diagnostic procedures for cystic fibrosis in Europe. J Cyst Fibros 2011;10:S53–S66.
- 137. Derichs N, Sanz J, Von Kanel T, Stolpe C, Zapf A, Tummler B, Gallati S, Ballmann M. Intestinal current measurement for diagnostic classification of patients with questionable cystic fibrosis: validation and reference data. Thorax 2010;65:594–599.
- Hug MJ, Tümmler B. Intestinal current measurements to diagnose cystic fibrosis. J Cyst Fibros 2004;3:157–158.
- 139. Mall M, Hirtz S, Gonska T, Kunzelmann K. Assessment of CFTR function in rectal biopsies for the diagnosis of cystic fibrosis. J Cyst Fibros 2004;3:165–169.
- 140. Mall M, Kreda SM, Mengos A, Jensen TJ, Hirtz S, Seydewitz HH, Yankaskas J, Kunzelmann K, Riordan JR, Boucher RC. The DeltaF508 mutation results in loss of CFTR function and mature protein in native human colon. Gastroenterology 2004;126:32–41.
- 141. Hirtz S, Gonska T, Seydewitz HH, Thomas J, Greiner P, Kuehr J, Brandis M, Eichler I, Rocha H, Lopes AI, et al. CFTR Cl- channel function in native human colon correlates with the genotype and phenotype in cystic fibrosis. Gastroenterology 2004;127:1085– 1095.
- 142. Clancy JP, Szczesniak RD, Ashlock MA, Ernst SE, Fan L, Hornick DB, Karp PH, Khan U, Lymp J, Ostmann AJ, et al. Multicenter intestinal current measurements in rectal biopsies from CF and non-CF subjects to monitor CFTR function. PLoS ONE 2013;8: e73905.
- 143. Rowe SM, Accurso F, Clancy JP. Detection of cystic fibrosis transmembrane conductance regulator activity in early-phase clinical trials. Proc Am Thorac Soc 2007;4:387–398.
- 144. Durmowicz AG, Witzmann KA, Rosebraugh CJ, Chowdhury BA. Change in sweat chloride as a clinical end point in cystic fibrosis clinical trials: the ivacaftor experience. Chest 2013;143:14–18.
- 145. Quinton P, Molyneux L, Ip W, Dupuis A, Avolio J, Tullis E, Conrad D, Shamsuddin AK, Durie P, Gonska T. β-adrenergic sweat secretion as a diagnostic test for cystic fibrosis. Am J Respir Crit Care Med 2012;186:732–739.
- 146. Mattar AC, Leone C, Rodrigues JC, Adde FV. Sweat conductivity: an accurate diagnostic test for cystic fibrosis?. J Cyst Fibros 2014;13:528–533.
- 147. Flume PA, Liou TG, Borowitz DS, Li H, Yen K, Ordoñez CL, Geller DE. Ivacaftor in subjects with cystic fibrosis who are homozygous for the F508del-CFTR mutation. Chest 2012;142: 718–724.
- 148. Davies JC, Wainwright CE, Canny GJ, Chilvers MA, Howenstine MS, Munck A, Mainz JG, Rodriguez S, Li H, Yen K, et al. Efficacy and safety of ivacaftor in patients aged 6 to 11 years with cystic fibrosis with a G551D mutation. Am J Respir Crit Care Med 2013;187:1219–1225.
- 149. Wine JJ, Char JE, Chen J, Cho HJ, Dunn C, Frisbee E, Joo NS, Milla C, Modlin SE, Park IH, et al. In vivo readout of CFTR function: ratiometric measurement of CFTR-dependent secretion by individual, identifiable human sweat glands. PLoS ONE 2013;8:e77114.